



IMPORTANCE OF CYTOGENETICS IN HEMATOLOGIC MALIGNANCIES

Marie Jarošová
and Sabina Sevcikova

IHOK FN Brno
and BMG UPF LF MU



What is cytogenetics?

- Study of number and structure of chromosomes, their properties, behavior during cell division, influence on phenotype
- Any changes in number or structure may lead to disease
- Changes lead to disrupted genes leading to proteins that do not function correctly
- Depending on size, location, and timing, structural changes in chromosomes can lead to birth defects, syndromes or even cancer



Cytogenetics

- Clinical cytogenetics (germline alterations)
 - All cells are effected
 - Changes are stable
- Tumor cytogenetics (acquired alterations)
 - Only a portion of cells is effected
 - Clonal evolution or regression
 - Role of sensitivity of any method

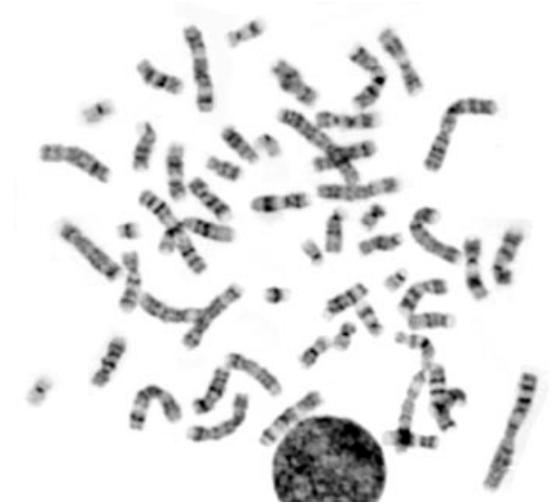


Tumor genetics

- A tumor is a genetic disease that arises as a consequence of accumulation of various genetic changes
- Every 3rd person will get a tumor disease
- In the Czech Republic, more than 70 thousand people are diagnosed with a tumor every year
- Tumor cells are characterized by chromosomal changes: numerical or structural changes of chromosomes

Tumor cytogenetics

- Studies of acquired chromosomal changes of tumor cells
- Analyses of numerical and structural changes of chromosomes
- Basic method – G- banding
- One analysis checks the entire genome



History of cytogenetics

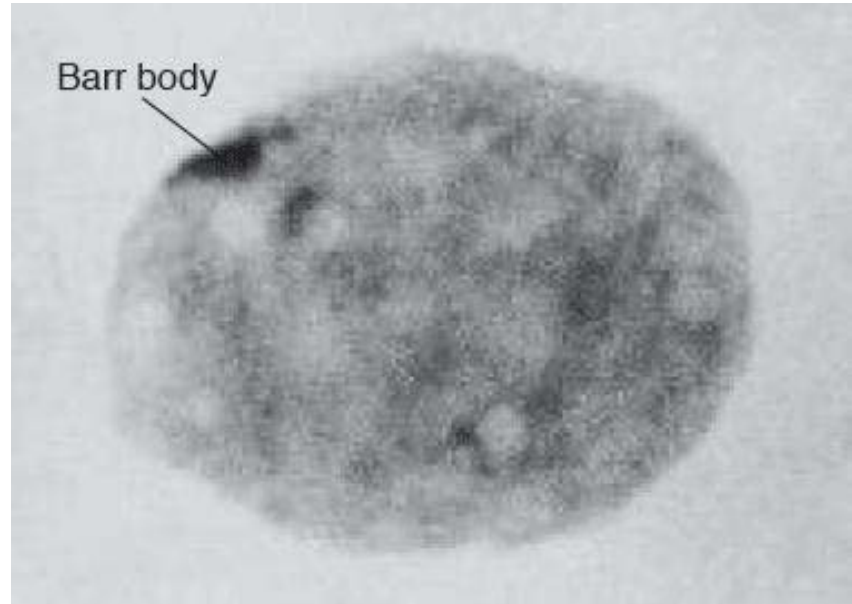


Early cytogenetics

- 1842 – Carl Nageli- cell division (anomaly), botanist
- 1879 – Walther Flemming – chromosomal movement during mitosis
- 1882 – published his work
- 1888 – Waldeyer - chromosome



1940s – the sex revolution



Dr. Barr and Dr. Bertram discovered sex-chromatin, now known as the Barr body, while working at The University of Western Ontario, in 1949.

1950s – the hypotonic revolution

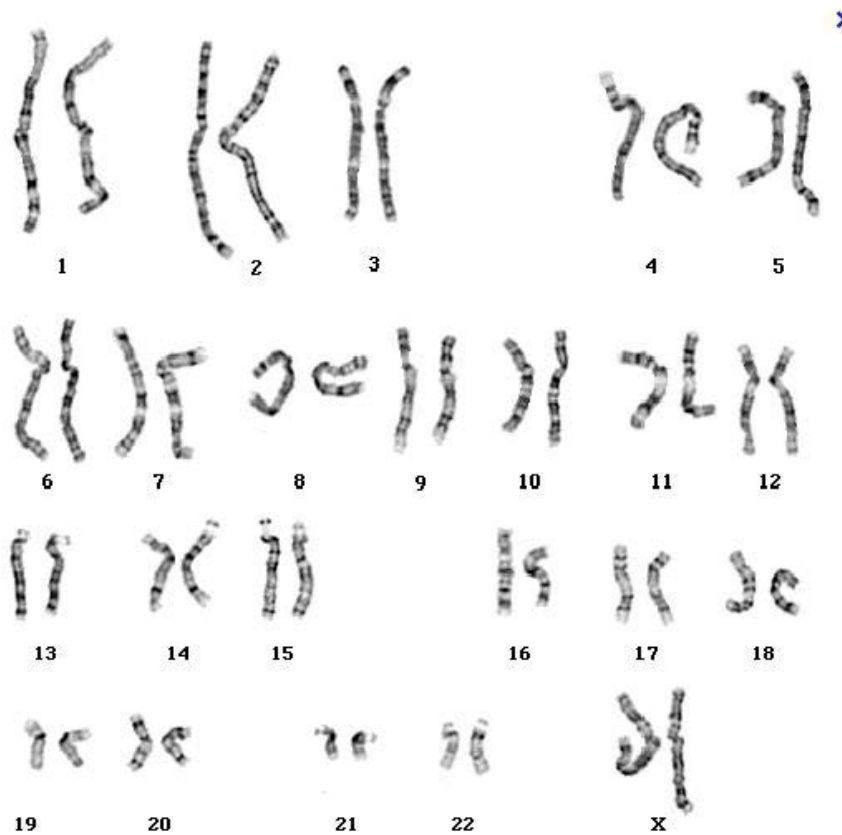
- T. C. Hsu discovered the utility of hypotonic solution in 1952
- Tijo and Levan reported the correct chromosome number in humans *in vitro*.
- This was confirmed within a year, *in vivo* by Ford and Hamerton (1956)



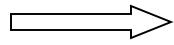
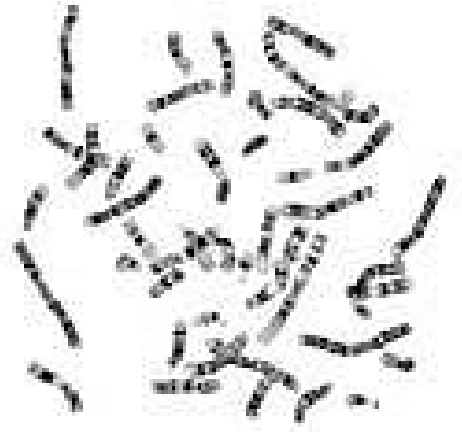
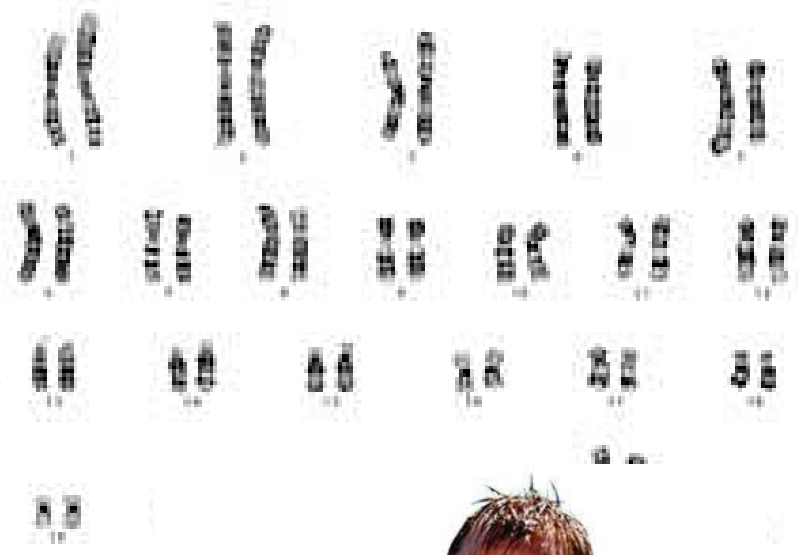
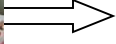
They could count the chromosomes but classification was rough and approximate.



1970s – the banding revolution



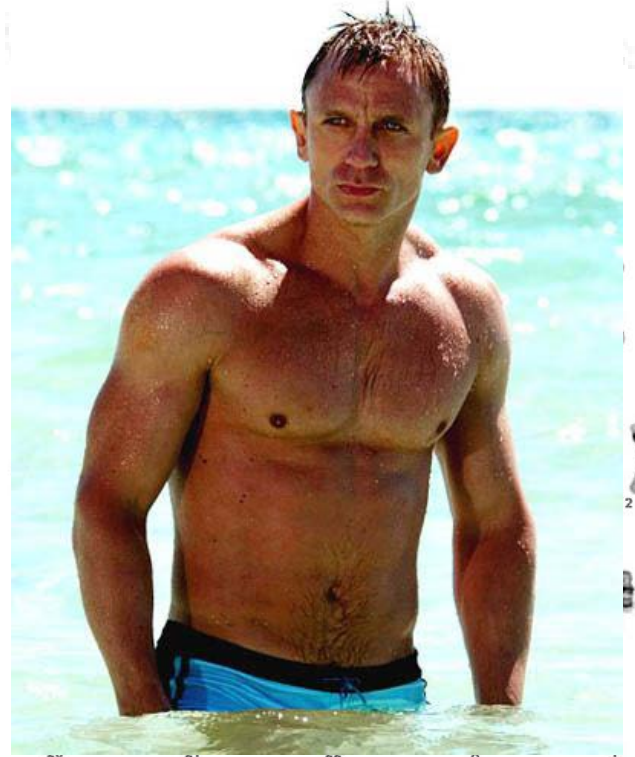
High Resolution G-banding,
Yunis, 1975



Human male G-bands



♂





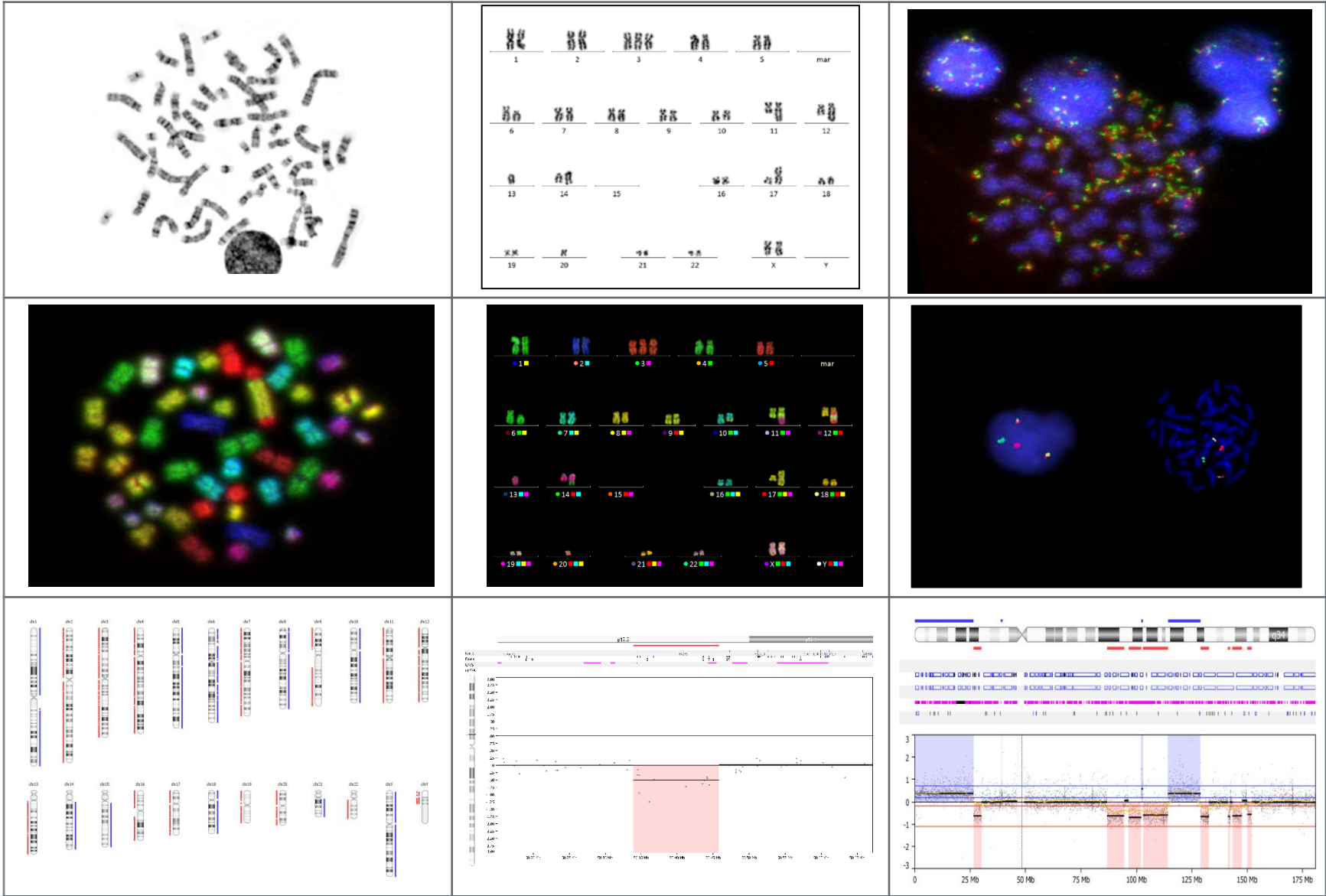
80s, 90s, 00s

- 80s – imaging – automated imaging
- 90s – colors – unique locus probes
- 00s – genomics – SNP arrays, next gen sequencing

