



LÉKAŘSKÁ FAKULTA MASARYKOVY UNIVERSITY  
Interní hematologická klinika LF MU a FN Brno  
Centrum molekulární biologie a genové terapie



# Analýza exprese microRNA

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University of California-San Diego a CEITEC MU

3/13 Moderní metody analýzy genomu



INVESTICE DO ROZVOJE VZDĚLÁVÁNÍ

Tato prezentace je spolufinancována  
Evropským sociálním fondem  
a státním rozpočtem České republiky

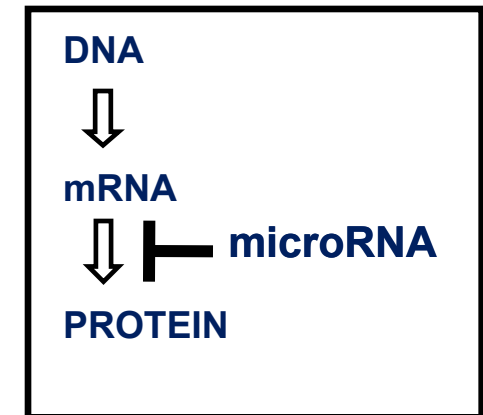
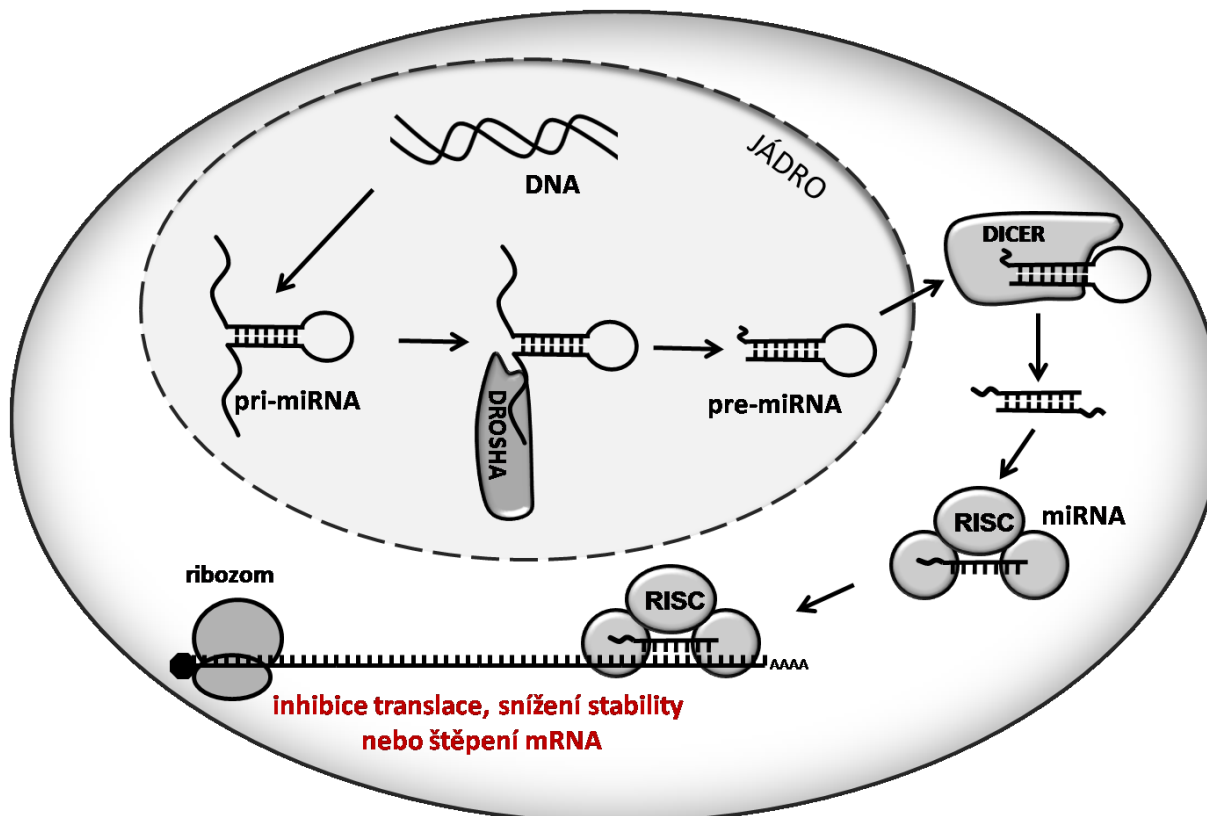
- Lidské miRNA geny: **cca 2000**

## microRNA (miRNA)

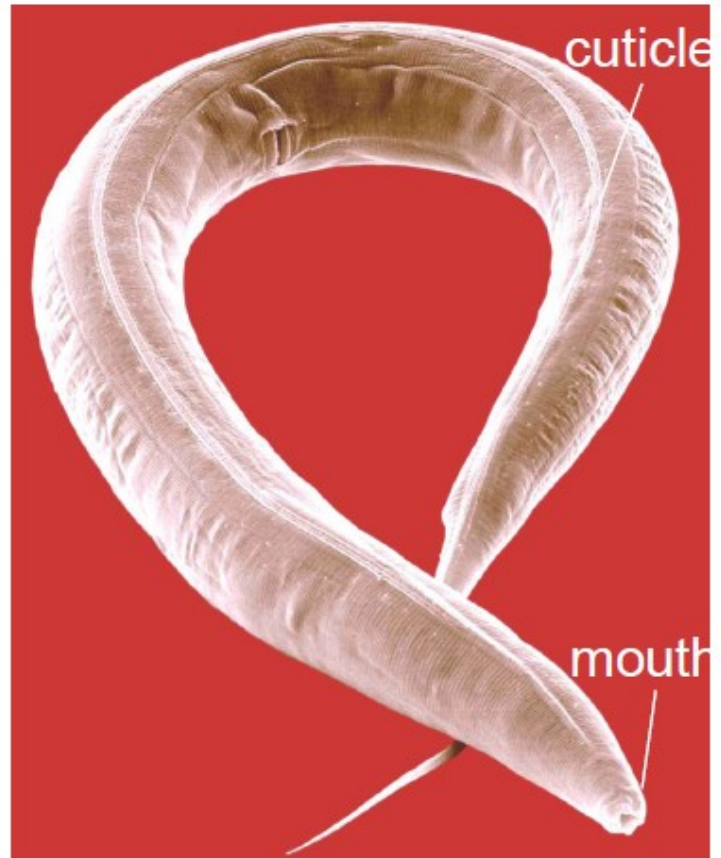
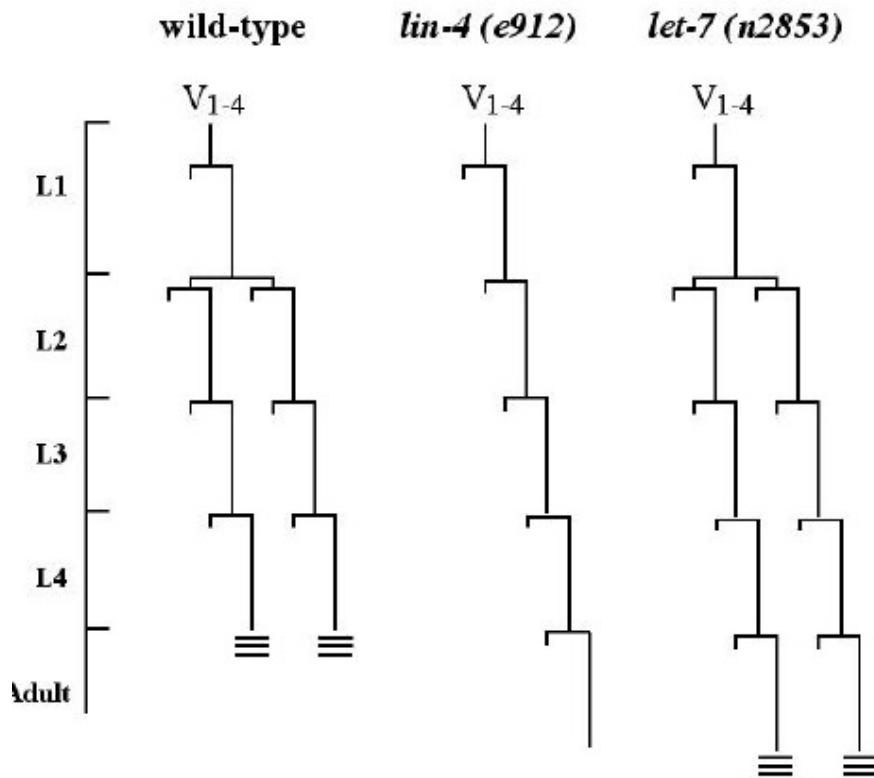
- ❑ krátké RNA molekuly  
~22 nukleotidů
- ❑ komplementární vazba k  
cílové mRNA
- ❑ inhibují translaci a snižují  
stabilitu mRNA



**Stovky evolučně  
konzervovaných microRNA**

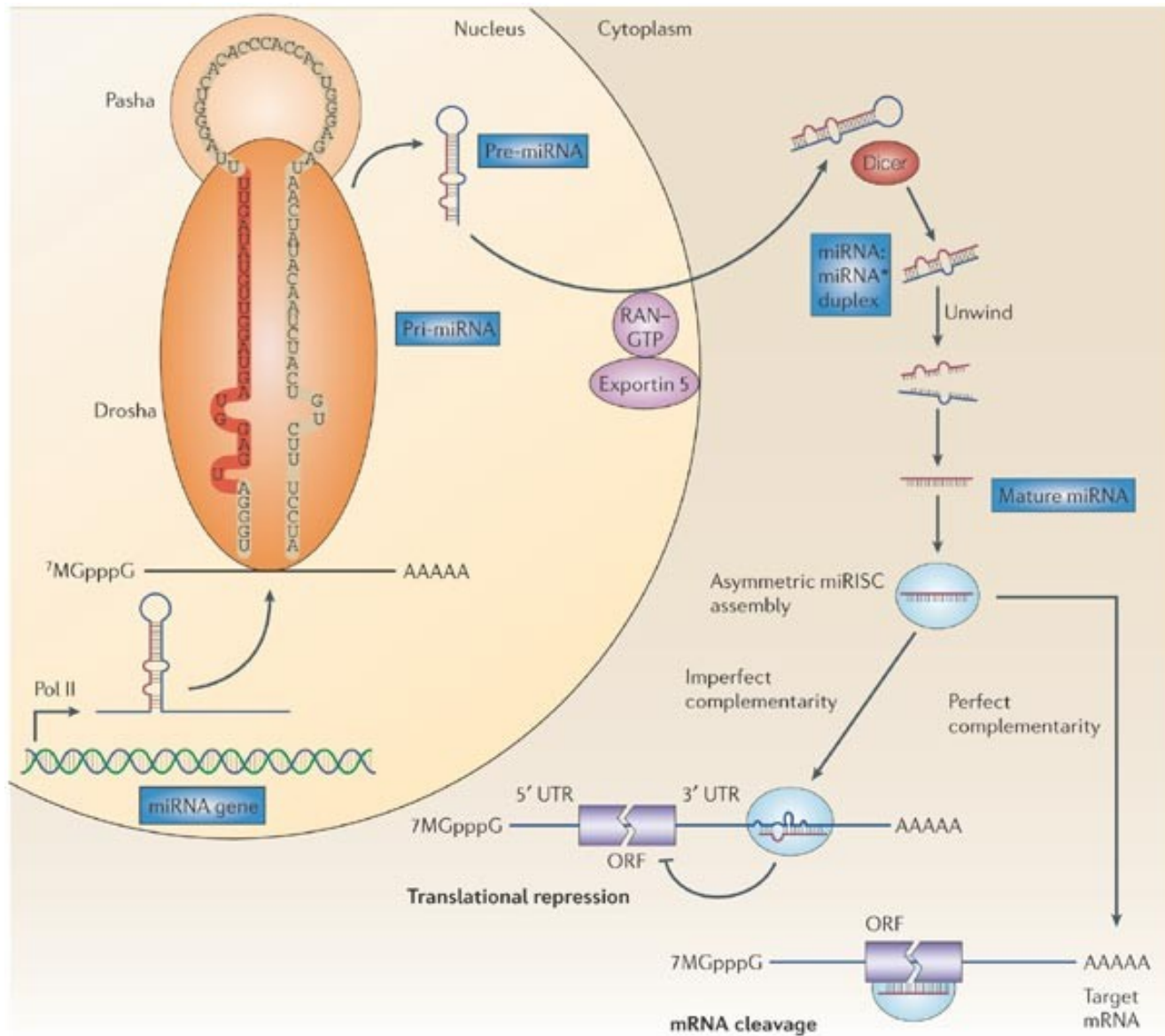


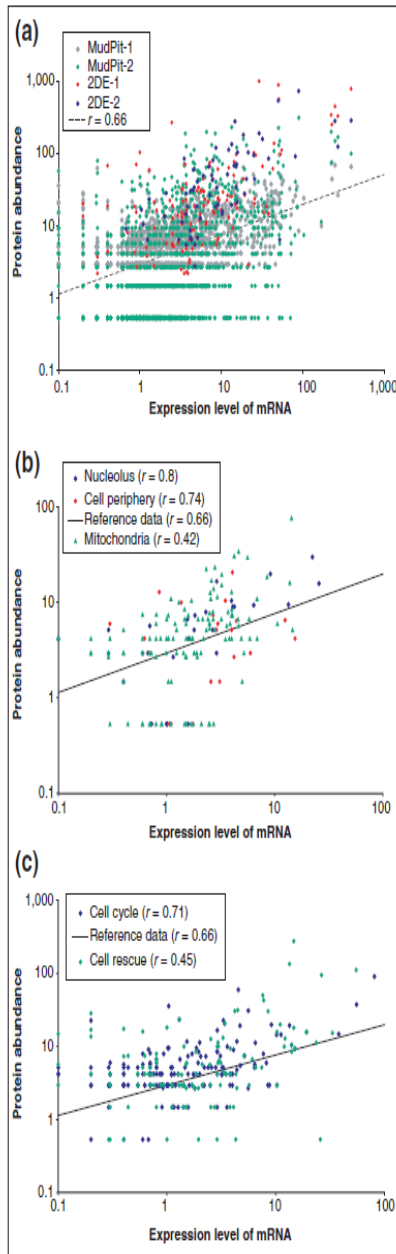
# MicroRNAs were discovered by V. Ambros and G. Ruvkun in *C. elegans*



Lee et al. 1993 *Cell*

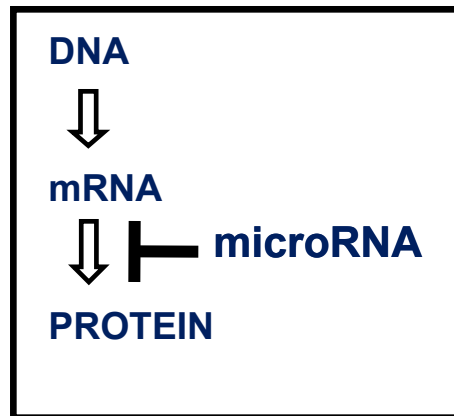
Reinhart et al. 2000 *Nature*





mRNA neznamená, že v buňce bude i protein

Historicky vždy velká neshoda mezi daty z expresních čipů a expresí proteinů (Western Blot)



## Specifika analýzy exprese microRNAs:

- velmi malé molekuly – 22nt – specifikum izolace, specifické značení i design sond
- malé zastoupení ve vzorku – separace microRNA
- v lidském genomu cca 2000 genů
- některé mají velmi podobnou sekvenci – rozdíl 1nt
- pre-miR, pri-miR, mature-miR
- málo se ví o jejich funkcích – obtížná interpretace výsledků
- zatím málo zkušeností a standardizace

Isolace

Microarrays

Identifikace miRNA (deep sequencing, cloning a Northern blot)

Real-Time PCR

## 1/ Izolace a stabilita microRNA

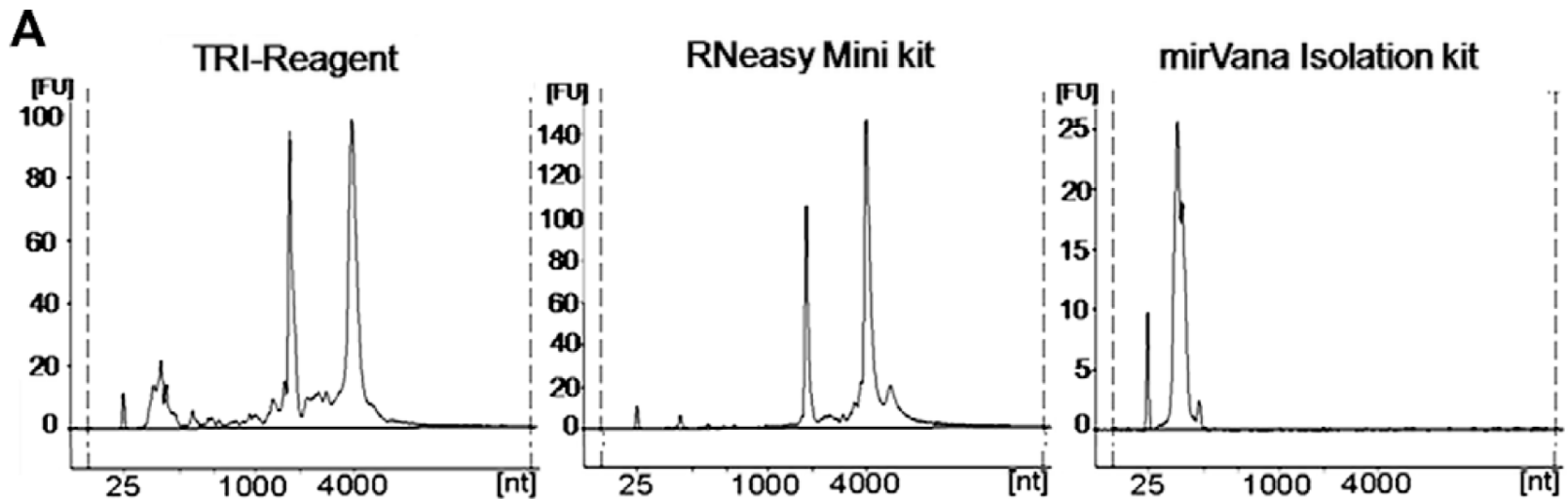
Problémy: velikost 22nt, celkově cca 0,01% z celkové RNA

### Izolace:

TRizol/TriReagent  
miRvana (Ambion)  
PureLink (Invitrogen)  
a další

### Obohacení:

PAGE  
FlashPAGE Fractionator  
(Ambion)



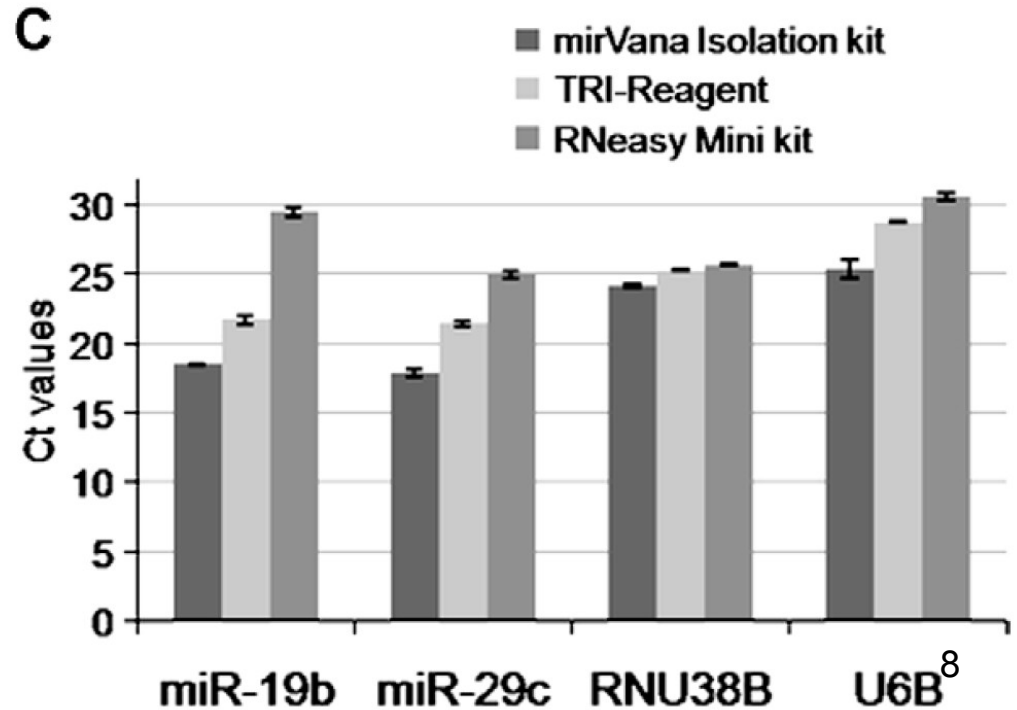
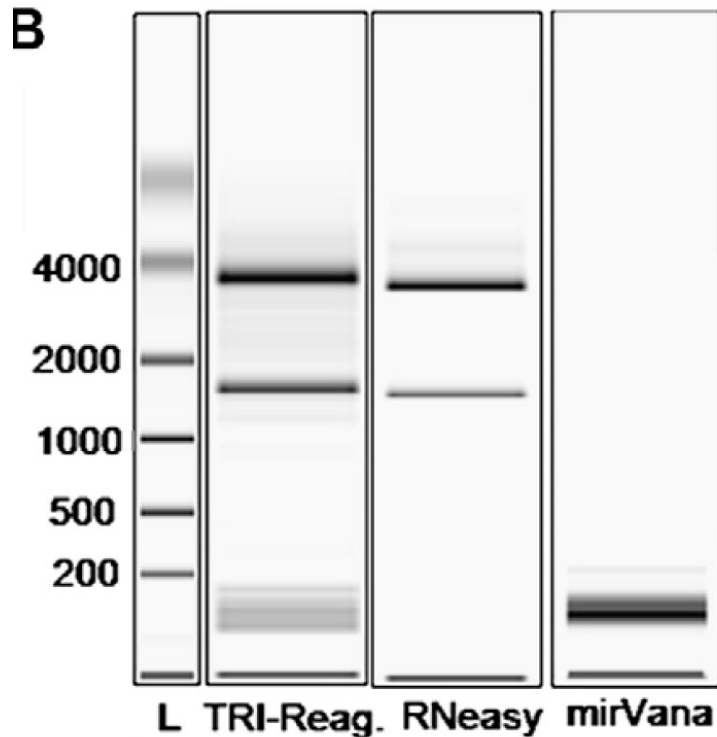
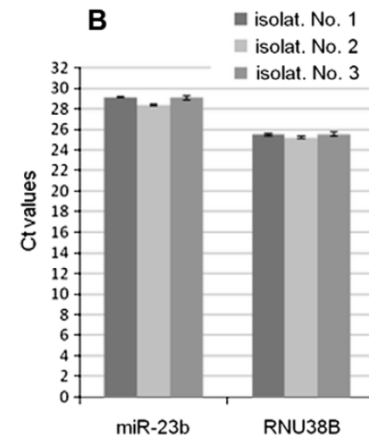
Mraz et al., 2009

