Of complexes and maintenance of genome stability

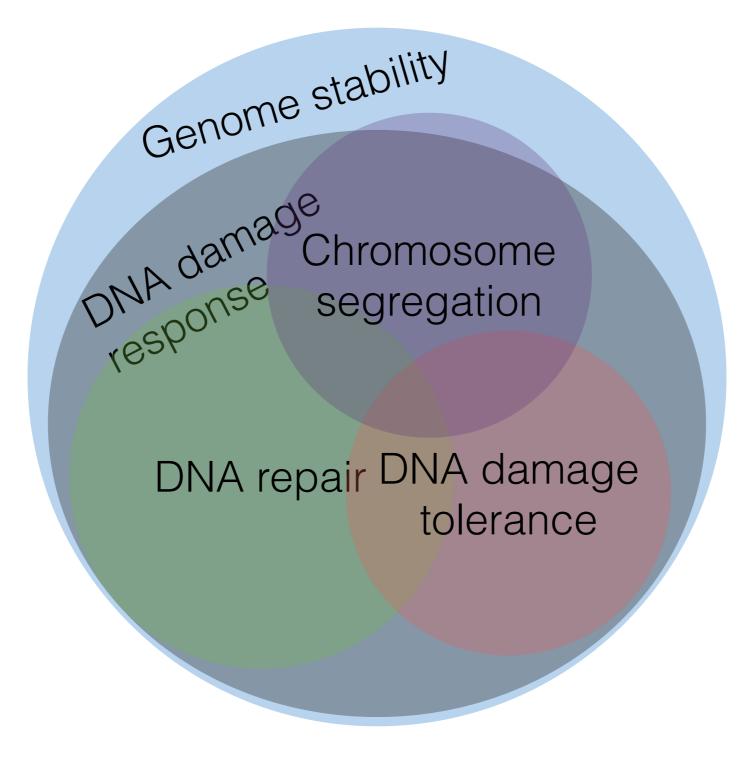
Marek Sebesta, PhD marek.sebesta@ceitec.muni.cz CSB, Ceitec, MU

11-Apr-19

Content

- 1. What is genome stability maintenance?
- 2. What are the challenges to genome stability?
- 3. How do cells know genome stability has been compromised?
- 4. How do cells maintain genome stability?
- 5. How to study genome stability maintenance? (Case study on Homologous recombination)

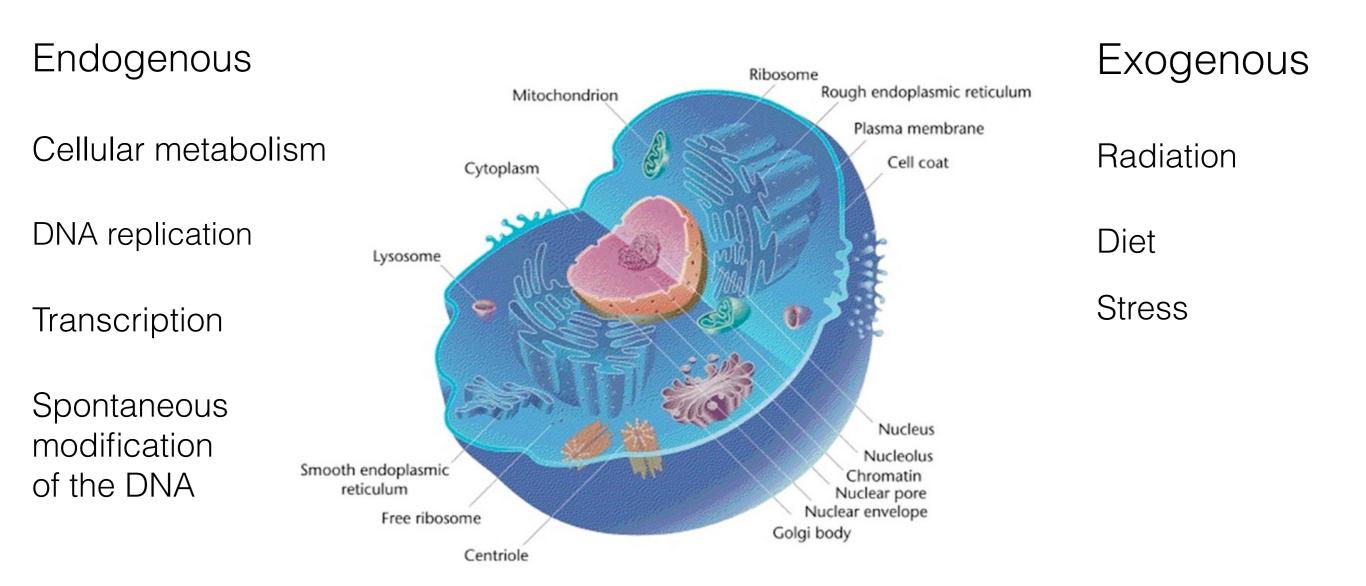
What is genome stability maintenance?

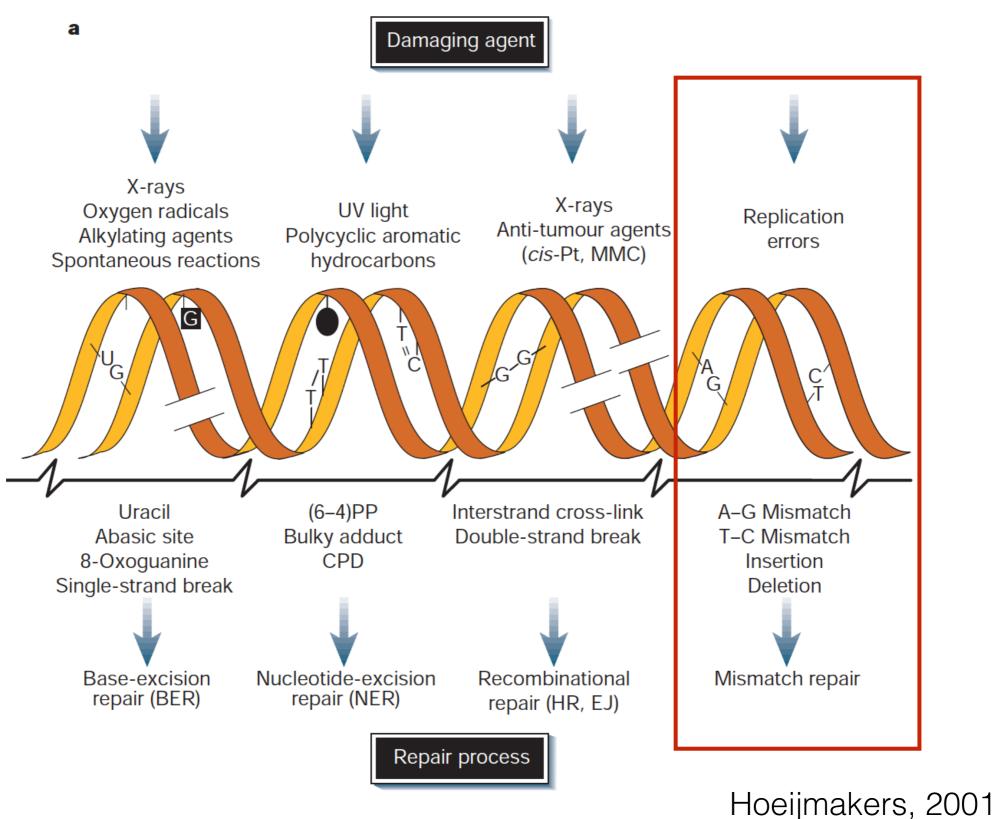


What is genome stability maintenance?

It is the ability of living organisms preserve its genetic material in time and across generations.