Cytogenetic methods

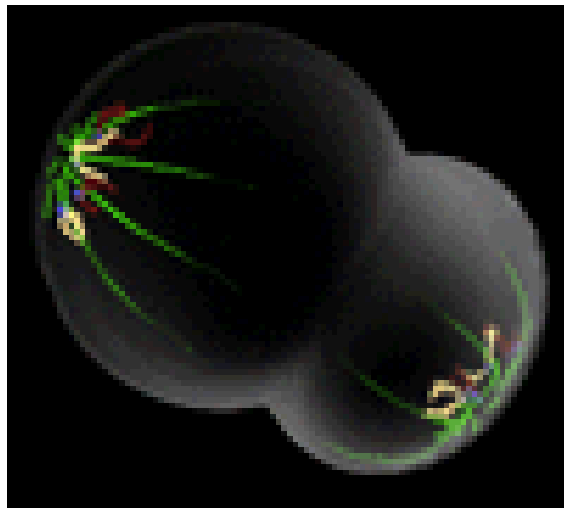
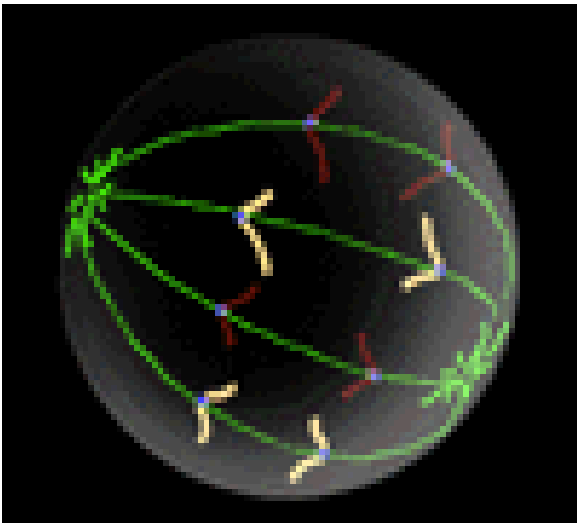
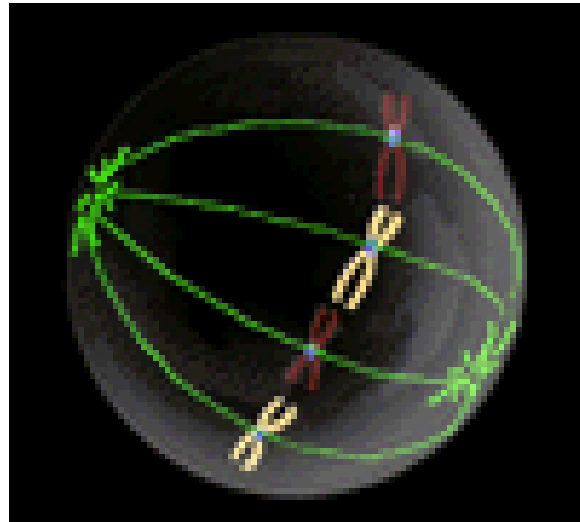
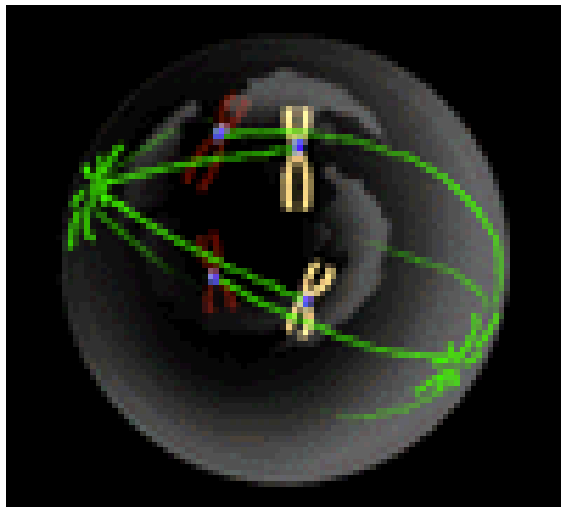
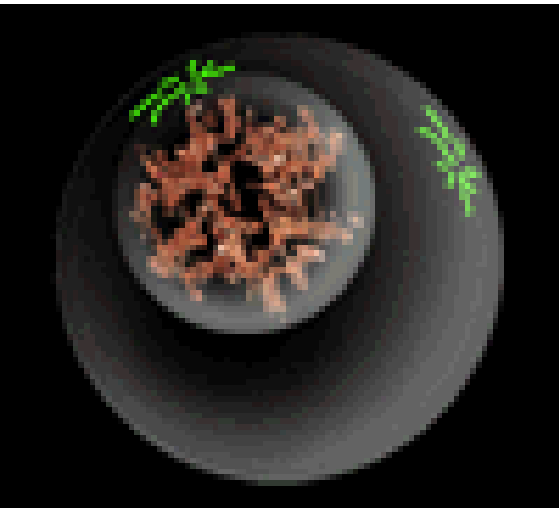
Cytogenetics

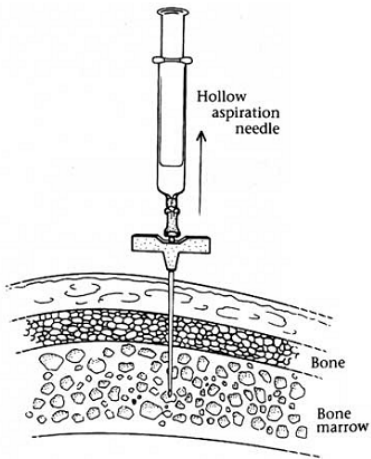
FISH



arrayCGH/SNP array

Metaphase



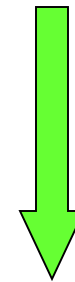


1-2ml

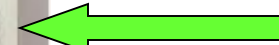


+

colcemide



cultivation
2/24/72 hrs



- ✓ bone marrow
- ✓ peripheral blood
- ✓ lymph node

COLCEMIDE BLOCKS MITOSIS IN METAPHASE

ABBOTT Laboratories, s.r.o.
Diagnostic Division

Abbott Office Park
Křivánská 100
150 00 Praha 5
Tel: +420 257 280 100
Fax: +420 257 280 100

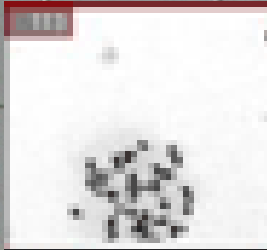


INFC

1	2	3	4	5	6	7
8	9	10	11	12	13	14
15	16	17	18	19	20	21
22	23	24	25	26	27	28
29	30	31				



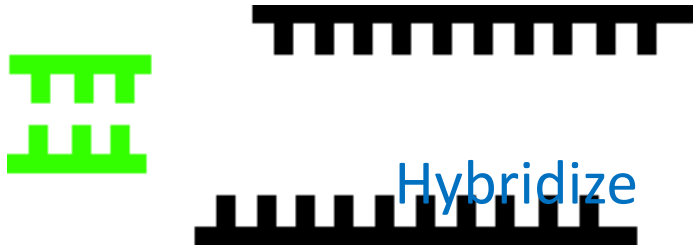
1	2	3	4	5	6	7	8	9	10
11	12	13	14	15	16	17	18	19	20
21	22	23	24	25	26	27	28	29	30
31	32	33	34	35	36	37	38	39	40
41	42	43	44	45	46	47	48	49	50
51	52	53	54	55	56	57	58	59	60
61	62	63	64	65	66	67	68	69	70
71	72	73	74	75	76	77	78	79	80
81	82	83	84	85	86	87	88	89	90
91	92	93	94	95	96	97	98	99	100





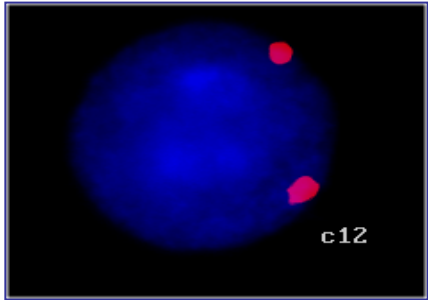
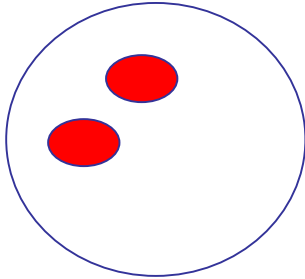
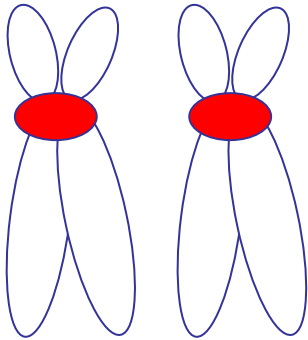
Molecular cytogenetics

Denature

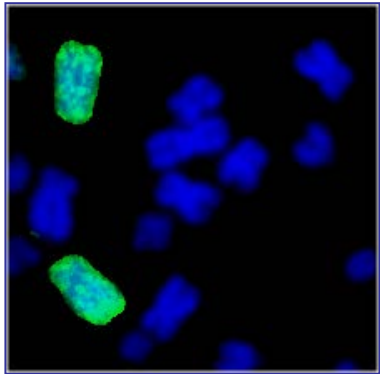
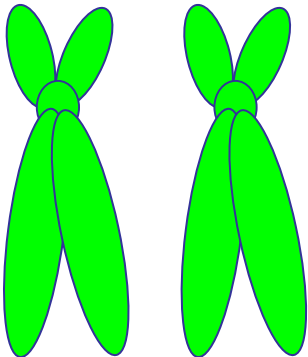


- Methods based on fluorescence in situ hybridization (FISH) – based on molecular as well as classical cytogenetics
- Methods use the basic property of single stranded DNA to bind together based on complementarity

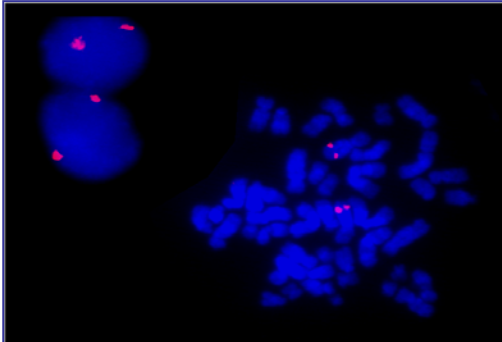
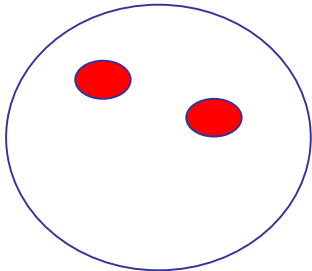
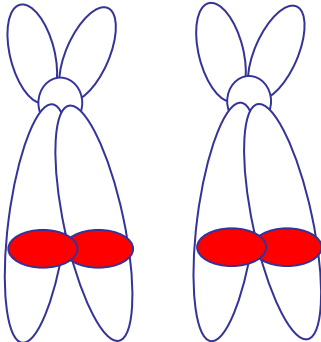
Types of probes



centromeric



Whole chromosome



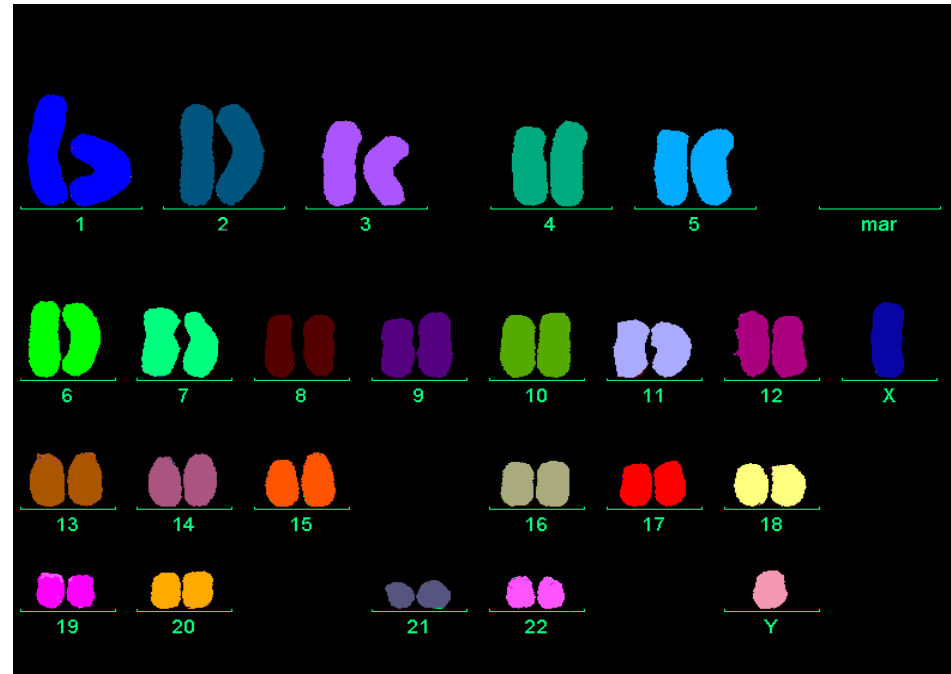
gene

Multicolor fluorescence in situ hybridization (M-FISH)

M-FISH is based on hybridization of 24 fluorescently labeled whole chromosome probes that allow the staining of all chromosomal pairs by different colors