## Izolace:

TRizol/TriReagent  
miRvana (Ambion)  
PureLink (Invitrogen)  
a další

## Izolace:

TRIReagent/TRIzol  
is the „gold standard“  
(Mraz et al., 2009)





# Obohacení:

## PAGE

### FlashPAGE Fractionator (Ambion)



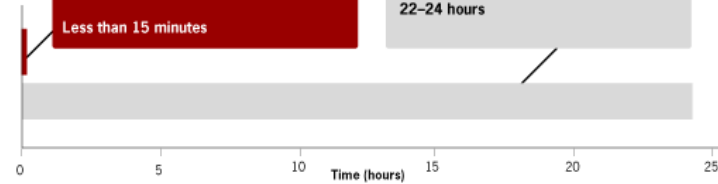
#### flashPAGE™ Protocol

1. Pipet flashPAGE™ Lower Running Buffer into the lower buffer chamber of the apparatus  
<30 seconds
2. Insert a "ready-to-use," pre-cast flashPAGE™ Gel Cartridge  
<5 seconds
3. Add flashPAGE™ Upper Running Buffer to the upper buffer chamber of the gel cartridge  
<30 seconds
4. Add your RNA or DNA sample (with flashPAGE™ A40 Dye Marker)  
<1 minute
5. Run gel at 70 V on any standard power supply  
~12 minutes
6. Collect PAGE-purified nucleic acid from lower buffer chamber\*  
<30 seconds

Less than 15 minutes

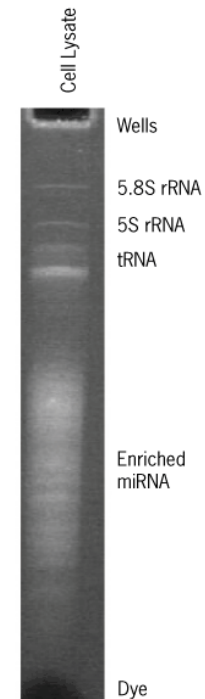
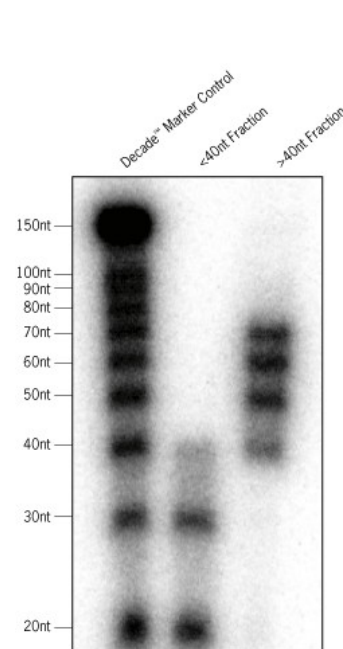
#### Traditional PAGE Purification

1. Prepare gel solutions  
30 minutes
  2. Cast gel  
2 hours
  3. Pre-run the gel  
30 minutes
  4. Load sample  
1 minute
  5. Electrophorese  
30-60 minutes
  6. Stain gel to visualize region of interest  
10 minutes
  7. Excise desired size fraction  
5 minutes
  8. Soak crushed gel with elution buffer  
overnight
  9. Collect first elution and elute again  
2 hours
- 22-24 hours



15min

20hours

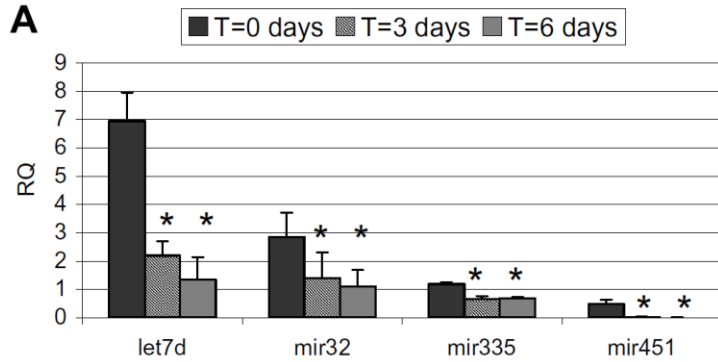


# Stabilita microRNA :

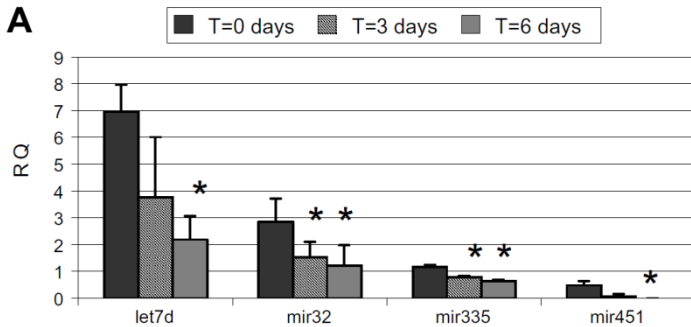
## Stabilita po izolaci

## Stabilita v FFPE (formalin-fixed parafin-embedded tissue)

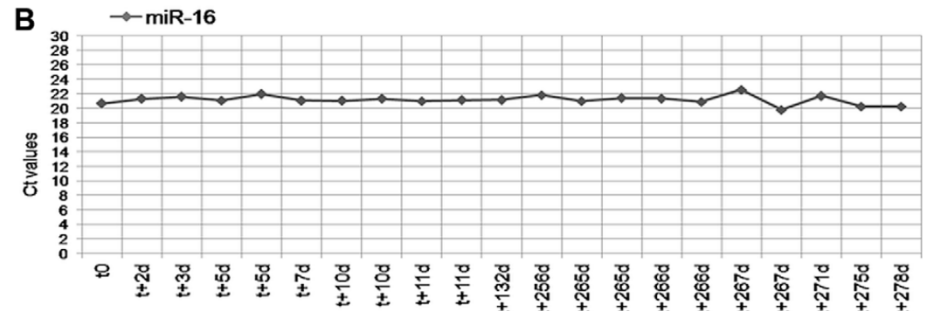
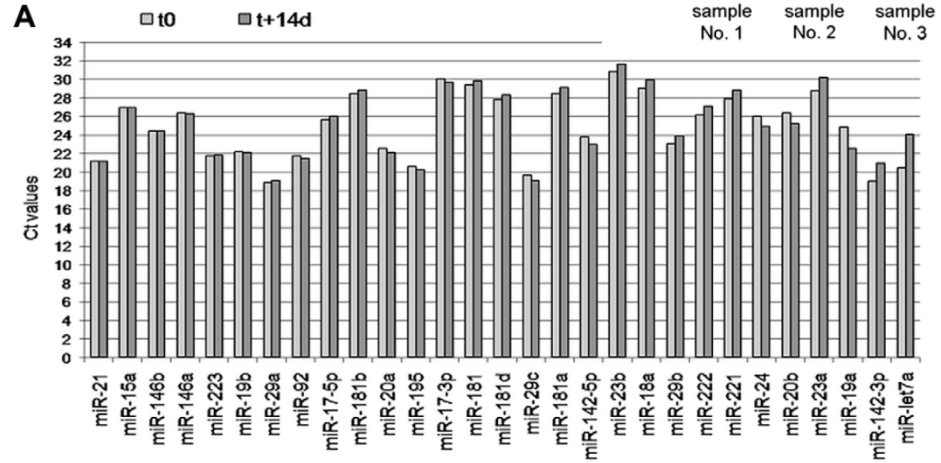
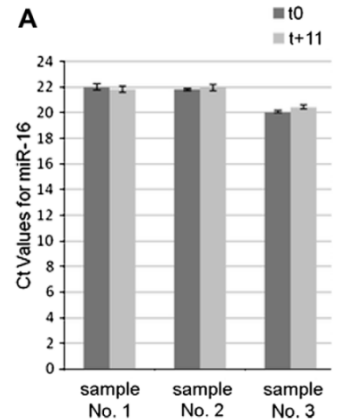
### RNA



### cDNA



Bravo et al., 2007



Mraz et al., 2009

# Stabilita microRNA :

## Stabilita v FFPE (formalin-fixed parafin-embedded tissue)

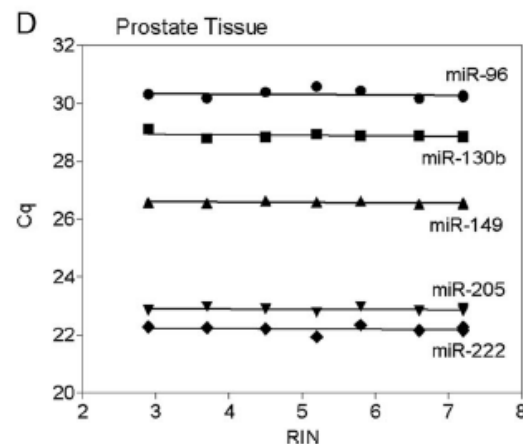
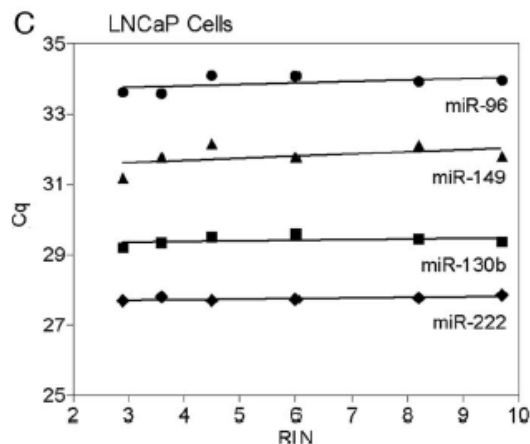
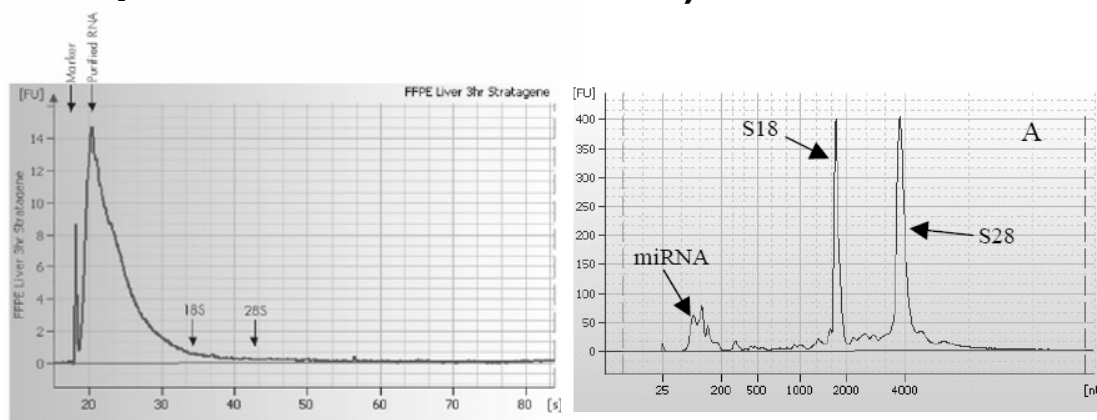
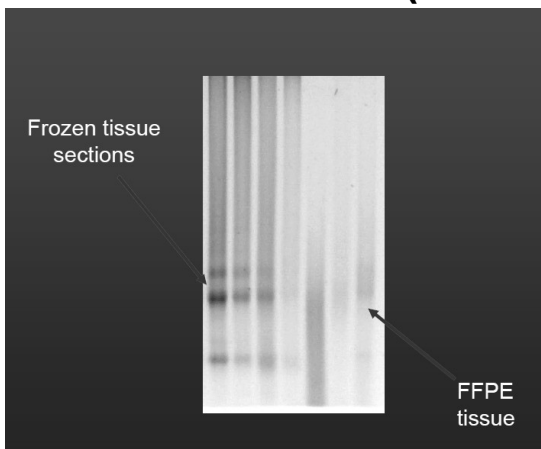
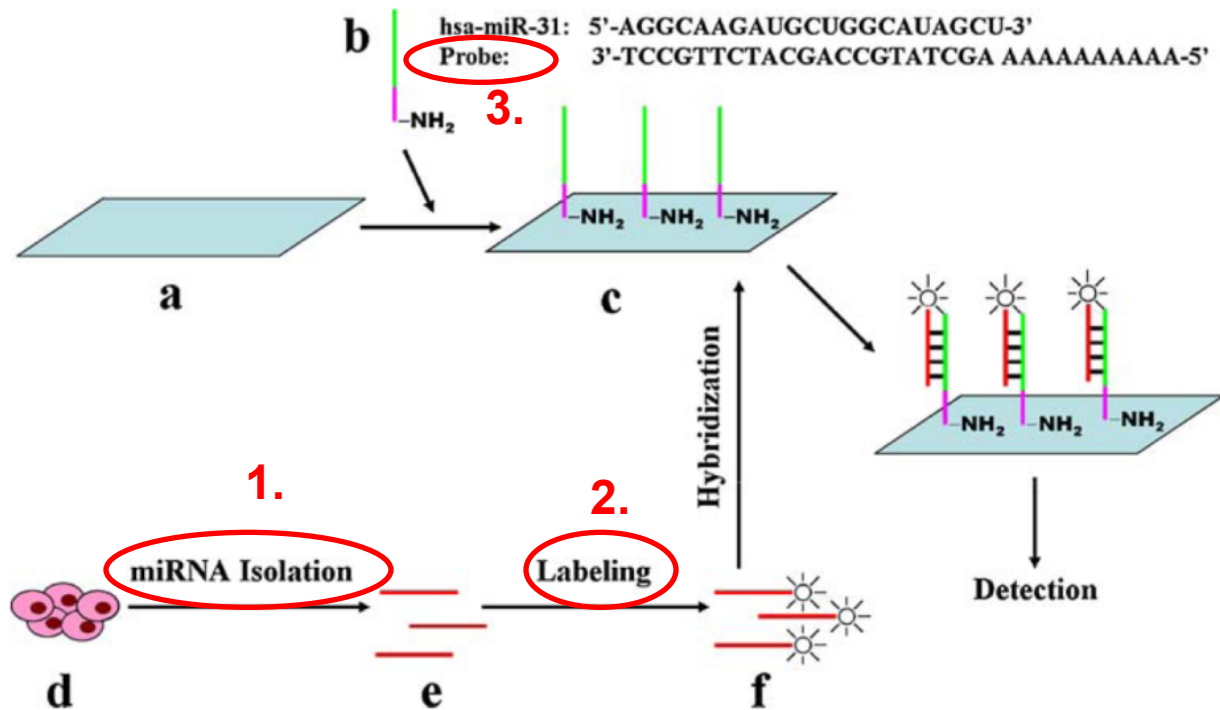


Fig. 2. Influence of RNA integrity on miRNA gene expression.

(A), miR-141, miR-155, miR-200c, and miR-210 in RNA samples from ccRCC cell line Caki-2. (B), miR-141, miR-155, miR-200c, and miR-210 in RNA samples from the renal tissue pool. (C), miR-96, miR-130b, miR-149, and miR-222 in RNA samples from PCa cell line LNCaP. (D), miR-96, miR-130b, miR-149, miR-205, and miR-222 in RNA samples from the prostate tissue pool. For further details, including regression line characteristics, 95% CIs of the slopes, and *P* values indicating significant deviations from 0, see Table 5 in the online Data Supplement.

## Expression microarrays pro microRNAs:

- velmi malé molekuly – 22nt – specifika izolace, specifické značení i design sond
- malé zastoupení ve vzorku – separace microRNA
- v lidském genu cca 2000 genů – menší počet sond na čipu
- některé mají velmi podobnou sekvenci – rozdíl 1nt
- pre-miR, pri-miR, mature-miR
- málo se ví o jejich funkcích – obtížná interpretace výsledků
- zatím málo zkušeností a standardizace



### 3/ Labeling – značení:

- ❑ Není možný labeling pomocí značených polyT při reverzní transkripci
- ❑ Přímé značení (direct labeling) – většinou nějaká fluorescenční barva
- ❑ Nepřímé značení (indirect labeling) – probíhá nějaká reverzní transkripce/PCR

#### Přímé značení:

Jednoduché, rychlé a „čím méně kroků tím méně vnesených chyb a variability“

#### 1/ Značení guaninu v microRNA

Fluochromem vážícím se na guanin jsou označeny miRNA (Ulysis Alexa Flour 546/647)

Všechny lidské miRNA obsahují guanin, ale v různém množství

Nemožnost usuzovat na vzájemnou expresi různých miRNA (různý obsah guaninu)

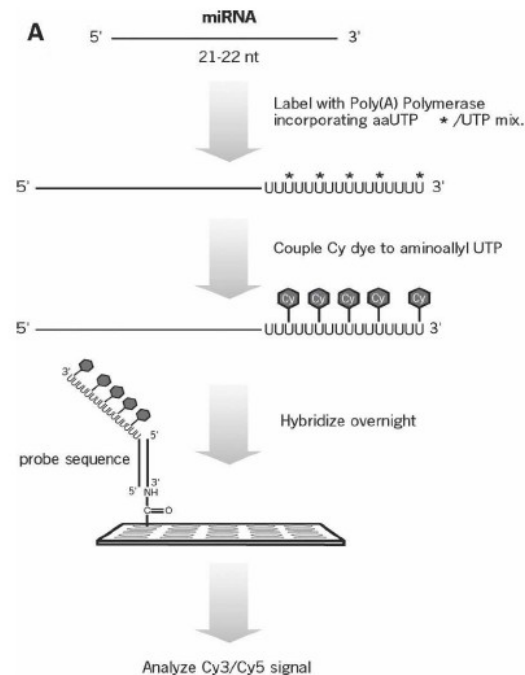
(Babak et al., 2004)

#### 2/ Značení pomocí Poly (A) polymerázy

Můžu se rozhodnout jak dlouhý bude poly(A)

a tím ovlivnit sílu signálu

(Shingara et al., 2005)



### 3/ značení chemickou metodou

3'OH skupina je oxidována na dialdehyd

Následuje reakce s Biotin-X-hydraxidem → Biotinilovaná miRNA

Vazba fluoresceční molekuly-quantum dot

(Liang et al., 2005)

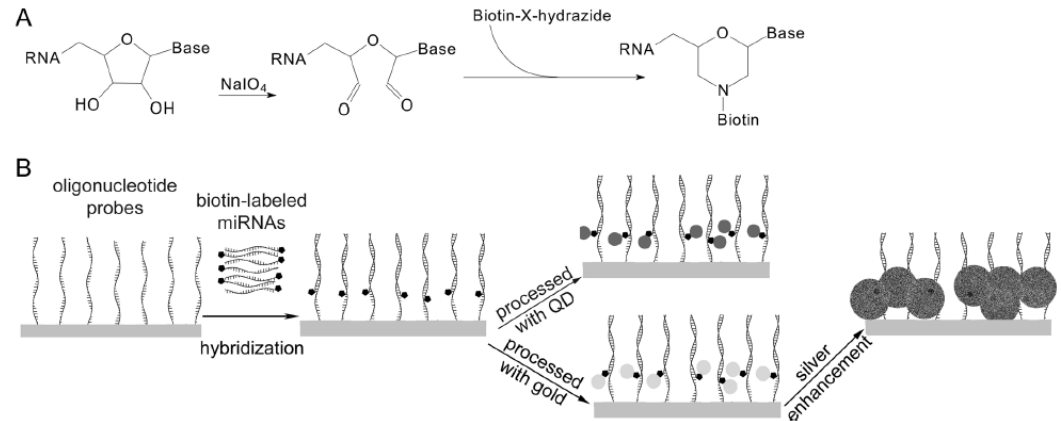


Figure 1. Schematic principles of the miRNA profiling microarray. (A) Principle of labeling miRNA at the 3' terminus with biotin. (B) Principle of the miRNA profiling microarray detected with QD or colorimetric method.

### 4/ značení pomocí T4 ligasy

Krátký značený oligonukleotid

je připojen T4 ligásou k 3'konci

Výhodou je přednostní vazba

na RNA o velikosti 18-30bp → total RNA

(Thomson et al., 2004; Castoldi et al., 2007)

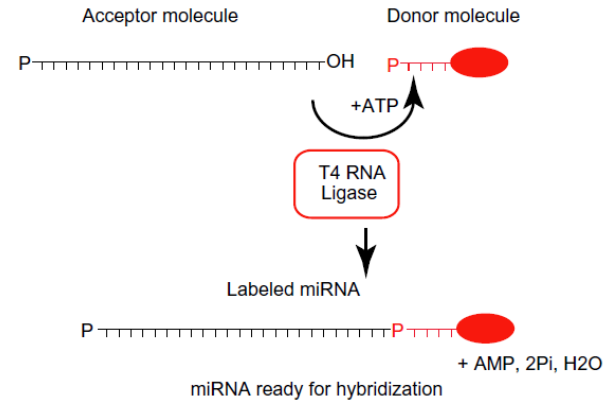


Fig. 2. Schematic representation of the miRNA labeling principle: a short Cy-dye labeled RNA-linker (donor molecule) is ligated to the single-stranded miRNA (acceptor molecule) by T4 RNA ligase in the presence of ATP.