All living mater is constantly exposed to environment that challenges genome stability

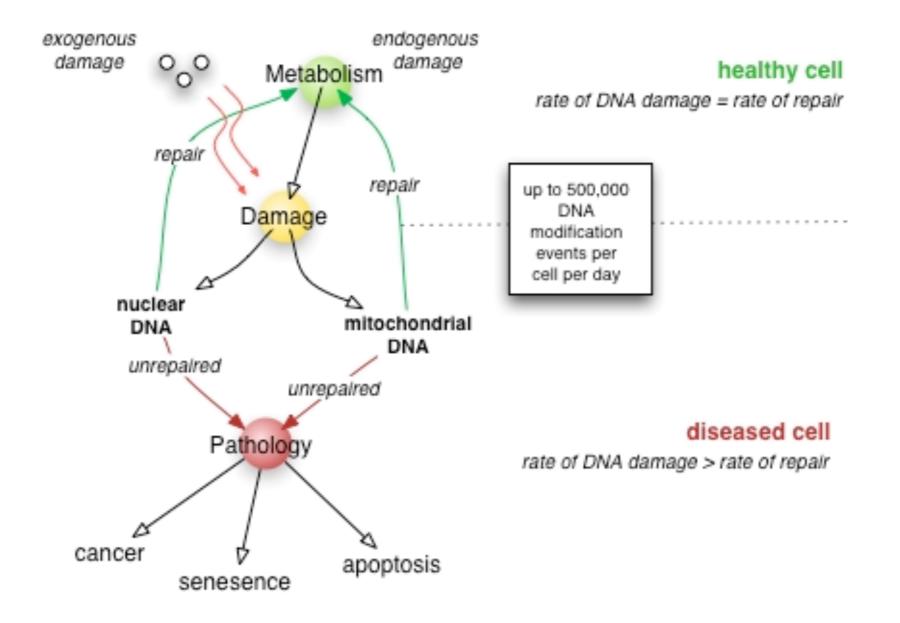




What is more prevalent? Exogenous or endogenous damage?

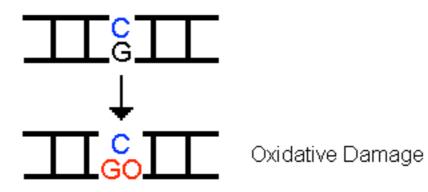
Even-though, historically, exogenous DNA damage was considered to be the prime cause of mutagenesis, recently, as the methodology has progressed, the cellular DNA metabolism pathways (replication and transcription) are being recognised as the more prevalent cause of mutations.

Inability to repair properly the damage may lead to cancer, senescence, or apoptosis.

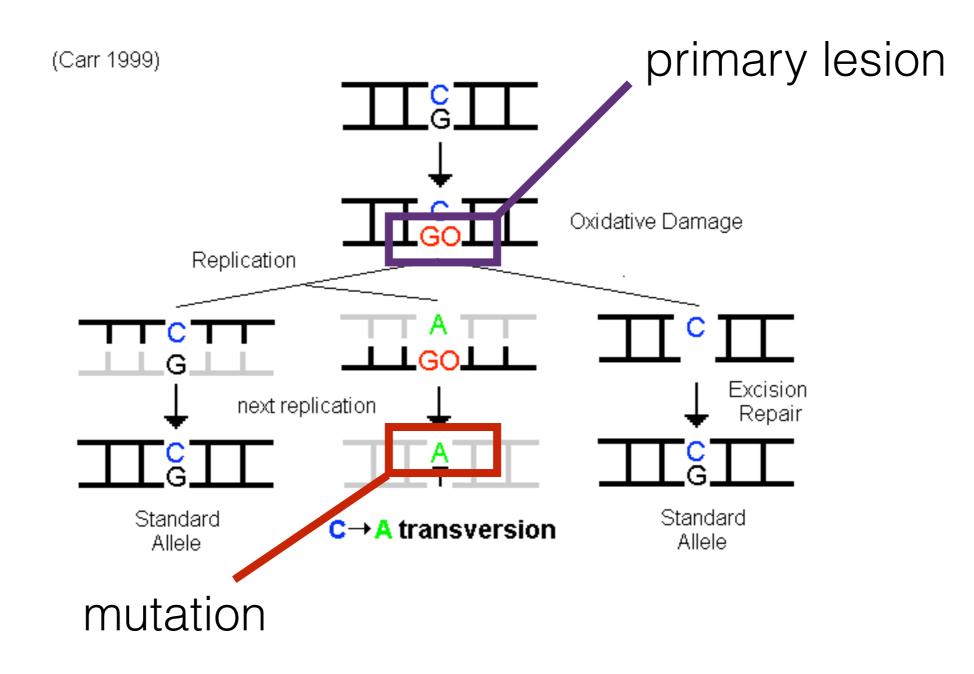


What is the difference between a primary lesion and a mutation?

(Carr 1999)



What is the difference between a primary lesion and a mutation?



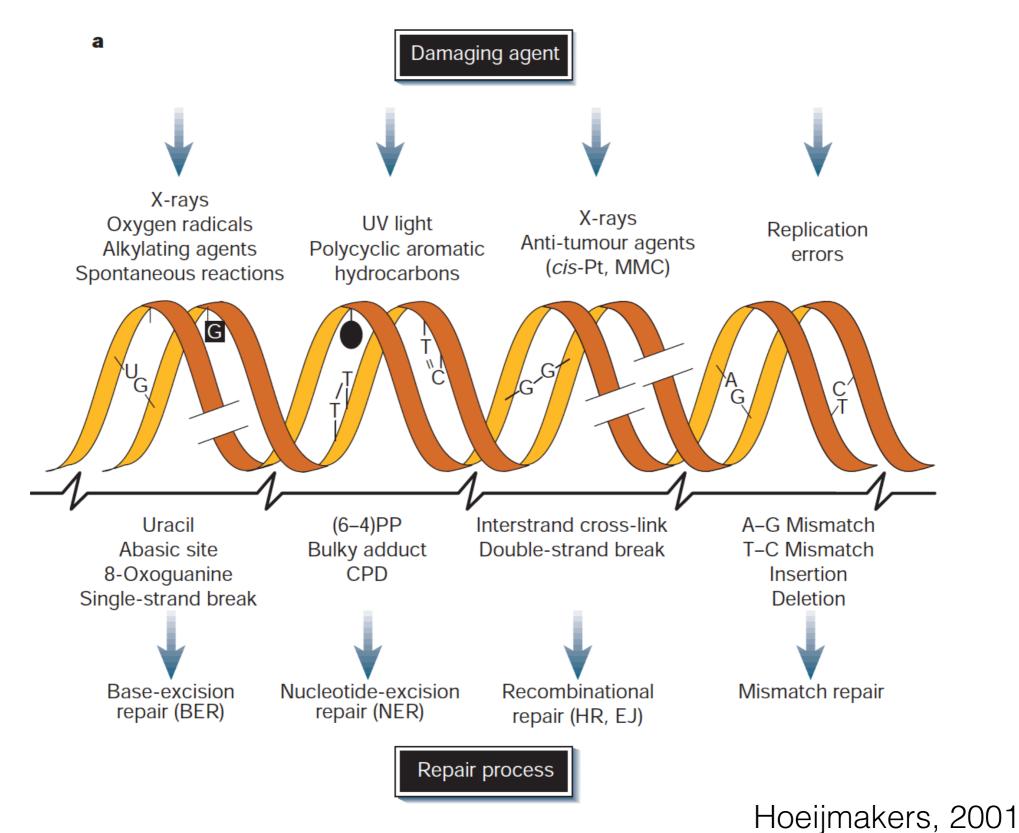
Transient summary I

Terms Genome stability, DNA damage response, DNA repair, DNA damage tolerance denote closely related, yet not interchangeable terms

Cells are continuously exposed to wide variety of DNA damage

Failure to properly deal with the damage may have fatal consequences to cells

How do cells know genome stability has been compromised?