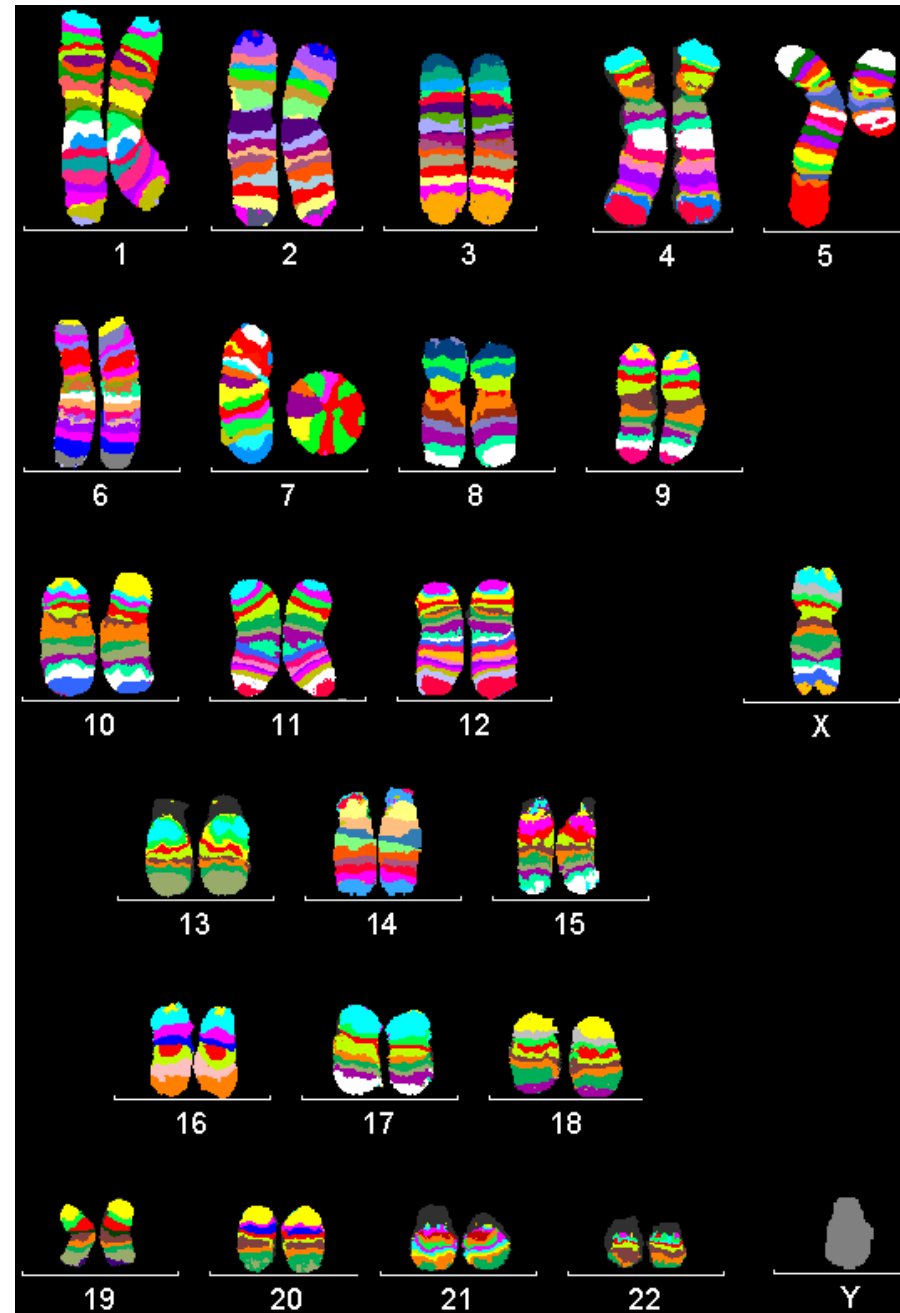
24 color karyotyping hybridization and detection kit

Chr.	FITC	Spectrum Orange	Texas Red	Cy5	DEAC
1				Red	
2					Blue
3			Red		
4	Green				
5		Yellow			
6	Green			Red	
7				Red	Blue
8			Red	Red	
9		Yellow		Red	
10	Green				Blue
11	Green		Red		
12	Green	Yellow			
13			Red		Blue
14		Yellow			Blue
15		Yellow	Red		
16	Green			Red	Blue
17	Green		Red	Red	
18	Green	Yellow		Red	
19			Red	Red	Blue
20		Yellow		Red	Blue
21	Green	Yellow	Red	Red	Blue
22	Green		Red		Blue
X	Green	Yellow			Blue
Y		Yellow	Red		Blue

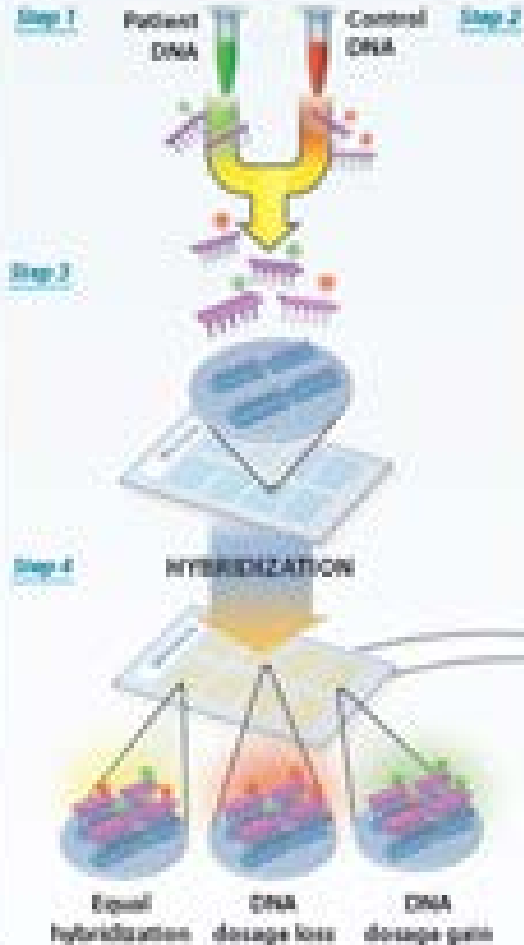


Mband FISH

- Combines paint probes specific for a region of a chromosome
- Banding covers the whole chromosome



Array CGH: The Complete Process

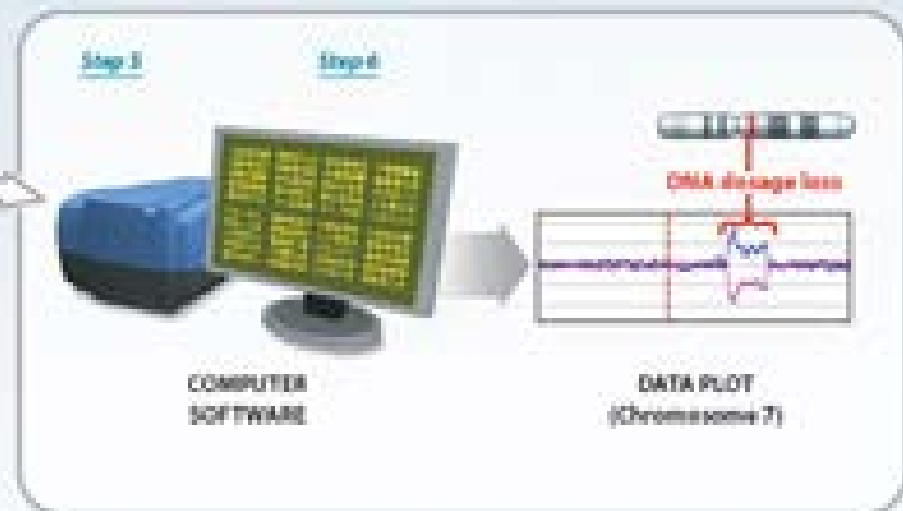


Steps 1-3 Patient and control DNA are labeled with fluorescent dyes and applied to the microarray

Step 4 Patient and control DNA compete to attach, or hybridize, to the microarray

Step 5 The microarray scanner measures the fluorescent signals.

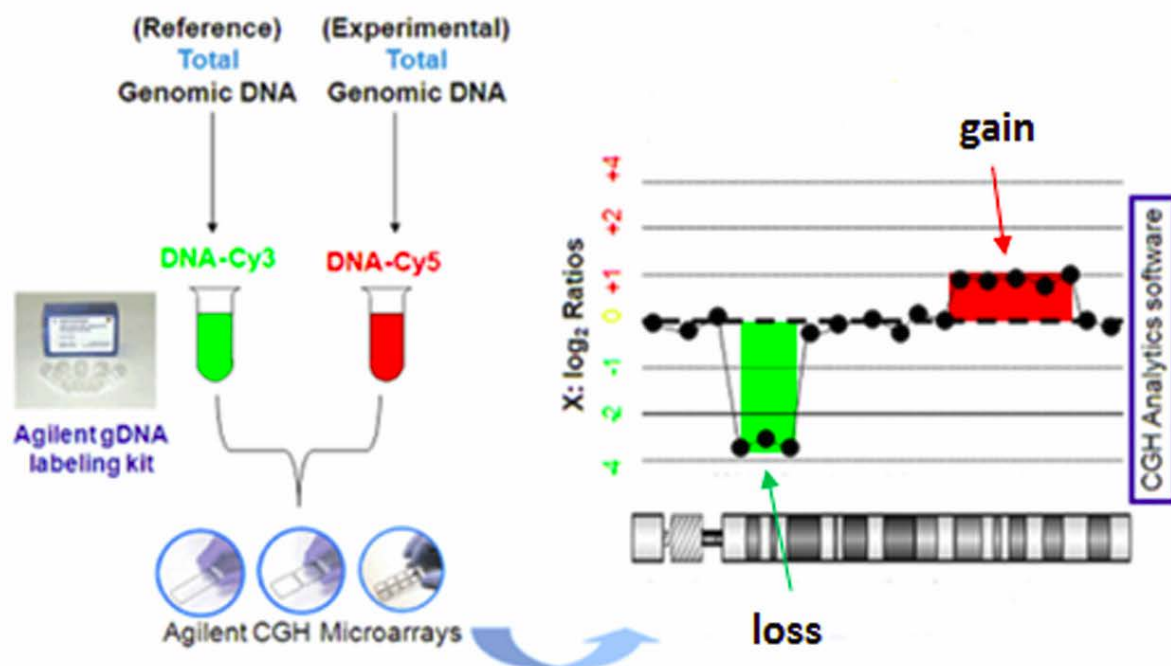
Step 6 Computer software analyzes the data and generates a plot.



COMPUTER SOFTWARE

DATA PLOT
(Chromosome 7)