## Nepřímé značení:

Značen je produkt reverzní transkripce či PCR

Výhody: cDNA je pak stabilní a lze uchovat, Pre-amplifikace a tím snadnější detekce méně exprimovaných miRNA

### 1/ značení reverzního transkriptu miRNA

Reverzní transkripce pomocí náhodných 8-merů značených 2 biotiny (3'-(N)8 – (A)12-biotin-(A)12-biotin-5' (Liu et al., 2004)

Reverzní transkripce pomocí náhodných neznačených 7-merů, následně označeny s pomocí terminální transferázy a biotin-dideoxy-UTP (Sun et al., 2004)

Nebezpečí chyb z nespecifické vazby primeru

### 2/ značení produktu RT-PCR

Výhoda: snadná pre-amplifikace

Dva adaptory

fluorescenčně-značený primer (k adaptoru)

(Miska et al., 2004)

Nevýhoda: antisense strand přiromen při hybridizaci

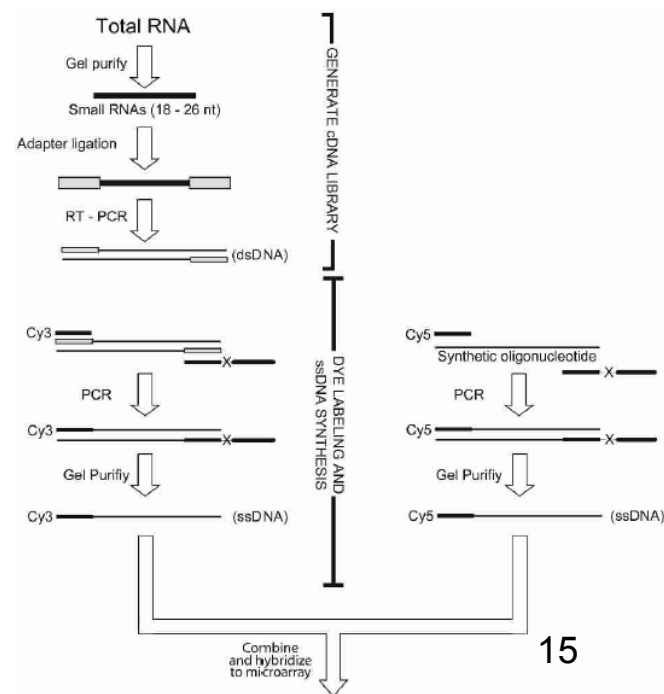
Rešením je různá délka sense a antisense ->PAGE

(Baskerville, 2005)

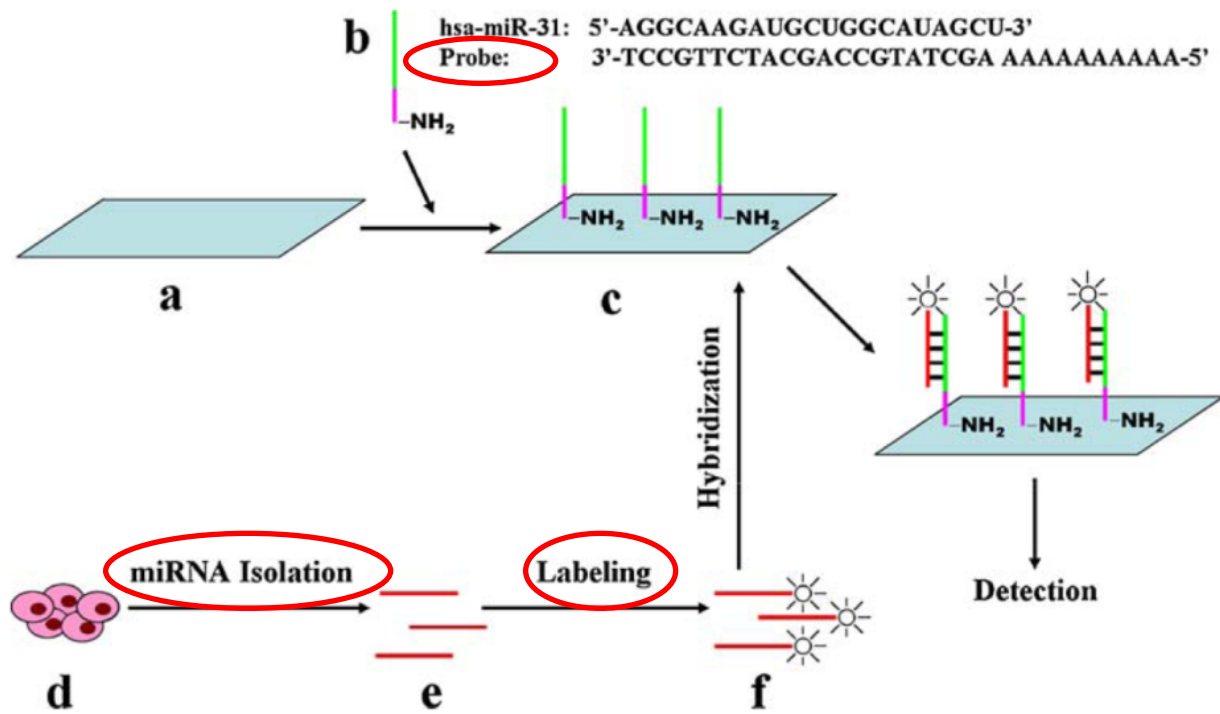
### 3/ značení in vitro transkriptu

Jeden z adaptorů je promotor T7 RNA polymerázy

(Barad et al. 2004)



### 3/ Microarrays/ Próby: Problémy: krátké RNA, malé rozdíly v sekvenci, Tm





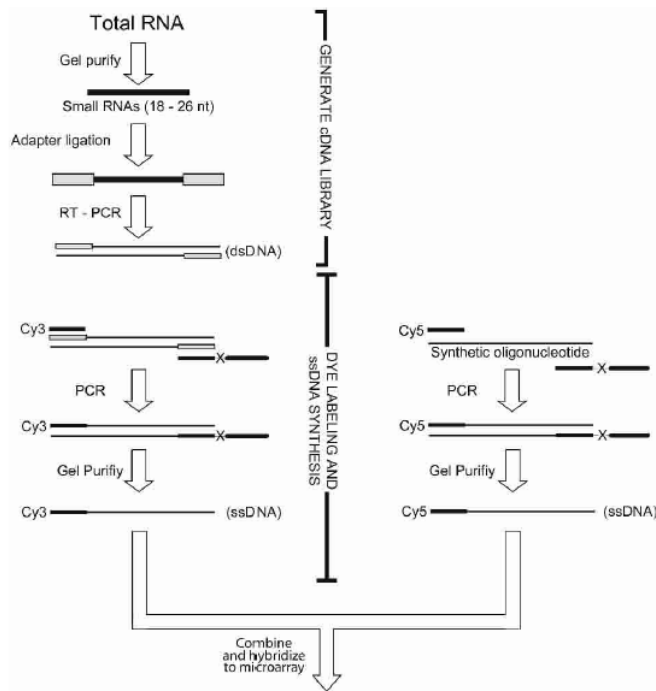
$T_m$  – melting temperature určité próby  
T – hybridizační teplota

$T_m < T$  .....nižší efektivita vazby miRNA  
 $T_m > T$  .....vyšší efektivita vazby miRNA

- ☐ Je třeba navrhnout próby tak, aby měly všechny podobnou  $T_m$
- ☐ To se u „dlouhých“ mRNA řeší vhodným výběrem oblasti genu k němuž bude sonda komplementární nebo délkou sondy
- ☐ navíc některé miRNA jsou téměř sekvenčně totožné

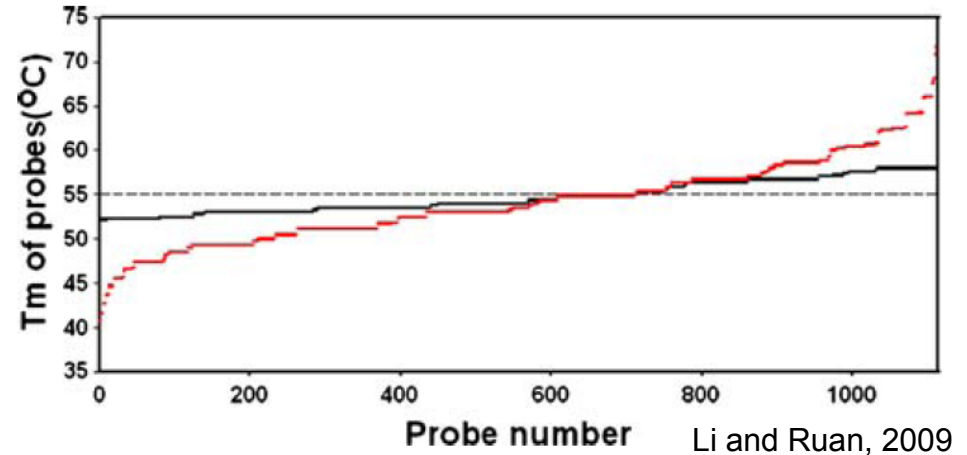
```
let-7b : TGAGGTAGTAGGTTGTGTGGTT : 22
let-7e : TGAGGTAGGAGGTTGTATAGT- : 21
let-7d : AGAGGTAGTAGGTTGCATAGT- : 21
let-7a : TGAGGTAGTAGGTTGTATAGTT : 22
let-7f : TGAGGTAGTAGATTGTATAGTT : 22
let-7i : TGAGGTAGTAGTTTGTGCT--- : 19
let-7g : TGAGGTAGTAGTTTGTACAGT- : 21
      tGAGGTAGtAG TTGt gt
```

# ÚPRAVA DÉLKY



**FIGURE 1.** Microarray sample preparation and reference oligonucleotide synthesis. Small RNAs were fractionated on a polyacrylamide gel, and oligonucleotide primers were then ligated to the 5' and 3' ends of the small RNA library (Lau et al. 2001). A cDNA library was generated through reverse transcription, and the product was amplified using PCR. Using a pair of modified oligonucleotide primers in a second PCR, the sense strand of the library was fluorescently labeled and the antisense strand was selectively lengthened (Williams and Bartel 1995). The sense strand of the asymmetric duplex was purified away from the antisense strand in a denaturing gel, and this purified dye-labeled ssDNA sample was used for hybridization and detection on the array. At each feature, the signal from the miRNA sample was compared to that from a reference sample, which had been generated by amplifying and labeling synthetic oligonucleotides using the same strategy as for the miRNA sample.

Baskerville and Bartel, 2005

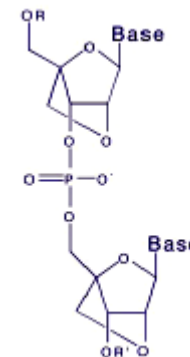


**Fig. 2** T<sub>m</sub> (melting temperature) distribution for microRNA probes for human, rat and mouse. *Red and black curves* represent the T<sub>m</sub> distributions of the raw and normalized probes, respectively

# ÚPRAVA SÍLY VAZBY NUKLEOTIDŮ

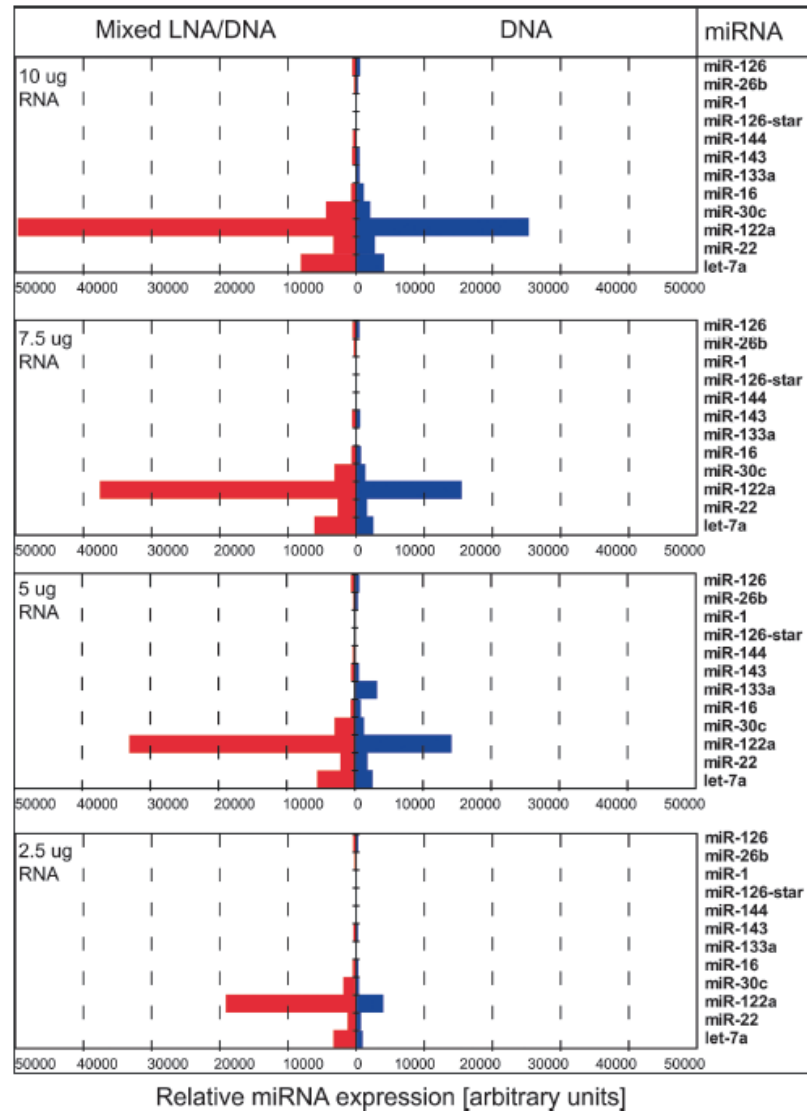
LNA próby (Locked Nucleic Acid)

ribózový kruh je „uzamčen“ methylenovým můstkem mezi atomy 2'-O a 4'-C



Použití LNA pro některé báze v průběh

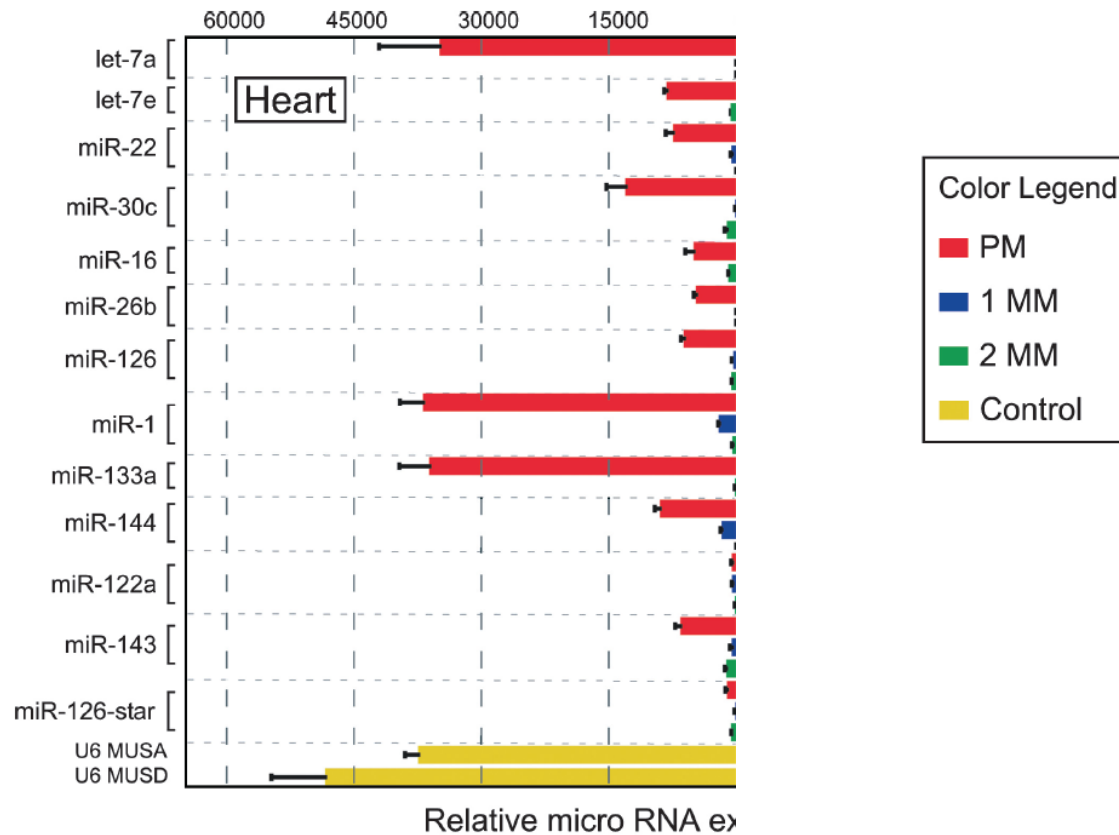
**SÍLA VAZBY:  
LNA vs DNA próba  
Tm až 72°C  
(Castoldi et al., 2006)**



**FIGURE 1.** Mixed DNA/LNA capture probes display increased sensitivity for miRNA detection. miRNA expression was assessed in murine liver using a test set of LNA-modified (*left*) or unmodified DNA oligonucleotide capture probes (*right*). Decreasing amounts of total RNA were used as input material for miRNA analysis. Data are presented as median intensity (four replicas per miRNA capture probe; a representative experiment is shown).

# SPECIFITA VAZBY: LNA vs DNA próba

(Castoldi et al., 2006)



# miRCURY LNA Array, Exiqon : 3 dny

## Protocol overview

### miRCURY™ LNA microRNA Power Labeling Kit

CIP treatment

Mix: RNA sample  
Spike-In miRNA kit



Labeling reaction

Mix: CIP'ed RNA sample  
Labeling buffer  
Hy3™ or Hy5™  
DMSO  
Enzyme



### miRCURY™ LNA microRNA Array Kit

Mix samples

Mix: Hy3™ labeled sample  
Hy5™ labeled sample  
Hybridization buffer  
Denature sample



Hybridize

Hybridize at 56°C for 16 hours



Stringency wash

Wash 2 min. in buffer A at 56°C  
Wash 2 min. in buffer B at 23°C  
Wash 2 min. in buffer C at 23°C  
Dry slides



Image acquisition

Scan slides (recommended scan at 5µm)  
Download relevant GAL files from  
[www.exiqon.com](http://www.exiqon.com)

# Co se nemusí podařit:

**Nekvalitní RNA**

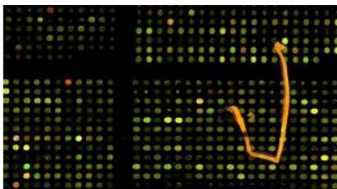
**Nepodaří se značení**

**Nepodaří se hybridizace**

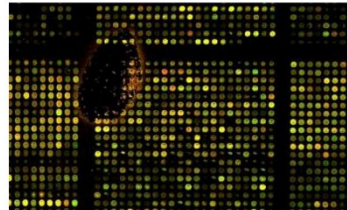
**Nepodaří se promývání**

**Technická variabilita čipů je větší než ta biologická**

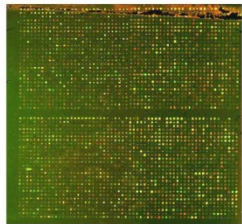
**Nepodaří se validace dat pomocí RT-PCR, atd**



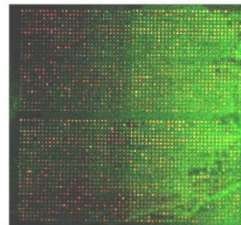
Fiber or scratch?



Bubble



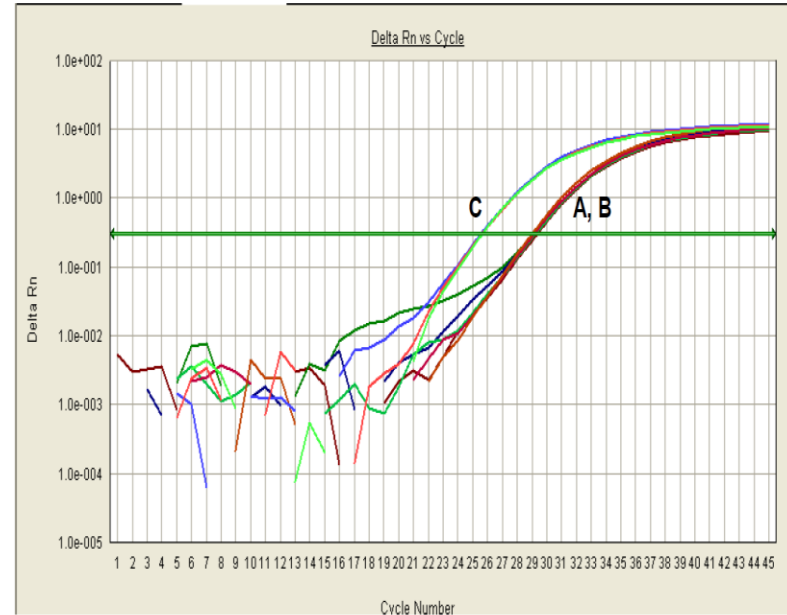
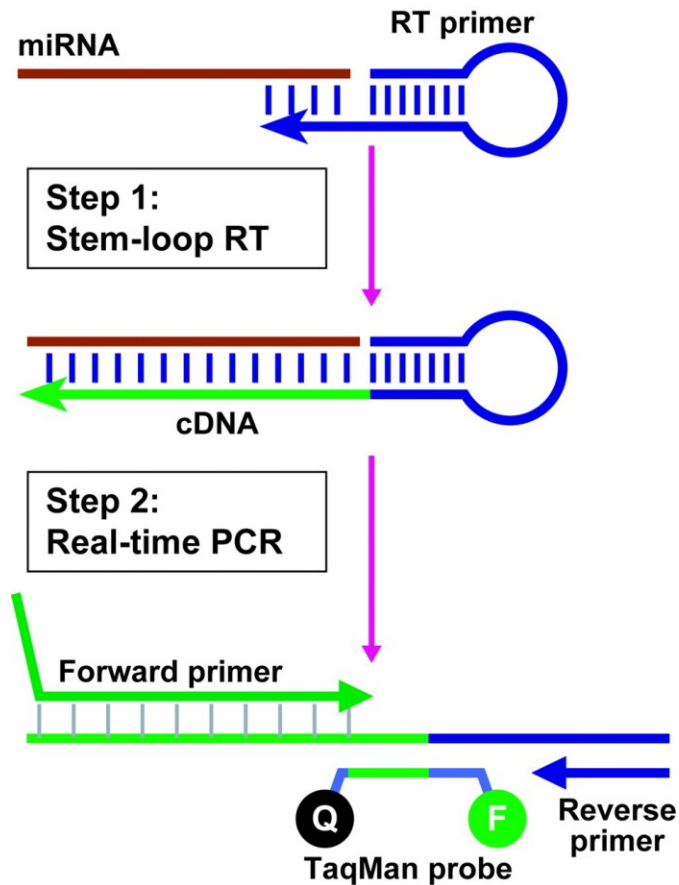
Edge effect



Background haze

Práce s miRNA čipy je velmi obtížná. Všeobecně nižší míra standardizace. Obtížná interpretace získaných dat z pohledu biologického smyslu např. deregulace několika miRNA (nádor vs. zdravá tkáň apod.)