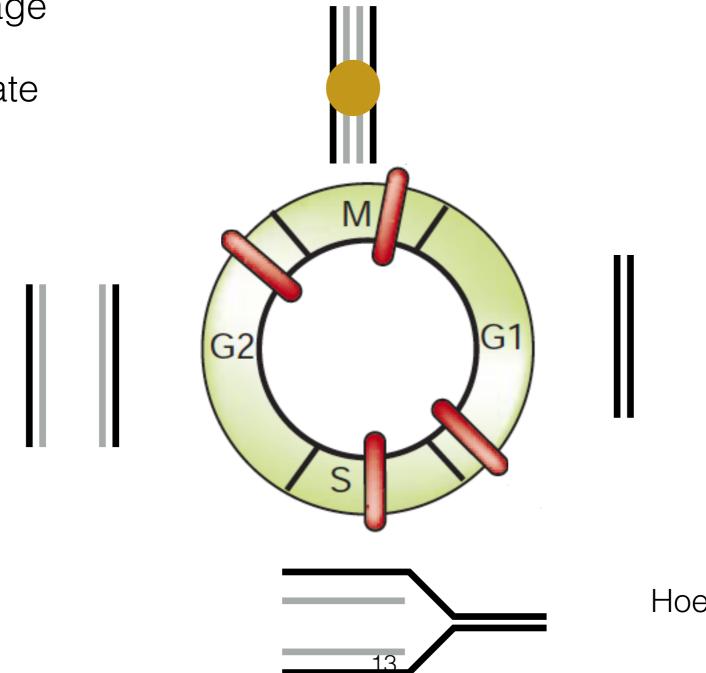


12

How do cells know genome stability has been compromised?

The challenges

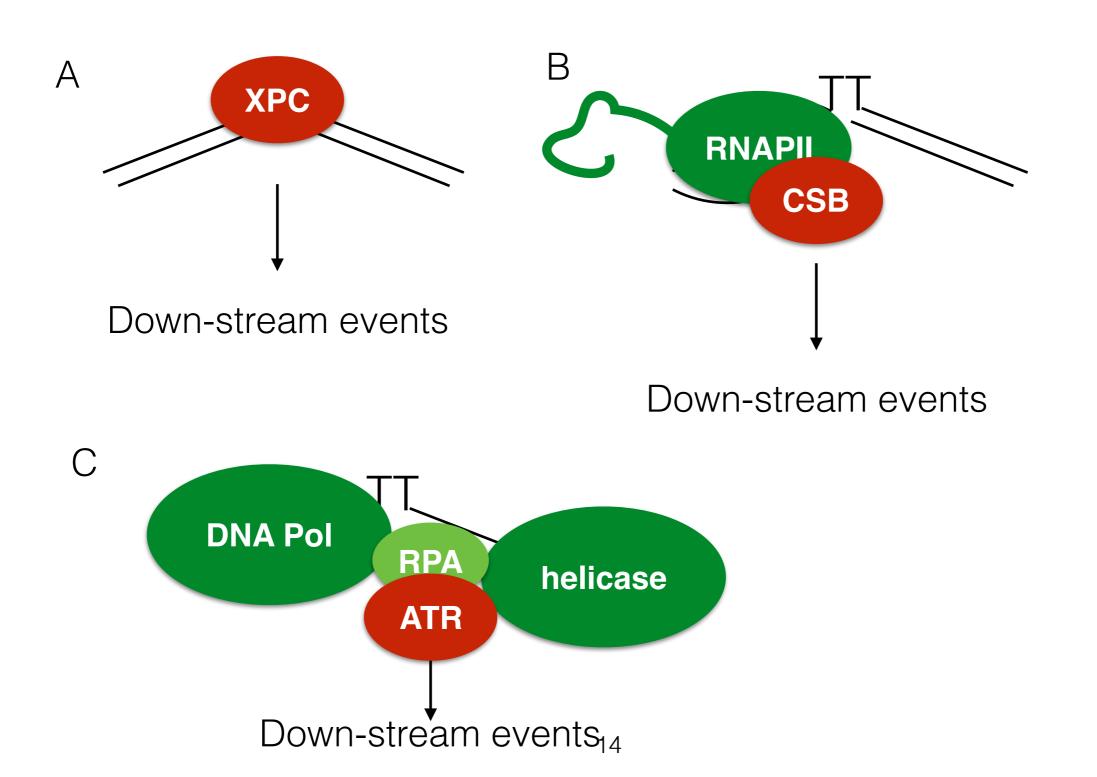
- different types of DNA damage
- cell-cycle stage
- metabolic state



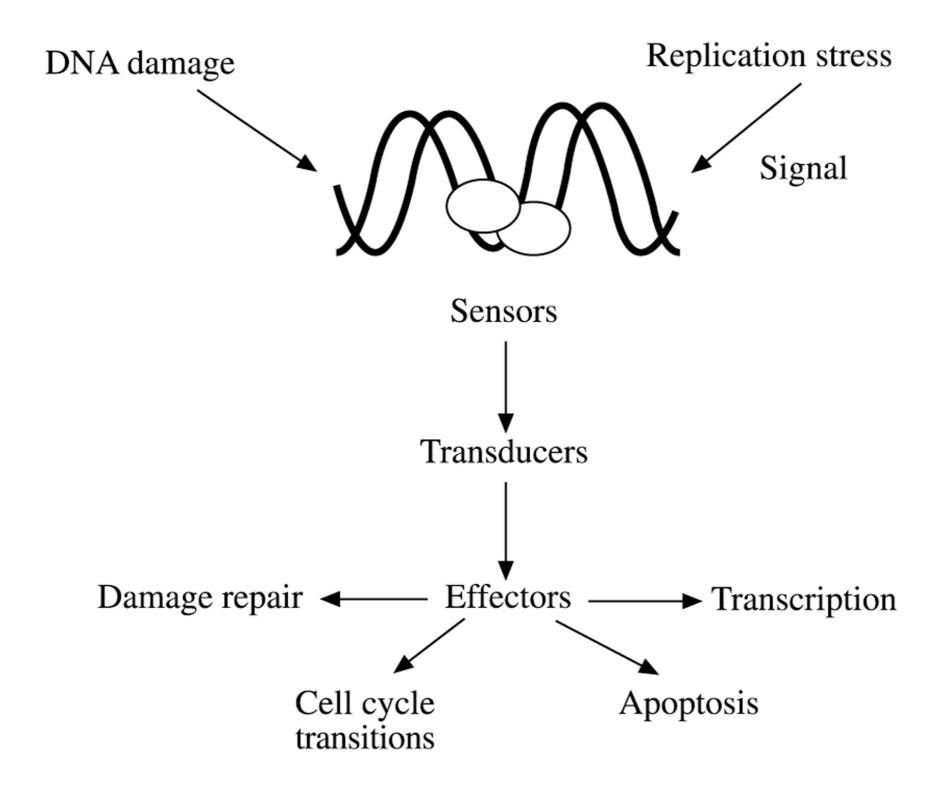
Hoeijmakers, 2001

How do cells know genome stability has been compromised?