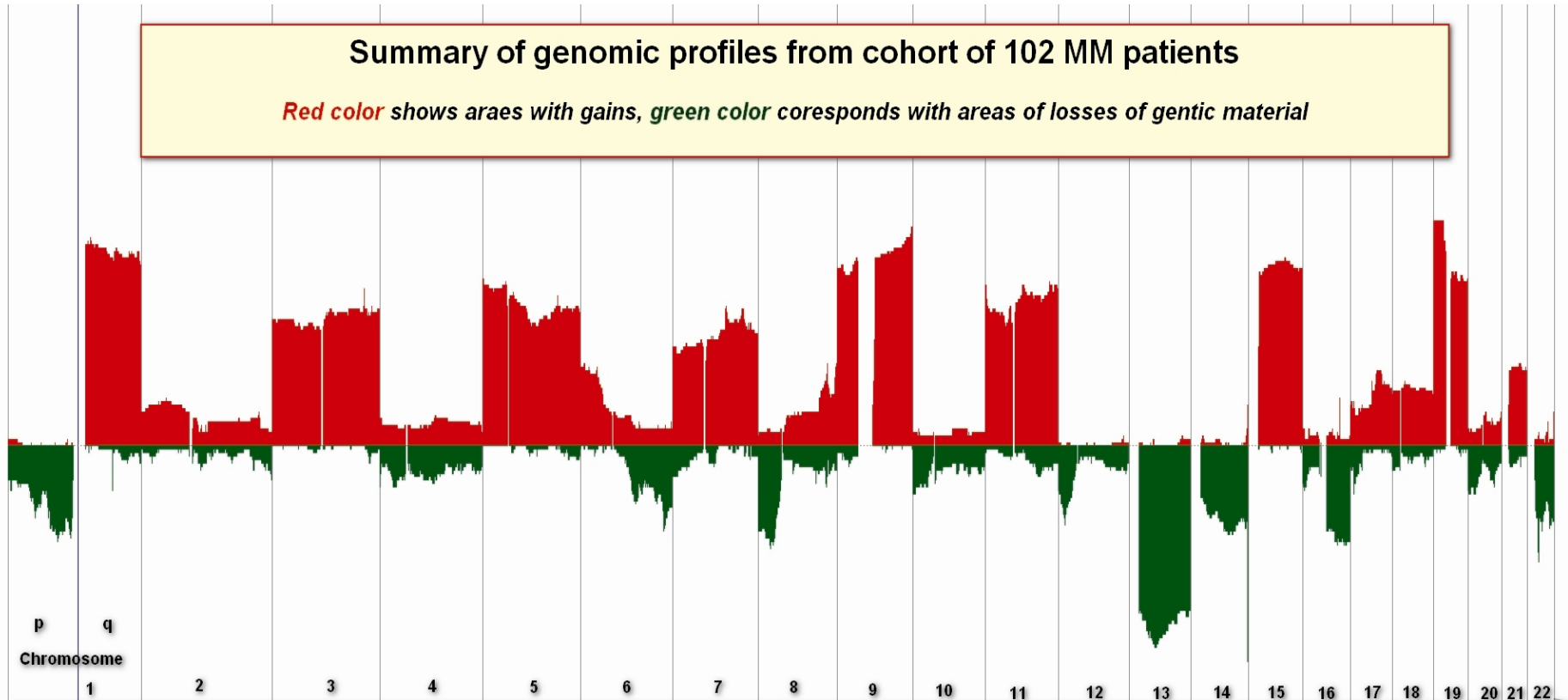
Figure 3



aCGH in MM

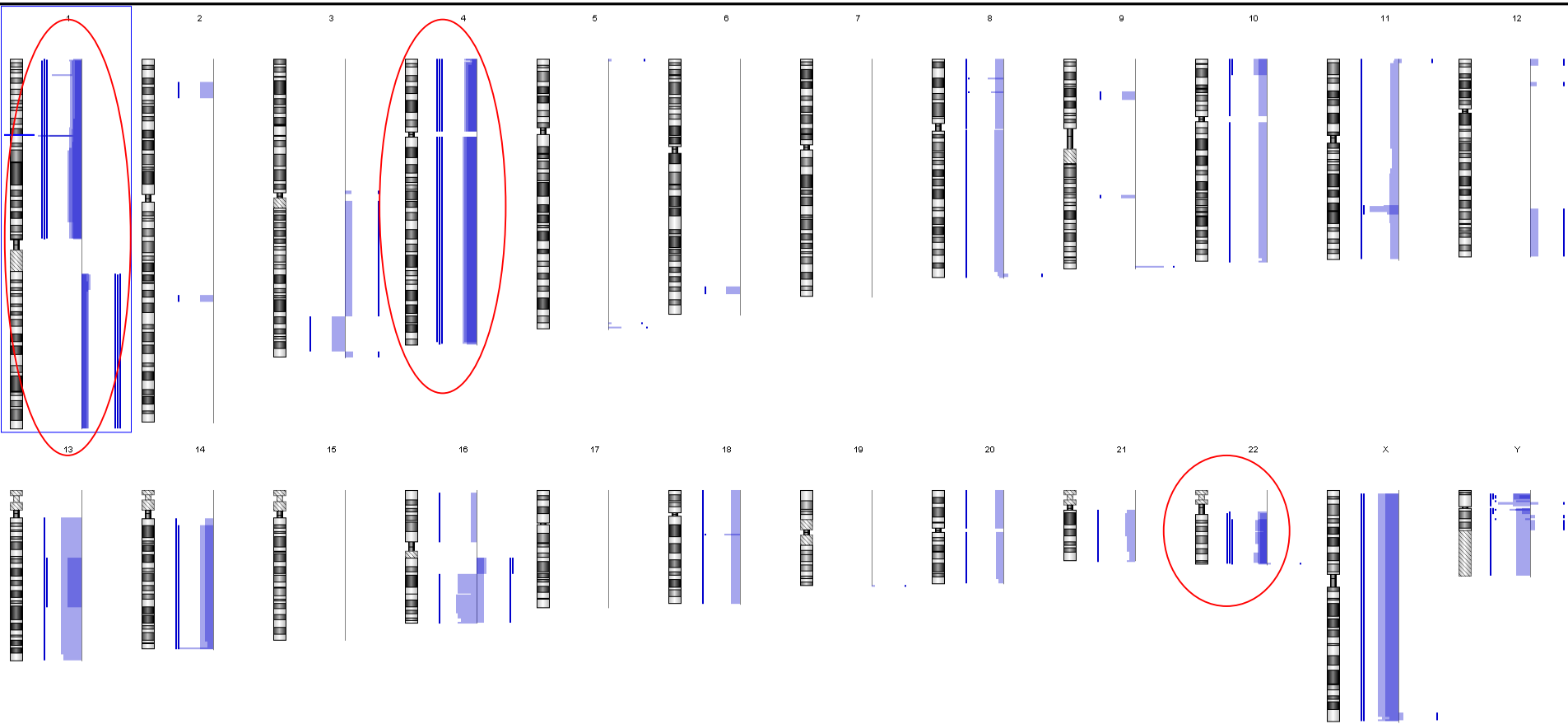
Summary of genomic profiles from cohort of 102 MM patients

Red color shows areas with gains, green color corresponds with areas of losses of genetic material



J. Smetana

Analysis





Cytogenetics in hematology

- Diagnosis
- Prognosis
- Treatment decisions



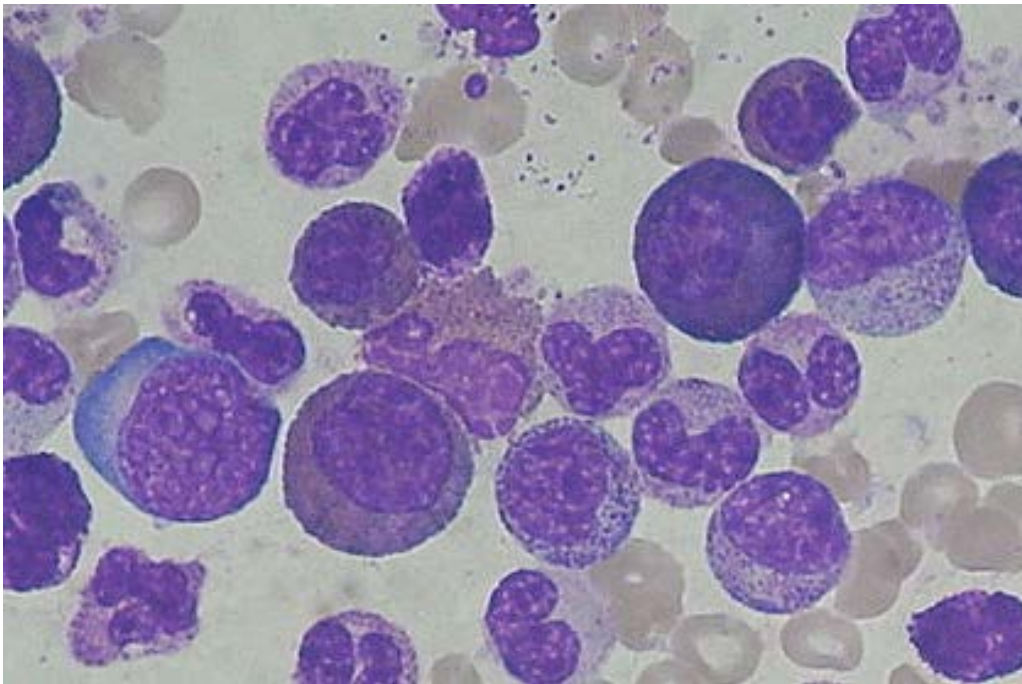
Genetic changes in hematological malignancies

- 90-95% of CML
- 60-80% of AML
- 60% of MDS
- 50-80% of CLL
- 60-90% of NHL
- 70-90% of MM
- 70-90% of ALL



Philadelphia chromosome (Ph)

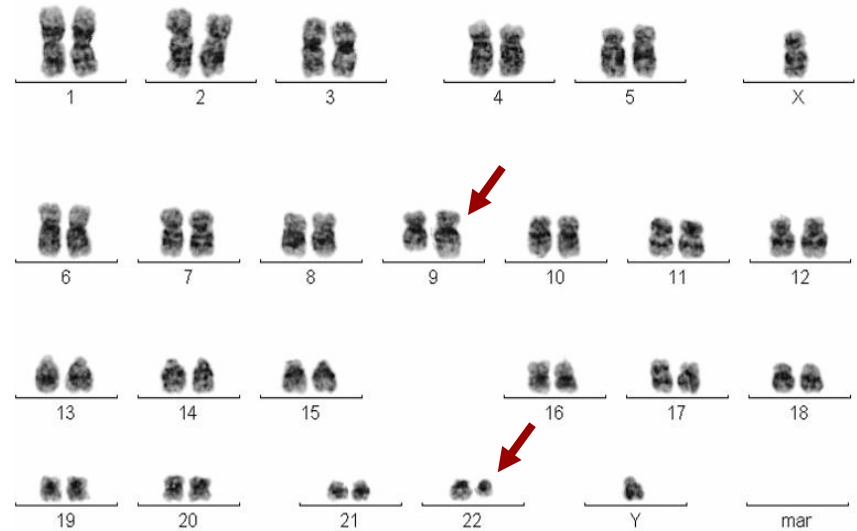
First specific chromosomal aberration linked to a tumor



Cytogenetics of CML

Diagnosis

90-95% Ph1 result of translocation
 $t(9;22)(q34;q21)$



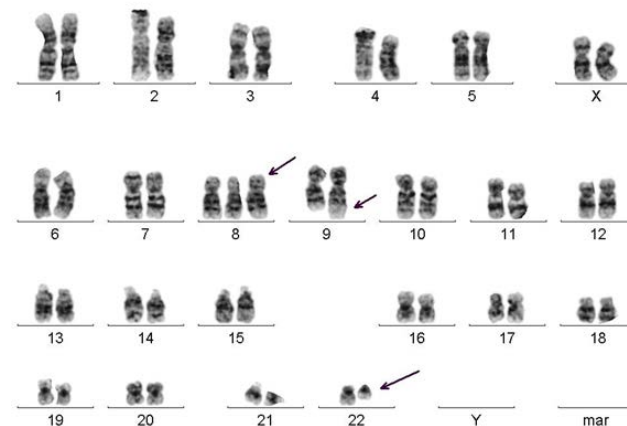
Prognosis

Additional chromosomal changes

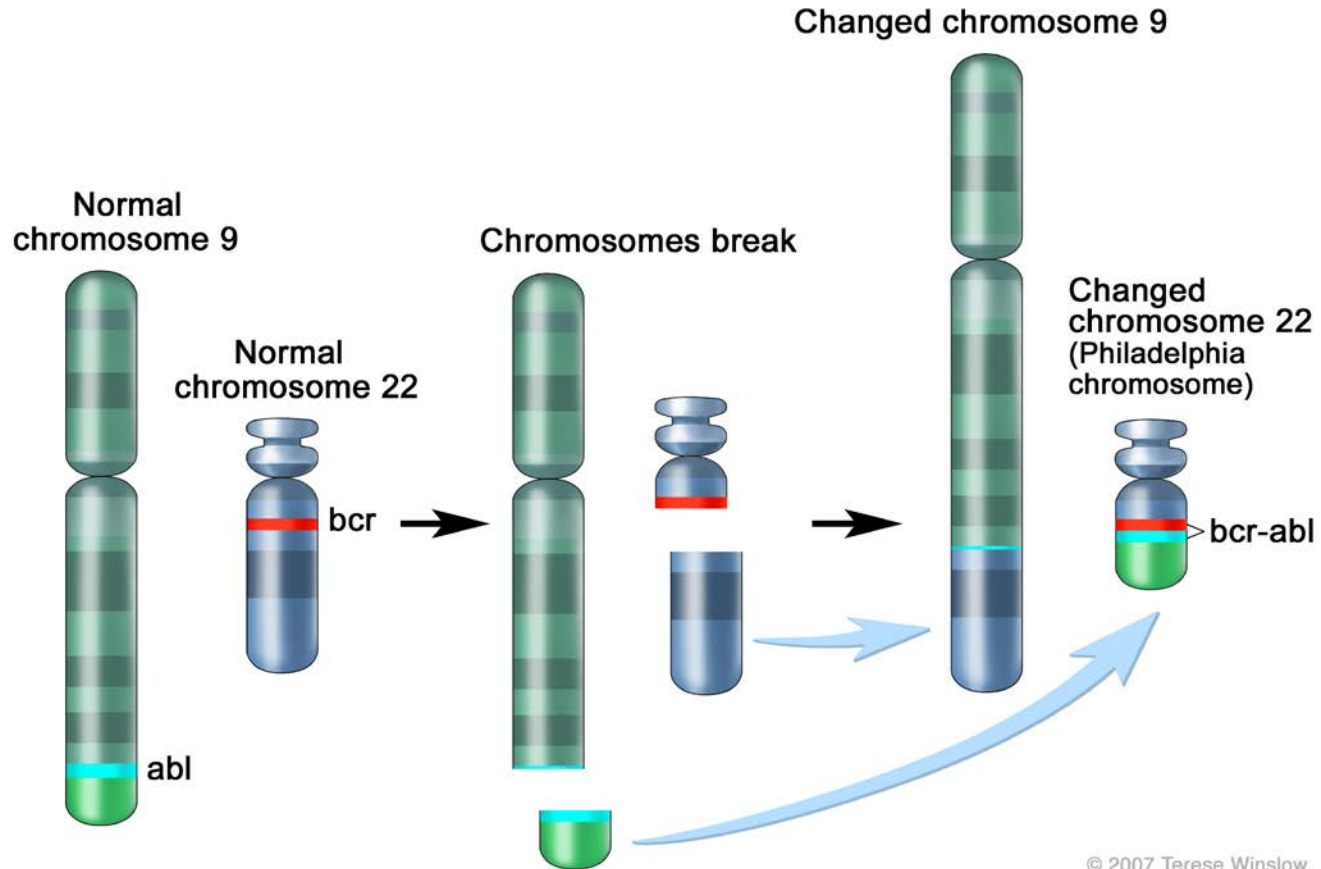
Diagnosis: ~12%

Accelerated phase: ~30%

Blast crisis : ~70%



Philadelphia chromosome



Philadelphia chromosome (Ph)

- 1960 – Peter Nowell and David Hungerford described abnormal chromosome in CML
- First genetic signature of cancer – growth advantage for abnormal cells?
- Cause or consequence?
- Janet Rowley in the 1972 – t(9,22)





Philadelphia chromosome (Ph)

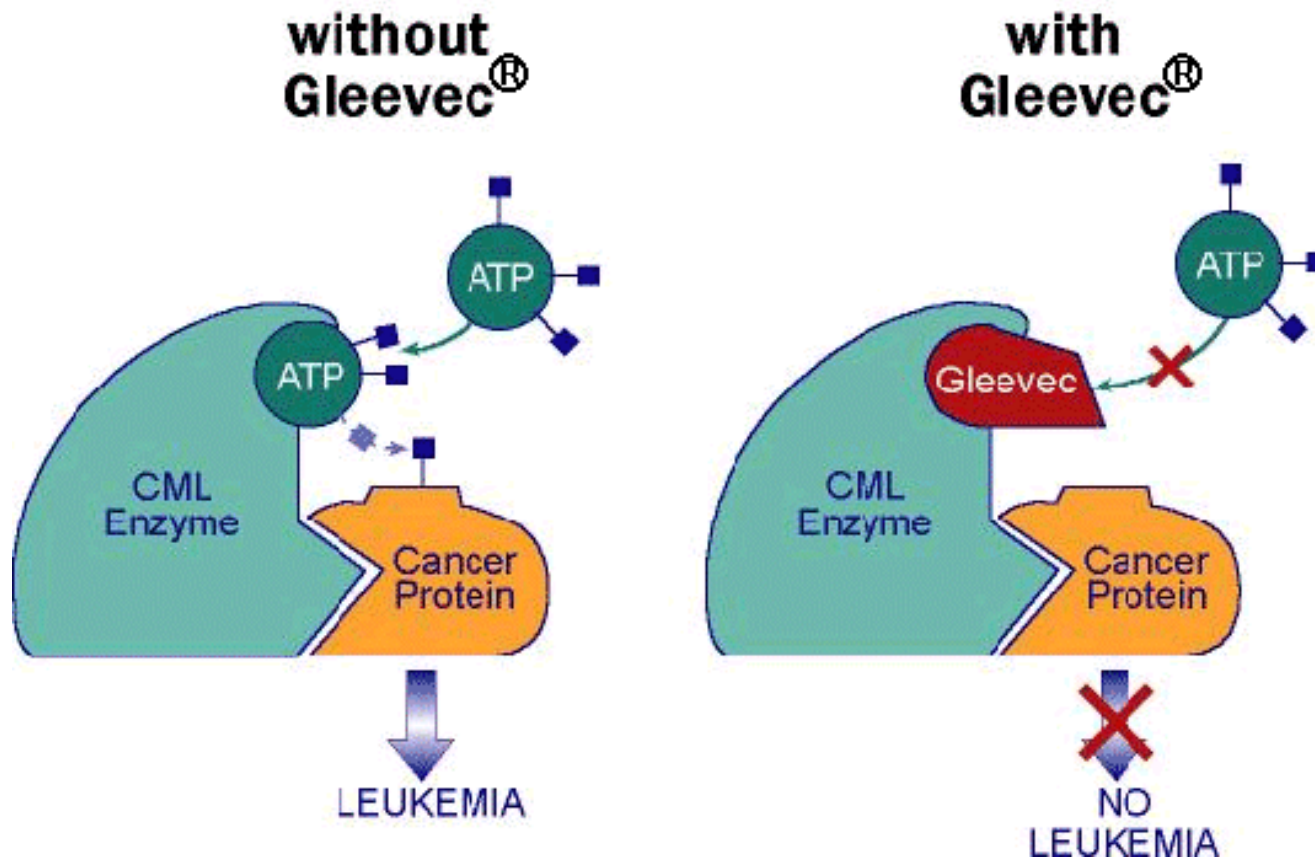
- 1983 abl (Heisterkamp)
- 1984 bcr (Groffen)
- 1990 bcr-abl cause of CML (Daley)
- 1990 - bcr-abl- abnormal tyrosine kinase activity (Lugo)
- Chronic phase, accelerated phase, blast crisis
- Bad prognosis



Gleevec - Imatinib mesylate (1993)

- Activity against CML colonies (Druker 1996)
- 2 years later – clinical study, 31 patients, 98% response
- Clinical trial phase III – 16 countries, 177 centers, 1100 patients – closed early
- All patients moved to Gleevec arm
- Survival 95%, 65% in blast crisis
- Molecular positivity still a problem
- Dasatinib, nilotinib....