# TaqMan-based real-time PCR quantification of mature miRNAs



# Konstrukce cDNA knihovny malých RNA

Cloning nebo deep sequencing  
Validace pomocí Northern blot

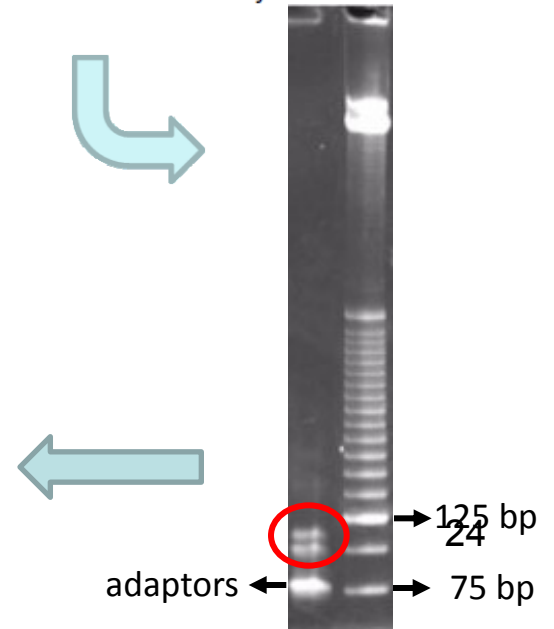
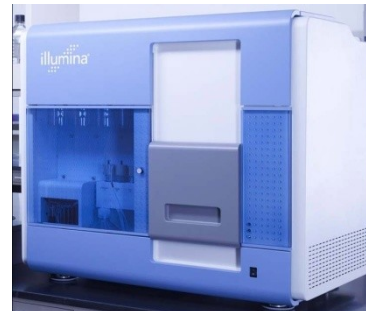
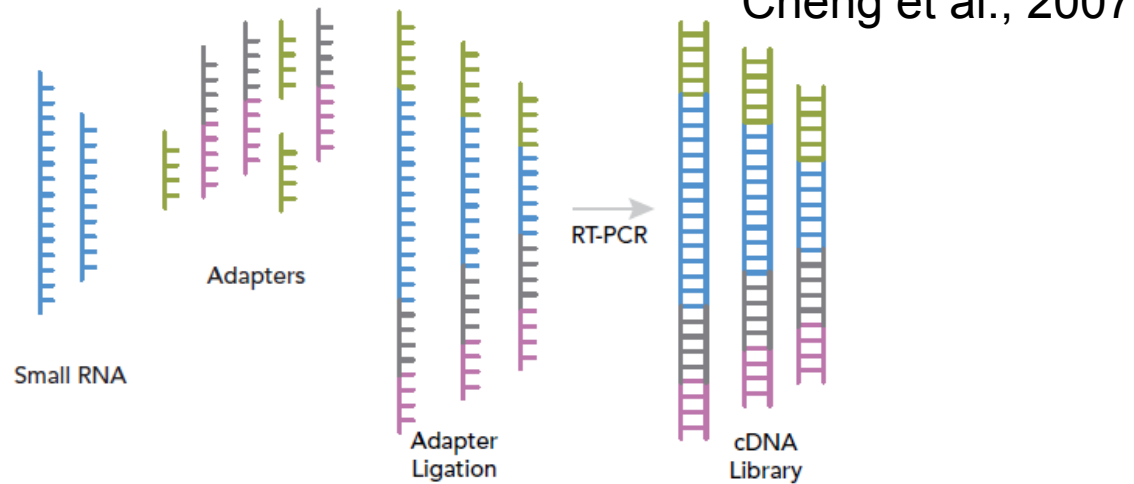
3' RNA adapter ligation

5' RNA adapter ligation

RT-PCR amplification

Purify small RNA library

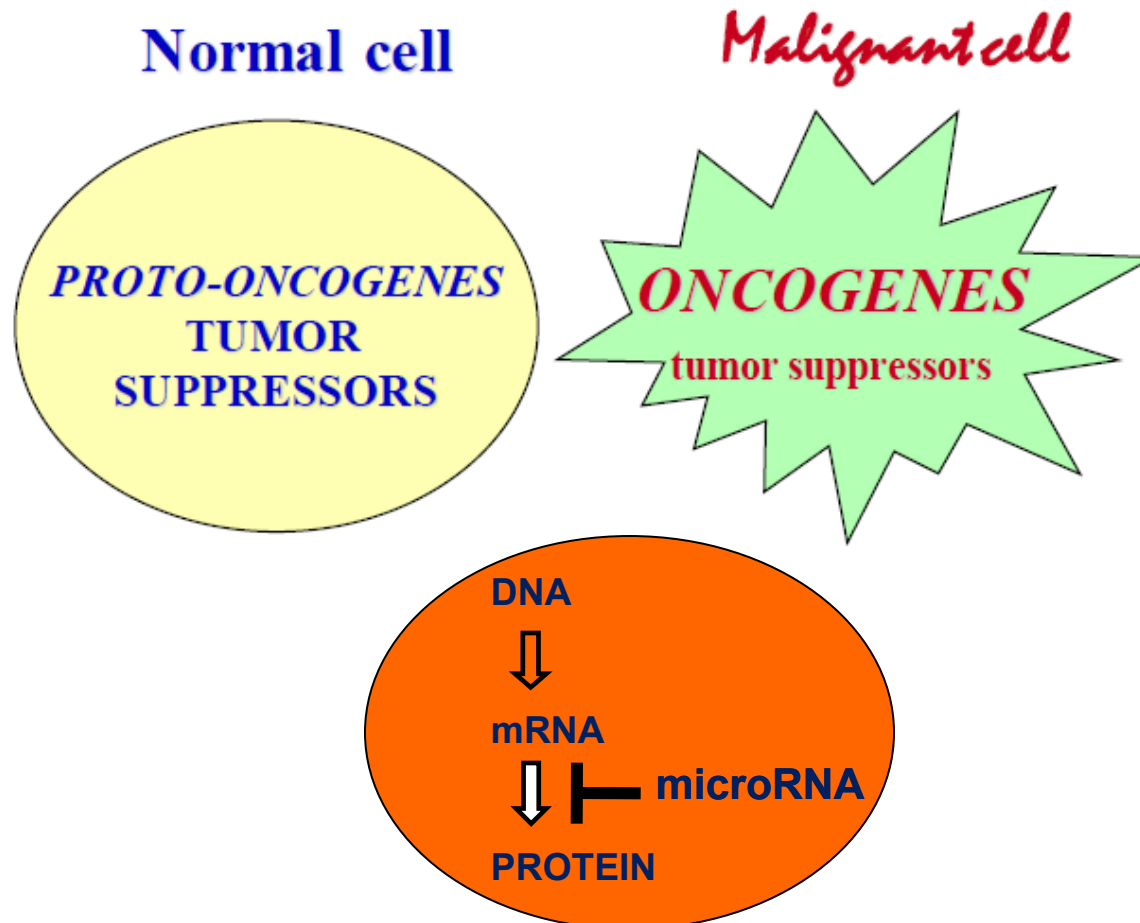
Cluster generation and sequencing (Cluster Station and Genome Analyzer )



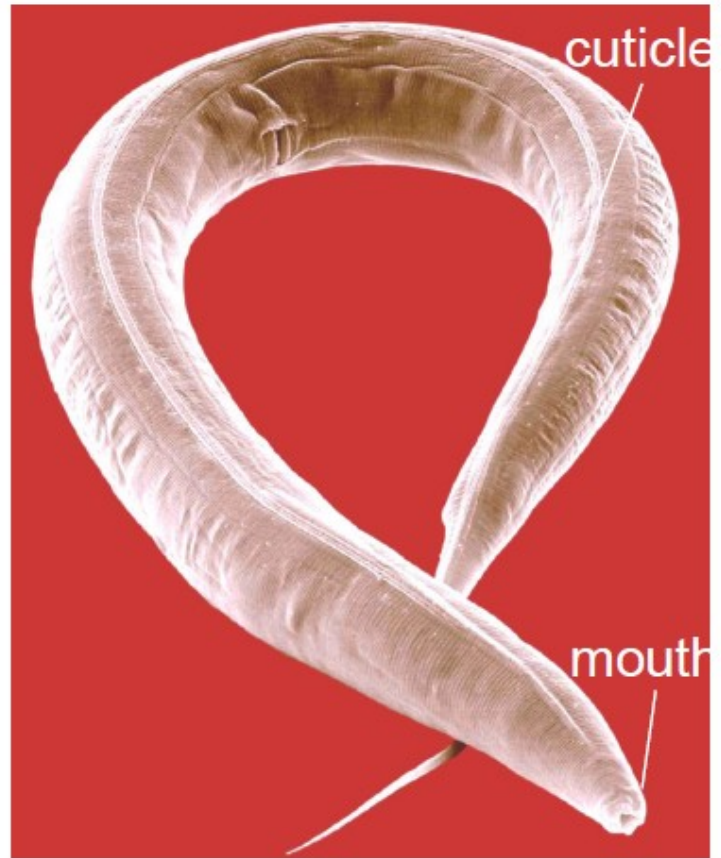
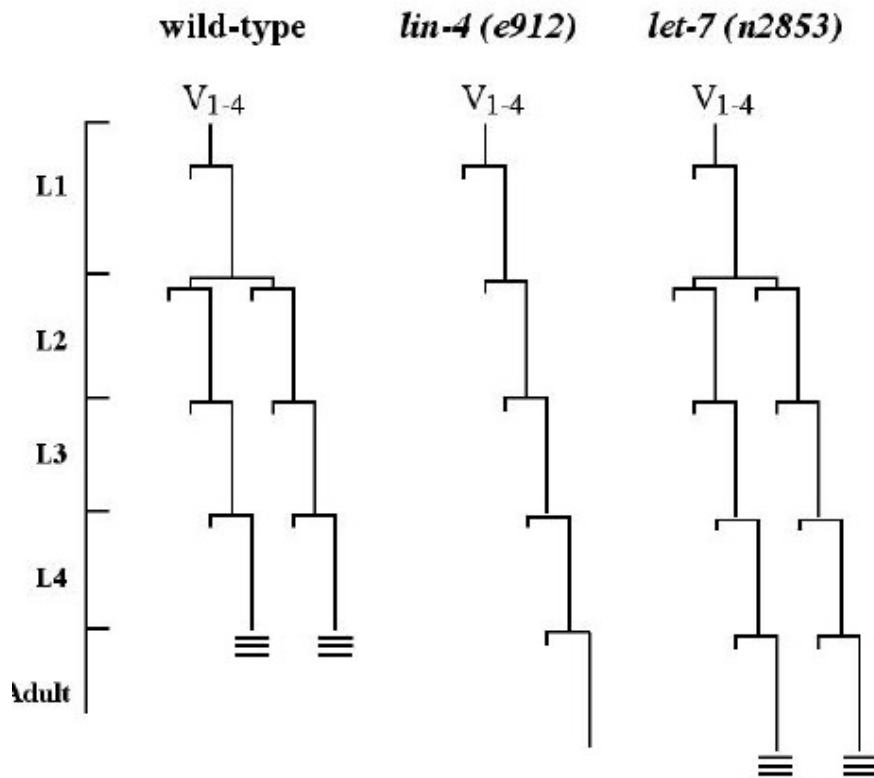


# Year 2K “Central dogma” of molecular oncology

Cancer is the GENETIC DISEASE with the most complex mechanism.  
Oncogenes and Tumor-suppressors are the two types of PROTEINS deregulated in cancer cells.



# MicroRNAs were discovered by V. Ambros and G. Ruvkun in *C. elegans*



Lee et al. 1993 *Cell*

Reinhart et al. 2000 *Nature*

# microRNA genes map to cancer loci

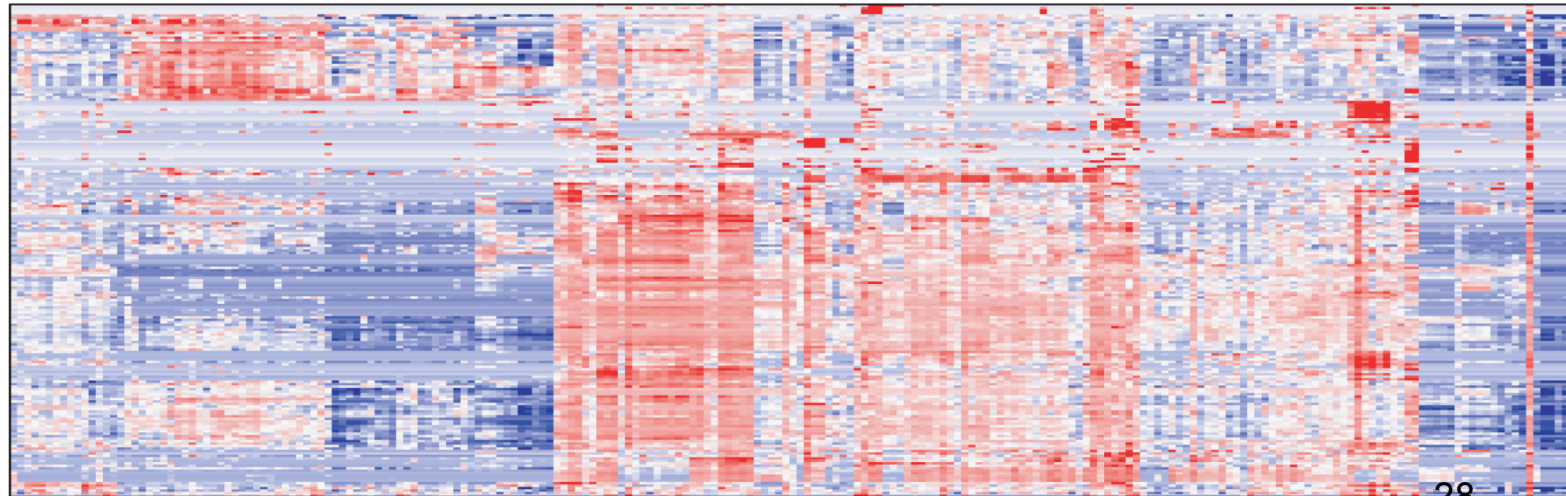
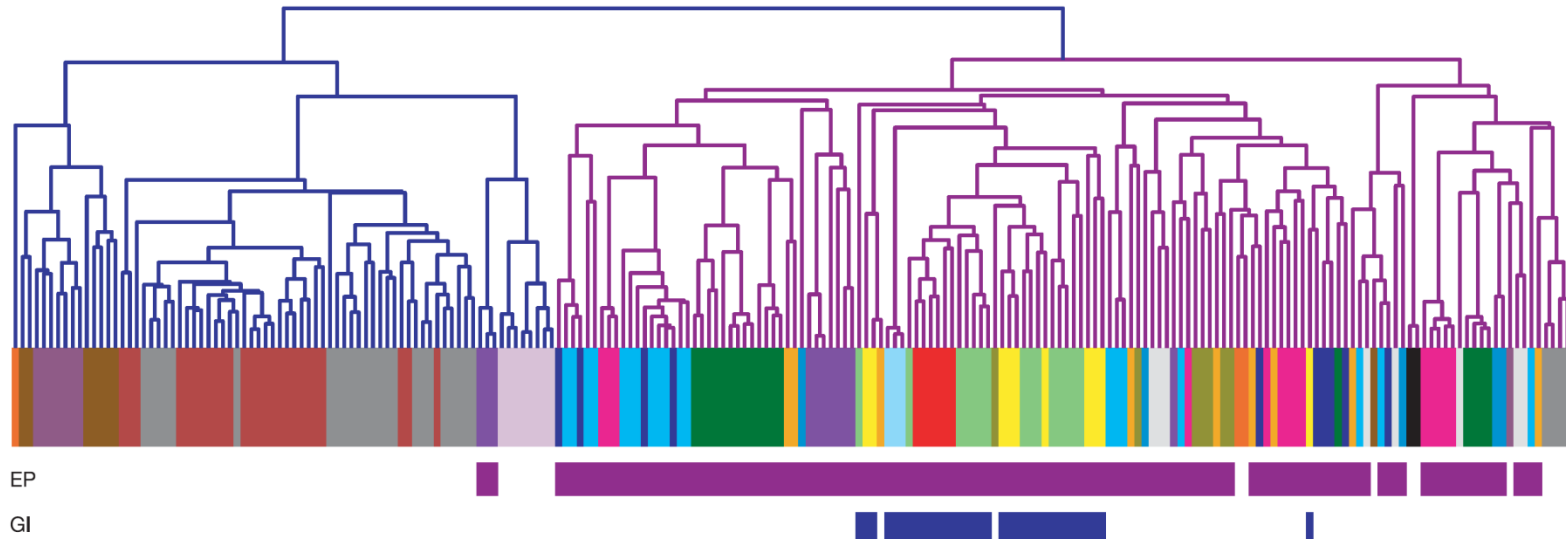
Table 2. Examples of miRNAs located in minimal deleted regions, minimal amplified regions, and breakpoint regions involved in human cancers

Chromosome	Location (defining markers)	Size, Mb	miR	Hystotype	Known OG/TS
3p21.1-21.2-D	ARP-DRR1	7	<i>let-7g/miR-135-1</i>	Lung, breast cancer	—
3p21.3(AP20)-D	GOLGA4-VILL	0.75	<i>miR-26a</i>	Epithelial cancer	—
3p23-21.31(MDR2)-D	D351768-D351767	12.32	<i>miR-26a; miR-138-1</i>	Nasopharyngeal cancer	—
5q32-D	ADRB2-ATX1	2.92	<i>miR-145/miR-143</i>	Myelodysplastic syndrome	—
9q22.3-D	D9S280-D9S1809	1.46	<i>miR-24-1/miR-27b/miR-23b; let-7a-1/let-7f-1/let-7d</i>	Urothelial cancer	PTC, FANCC
9q33-D	D9S1826-D9S158	0.4	<i>miR-123</i>	NSCLC	—
11q23-q24-D	D11S927-D11S1347	1.994	<i>miR-34a-1/miR-34a-2</i>	Breast, lung cancer	PPP2R1B
11q23-q24-D	D11S1345-D11S1328	1.725	<i>miR-125b-1/let-7a-2/miR-100</i>	Breast, lung, ovary, cervix cancer	—
13q14.3-D	D13S272-D13S25	0.54	<i>miR-15a/miR-16a</i>	B-CLL	—
13q32-33-A	stG15303-stG31624	7.15	<i>miR-17/miR-18/miR-19a/miR-20/ miR-19b-1/miR-92-1</i>	Follicular lymphoma	—
17p13.3-D	D17S1866-D17S1574	1.899	<i>miR-22; miR-132; miR-212</i>	HCC	—
17p13.3-D	ENO3-TP53	2.275	<i>miR-195</i>	Lung cancer	TP53
17q22-t(8;17)	miR-142s/c-MYC		<i>miR-142s; miR-142as</i>	Prolymphocytic leukemia	c-MYC
17q23-A	CLTC-PPM1D	0.97	<i>miR-21</i>	Neuroblastoma	—
20q13-A	FLJ33887-ZNF217	0.55	<i>miR-297-3</i>	Colon cancer	—
21q11.1-D	D21S1911-ANA	2.84	<i>miR-99a/let-7c/miR-125b</i>	Lung cancer	—

D, deleted region; A, amplified region; NSCLC, non-small-cell lung cancer; HCC, hepatocellular carcinoma; PTC, patched homolog (*Drosophila*); FANCC, Fanconi anemia, complementation group C; PPP2R1B, protein phosphatase 2, regulatory subunit A (PR 65),  $\beta$  isoform, miRNAs in a cluster are separated by a slash. For references, see Table 6.

# microRNA exprese je schopná rozlišit původ nádoru

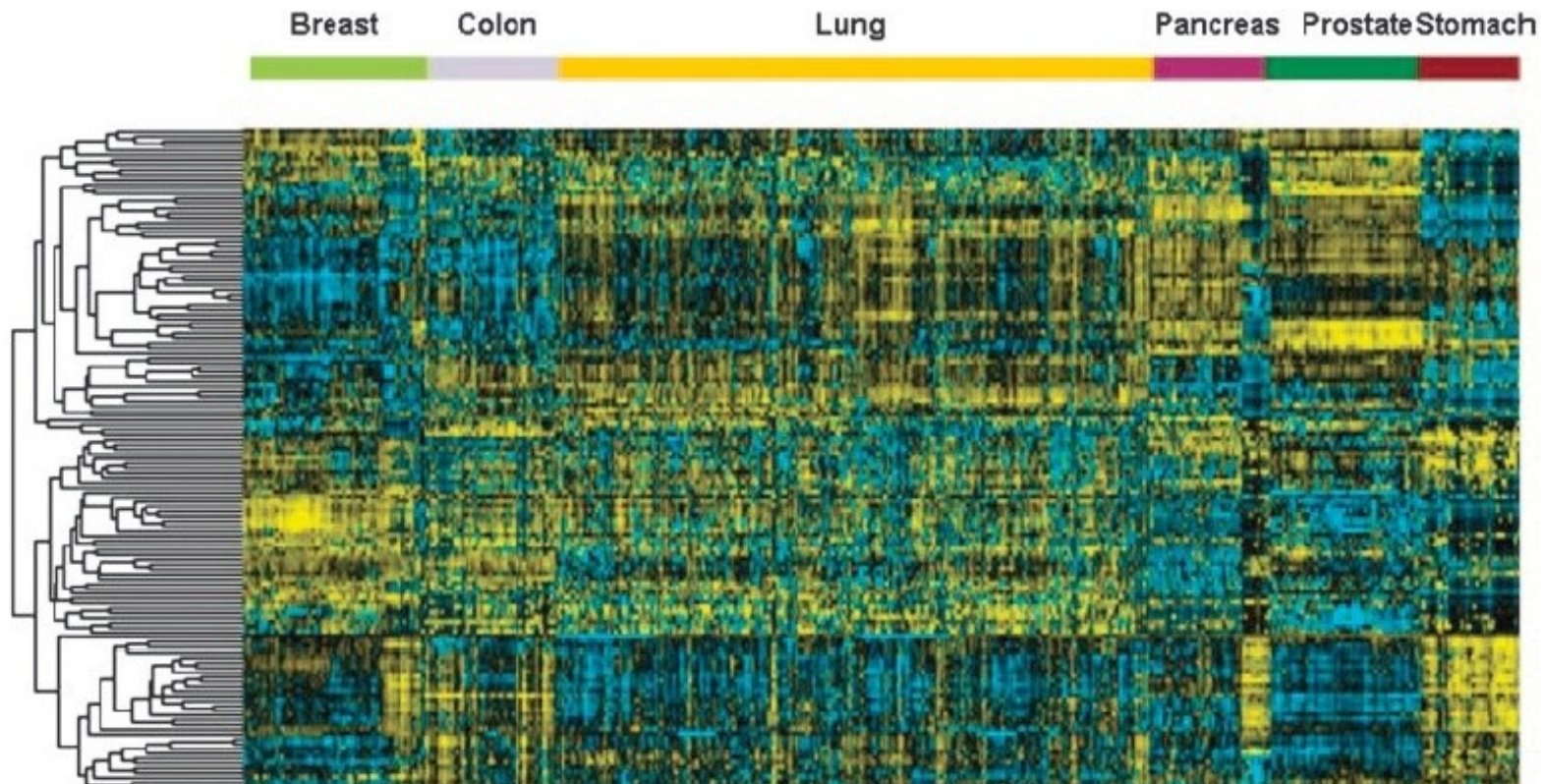
**a**



# A microRNA expression signature of human solid tumors defines cancer gene targets

Stefano Volinia<sup>\*††</sup>, George A. Calin<sup>\*‡</sup>, Chang-Gong Liu<sup>\*</sup>, Stefan Ambs<sup>§</sup>, Amelia Cimmino<sup>\*</sup>, Fabio Petrocca<sup>\*</sup>, Rosa Visone<sup>\*</sup>, Marilena Iorio<sup>\*</sup>, Claudia Roldo<sup>\*</sup>, Manuela Ferracin<sup>¶</sup>, Robyn L. Prueitt<sup>§</sup>, Nozumu Yanaihara<sup>§</sup>, Giovanni Lanza<sup>¶</sup>, Aldo Scarpa<sup>||</sup>, Andrea Vecchione<sup>\*\*</sup>, Massimo Negrini<sup>¶</sup>, Curtis C. Harris<sup>§</sup>, and Carlo M. Croce<sup>\*††</sup>