Cells possess context-specific sensors that recognise signals from the damaged DNA

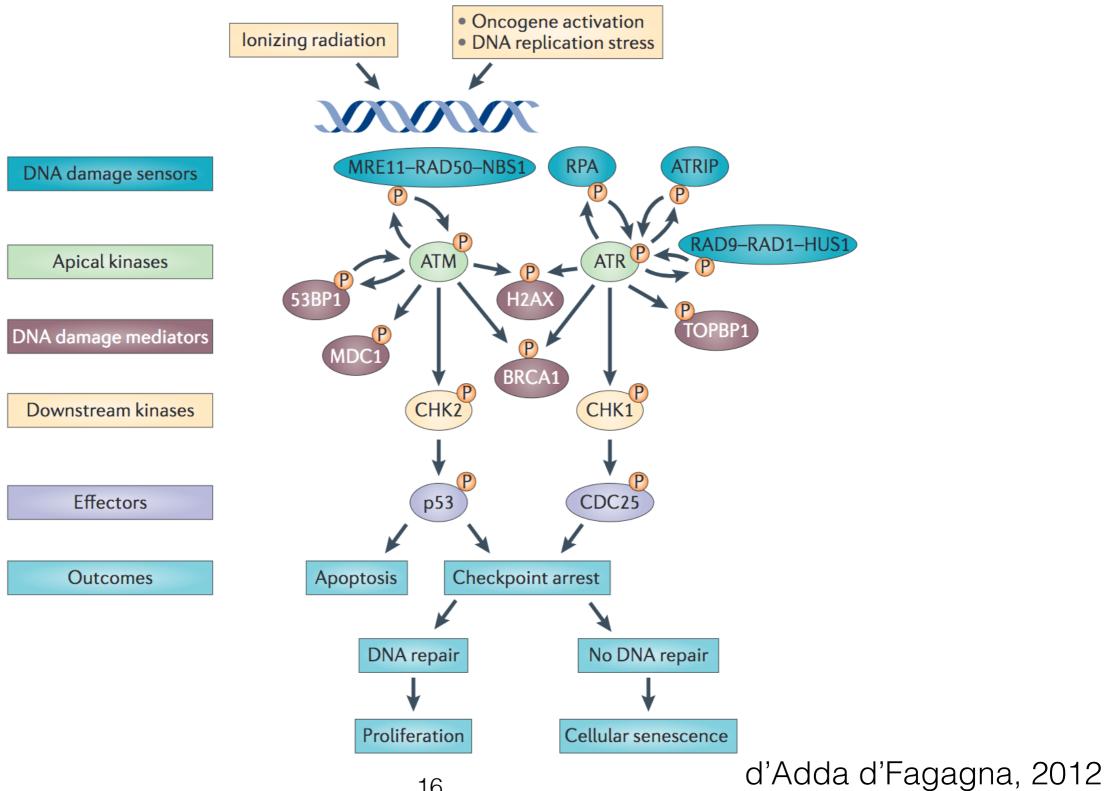


How do cells react to DNA damage?



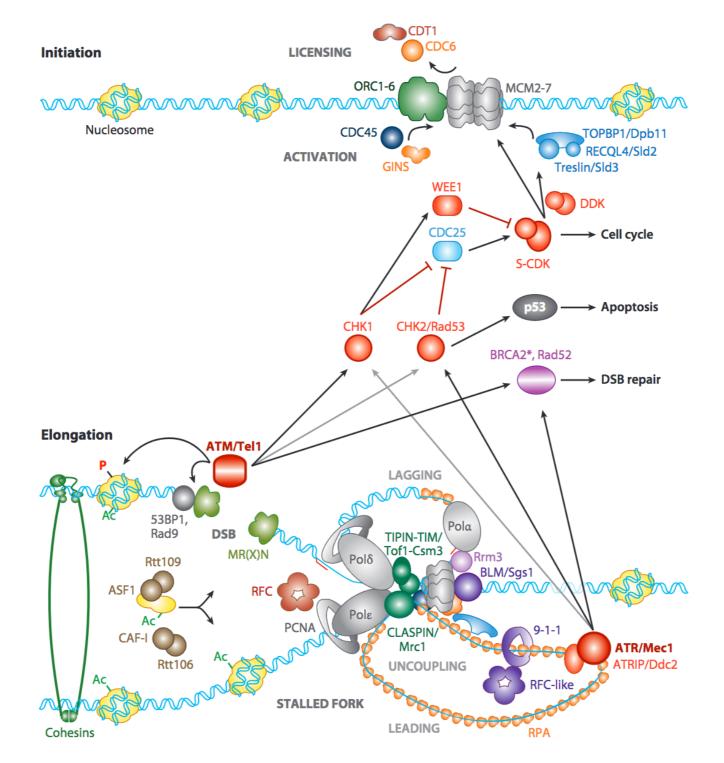
How do cells react to DNA damage?

A simplified picture



How do cells react to DNA damage?

A more comprehensive picture



Aguilera and García-Muse, 2013

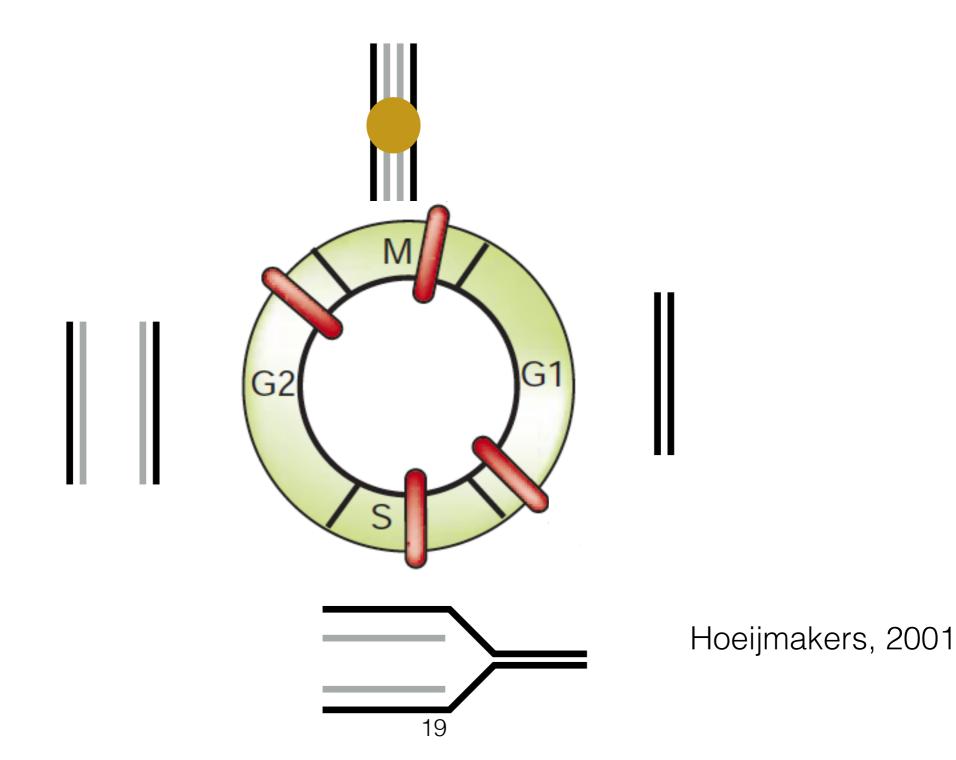
Transient summary II

Cells possess specific factors - sensors - that recognise insults to DNA structure, DNA breaks, or stalled machineries like transcription and replication.

