

**Institute for Microbiology, Medical Faculty of Masaryk University
and St. Anna Faculty Hospital in Brno**

**Miroslav Votava
Lenka Černohorská**

Agents of opportune infections

**The last (15th) lecture for 3rd-year students of dentistry
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Definition of nosocomial infections (NIs) – revision

Nosocomial (hospital-acquired) infections =
= infections occurring in connection with the stay
in a medical institution (as opposed to
community-acquired infections)

At least 5 % patients are afflicted, probably more

Exogenous NIs:

source = other patients, environment, personnel

vector = mostly personnel's unwashed hands

Endogenous NIs:

source = the patient himself/herself

Consequences of NIs – revision

- **Higher mortality (†)** – almost 40 % higher (a conservative estimate in this country is hundreds of unnecessary deaths per year)
- **Longer (by weeks) and more expensive hospitalization** (by tens of thousands, even more CZK per case)
- **Economic losses** circa 1.5 billions CZK/year
- **Additional ATB therapy** (both higher costs and toxicity)
- **Patients themselves = source of infection** for others

More than 1/3 of NIs can be prevented!

Main types of NIs – revision

- 1. Urinary tract infections in catheterized patients – up to 40 % of all NIs**
- 2. Respiratory tract infections – about 20 %**
 - Early ventilator-associated pneumonia
 - Late ventilator-associated pneumonia
 - Aspiration pneumonia
 - Other respiratory infections
- 3. Purulent infections of surgical wounds – about 20 %**
- 4. Blood-stream infections (sepsis by inserted intravenous catheters) – at least 15 %**

Etiology of nosocomial UTIs – revision

<i>Escherichia coli</i>	25 %
other enteric bacteria	20 %
enterococci	15 %
<i>Pseudomonas aeruginosa</i>	10 %
other G– nonfermenting rods	10 %
yeasts	5 %

Etiology of respiratory NIs I

– revision

Early VAP:

<i>Staphylococcus aureus</i>	25 %
<i>Streptococcus pneumoniae</i>	20 %
<i>Haemophilus influenzae</i>	15 %
enteric bacteria	10 %
other aerobically growing bacteria	5 %
anaerobes	1 %
<i>(monomicrobial etiology, agents originate in community)</i>	

Etiology of respiratory NIs II

– revision

Late VAP:

Gram–negative nonfermenting rods	40 %
<i>(P. aeruginosa, Acinetobacter baumannii)</i>	
enteric bacteria	30 %
<i>(klebsiellae, E. coli, enterobacters)</i>	
staphylococci	20 %
<i>(mainly S. aureus)</i>	
yeasts	5 %
<i>(some cases have polymicrobial etiology, agents are of hospital origin)</i>	

Etiology of respiratory NIs III – revision

Nosocomial aspiration pneumonia:

Older studies emphasized **anaerobes**

Newer studies are discovering the same etiology as in VAP and put more emphasis on **Gram-negative rods** (non-fermenting rather than enteric ones)

Pneumonia in febrile neutropenia:

Initial days: **G+ cocci** (staphylococci, pneumococci) occur twice more often than **G– rods** (enteric bacteria and pseudomonads)

Later on: ↓ **G+ cocci**, ↑ **candidae and aspergilli**

After allogenic transplantation of bone marrow: mainly **CMV**

Etiology of surgical wounds suppuration – revision

(depends on the terrain of the surgery)

Staphylococcus aureus

coagulase-negative staphylococci

Streptococcus pyogenes

enteric bacteria (*E. coli*)

bacteroids, prevotellae, peptostreptococci

Gram-negative non-fermenting rods

Clostridium perfringens

Etiology of sepsis during inserted i.v. catheter – revision

coagulase-negative staphylococci (>50 %)

– because of biofilm

enterococci – because of cephalosporins

Staphylococcus aureus

enteric bacteria (*E. coli*, *klebsiellae*)