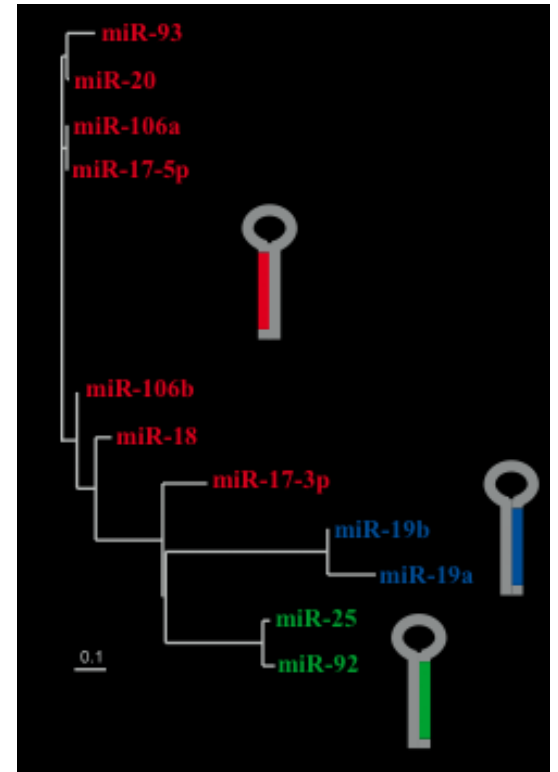
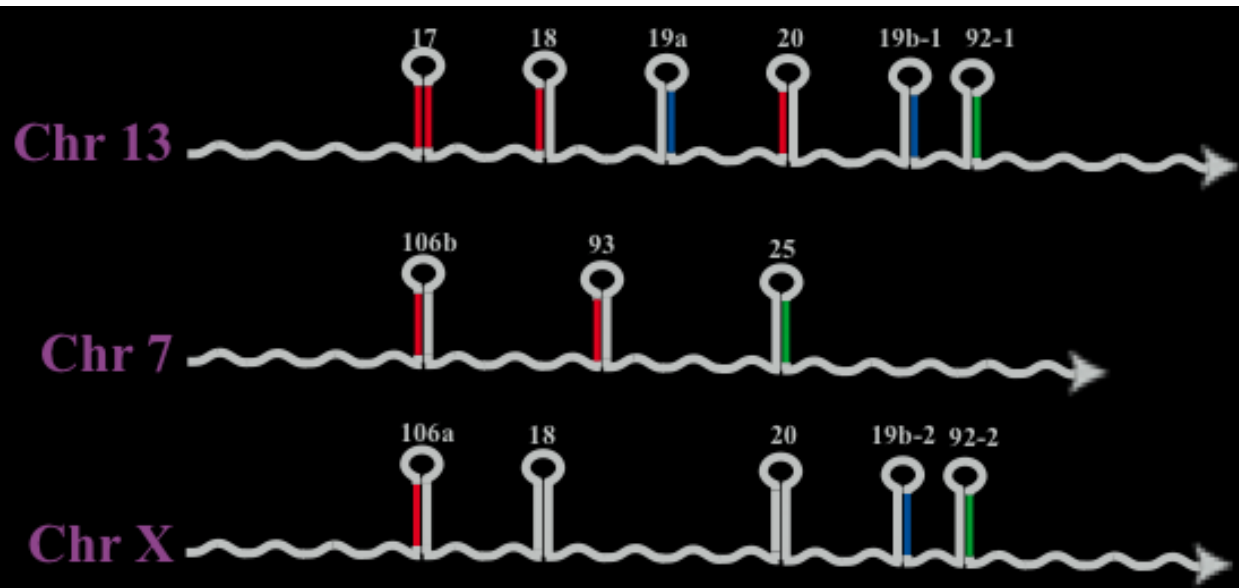
<sup>\*</sup>Department of Molecular Virology, Immunology, and Medical Genetics and Cancer Comprehensive Center, Ohio State University, Columbus, OH 43210; <sup>§</sup>Laboratory of Human Carcinogenesis, Center for Cancer Research, National Cancer Institute, National Institutes of Health, Bethesda, MD 20892; <sup>†</sup>Telethon Facility–Data Mining for Analysis of DNA Microarrays, Department of Morphology and Embryology, and <sup>¶</sup>Department of Experimental and Diagnostic Medicine and Interdepartmental Center for Cancer Research, University of Ferrara, 44100 Ferrara, Italy; <sup>||</sup>Department of Pathology, University of Verona, 37100 Verona, Italy; and <sup>\*\*</sup>Department of Histopathology, Sant'Andrea Hospital, and University of Rome "La Sapienza," 00185 Rome, Italy



**Table 2. The miRNAs shared by the signatures of the six solid cancers**

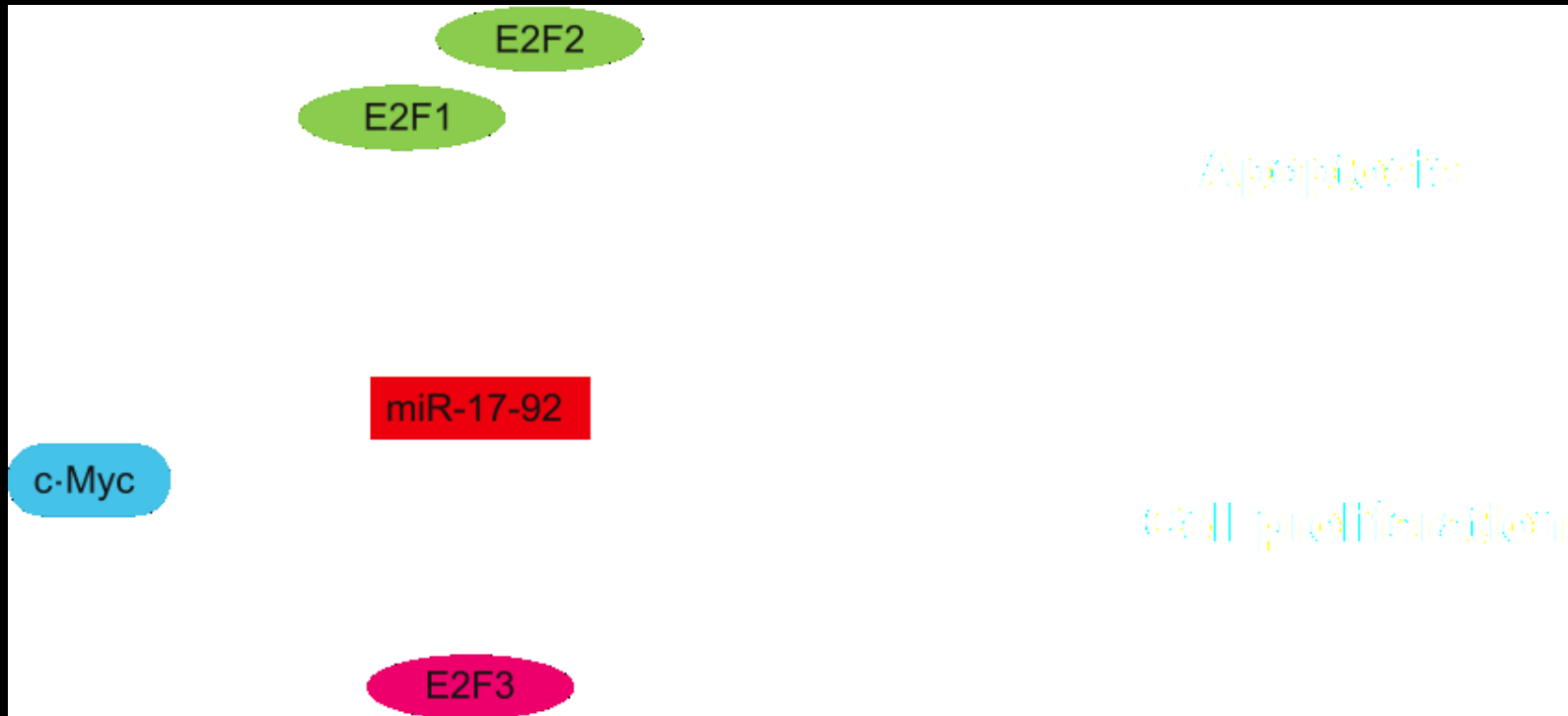
miR	<i>N</i>	Tumor type
miR-21	6	Breast, colon, lung, pancreas, prostate, stomach
miR-17-5p	5	Breast, colon, lung, pancreas, prostate
miR-191	5	Colon, lung, pancreas, prostate, stomach
miR-29b-2	4	Breast, colon, pancreas, prostate
miR-223	4	Colon, pancreas, prostate, stomach
miR-128b	3	Colon, lung, pancreas
miR-199a-1	3	Lung, pancreas, prostate
miR-24-1	3	Colon, pancreas, stomach
miR-24-2	3	Colon, pancreas, stomach
miR-146	3	Breast, pancreas, prostate
miR-155	3	Breast, colon, lung
miR-181b-1	3	Breast, pancreas, prostate
miR-20a	3	Colon, pancreas, prostate
miR-107	3	Colon, pancreas, stomach
miR-32	3	Colon, pancreas, prostate
miR-92-2	3	Pancreas, prostate, stomach
miR-214	3	Pancreas, prostate, stomach
miR-30c	3	Colon, pancreas, prostate
miR-25	3	Pancreas, prostate, stomach
miR-221	3	Colon, pancreas, stomach
miR-106a	3	Colon, pancreas, prostate

The list includes 21 commonly up-regulated microRNAs in 3 or more (*N*) types of solid cancers ( $P$  value =  $2.5 \times 10^{-3}$ ).



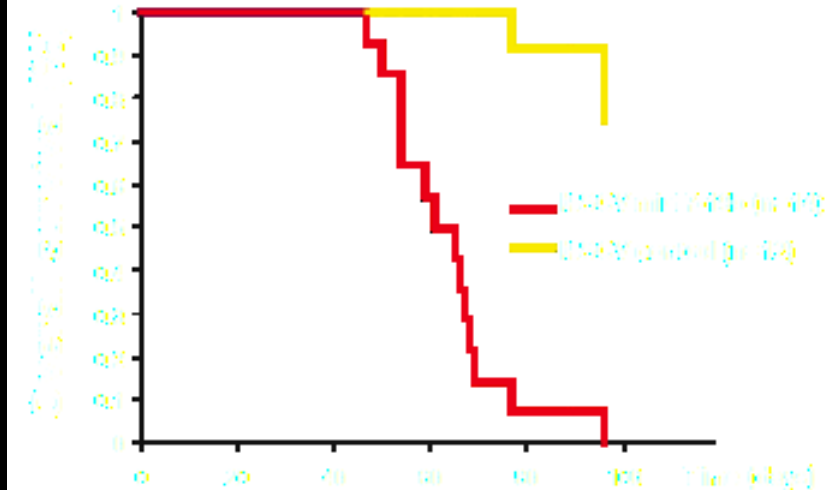
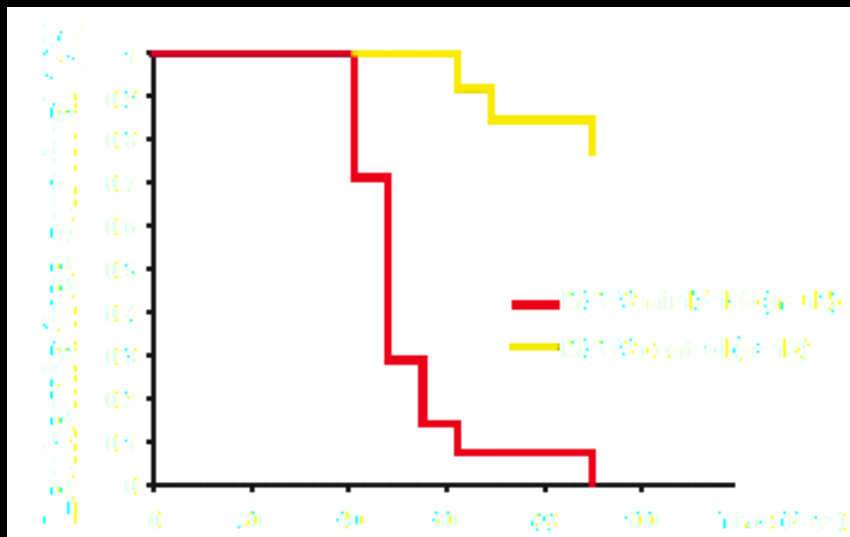
Annotations: ← Gene UTR Exon

# miR-17-92 is regulated by E2F transcription factors



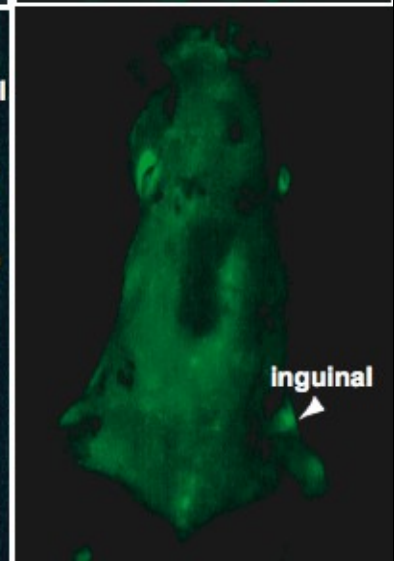
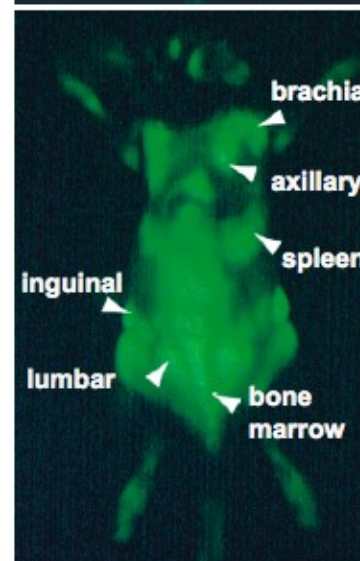
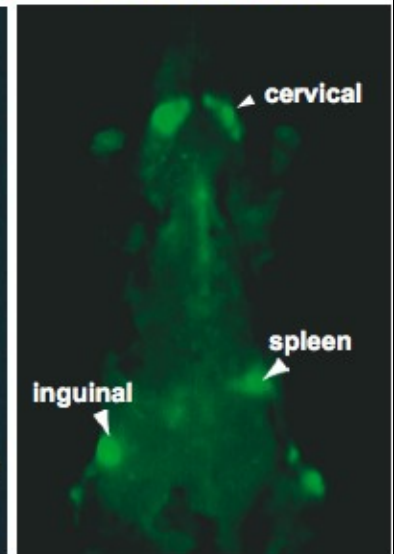
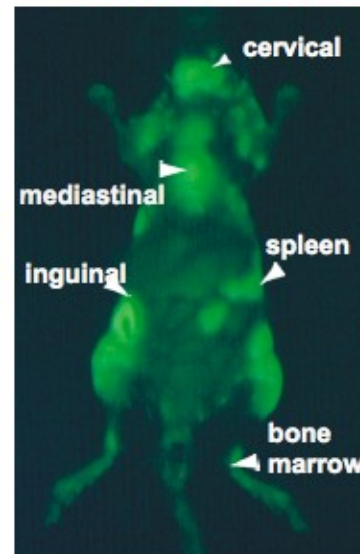


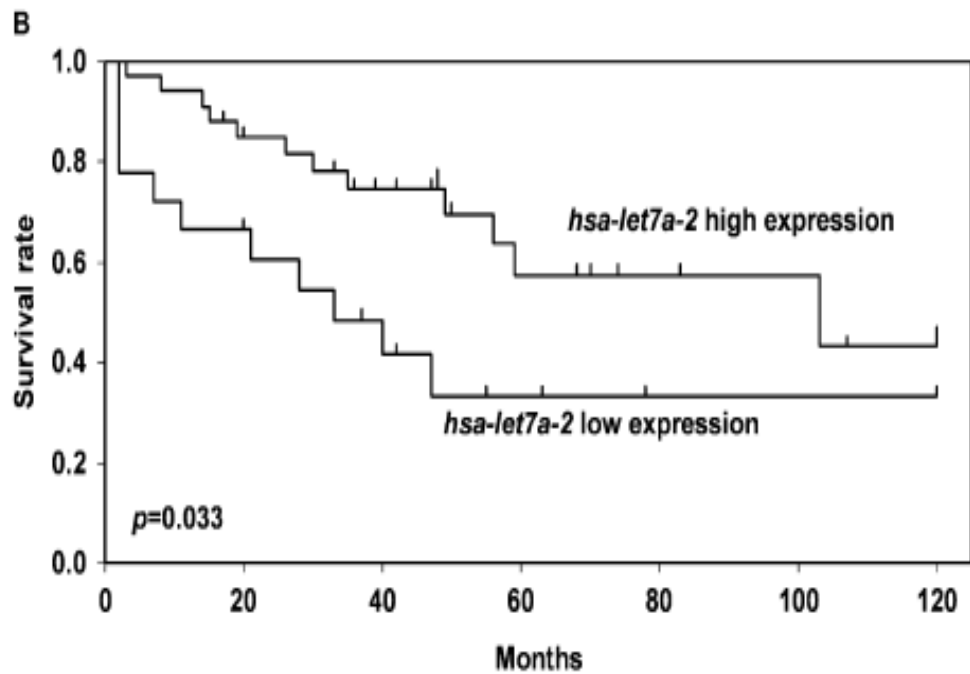
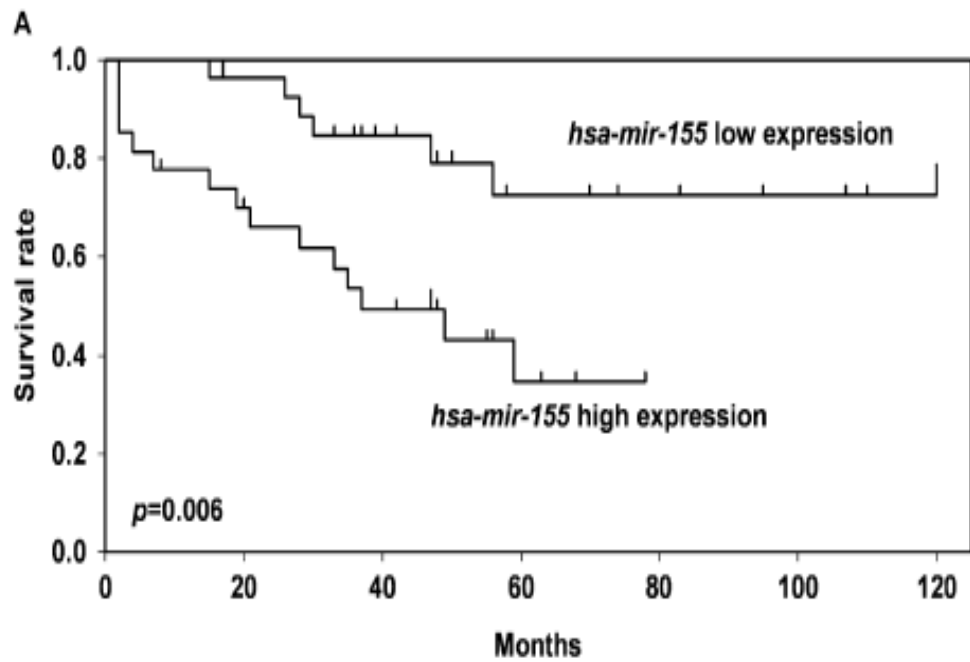
# Expression of miR17-19b Results in B-cell Lymphoma



**Eu-Myc/mir17-19b-1**

**Eu-Myc/MSCV**





## *A unique miRNA signature is associated with lung cancer prognosis*

**Table 5.** Postoperative survival of patients with lung adenocarcinoma in relation to clinicopathological characteristics and miRNA expression analyzed by microarray analysis

Variable	Subset	Hazard ratio (95% confidence interval)	p
→ Univariate analysis (n = 65)			
Age	age ≥ 67/age < 67	1.41 (0.67–3.06)	0.348
Sex	male/female	1.36 (0.64–2.93)	0.413
Stage	II–IV/I	2.51 (1.29–6.82)	0.010
Smoking history	current/former	1.32 (0.63–2.79)	0.456
→ <i>hsa-mir-155</i> (n = 55)	high/low	3.42 (1.42–8.19)	0.006
→ <i>hsa-let-7a-2</i> (n = 52)	low/high	2.35 (1.08–6.86)	0.033
→ Multivariate analysis (n = 55) <sup>a,b</sup>			
Age	age ≥ 67/age < 67	1.92 (0.71–5.17)	0.195
Sex	male/female	1.23 (0.47–3.22)	0.669
Stage	II–IV/I	3.27 (1.31–8.37)	0.013
Smoking history	current/former	1.49 (0.51–4.34)	0.457
→ <i>hsa-mir-155</i>	high/low	3.03 (1.13–8.14)	0.027

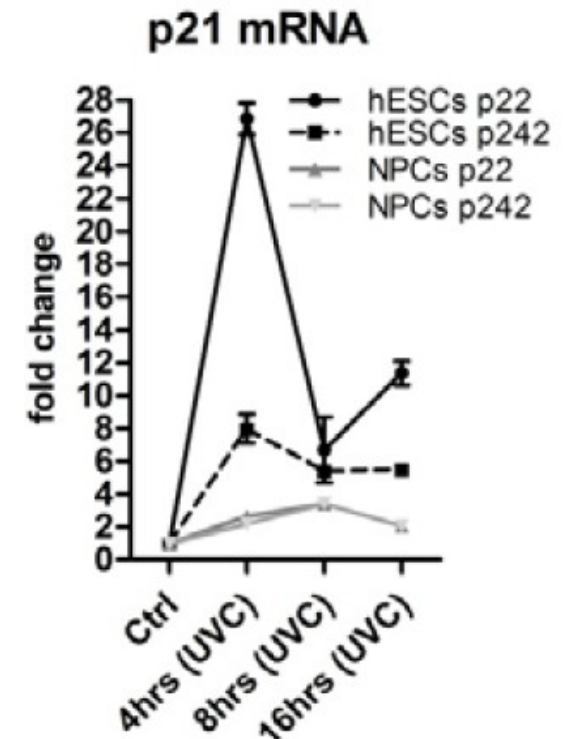
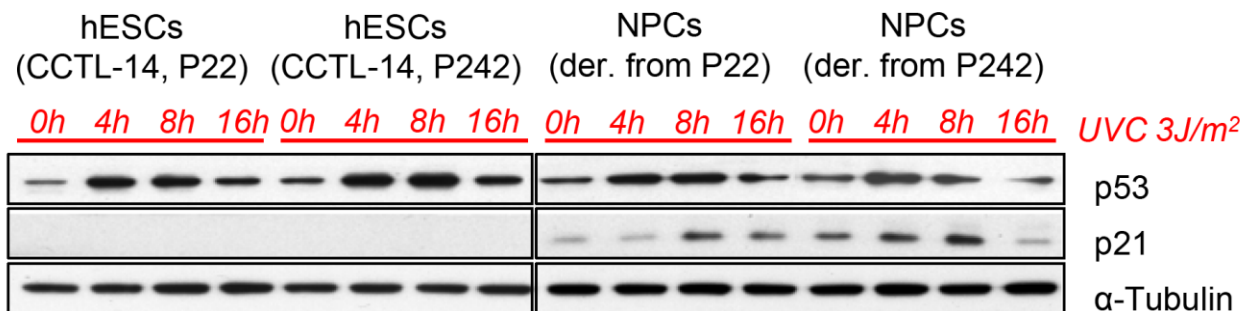
<sup>a</sup>Multivariate analysis, Cox proportional hazard regression model.  
<sup>b</sup>*hsa-let-7a-2* low/high was not statistically significant (p = 0.129).