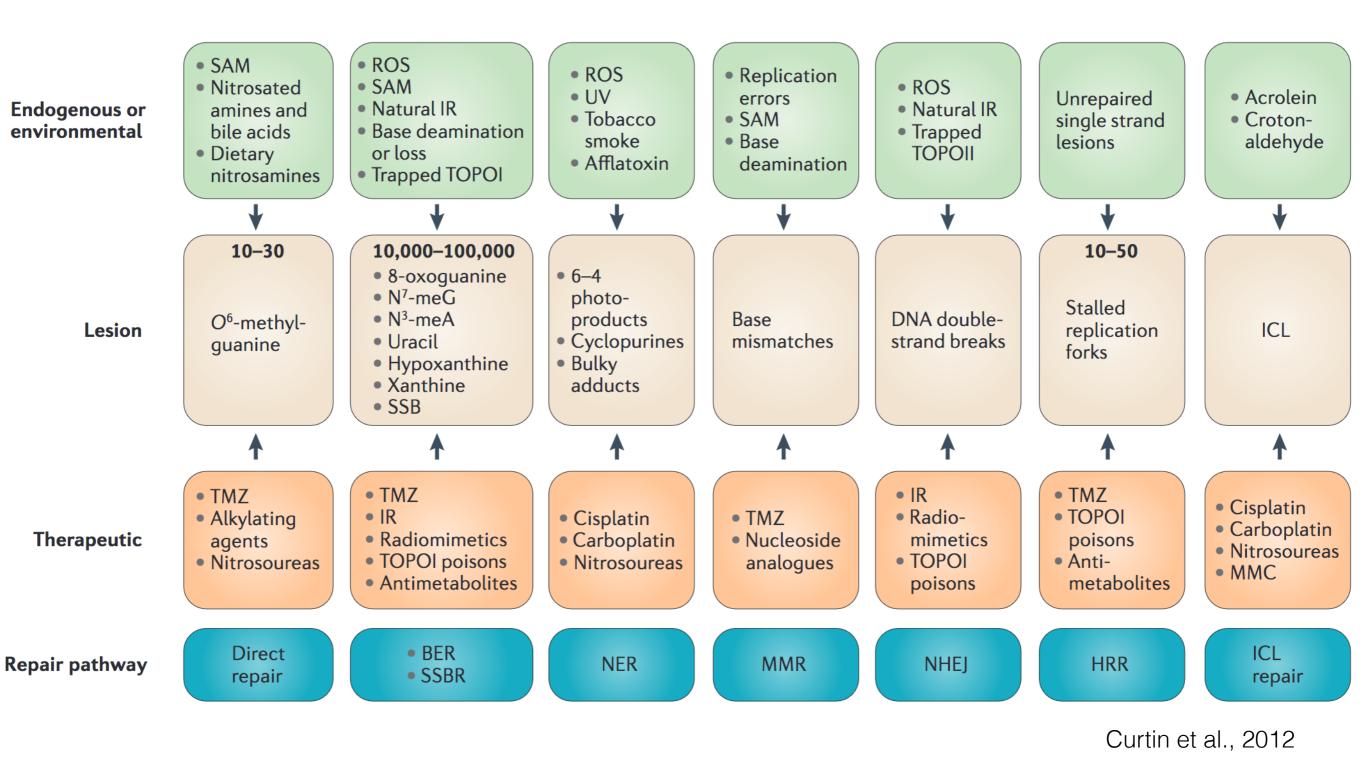
The sensors then activate complex signalling pathways that lead to halt of cell-cycle, as well as to decision as of which pathway is to be used; balancing the cell-cycle stage and other needs of the cell.

How do cell maintain genome stability?

DNA repair is prevalent outside the S-phase, in which DNA damage tolerance is preferred.

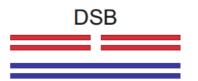


How do cell maintain genome stability?



How do cell maintain genome stability?

Double-stranded DNA breaks (DSB) repair





NHEJ: non-homologous end joining

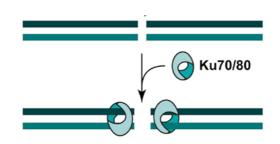
SSA: single strand annealing

SDSA: synthesis-dependent strandannealing

DSBR: DSB repair



How do cell maintain genome stability? Double-stranded DNA breaks (DSB) repair Non-homologous end joining

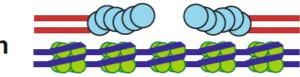


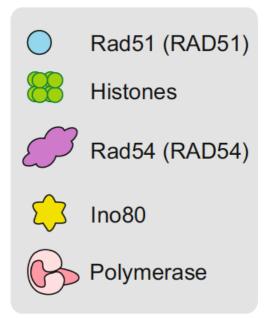
How do cell maintain genome stability? Double-stranded DNA breaks (DSB) repair Homologous recombination

Pre-synapsis

How do cell maintain genome stability? Double-stranded DNA breaks (DSB) repair Homologous recombination Synapsis

Homology search

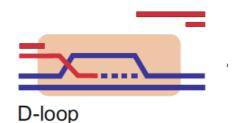




Sebesta and Krejci, 2016

How do cell maintain genome stability? Double-stranded DNA breaks (DSB) repair Homologous recombination

Post-synapsis



Transient summary III

Different types of DNA damage are repaired by specific repair pathway

The repair is generally error-free, except for NHEJ and SSA

In S-phase, cells activate tolerance mechanisms that allow timely completion of DNA replication

How to study genome stability maintenance? (Case study on Homologous recombination)



Review

www.microbialcell.com

Guidelines for DNA recombination and repair studies: Mechanistic assays of DNA repair processes

Hannah L Klein^{1,*}, Kenny K.H. Ang², Michelle R. Arkin², Emily C. Beckwitt^{3,4}, Yi-Hsuan Chang⁵, Jun Fan⁶, Youngho Kwon^{7,8}, Michael J. Morten¹, Sucheta Mukherjee⁹, Oliver J. Pambos⁶, Hafez el Sayyed⁶, Elizabeth S. Thrall¹⁰, João P. Vieira-da-Rocha⁹, Quan Wang¹¹, Shuang Wang^{12,13}, Hsin-Yi Yeh⁵, Julie S. Biteen¹⁴, Peter Chi^{5,15}, Wolf-Dietrich Heyer^{9,16}, Achillefs N. Kapanidis⁶, Joseph J. Loparo¹⁰, Terence R. Strick^{12,13,17}, Patrick Sung^{7,8}, Bennett Van Houten^{3,18,19}, Hengyao Niu^{11,*} and Eli Rothenberg^{1,*} How to study genome stability maintenance? (Case study on Homologous recombination)

Different strategies exist

Genetic tools

Allow us to identify genes and the relationships among, thereby building a pathway

Biochemical tools

Allow us to understand mechanisms and complex formations within a studied pathway

Microscopic tools

Give us a glimpse at spacial and temporal relationships of genes of interests

Structural tools

Allow us to understand molecular mechanisms at atomic resolution

Single molecule techniques

Allow us to understand behaviour at of single molecules as compared to bulk biochemical reactions

How to study genome stability maintenance? Step1: identify the genes

Molec. gen. Genet. 125, 197-216 (1973) © by Springer-Verlag 1973

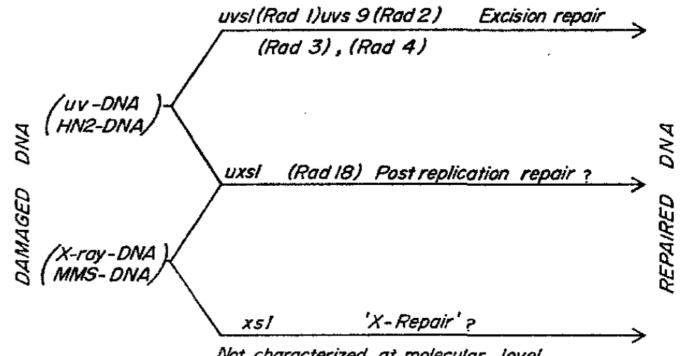
Interactions among Genes Controlling Sensitivity to Radiation and Alkylation in Yeast