Pseudomonas aeruginosa

***Acinetobacter* spp.**

***Candida* spp.**

Etiology of nosocomial virus infections – revision

influenza virus – mainly infants and older patients

RSV – newborns and suckling infants

adenoviruses – ophthalmologic wards

other respiratory viruses

CMV – after cytotoxic treatment

rubella virus – children (vaccination available now)

rotaviruses – mainly children

VHB – higher risk in longer hospitalization

HIV – in developing countries mostly

Predisposition to NIs – revision

Age – both extremes of age

Treatment – cytotoxic drugs, steroids, ATB

Underlying disease

hepatic disease

diabetes mellitus

cancer

renal failure

skin disorders

neutropenia

Trauma – incl. surgery and i.v. catheters

Prevention of NIs I – revision

Four main strategies:

- **Excluding sources of infection from the hospital environment**
- **Breaking the chain of infection from source to host**
- **Improving the host's ability to resist infection**
- **Investigating hospital infection**

Prevention of NIs II – revision

1. Exclusion of sources of infection from the hospital environment

- **Sterile** instruments, dressings, medicaments and intravenous fluids
- Using only **blood screened** for infectious agents
- **Clean linen, uncontaminated food**
- **Preventing contact with infected staff** – both acutely ill or carriers of pathogens

Prevention of NIs III – revision

2. Breaking the chain of infection

• Facilities

- ventilation systems & air flow (air-conditioning: legionellae, construction works: aspergilli)
- water systems (in particular warm water: legionellae)
- patient isolation
 - to protect a particularly susceptible patient
 - to prevent the spread of pathogens from a patient to others

• People

- facilitation of aseptic behavior of staff
- the most important is **effective hand washing**

Prevention of NIs IV – revision

3. Improving the host's resistance

- **Immunization**
 - influenza (older patients)
 - pneumococcal infections (before transplantation or splenectomy)
 - VHB (in seronegative persons before hemodialysis)
 - varicella (zoster Ig in immunocompromised exposed to VZV)
- **Appropriate ATB prophylaxis**
 - in „dirty“ surgery
 - In „super-clean“ surgery (orthopaedics, neurosurgery)
- **Reducing the risk of postoperative infection**
 - correct operating technique
 - care of invasive devices and intravenous fluids
 - correct nursing techniques (prevention of pressure sores) and active physiotherapy

Prevention of NIs V – revision

4. Investigating hospital infections

- **Surveillance** (= regular monitoring) – allows early recognition of any change in the number or type of hospital infection
- **Investigation of outbreaks** from epidemiological and microbiological point of view
- **Establishment and monitoring of procedures** designed to prevent infection

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Opportune infections: Definition and examples

Opportune infection = infection on a weakened terrain (in a lowered immunity)

Opportune infection is:

sometimes a **secondary one** during some other infection

often a **nosocomial one** = hospital-acquired

possibly a **iatrogenic one** = caused by a medical intervention

Examples of opportune infections during a HIV infection:

CMV retinitis, CMV or candidal oesophagitis CMV, herpes zoster, cryptosporidial or microsporidial enteritis, cryptococcal meningitis, toxoplasmatic encephalitis, colibacillary and other sepsis

Causes of lowered immunity

Defects of the **local barriers**

Defects of the **phagocytosis**

Defects of the **complement**

Defects of the **normal microflora**

Defects of the **humoral immunity**

Defects of the **cellular immunity**

Defects of local barriers – I

Burns

Other injuries incl. surgical wounds

Foreign bodies *in situ*

urinary catheters

i.v. catheters and cannulae

arteficial prostheses

Cystic fibrosis

Defects of local barriers – II

Burns

Not only a **direct impairment of barriers**, but also a **defect in neutrophil function** and burned tissue as a **culture medium** for bacteria

Little invasive infection limited to the eschar (scab)

Invasive infection → destruction of all skin layers → invasion of lymphatic vessels → sepsis