Gleevec: HOW IT WORKS



https://www.google.cz/search?q=gleevec+cml&dcr=0&source=Inms&tbm=isch&sa=X&ved=0ahUK EwjokaX99NnWAhXMDMAKHUfAAisQ_AUICigB&biw=1366&bih=604#imgrc=TNlp3Ot1Yx6sbM:



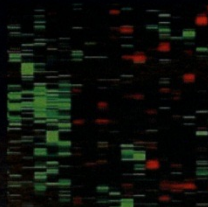
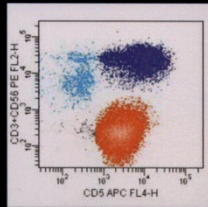
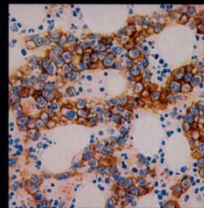
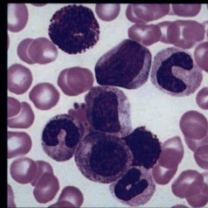
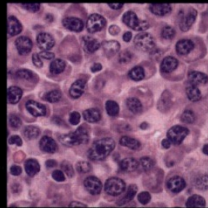
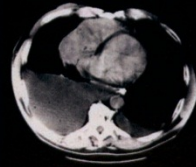
MRD in CML

	Type of Response	Definition
CHR	Complete Hematologic Response	Normal differential, WBC & platelets \leq ULN
MCyR	Major cytogenetic Response	0–35% Ph+marrow metaphases
CCyR	Complete Cytogenetic Response	0% Ph+marrow metaphases
MMR	Major Molecular Response	BCR-ABL/ABL \leq 0.1% (International Scale)
MR ^{4.0}		BCR-ABL/ABL \leq 0.001% (IS) “4-log reduction”
MR ^{4.5}		BCR-ABL/ABL \leq 0.003% (IS) “4.5-log reduction”
CMR	Complete Molecular Response	Undetectable BCR-ABL (test of sensitivity \geq 4.5 logs)

WHO Classification 2008

WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues

Edited by Steven H. Swerdlow, Elias Campo, Nancy Lee Harris, Elaine S. Jaffe, Stefano A. Pileri, Harald Stein, Jürgen Thiele, James W. Vardiman





WHO Classification

- Since 2008, cytogenetics is part of diagnosis and classification for many hematological malignancies
 - Cytogenetics is a part of WHO classification of AML
 - Together with cytomorphology stratifies MDS patients
 - Classification of lymphomas – histology, cytogenetics and FISH confirm classification
 - Is part of prognostic stratification of MM

WHO classification of AML

The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia

Daniel A. Arber,¹ Attilio Orazi,² Robert Hasserjian,³ Jürgen Thiele,⁴ Michael J. Borowitz,⁵ Michelle M. Le Beau,⁶ Clara D. Bloomfield,⁷ Mario Cazzola,⁸ and James W. Vardiman⁹

Acute myeloid leukemia (AML) and related neoplasms

AML with recurrent genetic abnormalities

AML with t(8;21)(q22;q22.1); *RUNX1-RUNX1T1*

AML with inv(16)(p13.1q22) or t(16;16)(p13.1;q22); *CBFB-MYH11*

APL with *PML-RARA*

AML with t(9;11)(p21.3;q23.3); *MLLT3-KMT2A*

AML with t(6;9)(p23;q34.1); *DEK-NUP214*

AML with inv(3)(q21.3q26.2) or t(3;3)(q21.3;q26.2); *GATA2, MECOM*

AML (megakaryoblastic) with t(1;22)(p13.3;q13.3); *RBM15-MKL1*

Provisional entity: AML with BCR-ABL1

AML with mutated *NPM1*

AML with biallelic mutations of *CEBPA*

Provisional entity: AML with mutated RUNX1

AML with myelodysplasia-related changes

Therapy-related myeloid neoplasms

AML, NOS

AML with minimal differentiation

AML without maturation

AML with maturation

Acute myelomonocytic leukemia

Acute monoblastic/monocytic leukemia

Pure erythroid leukemia

Acute megakaryoblastic leukemia

Acute basophilic leukemia

Acute panmyelosis with myelofibrosis

Myeloid sarcoma

Myeloid proliferations related to Down syndrome

Transient abnormal myelopoiesis (TAM)

Myeloid leukemia associated with Down syndrome

WHO myeloid neoplasm and acute leukemia classification

Blastic plasmacytoid dendritic cell neoplasm

Acute leukemias of ambiguous lineage

Acute undifferentiated leukemia

Mixed phenotype acute leukemia (MPAL) with t(9;22)(q34.1;q11.2); *BCR-ABL1*

MPAL with t(v;11q23.3); *KMT2A* rearranged

MPAL, B/myeloid, NOS

MPAL, T/myeloid, NOS

B-lymphoblastic leukemia/lymphoma

B-lymphoblastic leukemia/lymphoma, NOS

B-lymphoblastic leukemia/lymphoma with recurrent genetic abnormalities

B-lymphoblastic leukemia/lymphoma with t(9;22)(q34.1;q11.2); *BCR-ABL1*

B-lymphoblastic leukemia/lymphoma with t(v;11q23.3); *KMT2A* rearranged

B-lymphoblastic leukemia/lymphoma with t(12;21)(p13.2;q22.1); *ETV6-RUNX1*

B-lymphoblastic leukemia/lymphoma with hyperdiploidy

B-lymphoblastic leukemia/lymphoma with hypodiploidy

B-lymphoblastic leukemia/lymphoma with t(5;14)(q31.1;q32.3) *IL3-IGH*

B-lymphoblastic leukemia/lymphoma with t(1;19)(q23;p13.3); *TCF3-PBX1*

Provisional entity: B-lymphoblastic leukemia/lymphoma, BCR-ABL1-like

Provisional entity: B-lymphoblastic leukemia/lymphoma with iAMP21

T-lymphoblastic leukemia/lymphoma

Provisional entity: Early T-cell precursor lymphoblastic leukemia

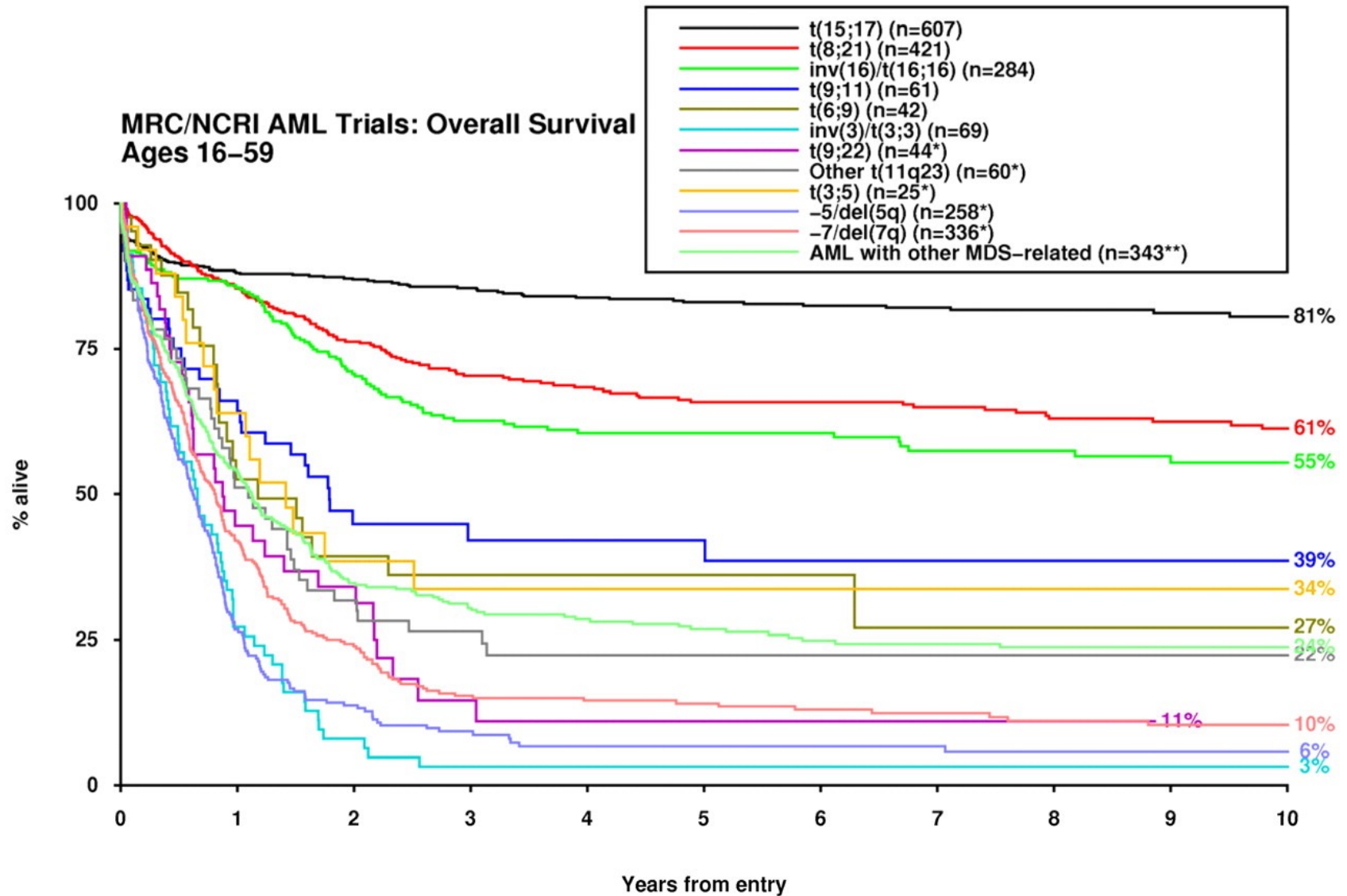
Provisional entity: Natural killer (NK) cell lymphoblastic leukemia/lymphoma

WHO prognostic stratification of AML

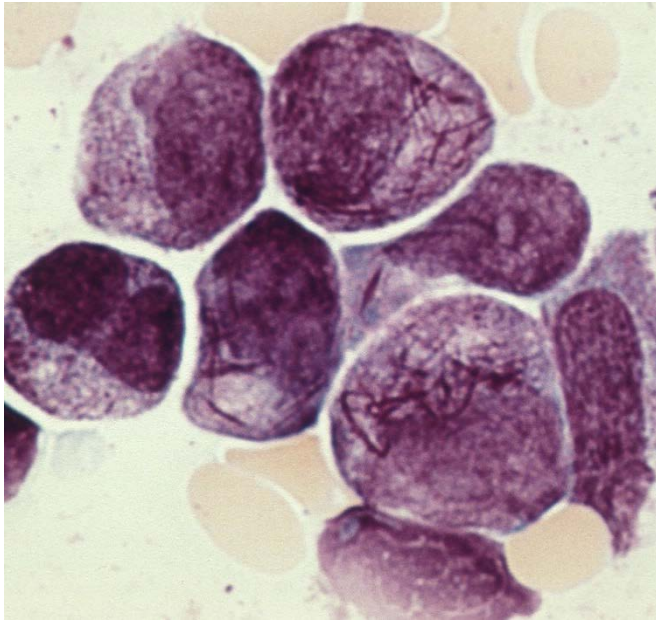
Genetic group	Subsets
Favorable	t(8;21)(q22;q22); <i>RUNX1-RUNX1T1</i> inv(16)(p13.1q22) or t(16;16)(p13.1;q22); <i>CBFB-MYH11</i> Mutated <i>NPM1</i> without <i>FLT3-ITD</i> (normal karyotype) Mutated <i>CEBPA</i> (normal karyotype)
Intermediate-I*	Mutated <i>NPM1</i> and <i>FLT3-ITD</i> (normal karyotype) Wild-type <i>NPM1</i> and <i>FLT3-ITD</i> (normal karyotype) Wild-type <i>NPM1</i> without <i>FLT3-ITD</i> (normal karyotype)
Intermediate-II	t(9;11)(p22;q23); <i>MLLT3-MLL</i> Cytogenetic abnormalities not classified as favorable or adverse†
Adverse	inv(3)(q21q26.2) or t(3;3)(q21;q26.2); <i>RPN1-EV11</i> t(6;9)(p23;q34); <i>DEK-NUP214</i> t(v;11)(v;q23); <i>MLL</i> rearranged -5 or del(5q); -7; abn(17p); complex karyotype‡

Döhner H. et al. for the ELN, Blood 2010

Stratification based on cytogenetics



APL $t(15;17)(q22;q12)$ / *PML-RARA*

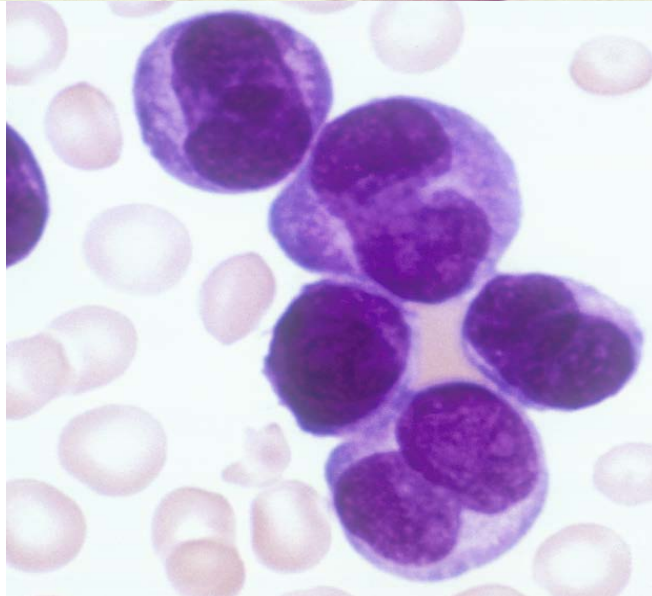


15/17 TRANSLOCATION, A CONSISTENT CHROMOSOMAL CHANGE IN ACUTE PROMYELOCYTIC LEUKAEMIA

SIR,—We have described a similar chromosomal abnormality in two patients with acute promyelocytic leukaemia

Department of Medicine,
Franklin McLean Memorial
Research Institute,
University of Chicago,
Chicago, Illinois 60637, U.S.A.