Martin Brendel and Robert H. Haynes

Department of Biology, York University, Toronto, Canada

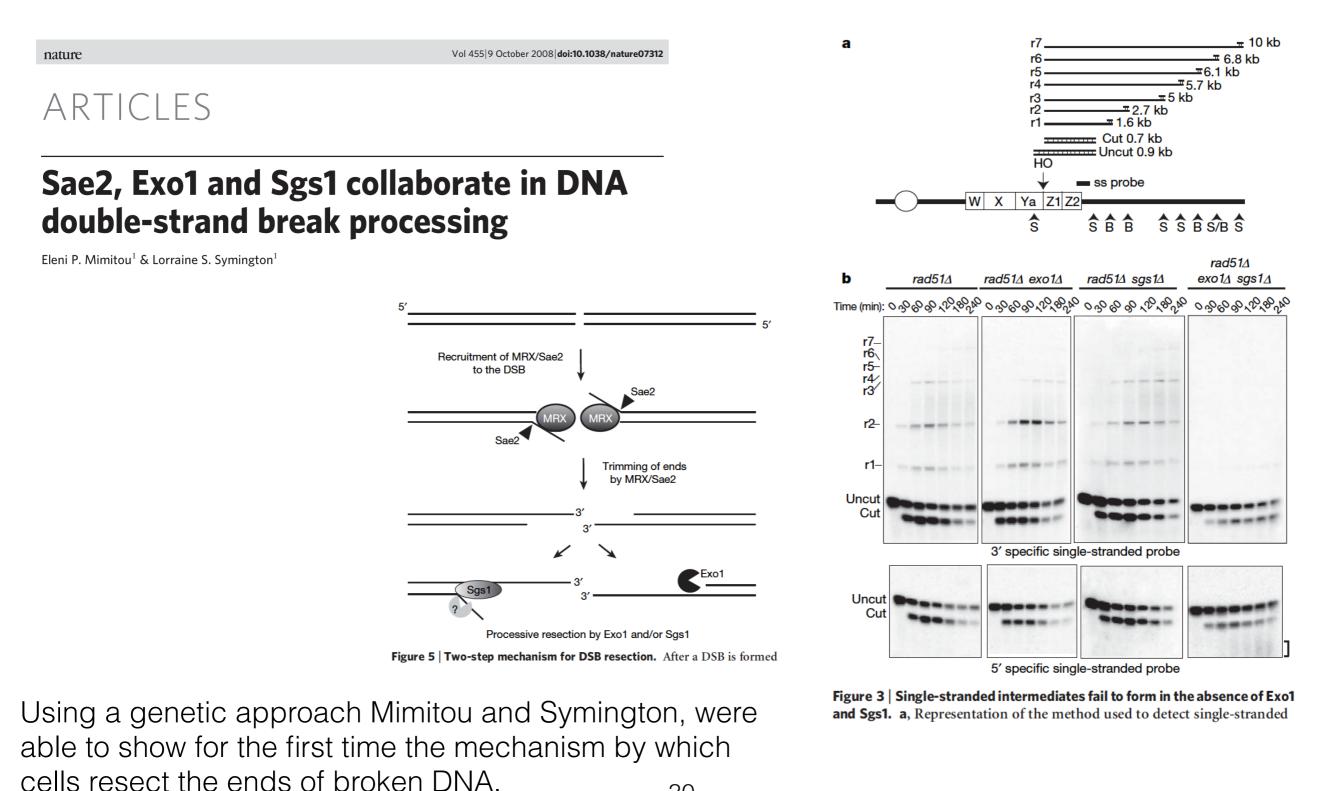
Received March 27, 1973

Using a thorough genetic analysis of the isolated mutants, they were able to build a first model of multiple pathways dealing with DNA damage.



Not characterized at molecular level

How to study genome stability maintenance? Step1: identify the genes



How to study genome stability maintenance? Step2: purify and study the proteins alone

Catalysis of ATP-Dependent Homologous DNA Pairing and Strand Exchange by Yeast RAD51 Protein

Patrick Sung

Fig. 1. Overproduction A В and purification of RAD51 **kD** <u>kD</u> Strand (-) protein. (A) Immunoblot Synapsis exchange 97analysis. The nitrocellu-(+) 97 lose blot of a 9% denatur-66ing polyacrylamide gel 66 was probed with affinity-- RAD51 Viral ss Linear ds Joint molecule Nicked circular Displaced ss purified antibodies to rad51* ← RAD51 43-RAD51. Lane 1. extract ← rad51* 43 from the rad51 deletion RAD51 veast strain YR51 Δ -1: 60 10 30 60 Time (min) 60 RAD51 31lane 2, extract from the Time (min) 60 10 30 60 60 yeast strain LP2749-9B 31 harboring the 2µ multi-12345 jm∟ copy vector pSCW231, jm_E ncnc which contains the ADC1 promoter but lacks the RAD51 gene; 21 . ds lane 3, extract from strain LP2749-9B harboring the 2µ plasmid 2 3 pR51.2, which contains the RAD51 gene under the control of its Displaced ssown promoter; lane 4, extract from strain LP2749-9B harboring Displaced ssthe 2µ plasmid pR51.1, which contains the RAD51 gene under the control of the ADC1 promoter; and lane 5, 10 ng of purified RAD51 protein. (B) Purity analysis by SDS-PAGE. A 9% denaturing polyacrylamide gel was 5 2 3 4 1 2 3 5 stained with Coomassie blue. Lane 1, molecular size markers; lanes 2 and 3, 1 µg and 3 µg of purified RAD51 5' Labeled protein. Molecular sizes are indicated on the left (in kilodaltons). 3' Labeled

Using a purified protein, Patrick Sung was able to show that Rad51 is a bona fide recombinase.

How to study genome stability maintenance? Step2: purify and study the proteins in assemblies

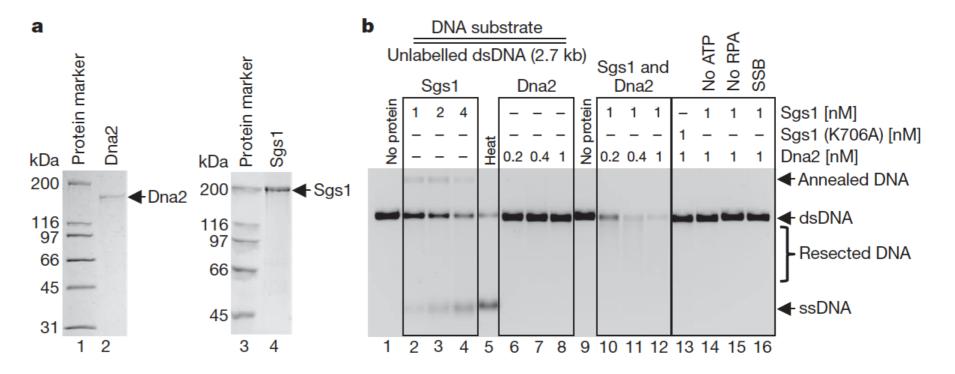
nature

Vol 467 2 September 2010 doi:10.1038/nature09355

LETTERS

DNA end resection by Dna2–Sgs1–RPA and its stimulation by Top3–Rmi1 and Mre11–Rad50–Xrs2

Petr Cejka^{1,2}, Elda Cannavo^{1,2}, Piotr Polaczek³, Taro Masuda-Sasa³, Subhash Pokharel³, Judith L. Campbell³ & Stephen C. Kowalczykowski^{1,2}



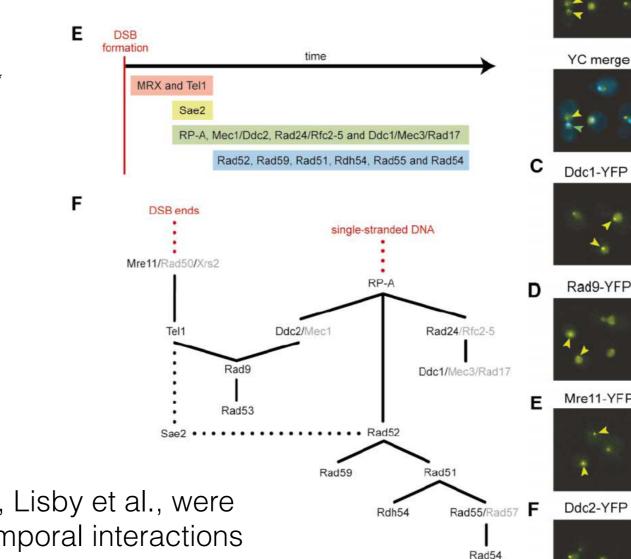
Using purified proteins, Cejka et al., were able to reconstitute end resection *in vitro*.