Agents: mainly *P. aeruginosa*, furthermore ***S. aureus***, enterobacteria, ***S. pyogenes*** and other streptococci and enterococci

Defects of local barriers – III

Other injuries

See one of previous lectures

Surgical wounds - I

Staphylococcus aureus

coagulase-negative staphylococci (mainly
Staphylococcus epidermidis)

enterobacteria (*Escherichia coli*, *Proteus mirabilis*)

Streptococcus pyogenes

anaerobes (*Peptostreptococcus anaerobius*, *P. micros*, *Bacteroides fragilis*)

Defects of local barriers – IV

Surgical wounds – II

Infection *during* a surgery:

preparing the operating field

terrain (clean versus contaminated one)

level of **asepsis**

surgical **technique**

Infection *after* a surgery:

slip (e.g breach of asepsis) during **dressings**

In general: **dangerous is the non-sterile touch**,
much less the non-sterile environment (but be
careful during implantation of artificial devices)

Defects of local barriers – V

Foreign bodies in situ (coated by the biofilm)

Urinary catheters: after >48 hrs quite commonly **G– rods** (periurethral and bowel flora – but transmission by hands also possible)

I.v. cannulae: >50 % *S. epidermidis* & other coagulase-negative **staphylococci** (from the skin), because of the biofilm resistant to ATB, infection not eradicable except by the removing the cannula

Artificial valves, joints, pacemakers, vascular prostheses, shunts: dtto

Cystic fibrosis

Impairment of ciliary function & production of mucus

After *S. aureus* and *H. influenzae* ***Pseudomonas aeruginosa*** (mucous strains producing alginate slime) and ***Burkholderia cepacia*** continue

Defects of phagocytosis – I

Neutropenia

in haematologic malignancies

in recipients of bone marrow

in recipients of transplanted solid
organs

Chronic granulomatous disease

Asplenia

Defects of phagocytosis – II

Neutropenia:

No circulating leukocytes → sepsis (the first sign or the result of cytotoxic chemotherapy)

Febrile neutropenia

Agents:

formerly G– rods (*E. coli*, *P. aeruginosa* etc.)

now G+ cocci (staphylococci, streptococci, enterococci) and yeasts (in particular if the neutropenia persists)

Defects of phagocytosis – III

Chronic granulomatous disease (CGD):

Defect of NADPH-oxidase → none H_2O_2

leukocytes in CGD cannot kill staphylococci
and other catalase-positive bacteria →
these microbes cause chronic deep
abscesses

Asplenia:

Encapsulated bacteria (pneumococci,
hemophilli) are not cleared from blood

Defects of complement

Relatively rare conditions

Resistance lowered against infections caused by neisseriae (mainly by **meningococci**)

Defects of normal flora

Problems mostly in **intestinal** microflora

After some **antibiotics** (clindamycin, cefoxitin, amoxicillin/clavulanic acid) ***Clostridium difficile*** multiplies excessively

Defects of humoral immunity

In particular agents of **pyogenic infections** and enteroviruses

Defects of cellular immunity

In particular infections by **intracellular microbes**:

viruses (mainly CMV, VZV, HSV)

mycobacteria (not only *M. tbc*, but also atypical as *M. avium-intracellulare*), *Nocardia asteroides*, *Listeria monocytogenes*

fungi (candidae, aspergilli, *Pneumocystis jirovecii*, *Cryptococcus neoformans*)

parasites (*Toxoplasma gondii*, *Cryptosporidium parvum*, *Isospora belli*, *Strongyloides stercoralis* and other)

Homework 15

The third part of the cycle mentioned in the 13th lecture



Answer and questions

The solution of the homework and possible questions please mail (on Monday Dec. 12th at 6.30. at the latest) to the address

mvotava@med.muni.cz

**Thank you for your attention
Merry Christmas!
Good luck in the examination!**