JANET D. ROWLEY
HARVEY M. GOLOMB
CHARLOTTE DOUGHERTY



$t(15;17)(q22;q12)$



Targeted treatment of APL

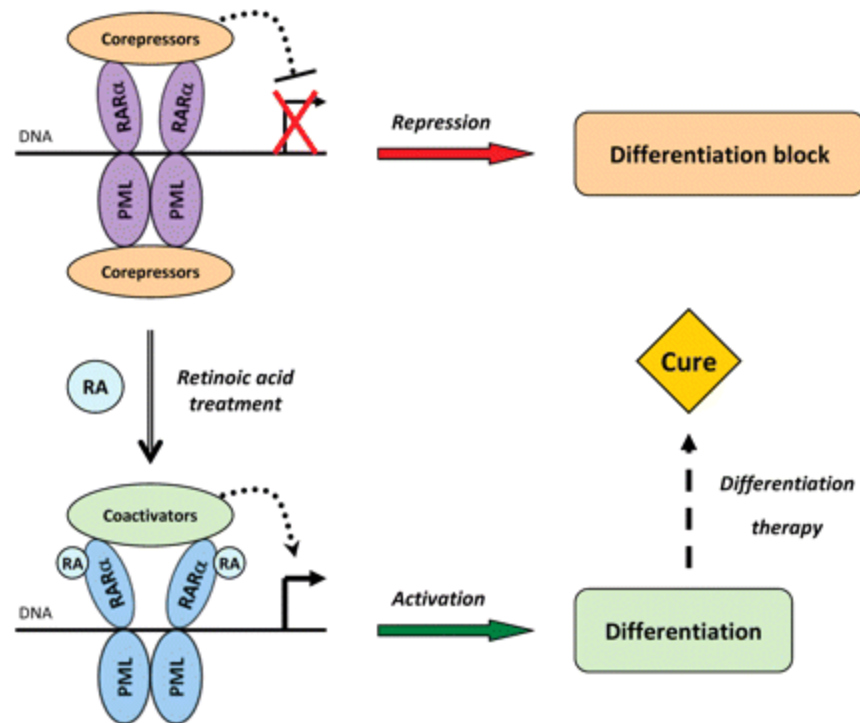
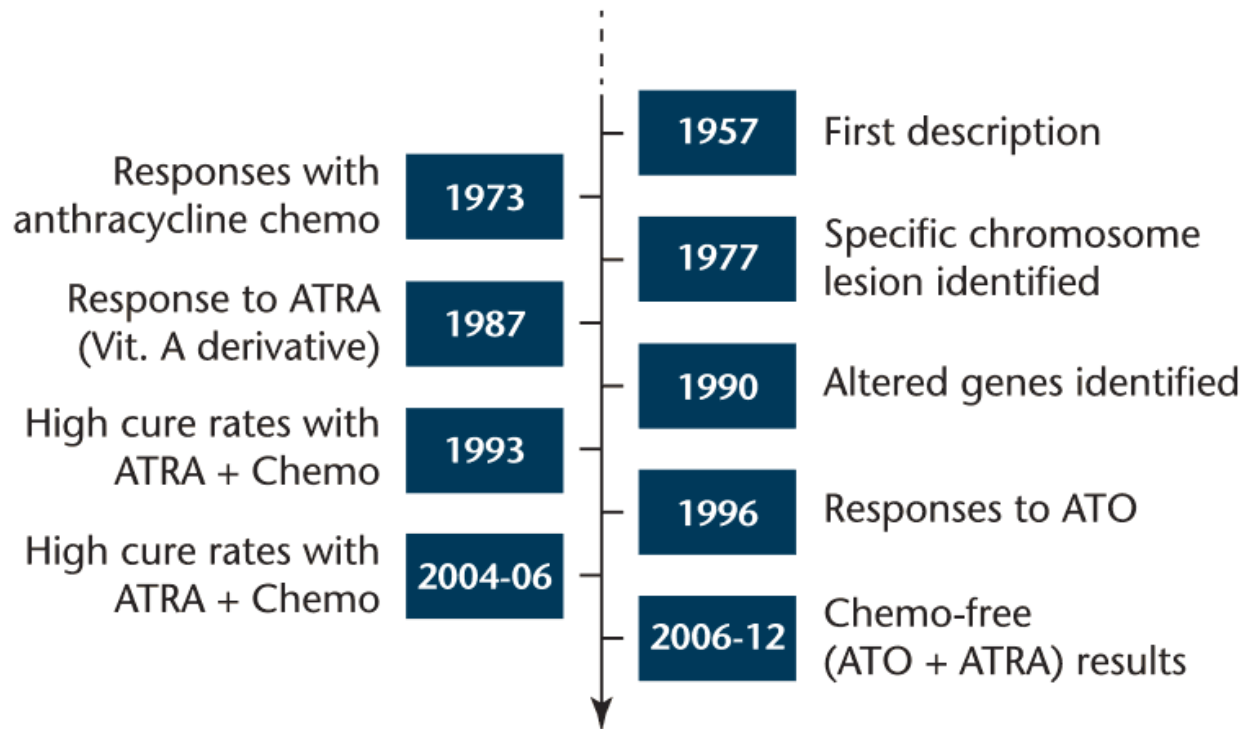


Figure 1. History of APL

Acute Promyelocytic Leukemia: From Highly **Fatal** to Highly **Curable**



ATO = arsenic trioxide; ATRA = all-transretinoic acid; Chemo = chemotherapy

Adapted from Dr. Lo-Coco's presentation at AMHOQ, 2013

ALL

- heterogeneous disease with monoclonal proliferation and expansion of lymphoid cells in BM, PB and other organs
- Cytogenetics- prognostic significance
- Immunophenotyping – diagnostic significance

TABLE 2: WHO 2008 classification of acute lymphoblastic leukemia (ALL)

Precursor lymphoid neoplasms

B-cell lymphoblastic leukemia/lymphoma, not otherwise specified

B-cell lymphoblastic leukemia/lymphoma with recurrent genetic abnormalities

B-cell lymphoblastic leukemia/lymphoma with t(9;22)(q34;q11.2); *BCR-ABL1*

B-cell lymphoblastic leukemia/lymphoma with t(v;11q23); *MLL* rearranged

B-cell lymphoblastic leukemia/lymphoma with t(12;21)(p13;q22);

TEL-AML1 (ETV6-RUNX1)

B-cell lymphoblastic leukemia/lymphoma with hyperploidy

B-cell lymphoblastic leukemia/lymphoma with hypodiploidy (hypodiploid ALL)

B-cell lymphoblastic leukemia/lymphoma with t(5;14)(q31;q32); *IL3-IGH*

B-cell lymphoblastic leukemia/lymphoma with t(1;19)(q23;p13.3);

E2A-PBX1 (TCF3-PBX1)

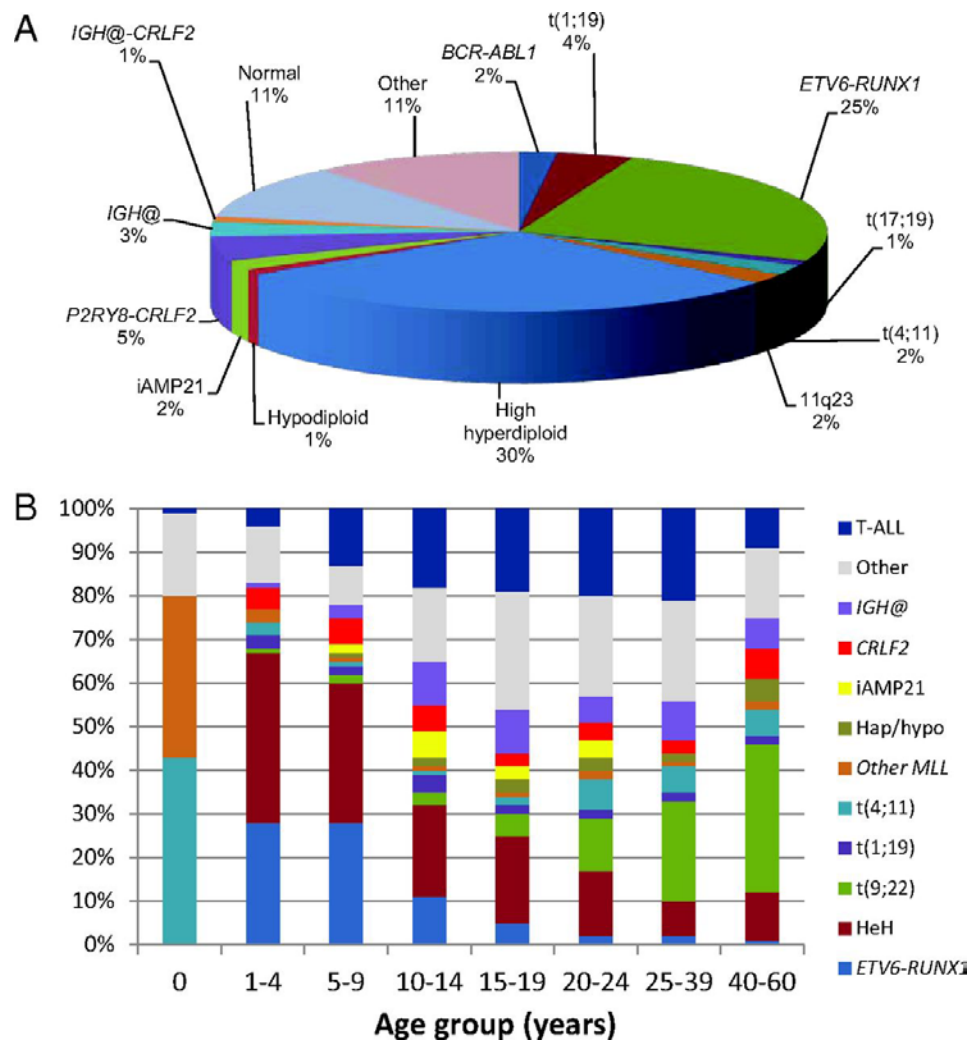
T-cell lymphoblastic leukemia/lymphoma

WHO = World Health Organization

Swerdlow SH, Campo E, Harris NL, et al (eds): WHO classification of tumours of haematopoietic and lymphoid tissues. Lyon, France: IARC Press; 109-138, 2009.

Pediatric ALL

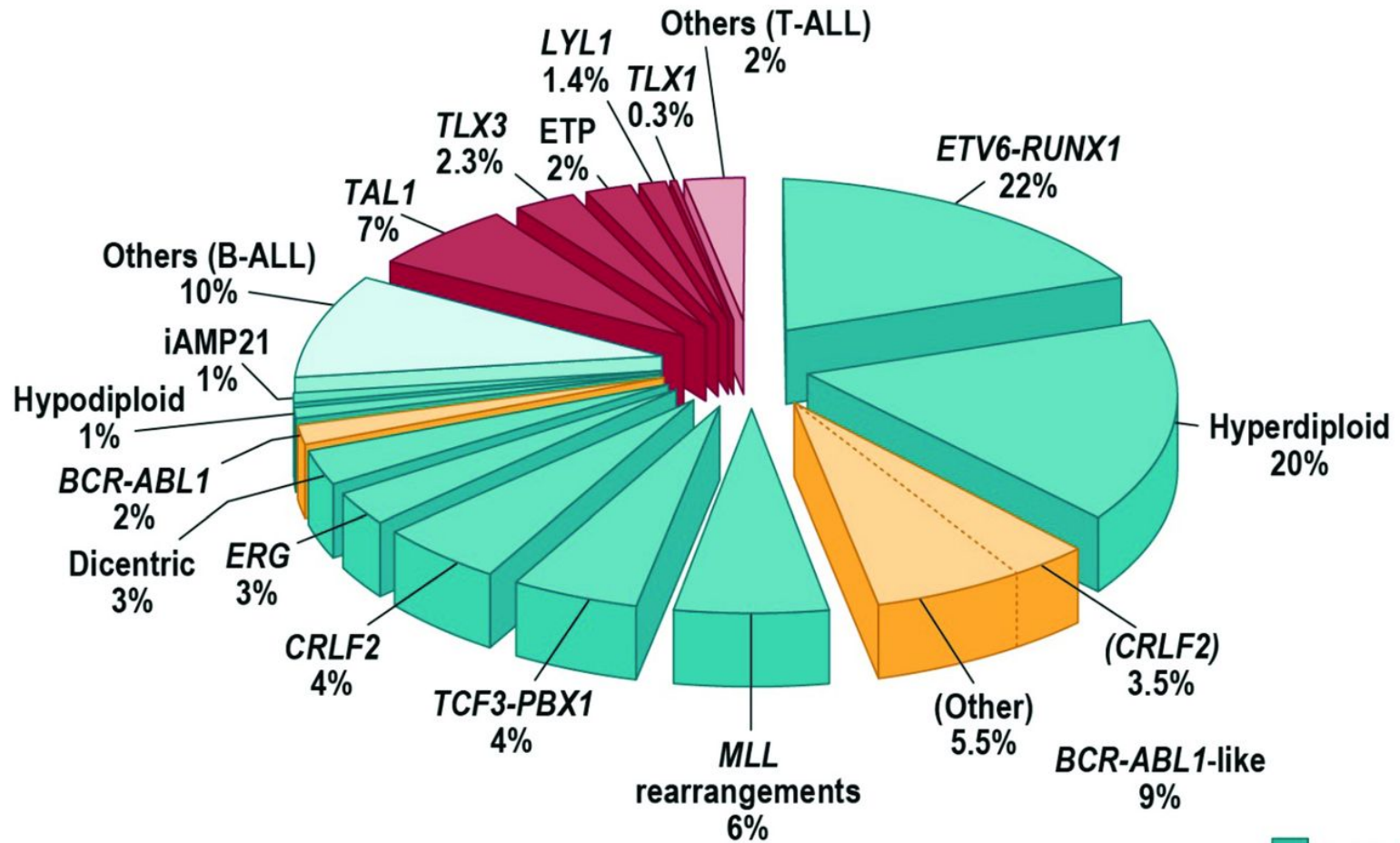
cca 30% of all pediatric tumors



Christine J. Harrison Hematology 2013;2013:118-125

Distribution of cytogenetic abnormalities from data collected from UK childhood ALL treatment trials.