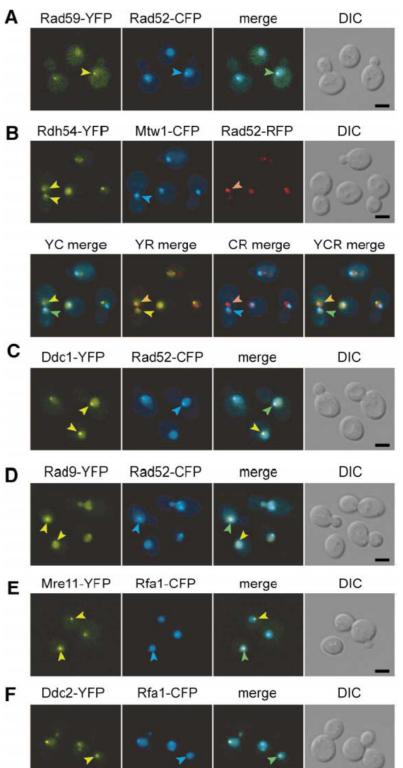
How to study genome stability maintenance? Step3: study the proteins in time and space

Cell, Vol. 118, 699-713, September 17, 2004, Copyright ©2004 by Cell Press

Choreography of the DNA Damage Response: Spatiotemporal Relationships among Checkpoint and Repair Proteins

Michael Lisby,^{1,3} Jacqueline H. Barlow, Rebecca C. Burgess,² and Rodney Rothstein*





Using life-cell microscopy, Lisby et al., were able to study the spatiotemporal interactions among recombination factors. How to study genome stability maintenance? Step4: study the role of protein complex formation?

Protein Group Modification and Synergy in the SUMO Pathway as Exemplified in DNA Repair

Ivan Psakhye¹ and Stefan Jentsch^{1,*}

¹Department of Molecular Cell Biology, Max Planck Institute of Biochemistry, Am Klopferspitz 18, 82152 Martinsried, Germany

*Correspondence: jentsch@biochem.mpg.de http://dx.doi.org/10.1016/j.cell.2012.10.021

Α Light Heavy Lys0Arg0 Lys8Arg10 Untreated MMS 0.2% HisSUMO Ni-NTA PD Peptide intensity (log₁₀) RFA2 Slice 1 AD52 250-2 RAD9 150 -100-RAD59 75 -50 -37 -8 25 g 20 -15 Tryptic digestion, LC-MS/MS, -5 10 MASCOT/MaxQuant data processing SILAC ratio (log₂ MMS-treated/untreated)

Using SILAC approaches, Psakhye and Jentsch showed that majority of HR proteins are Sumoylated upon DSBs induction. How to study genome stability maintenance? Step4: study the role of protein complex formation?

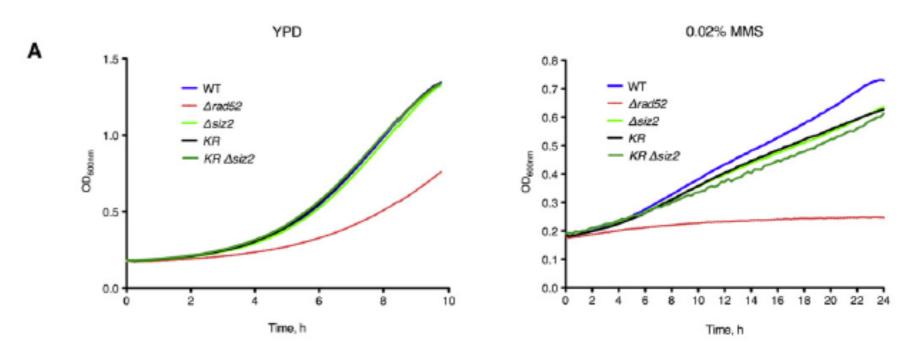
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This Sumo-SIM mediated interactions are trigger timely completion of HR.

How to study genome stability maintenance? Step4: study the role of protein complex formation?

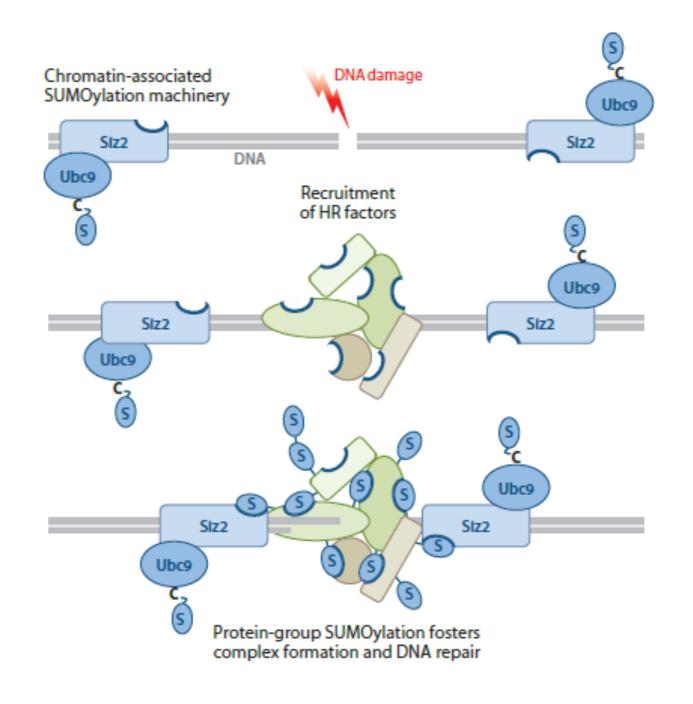
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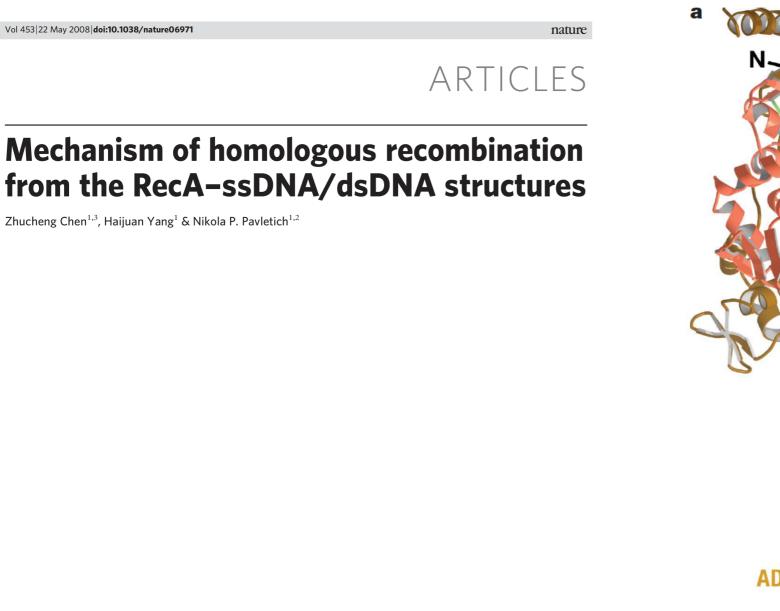
¹Department of Molecular Cell Biology, Max Planck Institute of Biochemistry, Am Klopferspitz 18, 82152 Martinsried, Germany *Correspondence: jentsch@biochem.mpg.de

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This Sumo-SIM mediated interactions are trigger timely completion of HR.