(Yanaihara et al, Cancer Cell, 2006)

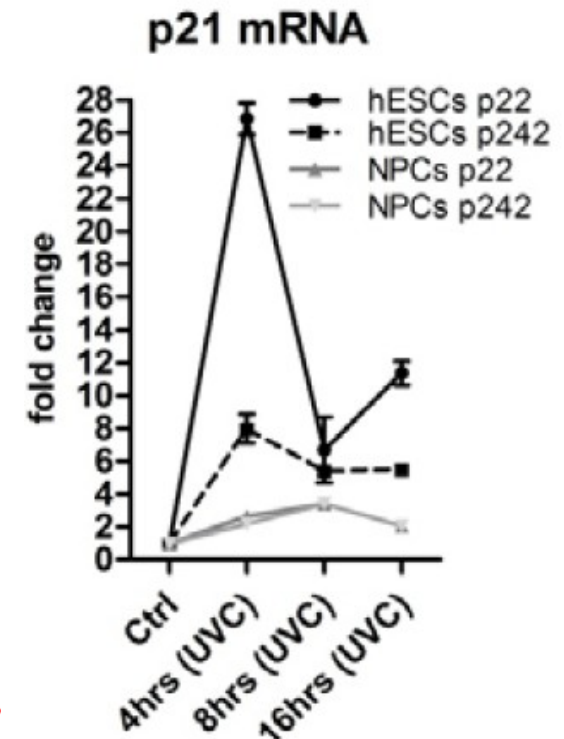
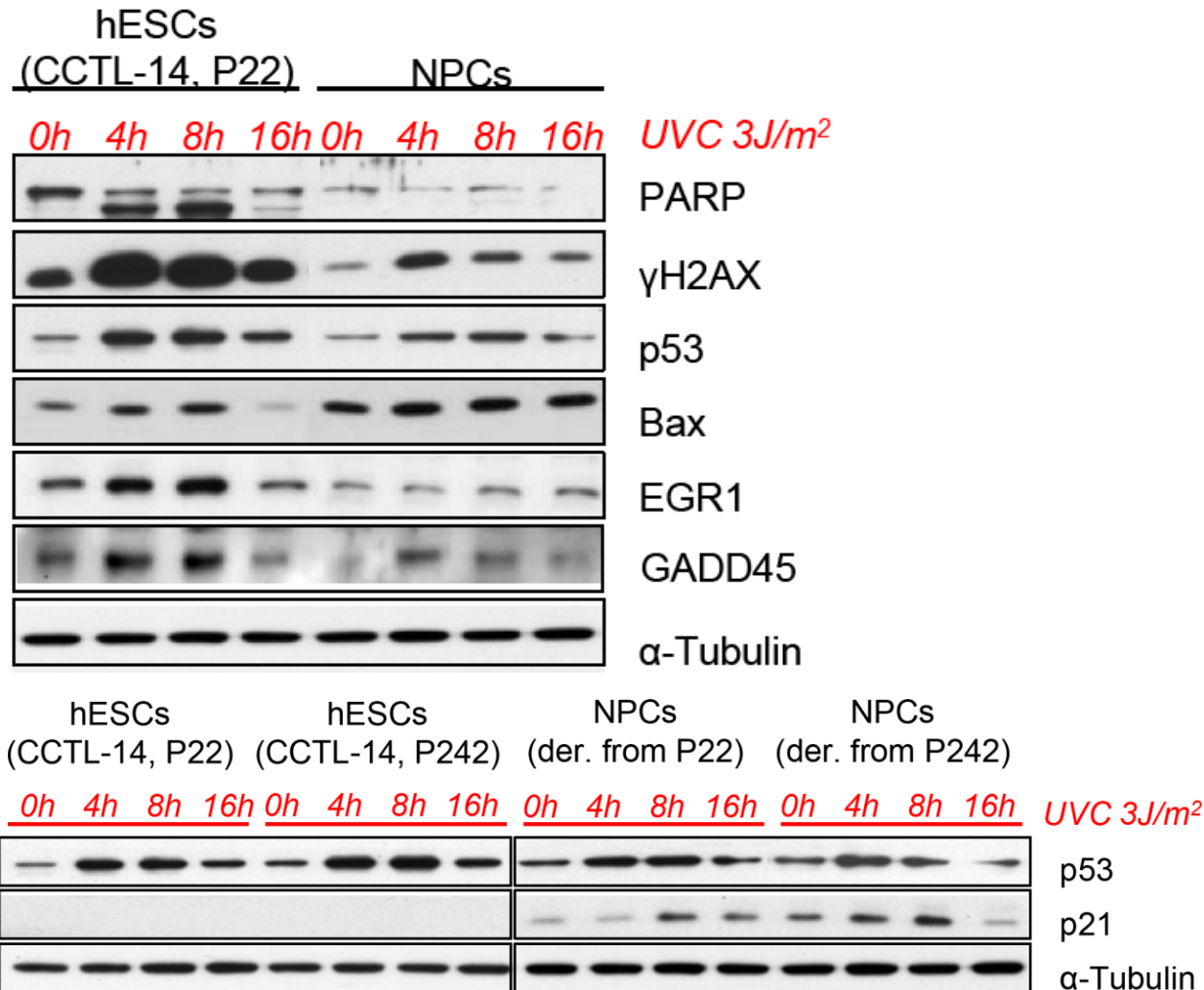
# **MICRORNAS EXPRESSION IN THE REGULATION OF P21 IN HUMAN EMBRYONIC STEM CELLS**

*Dasa Dolezalova*  
*Marek Mraz*

# MicroRNAs expression in the regulation of p21 in human embryonic stem cells

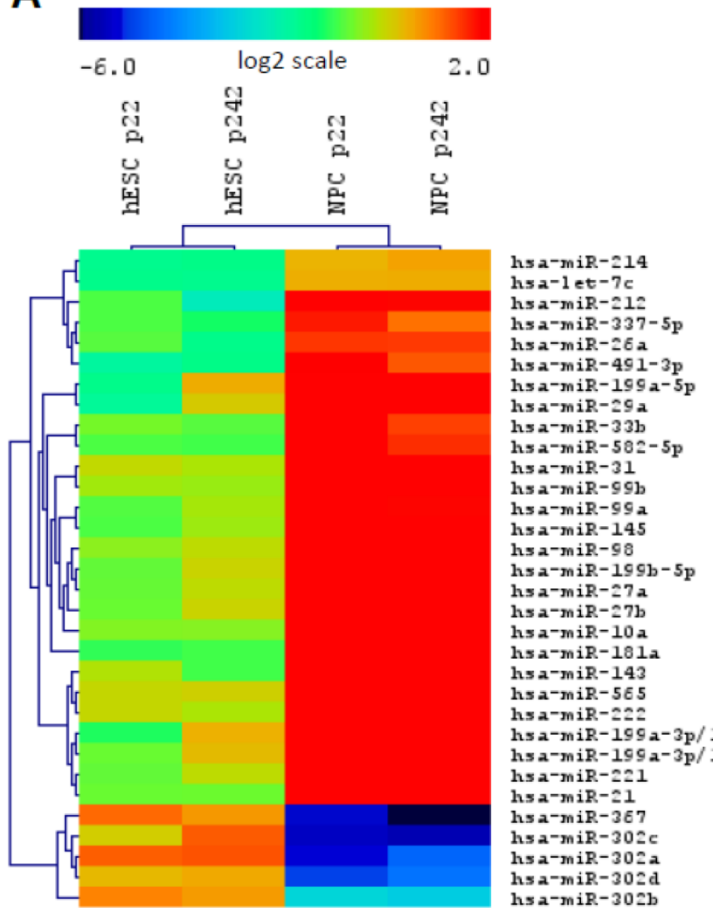


# MicroRNAs expression in the regulation of p21 in human embryonic stem cells



# Differences in microRNA expression in hESC vs NPCs

A



hsa-miR-106a  
 hsa-miR-106b  
 hsa-miR-17  
 hsa-miR-20a  
 hsa-miR-20b  
 hsa-miR-93  
 hsa-miR-302a  
 hsa-miR-302b  
 hsa-miR-302c  
 hsa-miR-302d

```

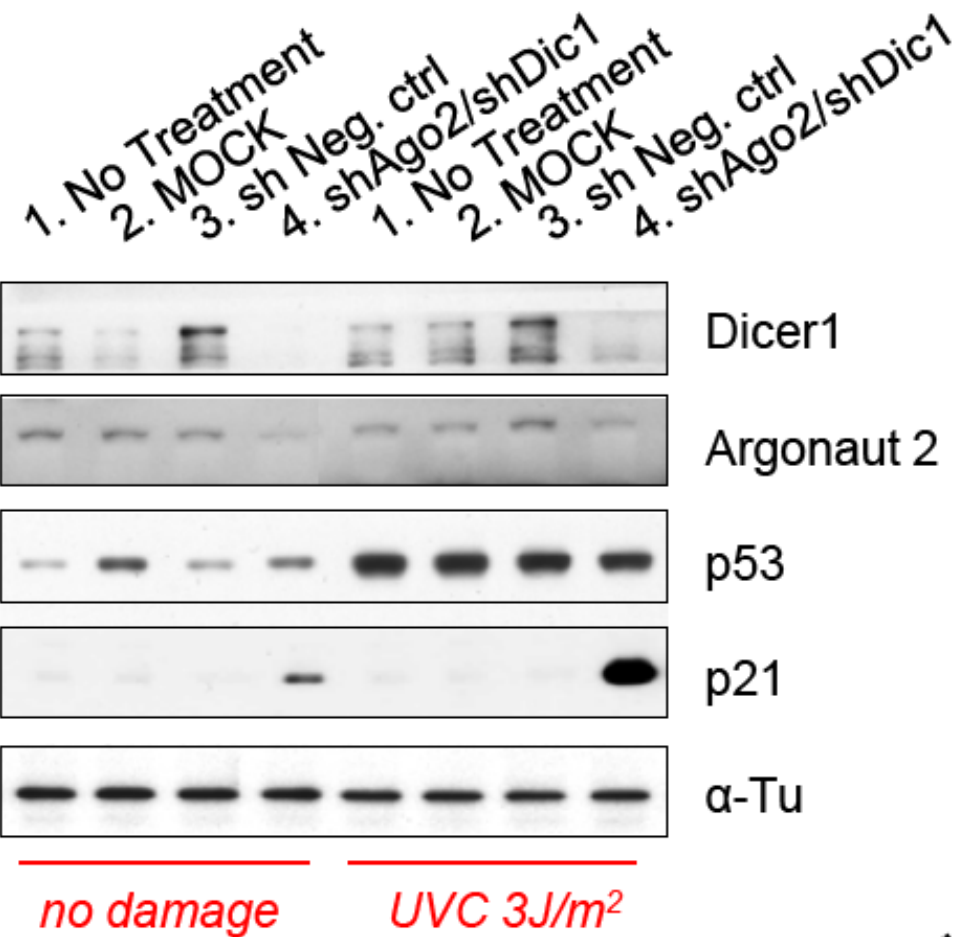
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CAAAGUGCUGUUC-GUGCAGGUAG
-UAAGUGCUUCCAUGUUUUGGUGA
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-UAAGUGCUUCCAUGUUUGAGUG-
    
```

seed sequence

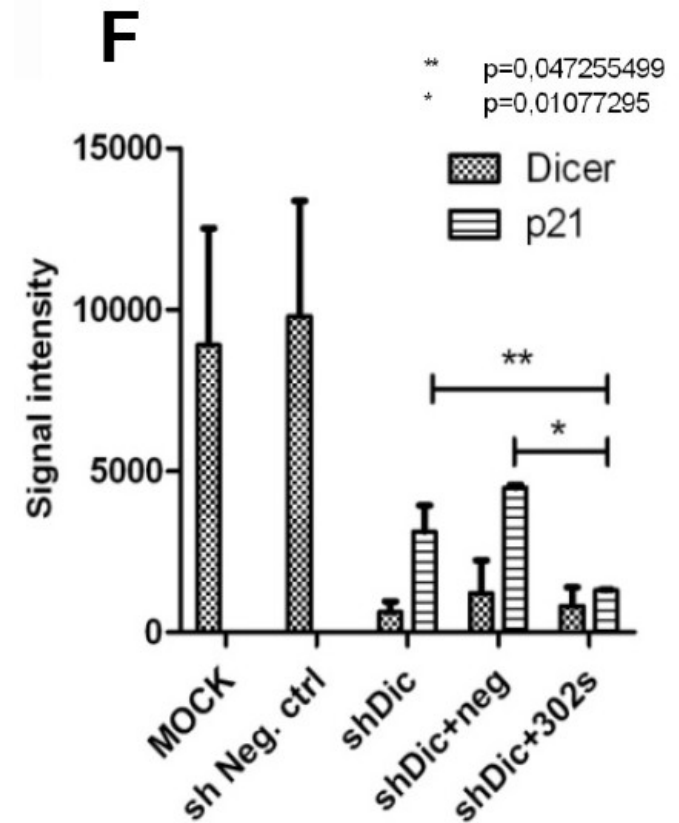
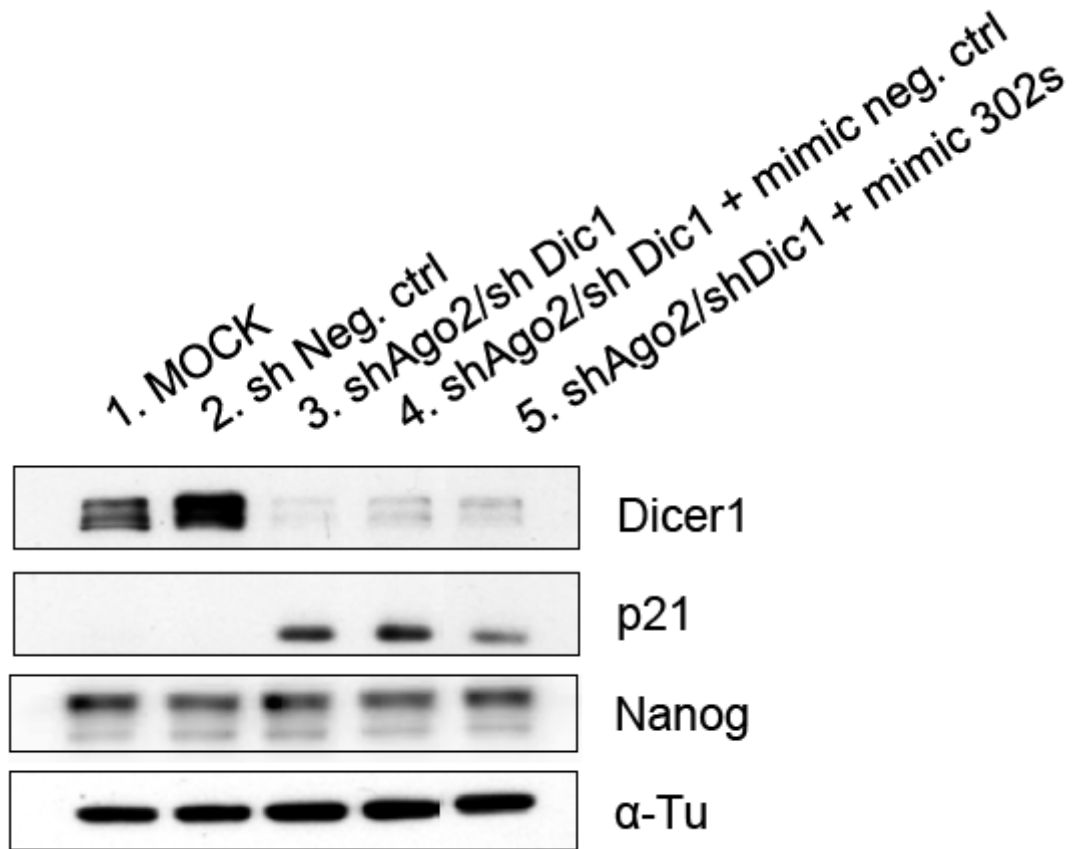




# MicroRNA pathway regulates p21

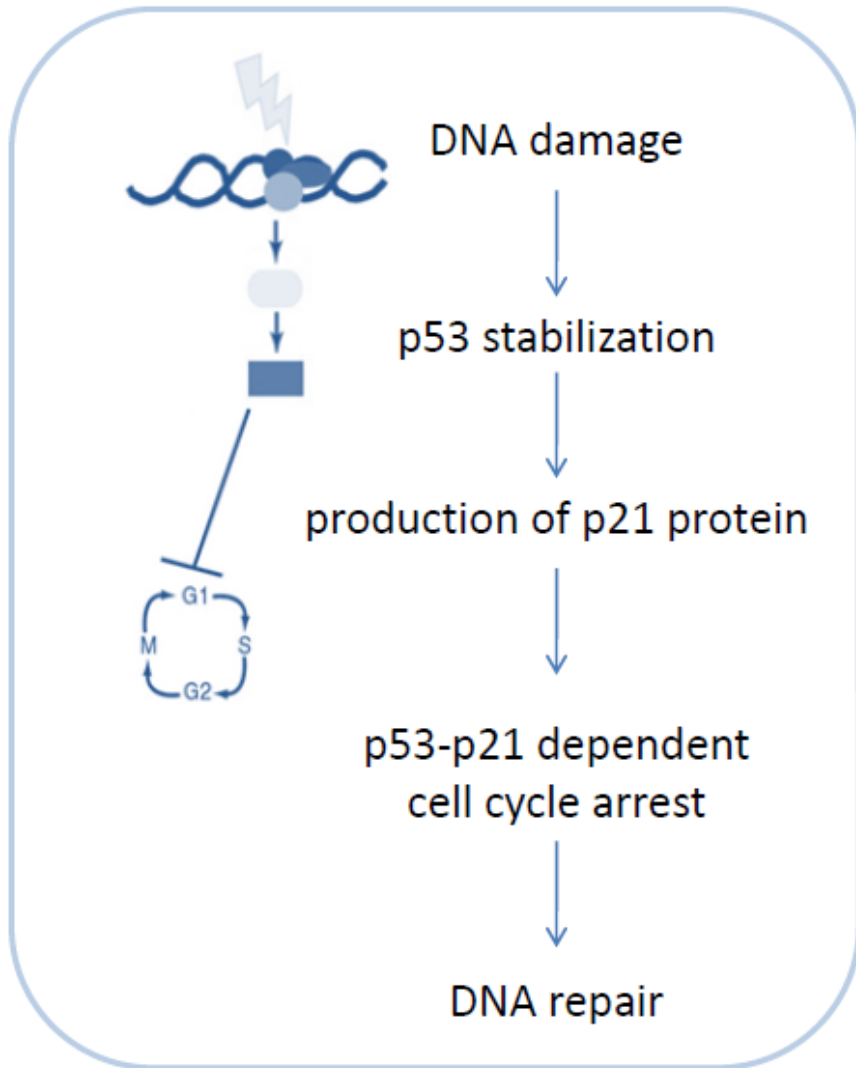


# MicroRNA pathway regulates p21: miR-302s contribute

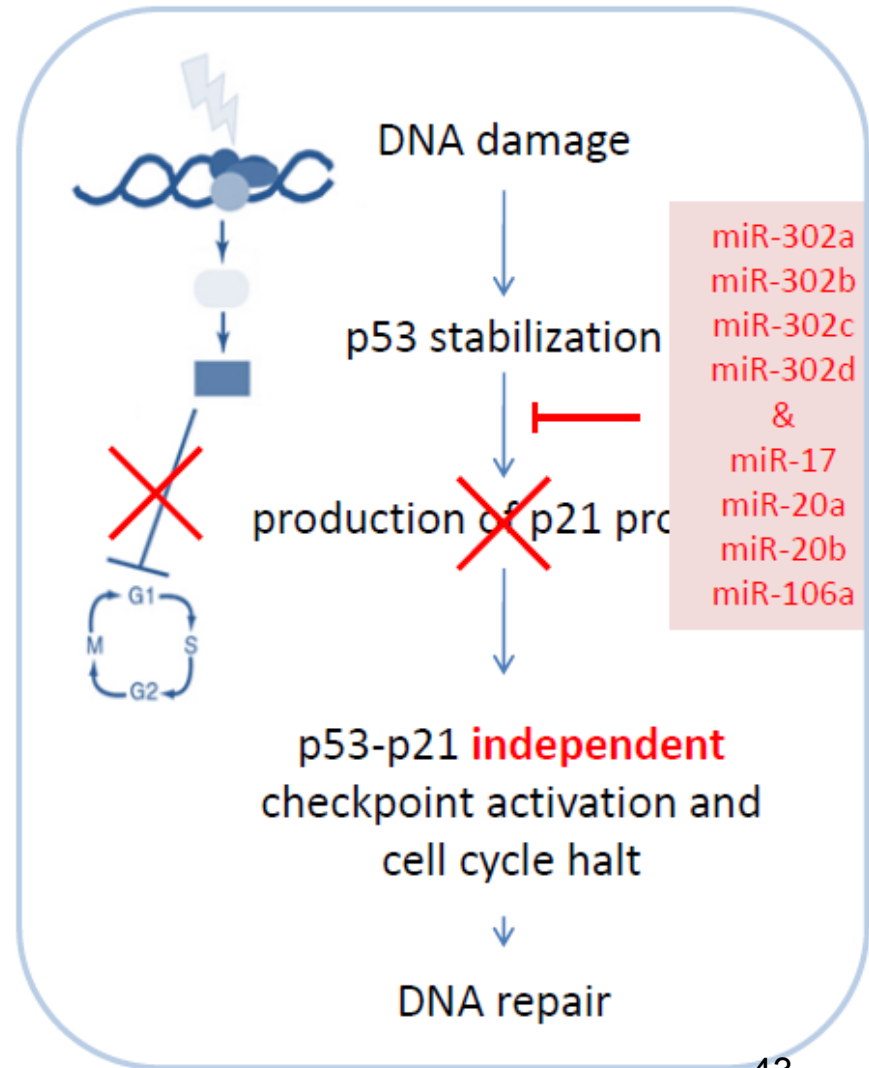


# Conclusion

## Differentiated cells (NPC)



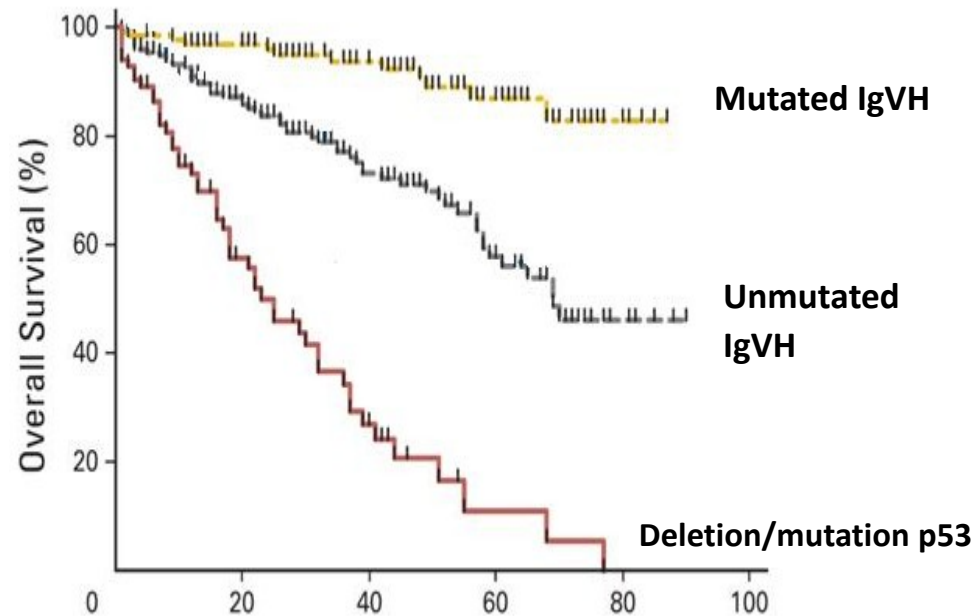
## Embryonic stem cells (hESCs)



