













Regulation of transcription occurs via specific interaction of both general and tissue specific transcription factors (TFs) with promoter and/or enhancer sequences.

The scheme above shows simplified subsequent formation of the complex of TFs involved in the regulation of transcription. Interaction of general TFIID with the TATA box induces distortion of the DNA structure (see the next slide).



Induction of structural changes upon interaction of TFIID with DNA. This may be important for the assembly of other TFs involved in the formation of transcription initiation complex.

This change of confirmation provides a kind of "signature" that is recognized by other proteins and NA polymerase to recognize the proper binding site. However, there are also TATA box-less promoter, where probably other types of "signatures" occur.



The scheme showing the formation of the transcription initiation complex and the interaction of both positive (open symbols) and negative (solid symbols) factors.

These proteins bind to the regulatory sequences that might be hundreds or even thousands of base pairs away from the promoter. These protein interact with each other and with the RNA polymerase, integrating thus many signals into a "yes" or "no" response of the basal promoter, i.e. the region adjacent to the TATA box and recognized by the RNA polymerase.

The individual positive or negative factors are complex and their activity might be regulated by their phosphorylation status or via their interaction with other proteins (i.e. momomeric or dimeric) etc..



There is a whole family of transcription activating factors (TAFs) that interact with signalling molecules, e.g. steroid hormones, thyroid hormones or retinoic acid and in a response to the signal transfer to the nucleus where they regulate transcription.

One of the type of TAF are leucine zipper or bZIP type TAFs. These TAFs are dimeric, with leucine-rich hydrophobic face formed by the Leu that occurs every 7^{th} aa.

That allows the factor to take the proper configuration, which provides the dimer with the ability to bind DNA via charged aa.



An example of the "microprocessor"-like acting promoter is a promoter of the *endo16* gene from the sea urchin.

There have been identified several gene regulatory modules in the *endo16* gene that have positive or negative regulatory role. These modules were identified via formation of deletion mutants of the transcriptional fusions with reporter gene.

The analysis has revealed that the module A has a positive function and must interact with its cognate TAFs for transcription to occur.

Module G enhances the expression when the A and B are active.

C, D, E and F are responsible for the specificity of the expression of endo16 during sea urchin development.

Each of the modules has several protein interaction sites, some of them general, other unique. Site for the protein SpGCF1 is present in many modules and is probably responsible for looping of chromatin, allowing thus bringing of distal regulatory modules close to the basal promoter.

This type of regulation, i.e. based on the different activities of diverse regulatory sequences is sometimes called *combinatorial* and is common for development of many living creatures.

In the combinatorial type of regulations, some modules may act synergistically, some of them antagonistically, some may have both positive and negative roles (e.g. the module B, see the figure). This variability allows very precise and responsive regulations towards changing environmental conditions.



An example of the combinatorial gene regulation is the regulation of β -globin type of hemoglobin chains of humans.

As discussed in the Lesson 5 (course Bi8940 Developmental biology), the type of hemoglobin produced by the fetus changes during development. The hemoglobin present in the liver-produced hemoglobin is composed of two α - and two β -type chains. The β -type hemoglobin chains are of several developmental types, produced by ϵ , $\gamma 1$, $\gamma 2$ and β (in this order). In addition, there is minor adult type of β -type hemoglobin, called δ globin.

The genes for the β -type chains are aligned on the chromosome in the order, in which they are expressed during development (see the figure).

For the expression of individual cell types is distinctive an upstream regulatory sequence called locus control region (LCR). LCR is located about 50 kbp away from the most proximal ϵ gene.

The LCR structure is different in erytrocyte precursor cells in comparison to other cells that could be demonstrated by the changes in the sensitivity to low concentrations of DNase, suggesting low amount of nucleosomes bound.

For the expression of the particular genes, the interaction of their regulatory sequences with LCRs is necessary. Because of LCR can interact only with one regulatory sequence at a time, only one type of genes for the particular β -type chain is activated (the first interaction of LCR with ϵ gene, which is later in development replaced by the other one, is shown by the double-headed arrow).

The underlying molecular mechanisms of the specific pattern of the LCR movement from the most proximal towards the most distal gene cannot be satisfactory explained.

Probably, acetylation of H3 histones might play a role and possibly, other genes outside of the β -type chain family are involved in the regulation of LCR activity. That seems to be confirmed by the identification of other human genes with similar structure, suggesting common regulatory mechanisms via LCRs.







GUS reporter in mouse embryos







Genová exprese

Translační fůze kódující oblasti analyzovaného genu s repotérovým genem

příprava transgenních organismů nesoucích tuto rekombinantní DNA a jejich histologická analýza

oproti transkripční fůzi umožňuje analyzovat např. intracelulární lokalizaci genového produktu (proteinu) nebo jeho dynamiku















Microarray expression profiles of 19 fluorescently sorted GFP-marked lines were analyzed (3–9, 23, 24). The colors associated with each marker line reflect the developmental stage and cell types sampled. Thirteen transverse sections were sampled along the root's longitudinal axis (red lines) (10). CC, companion cells.