Frequency of cytogenetic subtypes of pediatric ALL

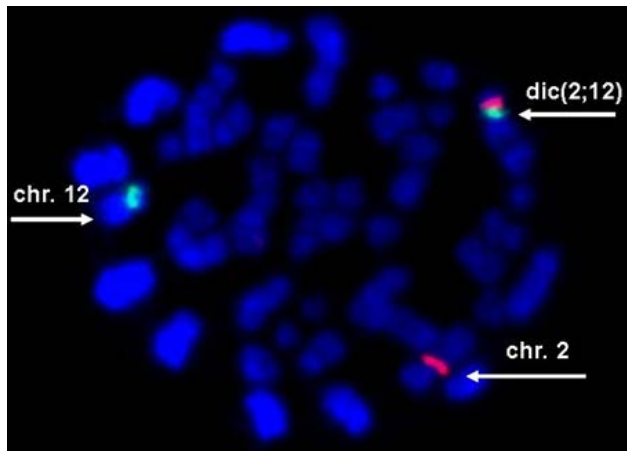
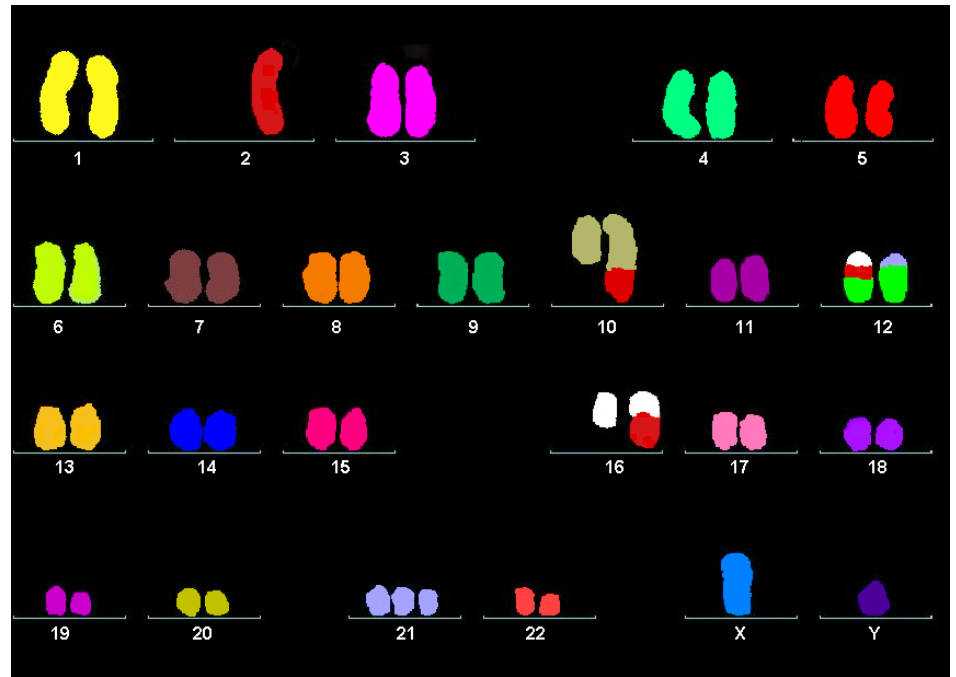
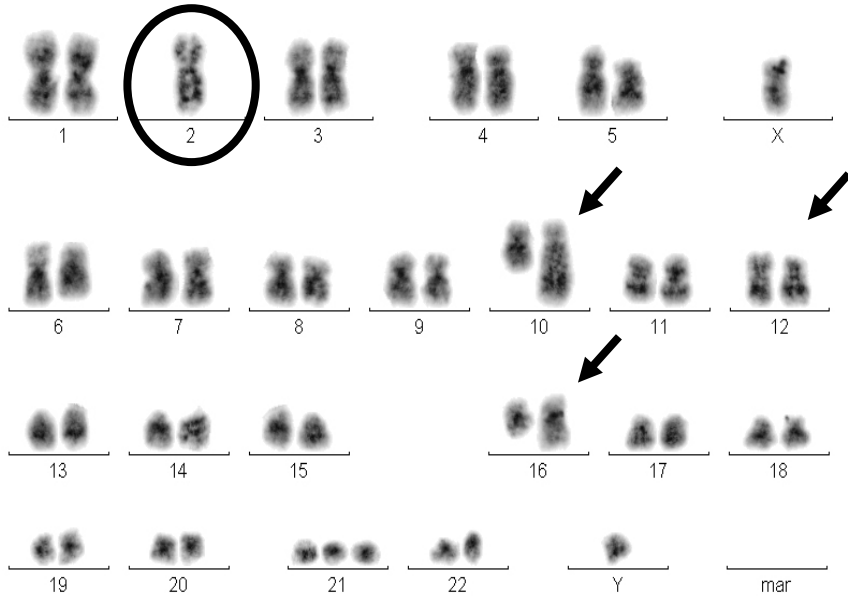


Charles G. Mullighan Hematology 2012;2012:389-396



Boy b 1998, dg. BCP-ALL 2003

46,XY,dic(2;12)(?;p?12)t(2;16)(?;q?),der(10)t(2;10)(q?12;q?12),t(12;21)(p13;q22),+21



WHO classification

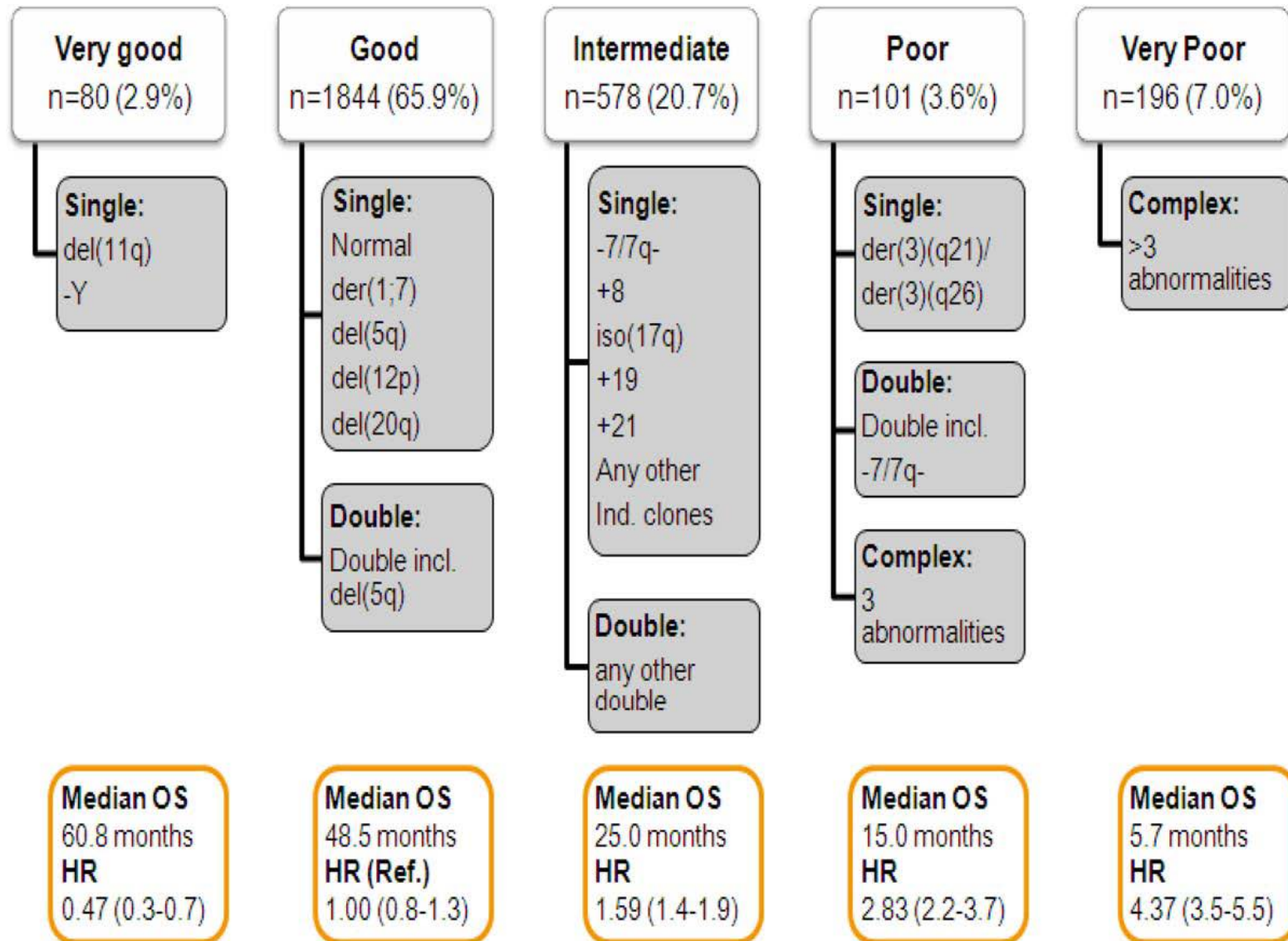
- Refractory cytopenia with unilineage dysplasia (RCUD)
- Refractory anemia with ringed sideroblasts (RARS)
- Refractory cytopenia with multilineage dysplasia (RCMD)
- Refractory anemia with excess blasts-1 (RAEB-1)
- Refractory anemia with excess blasts-2 (RAEB-2)
- Myelodysplastic syndrome, unclassified (MDS-U)
- Myelodysplastic syndrome associated with isolated del(5q)

Clinical heterogeneity is mirrored in heterogeneity of acquired genetic changes

Chromosomal changes in MDS

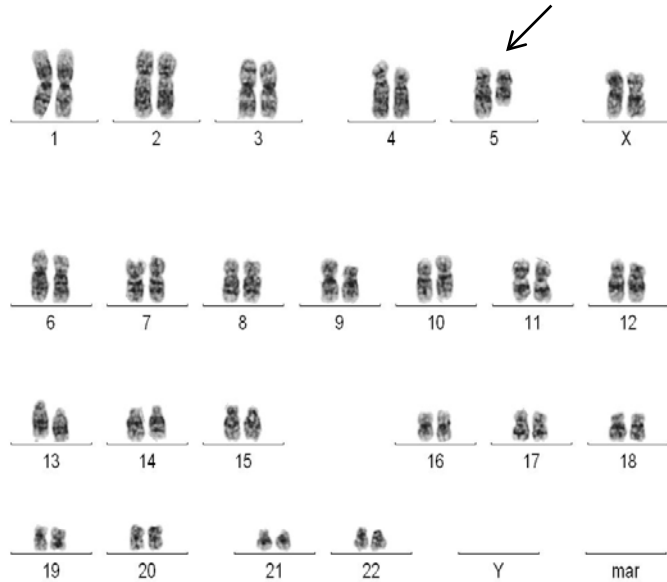
- de novo MDS 40-60%
- t-MDS or secondary MDS 90%
- SNPs+arrayCGH 70%

Prognostic stratification of MDS



5q- SYNDROME

46,XX,del(5)(q31)