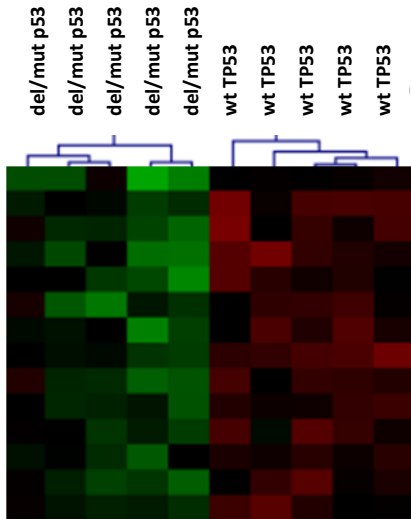
## Chronic lymphocytic leukemia (CLL)

- ❑ Prototype disease
- ❑ Most frequent leukemia in adults
- ❑ Extremely variable survival (months vs. dozens of years)
- ❑ No unifying genetic aberrations (most frequently del 13q14)

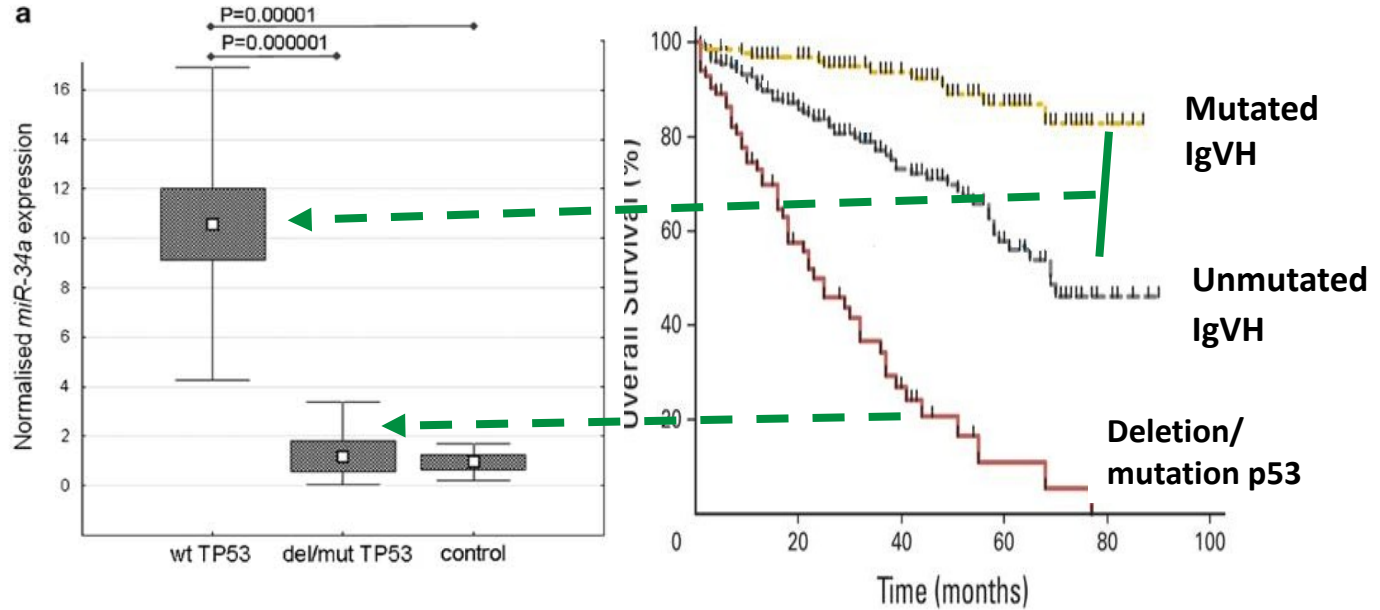
**miR-15-16 are deleted in  
60% of CLL cases  
(Calin et al., 2002)**



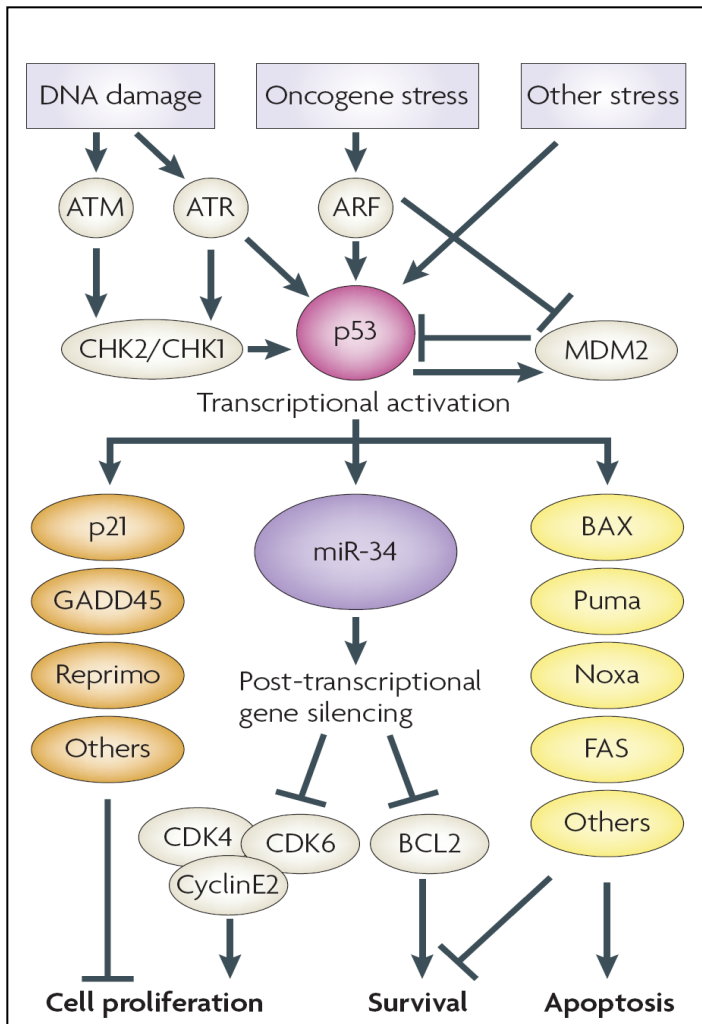
# (i) microRNAs and DNA-damage response in CLL



## *miR-34a*



# (i) microRNAs and DNA-damage response in CLL



He et al., 2007

□ **miR-34a is transcriptionally activated by p53 protein**

(He et al., 2007)

□ **miR-34a is abnormally expressed in leukemia patients**

Mraz et al., Leukemia 2009

Mraz et al., Leuk Lymphoma, 2009

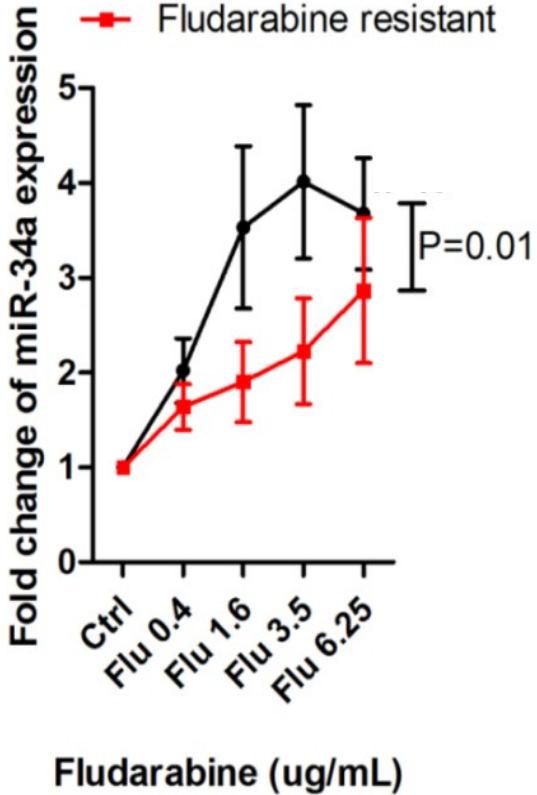
Asslaber et al., Blood, 2010

# miR-34a clinical assay

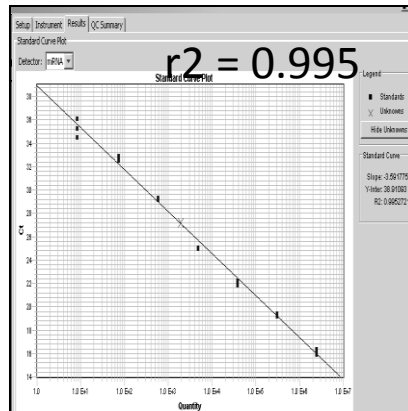
University of California, Mayo Clinic, University of Salzburg, European Research Initiative on CLL

## LAB RESEARCH

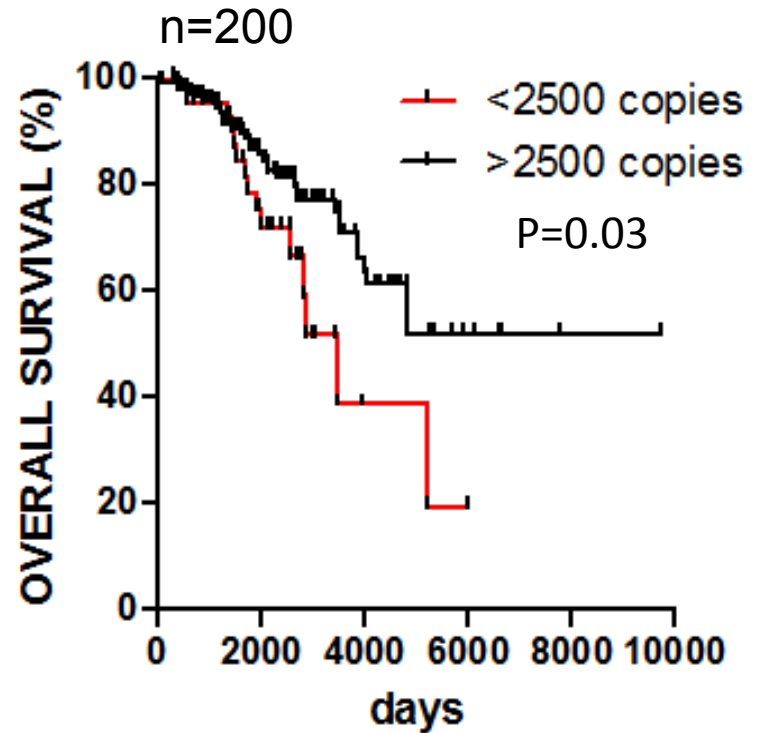
● Fludarabine sensitive  
■ Fludarabine resistant



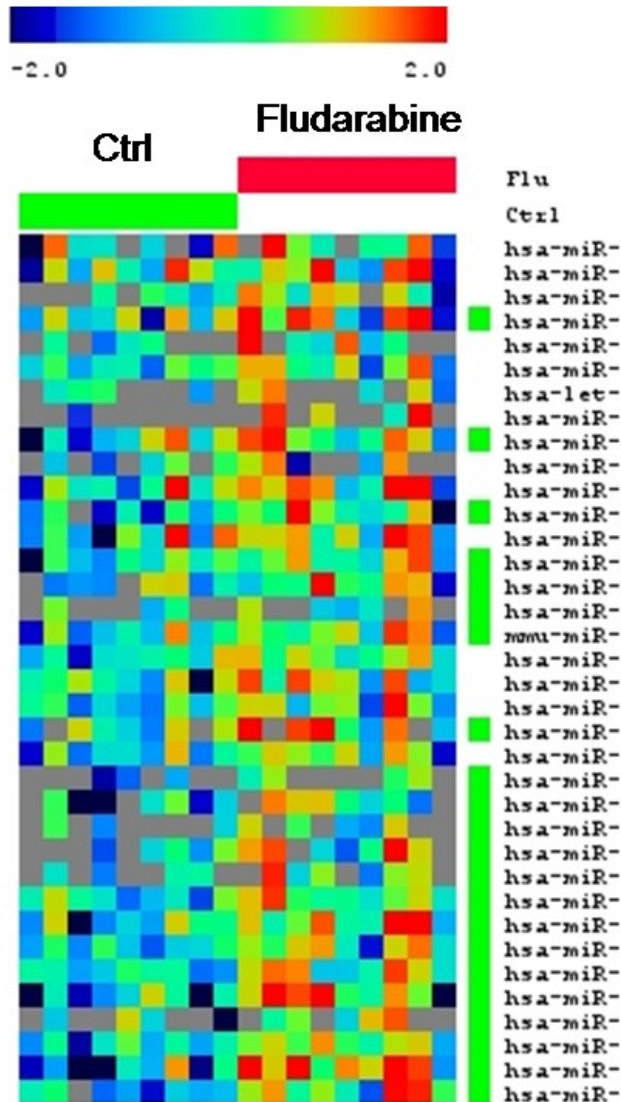
## DIAGNOSTIC TEST



## CLINICAL TRIALS

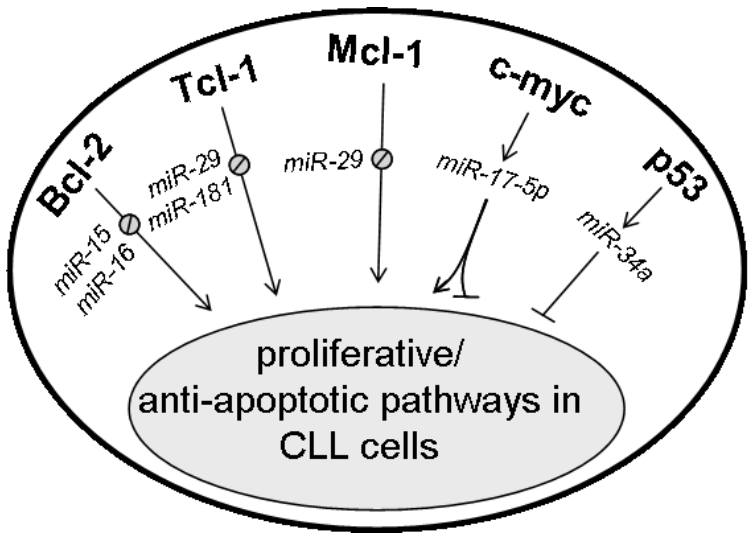


# DNA damage response





- Definování expresního profilu miRNA u pacientů s CLL a delecí/mutací p53 genu.



**Mraz et al., Leukemia 2009**  
**Mraz et al., Leukemia&Lymphoma, 2009**  
**Mraz et al., BBRC, 2009**  
**Mraz et al., 2012**  
**Mraz et al., 2013**

## Je to k něčemu dobré i v medicíně?

**Tab. 1**  
 Exprese microRNA u pacientů s nepříznivními prognostickými markery (nemutované IgVH a delece/mutace p53)  
 (dle <sup>1</sup> Calin, 2005; <sup>2</sup> Mraz, 2009b)

MicroRNA	Chromozomální oblast	Exprese miRNA u CLL vzorků s:	
		nemutovaným IgVH (ZAP70+) vs. mutovaným IgVH (ZAP70-)	delecí/mutací p53 (bez ohledu na IgVH) vs. wild-type p53 (bez ohledu na IgVH)
miR-15a	13q14	VYSOKÁ <sup>1</sup>	NEZMĚNĚNA <sup>2</sup>
miR-16-1	13q14	VYSOKÁ <sup>1</sup>	NEZMĚNĚNA <sup>2</sup>
miR-16-2	3q26	VYSOKÁ <sup>1</sup>	NEZMĚNĚNA <sup>2</sup>
miR-23b	9q22	VYSOKÁ <sup>1</sup>	NEZMĚNĚNA <sup>2</sup>
miR-24-1	9q22	VYSOKÁ <sup>1</sup>	NEZMĚNĚNA <sup>2</sup>
miR-29a-2	7q32	NÍZKÁ <sup>1</sup>	NEZMĚNĚNA <sup>2</sup>
miR-29b-2	1q32	NÍZKÁ <sup>1</sup>	NEZMĚNĚNA <sup>2</sup>
miR-29c	1q32	NÍZKÁ <sup>1</sup>	NÍZKÁ <sup>2</sup>
miR-146	5q34	VYSOKÁ <sup>1</sup>	NEZMĚNĚNA <sup>2</sup>
miR-155	21q21	VYSOKÁ <sup>1</sup>	NEZMĚNĚNA <sup>2</sup>
miR-195	17p13	VYSOKÁ <sup>1</sup>	NEZMĚNĚNA <sup>2</sup>
miR-221	Xp11.3	VYSOKÁ <sup>1</sup>	NEZMĚNĚNA <sup>2</sup>
miR-223	Xq12	NÍZKÁ <sup>1</sup>	NEZMĚNĚNA <sup>2</sup>
miR-34a	1p36	NEZMĚNĚNA <sup>2</sup>	NÍZKÁ <sup>2</sup>
miR-17-5p	13q31	NEZMĚNĚNA <sup>2</sup>	NÍZKÁ <sup>2</sup>
miR-142		NÍZKÁ <sup>2</sup>	NEZMĚNĚNA <sup>2</sup>

- potentially useful as **therapeutic targets**

nature Vol 452 | 17 April 2008 | doi:10.1038/nature06783

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LETTERS

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**LNA-mediated microRNA silencing in non-human primates**

Joacim Elmén<sup>1\*</sup>, Morten Lindow<sup>1\*</sup>, Sylvia Schütz<sup>2</sup>, Matthew Lawrence<sup>3</sup>, Andreas Petri<sup>1</sup>, Susanna Obad<sup>1</sup>, Marie Lindholm<sup>1</sup>, Maj Hedtjärn<sup>1</sup>, Henrik Frydenlund Hansen<sup>1</sup>, Urs Berger<sup>4</sup>, Steven Gullans<sup>3</sup>, Phil Kearney<sup>1</sup>, Peter Sarnow<sup>2</sup>, Ellen Marie Straarup<sup>1</sup> & Sakari Kauppinen<sup>1,5</sup>

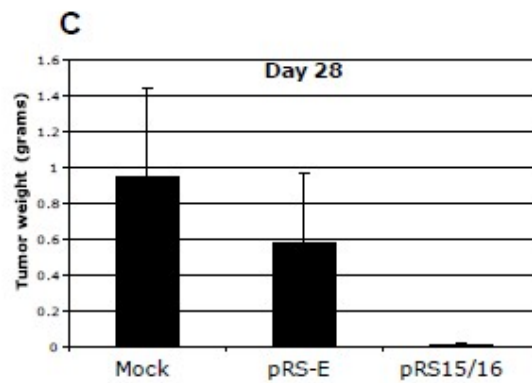
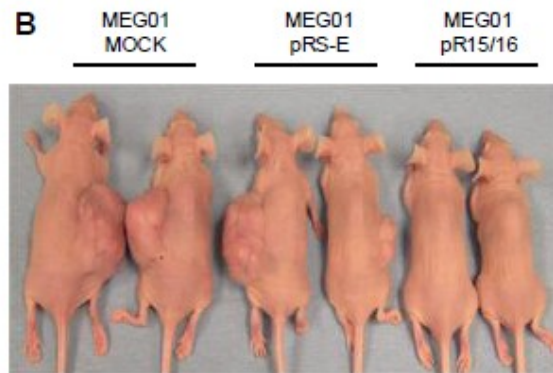
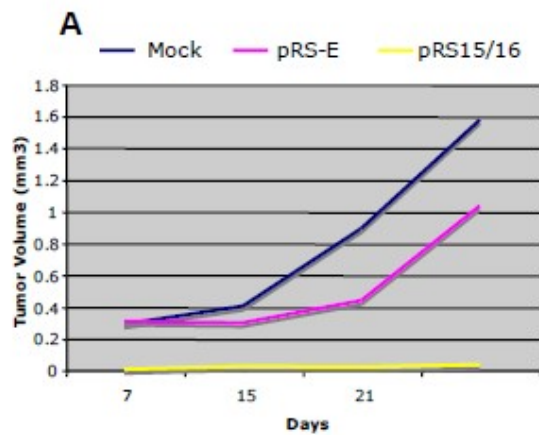
**PCR based therapeutics?!**

# blood

2012 120: 1678-1686  
Prepublished online July 13, 2012;  
doi:10.1182/blood-2012-02-410647

## **LNA-mediated anti-miR-155 silencing in low-grade B-cell lymphomas**

Yong Zhang, Aldo M. Roccaro, Christopher Rombaoa, Ludmilla Flores, Susanna Obad, Stacey M. Fernandes, Antonio Sacco, Yang Liu, Hai Ngo, Phong Quang, Abdel Kareem Azab, Feda Azab, Patricia Maiso, Michaela Reagan, Jennifer R. Brown, To-Ha Thai, Sakari Kauppinen and Irene M. Ghobrial





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Centrum molekulární biologie a genové terapie



**Děkuji za pozornost**

Marek Mráz  
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INVESTICE DO ROZVOJE VZDĚLÁVÁNÍ

Tato prezentace je spolufinancována  
Evropským sociálním fondem  
a státním rozpočtem České republiky

# MicroRNA-650 Expression Is Influenced by Immunoglobulin Gene Rearrangement and Affects the Biology of Chronic Lymphocytic Leukemia

Marek Mraz<sup>1,2</sup>, Dasa Dolezalova<sup>3</sup>, Karla Plevova<sup>1,2</sup>, Katerina Stano Kozubik<sup>1,2</sup>,  
Veronika Mayerova<sup>1,2</sup>, Katerina Cerna<sup>1,2</sup>, Katerina Musilova<sup>1,2</sup>, Boris Tichy<sup>1,2</sup>, Sarka  
Pavlova<sup>1,2</sup>, Marek Borsky<sup>2</sup>, Jan Verner<sup>1,2</sup>, Michael Doubek<sup>1,2</sup>, Yvona Brychtova<sup>2</sup>,  
Martin Trbusek<sup>1,2</sup>, Ales Hampel<sup>3</sup>, Jiri Mayer<sup>1,2</sup>, Sarka Pospisilova<sup>1,2</sup>