How to study genome stability maintenance? Step5: study the molecular mechanisms by the means of structural biology



RecA¹ RecA² RecA³ RecA⁴ ADP-AIF,-Mg RecA⁶

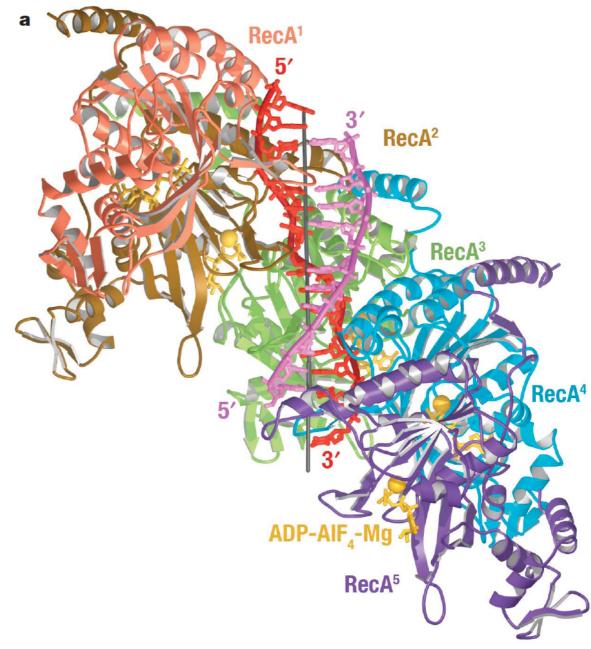
b

Crystal structure of presynaptic filament.

How to study genome stability maintenance? Step5: study the molecular mechanisms by the means of structural biology

Vol 453/22 May 2008/doi:10.1038/nature06971 ACTICLES ARTICLES Mechanism of homologous recombination from the RecA-ssDNA/dsDNA structures

Zhucheng Chen^{1,3}, Haijuan Yang¹ & Nikola P. Pavletich^{1,2}



Crystal structure of postsynaptic filament.

How to study genome stability maintenance? Step5: study the molecular mechanisms by the means of structural biology

Mechanism of homologous recombination from the RecA-ssDNA/dsDNA structures	
	ARTICLES
Vol 453 22 May 2008 doi:10.1038/nature06971	nature

Zhucheng Chen^{1,3}, Haijuan Yang¹ & Nikola P. Pavletich^{1,2}

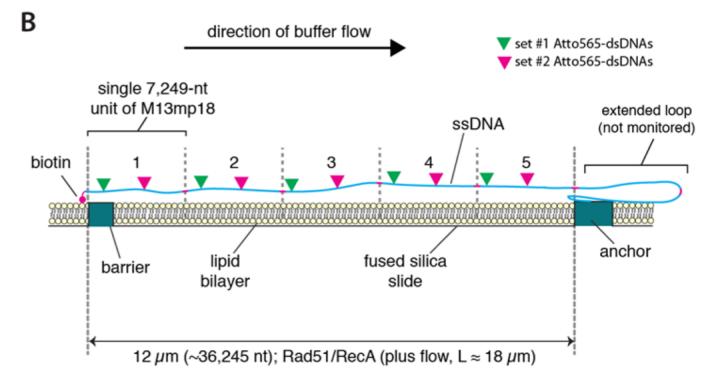
By comparing the two structure a detailed, molecular mechanism of the strand exchange reaction can be inferred.

How to study genome stability maintenance? Step6: study the molecular mechanisms by the means of single-molecule techniques.

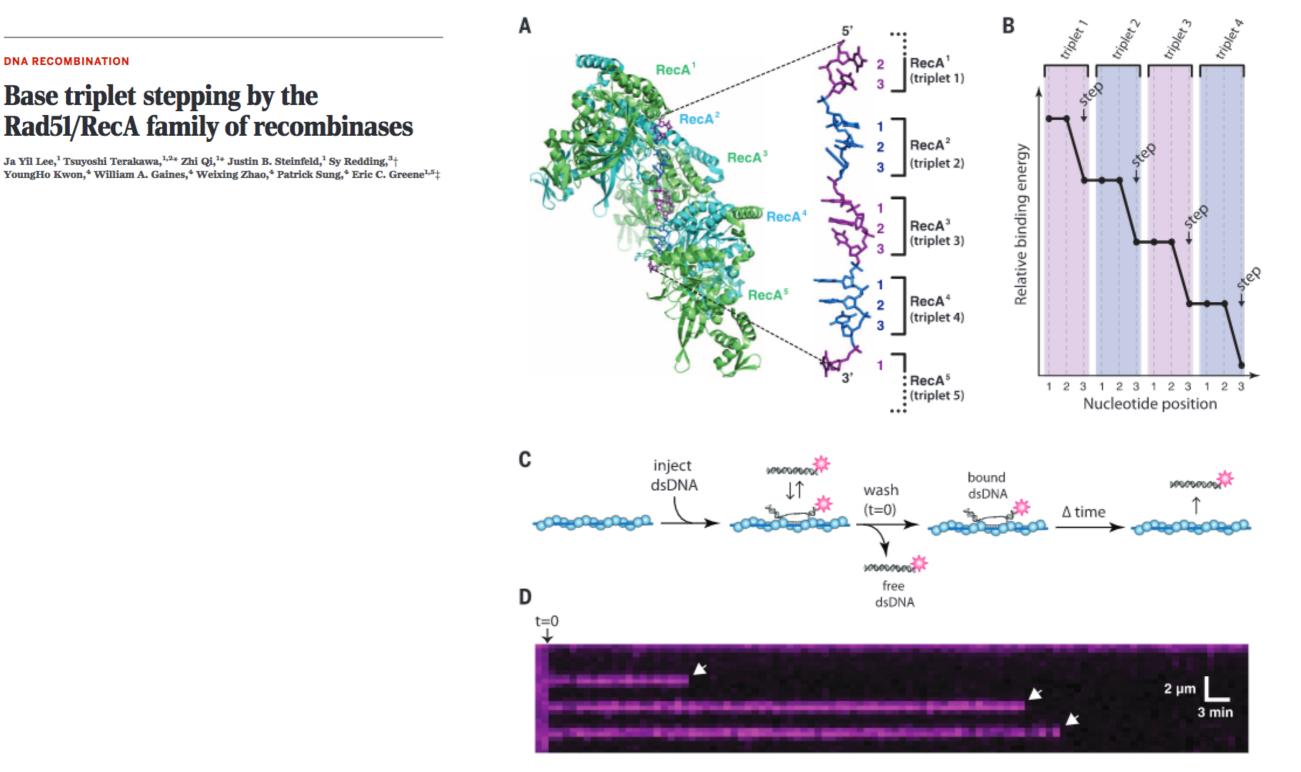
DNA RECOMBINATION

Base triplet stepping by the Rad51/RecA family of recombinases

Ja Yil Lee,¹ Tsuyoshi Terakawa,^{1,2*} Zhi Qi,^{1*} Justin B. Steinfeld,¹ Sy Redding,³[†] YoungHo Kwon,⁴ William A. Gaines,⁴ Weixing Zhao,⁴ Patrick Sung,⁴ Eric C. Greene^{1,5}[‡]



How to study genome stability maintenance? Step6: study the molecular mechanisms by the means of single-molecule techniques.



Transient summary IV

There are different techniques that allow us understand any given pathway

The techniques must be combined, in order to get a full picture of the pathway

Use whatever technique at hand that will help you answer your scientific question

Summary

Maintenance of genome stability is a complex endeavour that requires interplay of multiple pathways

Cells use sophisticated mechanism in deciding which pathway to use at any given moment

Majority of factors responsible for maintaining genome stability acts in complexes, let those be dynamic or not