(A) The majority of enriched GO terms (hierarchically clustered) are associated with individual cell types (blue bar). A smaller number are present across multiple cell types (green bar). (fig. S2) (**B**) GO category enrichment for hair cells confirms a previous report (15). Enriched cis-elements and an enriched TF family were also identified. (C) From the top 50% of varying probe sets, 51 dominant radial patterns were identified. Pattern expression values were mean-normalized (rows) and log₂ transformed to yield relative expression indices for each marker line (columns). Marker line order is the same for all figures; see table S1 for marker line abbreviations. (**D**) Pattern expression peaks were found across one to five cell types. (E to G) Patterns where expression is enriched in single and multiple cell types support transcriptional regulation of auxin flux and synthesis. In all heat maps with probe sets, expression values were mean-normalized and log₂ transformed. Expression is false-colored in representations of a root transverse section, a cut-away of a root tip, and in a lateral root primordium. (E) Auxin biosynthetic genes (CYP79B2, CYP79B3, SUPERROOT1, and SUPERROOT2) are transcriptionally enriched in the QC, lateral root primordia, pericycle, and phloem-pole pericycle ($P = 1.99E^{-11}$, pattern 5). All AGI identifiers and TAIR descriptions are found in table S14. (F) Auxin amido-synthases GH3.6 and GH3.17 that play a role in auxin homeostasis show enriched expression in the columella, just below the predicted auxin biosynthetic center of the QC (P =8.82E⁻⁴, pattern 13). (G) The expression of the auxin transporter, PIN-FORMED2, and auxin transport regulators (PINOID, WAG1) are enriched in the columella, hair cells, and cortex ($P = 1.03E^{-4}$, pattern 31).



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Schematic flowchart of the Human Protein Atlas. For each gene, a signature sequence (PrEST) is defined from the human genome sequence, and following RT-PCR, cloning and production of recombinant protein fragments, subsequent immunization and affinity purification of antisera results inmunospecific antibodies. The produced antibodies are tested and validated in various immunoassays. Approved antibodies are used for protein profiling in cells (immunofluorescence) and tissues (immunohistochemistry) to generate the images and protein expression data that are presented in the Human Protein Atlas (Ponten et al., *J Int Med*, 2011).































	R Bi	esults c iologica	of –o Illy F	m Re	ic le	s Va	St ant	uc t C	lies Conc	v: clu	s Isi	on	S
		Transcriptiona regulated gen	l profilin nes	gу	ield	ed	more	e the	en <mark>7K</mark>	diff	eren	tially	
											Ddi	i et al., <i>unp</i>	ublish
gene			locus	sample_1	1 sample,	2 status	value_1	value_2	log2(fold_change)	test_stat	p_value	q_value si	ignifica
AT1G07795			1:2414285-2414987	WT	MT	OK	0	1,1804	1.79769e+308	1.79709e+ 308	6 88885e-05	0,00039180 1 y	**
HRS1			1:4556891-4558708	WT	MT	ок	0	0,696583	1.79769e+308	1.79769e+ 308	6.61994e-05	4.67708e- 05 y	**
ATMLO14			1.9227472-9232296	WT	MT	ок		0.514809	1.79769+308	1.79709e+ 308	9.74219e-05	0,00053505	
			1 0400683 0401700	WT	MT	~		0.077044	1 70780300	1.79769e+	1 2492+ 00	3.50131e-	
										1.79769e+			
AT1G27070			1:9070420-9082370 1:22159735-	WT	MT	OK	0	2,0829	1.79709e+308	1.79709e+	9.70039e-00	0.047e-00 y 9.84992e-	es
AT1360095			22102419	WT	MT	OK	0	0,688588	1.79769e+308	308 1.79769e+	9.95901e-08	07 y	85
T1G03020			1:098200-098515	WT	MT	OK	0	1,78859	1.79769e+308	308	0,00913915	0,0277958y	es
T1G13609			1:4662720-4663471	WT	MT	OK	0	3,55814	1.79769e+308	308	0,00021683	0,00108079y	85
T1G21550			1:7553100-7553876	WT	MT	OK	0	0,562868	1.79769e+308	1./9/09e+ 308	0.00115582	1 0.00471497 y	es
T1G22120			1:7808308-7809832	WT	MT	OK	0	0,617354	1.79769e+308	1.79769e+ 308	2.48392e-08	1.91089e- 05 y	25
T1G31370			1:11238297- 11239383	WT	MT	OK	0	1.46254	1.79789e+308	1.79769e+ 308	4.63523e-05	0.00028514 3v	**
PLIMID			1:13253397-	WT	MT	OK		0 501031	1 797694-108	1.79769e+	7 878554.04	5.40003e-	
			1:18010728-							1.79789e+		0.00037473	
411048700			1:21746209-	WI	MI	UK	0	0,000020	1./9/09e+308	1.79769e+	0.5391/e-05	٥y	es
AT1G59077			21833195 1:22121549-	WT	MT	OK	0	138,885	1.79709e+308	308 1.79769e+	0,00122785	0,00496816 y	*5
AT1G80050			22123702	WT	MT	OK	0	0,370087	1.79769e+308	306	0,00117953	0,0048001 y	25
474G15242			4.5705786-5706997	WT	MT	OK	0.00930712	17,9055	10,9098	-4,4052	31.05673e-05	7.13983e-05	yes
T5G33251			12500433	WT	MT	OK	0.0496375	52,2837	10,0349	-9,811		0	yes
T4011610			4:7421055-7421738	WT	MT	OK	0,0195111	15,8518	9,66612	-3,9004	39.60217e-05	0,000528904	yes
NITO ILOLO			1-22100651										
AT1060020			1:22100651- 22105276	WT	MT	OK	0.0118377	7,10023	9,24011	-7,5038	20.195044-14	1.4000e-12	yes

Excample of an output of transcriptional profiling study using Illumina sequencing performed in our lab. Shown is just a tiny fragment of the complete list, copmprising about 7K genes revealing differential expression in the studied mutant.





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