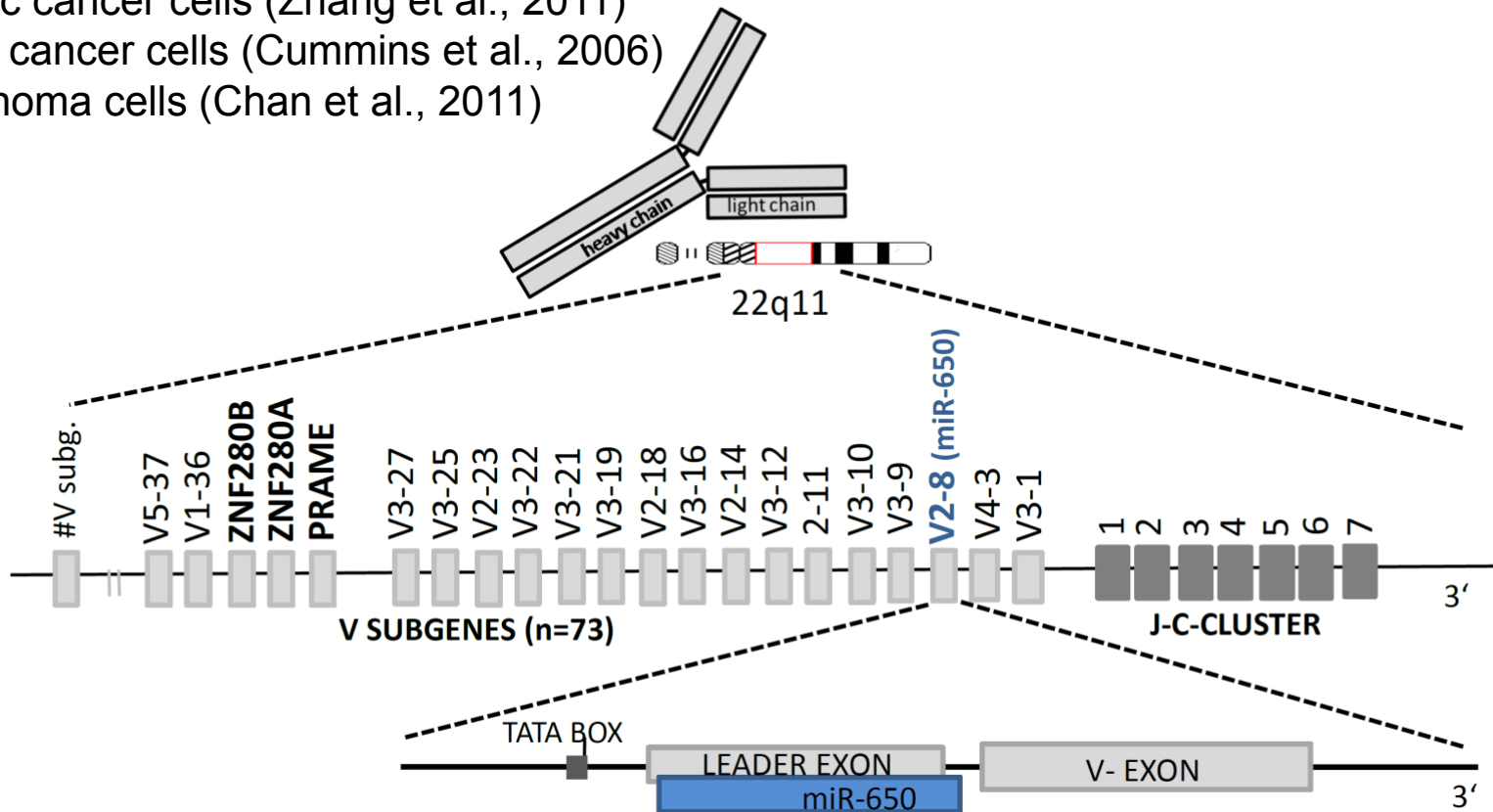
***Blood***, epub ahead of print

# IgL locus structure

miR-650 identified by miRAGE cloning approach in colorectal cells (Cummins et al., 2006)

miR-650 is expressed in:

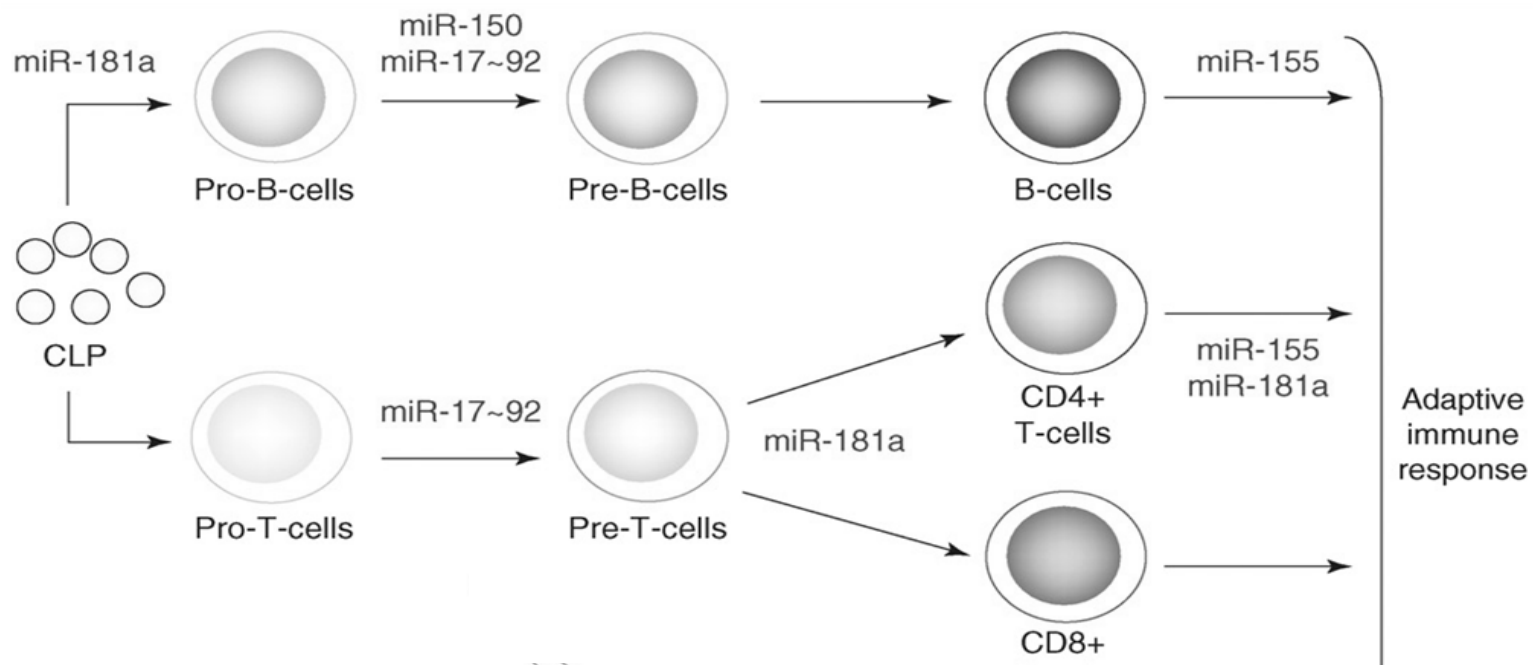
- breast cancer cells (Chien et al., 2011)
- gastric cancer cells (Zhang et al., 2011)
- colon cancer cells (Cummins et al., 2006)
- melanoma cells (Chan et al., 2011)



# MicroRNAs regulate production of immunoglobulins

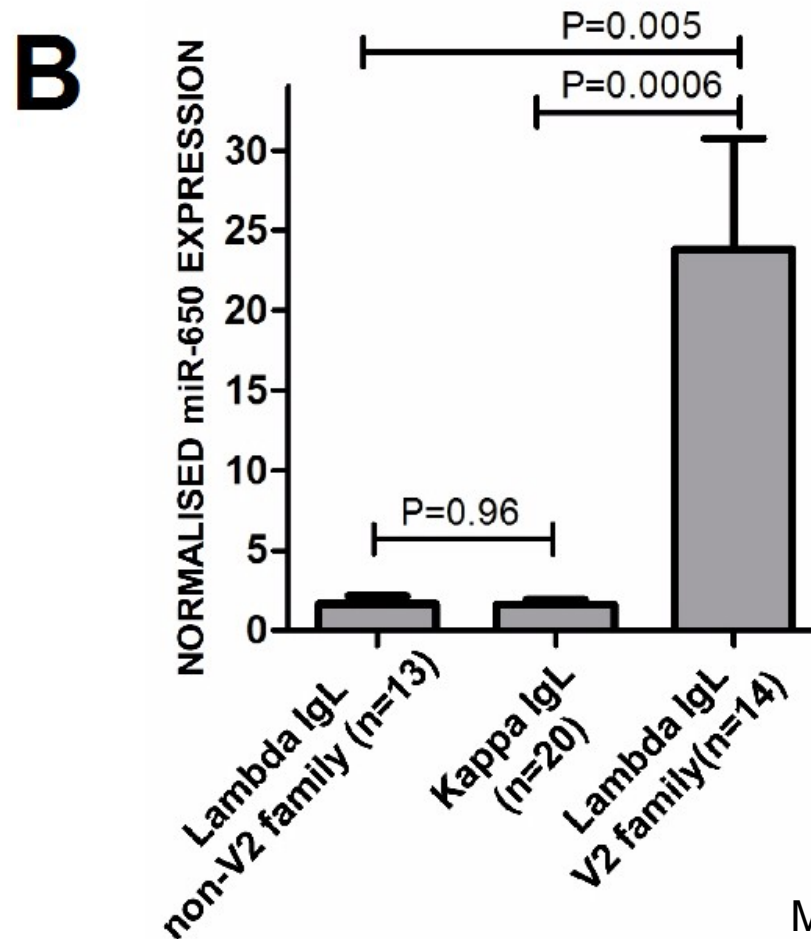


- Associated with IgG production and V(D)J recombination (*Koralov, 2008; Vigorito, 2007*)



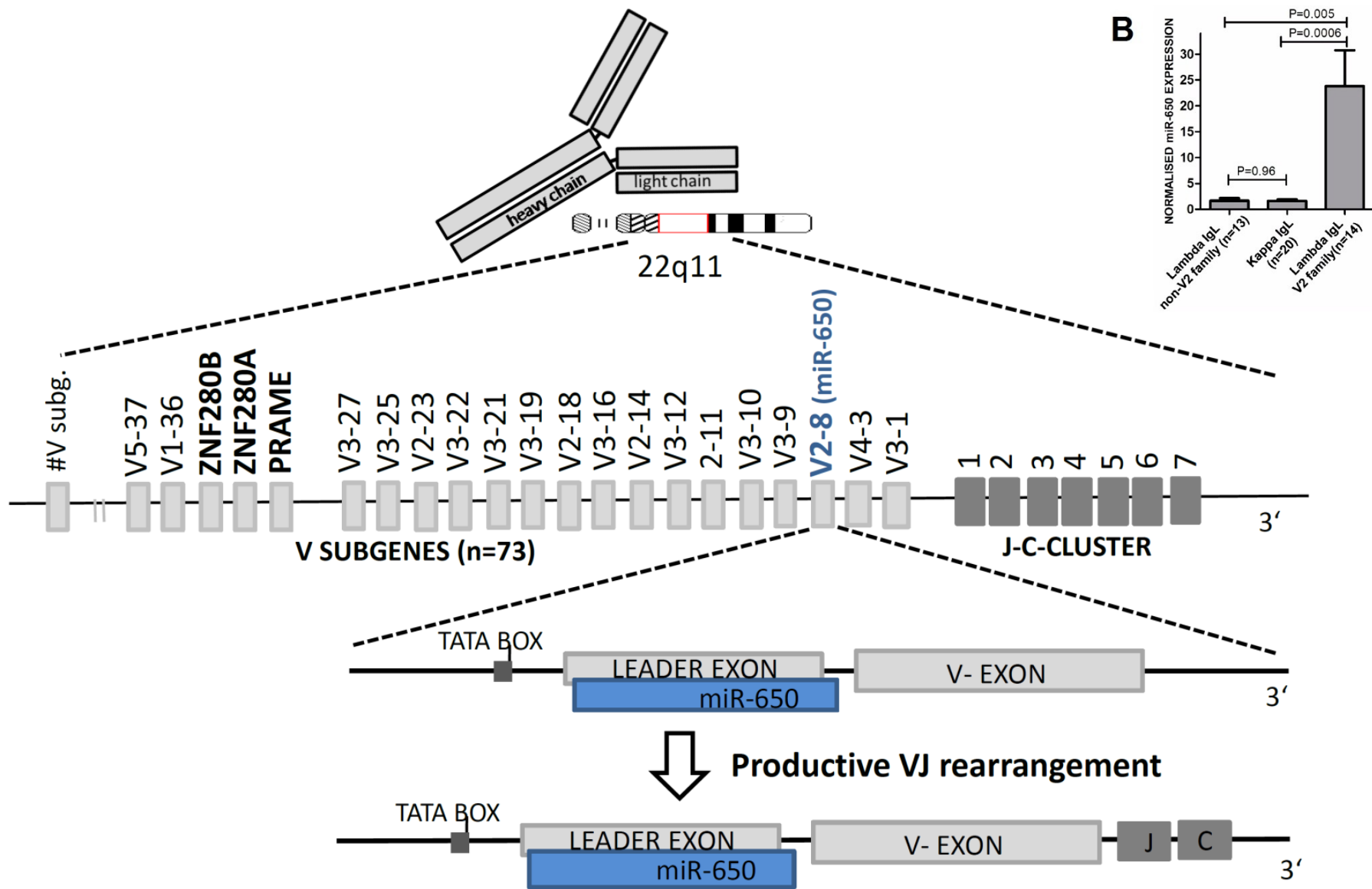
Lindsay, 2008

High expression of miR-650 in cells utilizing:  
V2-8, V2-14, V2-5, V2-18, V2-23 subgenes for IgL



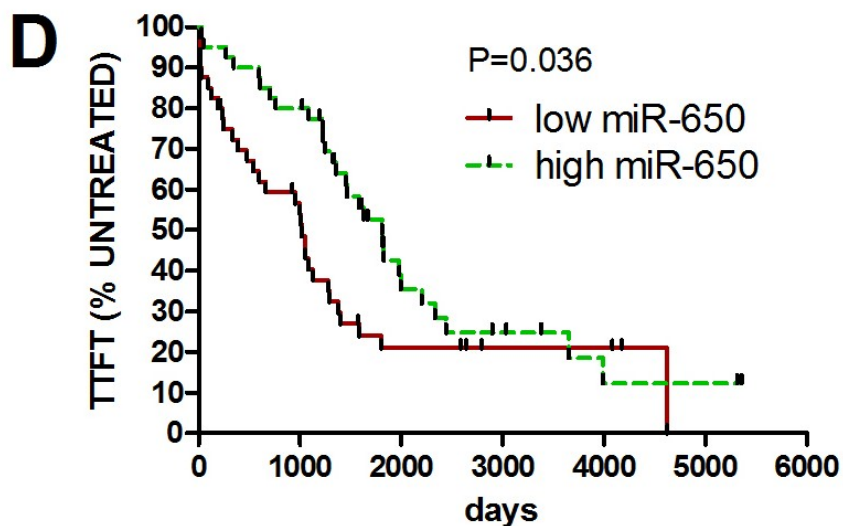
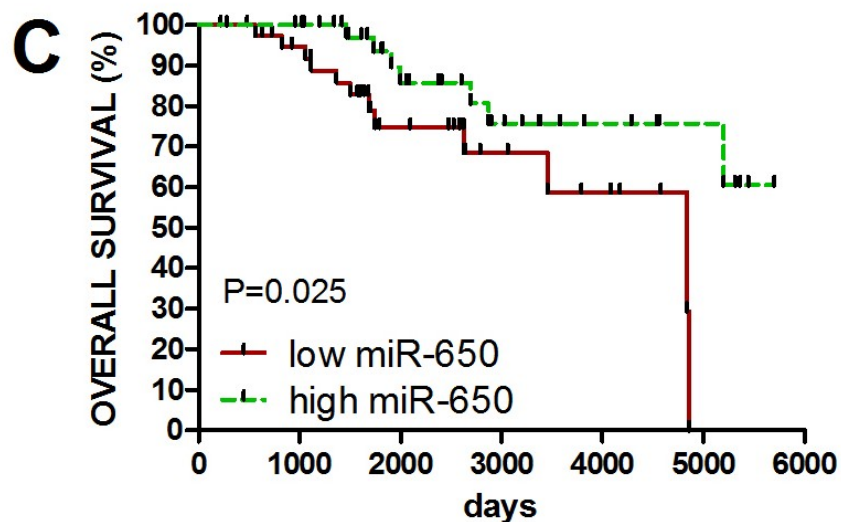


# Lambda locus rearrangement utilizing V2 family is coupled with activation of miR-650 expression



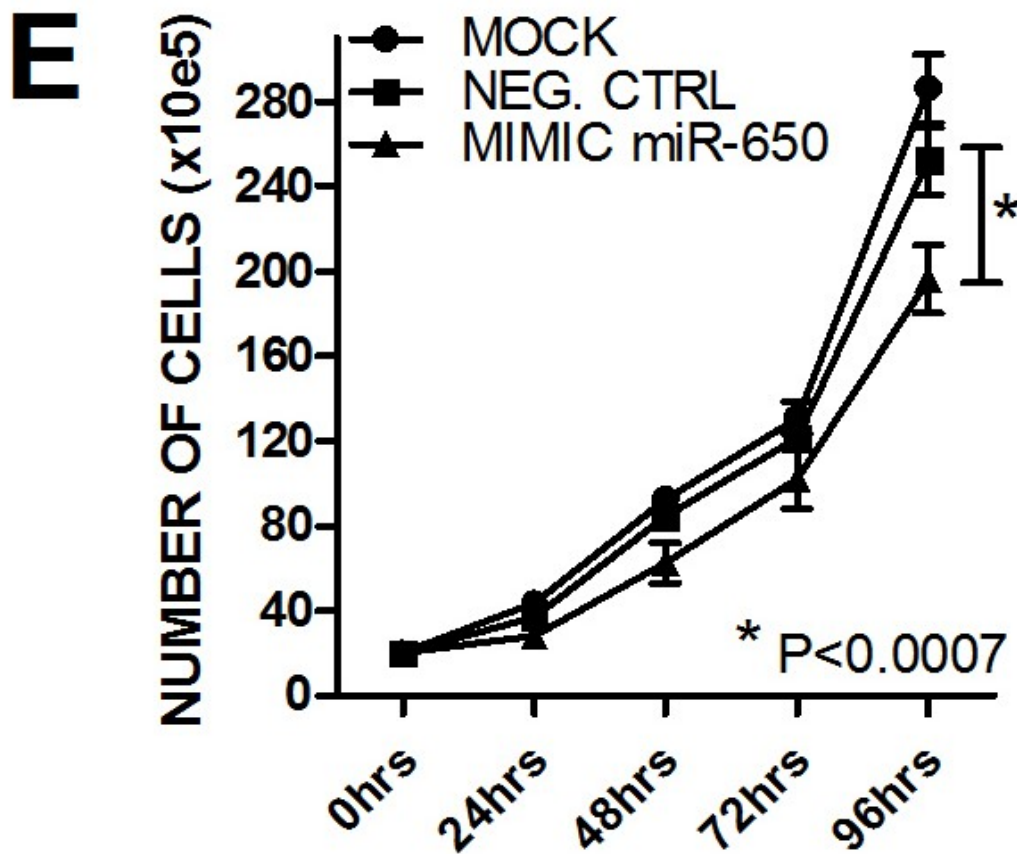
# Expression miR-650 is associated with CLL prognosis

higher expression is favorable



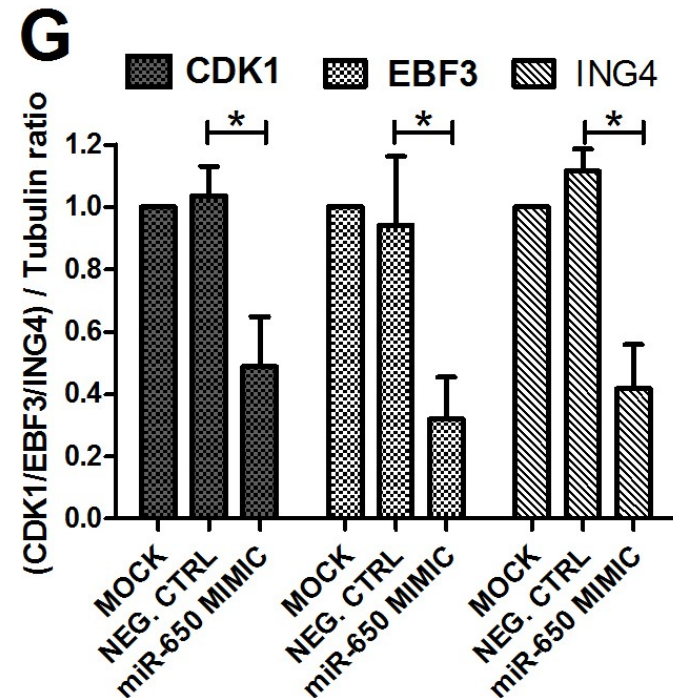
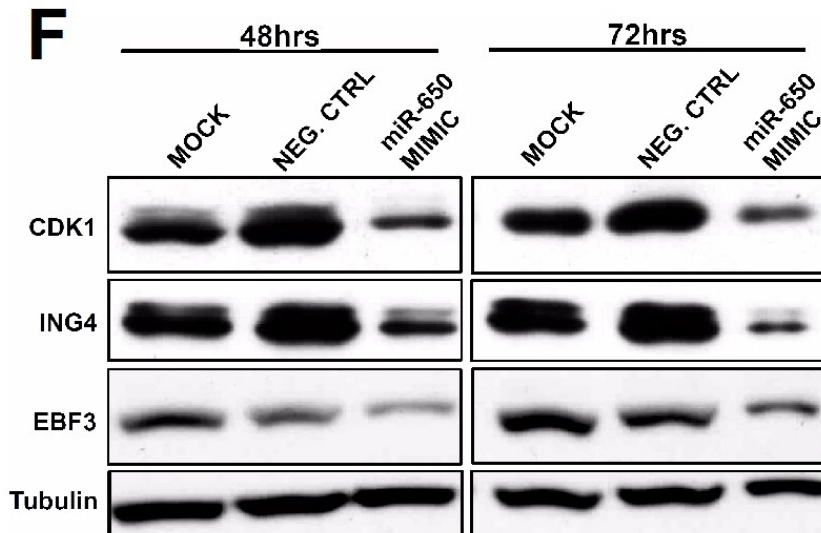
# miR-650 reduces the proliferative capacity of B-cell proliferation

miR-650 inhibits cell cycle progression by regulating p16INK4-mediated pathway (Chien et al., 2011)



# miR-650 regulates CDK1, ING4 and EBF3 in B-cells

- ~50% of predicted targets of miR-650 are expressed in B-cells with numerous being important in B-cells physiology (Gene Ontology and Functional analysis by DAVID tool).
- miR-650 regulates the Cyclin Dependent Kinase 1 (CDK1) in breast cancer cells (Chien et al., 2011)
- Inhibitor of Growth 4 (ING4) in gastric cancer cells (Zhang et al., 2011)
- The EBF3 protein is a putative target with the highest score (TargetScan)



# Conclusion (I)

## miR-650

- ❑ V2 family of subgenes for lambda IgL includes homologues of miR-650
- ❑ coupled activation of miR-650 expression with IgL expression
- ❑ miR-650 is associated with CLL prognosis
- ❑ miR-650 regulates proliferation of B-cells
- ❑ targets for miR-650:

CDK1 (Cyclin Dependent Kinase 1)

ING4 (Inhibitor of Growth 4)

EBF3 (early B cell factor)

**DISCRIPTION OF A UNIQUE MECHANISM OF MICRORNA EXPRESSION  
COUPLED WITH IMMUNOGLUBULIN EXPRESSION**