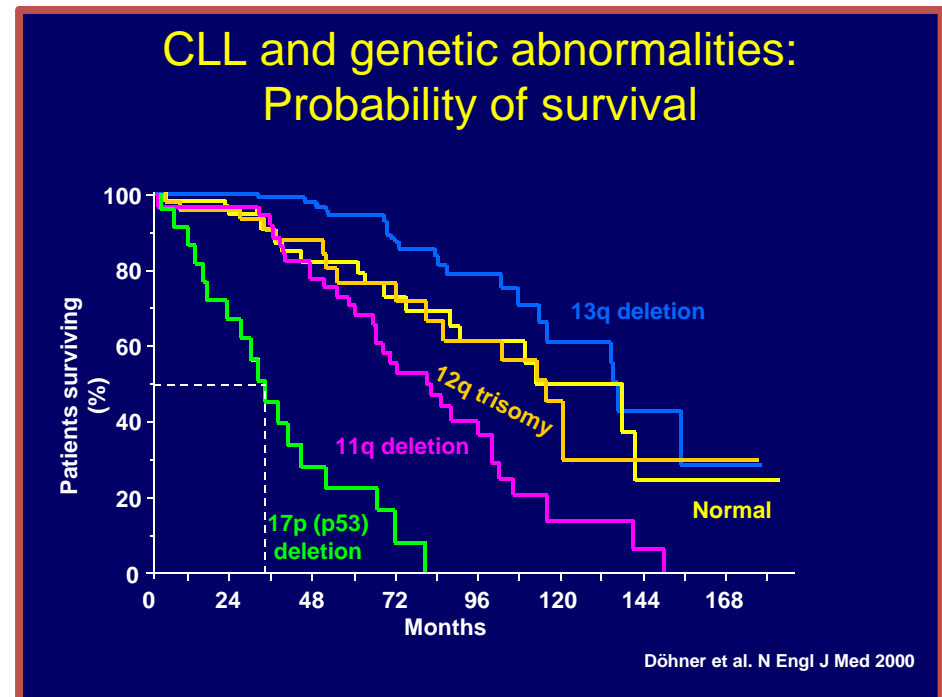


- 10% of patients
- good prognosis
- 5-16 % progress into AML

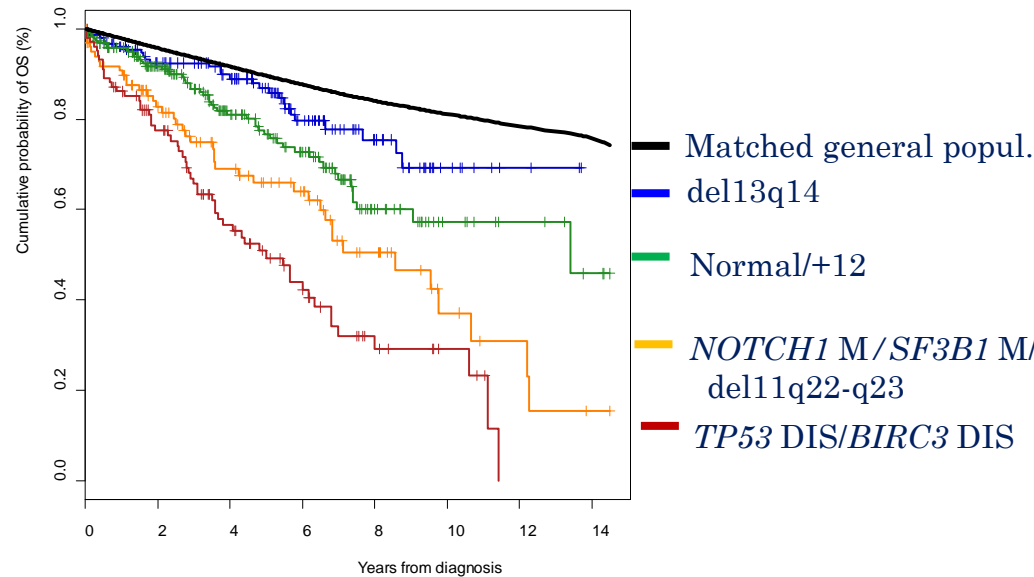
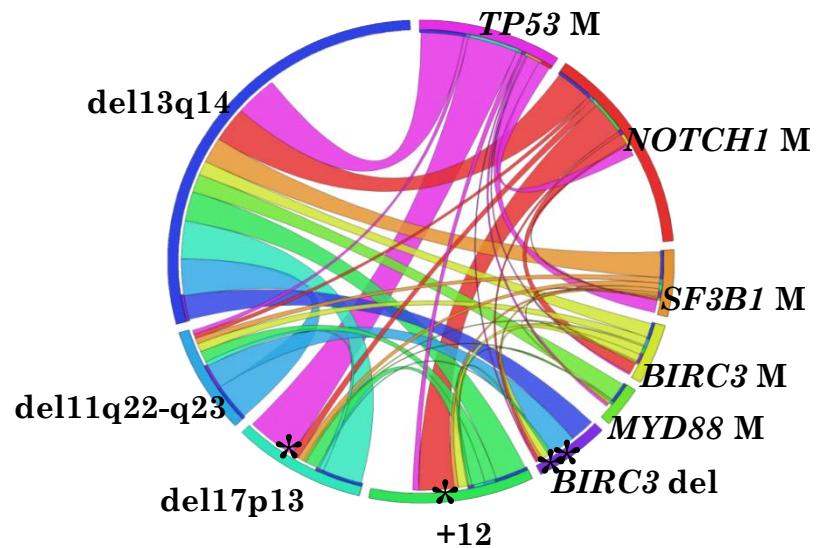
Prognostic significance of cytogenetics in CLL

Döhner H, Stilgenbauer S, Benner A, Leupolt E, Krober A, Bullinger L, Dohner K, Bentz M, Lichter P: Genomic aberrations and survival in chronic lymphocytic leukemia.

N Engl J Med 2000; 343:1910-1916.



Mutational and cytogenetic model of CLL



CLL – prognostic and treatment stratification

Category	Associated genetic factors	Therapeutic strategies
Very high risk	del(17p) [*] / <i>TP53</i> mutation and/or <i>BIRC3</i> mutation	p53-independent drugs, BTK inhibitors, allogeneic stem cell transplantation
High risk	del(11q) [*] / <i>ATM</i> mutation and/or <i>NOTCH1</i> mutation and/or <i>SF3B1</i> mutation	FCR
Intermediate risk	Trisomy 12 Normal karyotype and FISH	Not recommended
Low risk	Isolated del(13q) [*]	Not recommended

NON HODGKIN LYMPHOMA

- Heterogenous group of tumors of the lymphatic tissues
- Arise from genetic changes in originally normal cells
- Classification – histopathology WHO 2008
- Cytogenetics and FISH used for classification

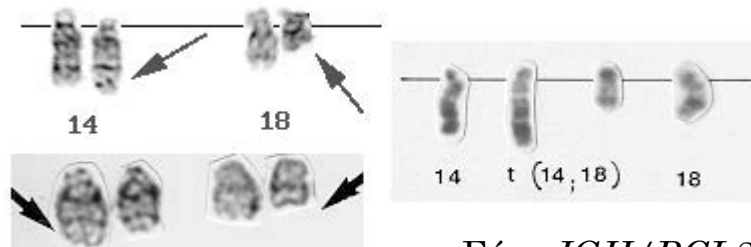
Follicular lymphoma (FL)

indolent B cell lymphoma

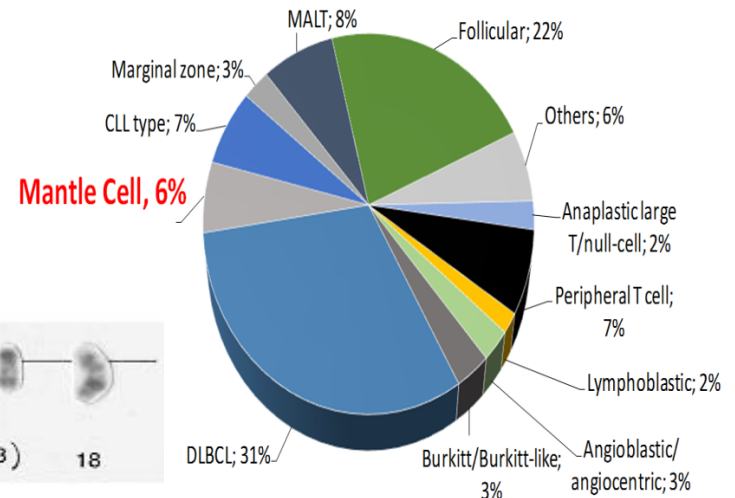
~20% of all lymphomas

Clinically heterogenous, OS up to 20 years

90% of patients $t(14;18)(q32;q21)$

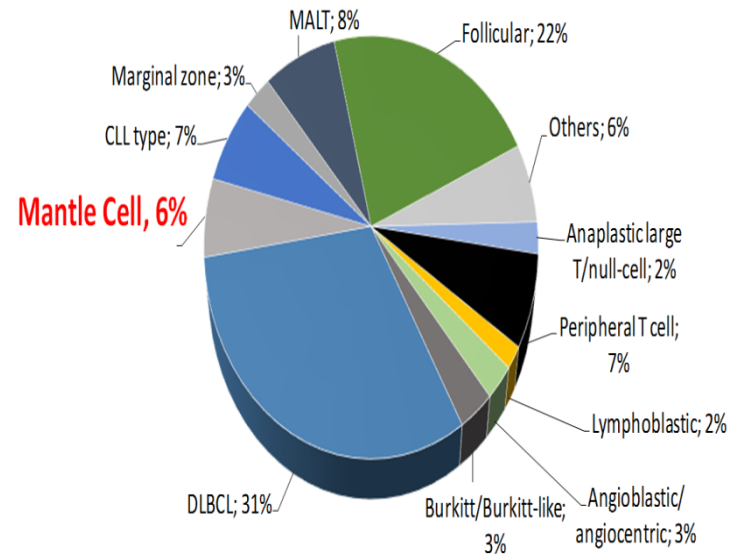


Fúze $IGH/BCL2$



MANTLE CELL LYMPHOMA

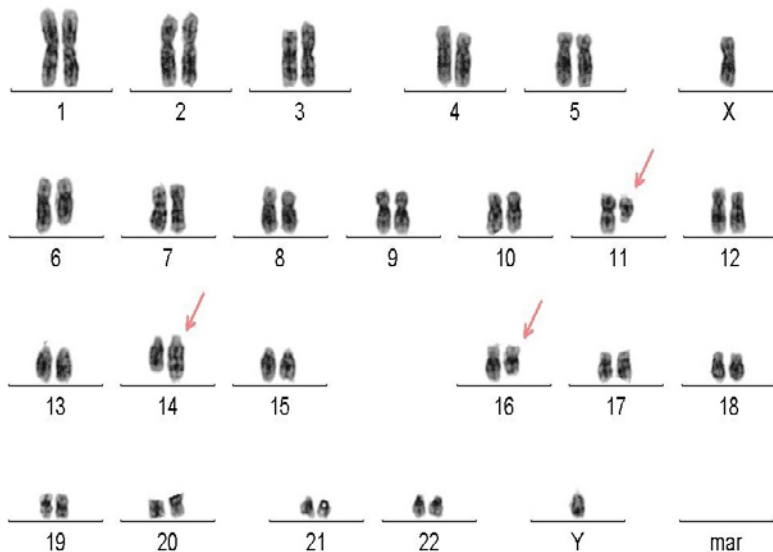
- Aggressive disease (OS 3-5 years)
- ~ 6 % všech NHL
- Diagnostics:
 - Morphology
 - Immunohistochemistry
 - Immunophenotyping
 - Genetics:
 - cytogenetics
 - FISH
 - Molecular genetics - PCR



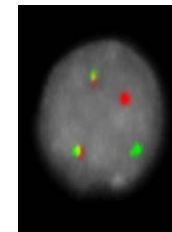
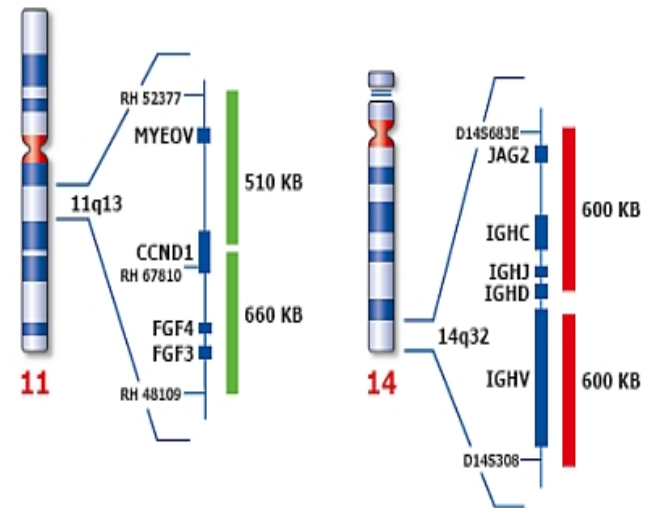
MANTLE CELL LYMPHOMA

- Conventional cytogenetics

t(11;14)



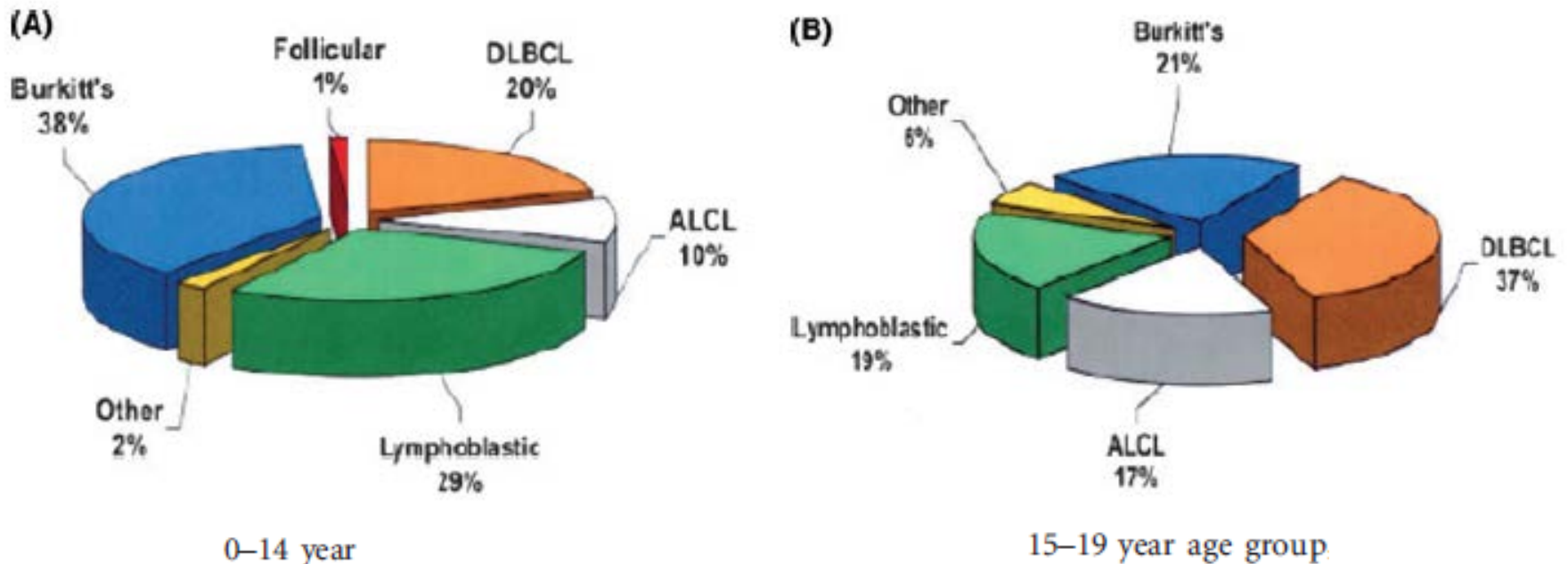
- FISH
dual color dual fusion probes



IgH/CCND1 DC
DF Kreatech

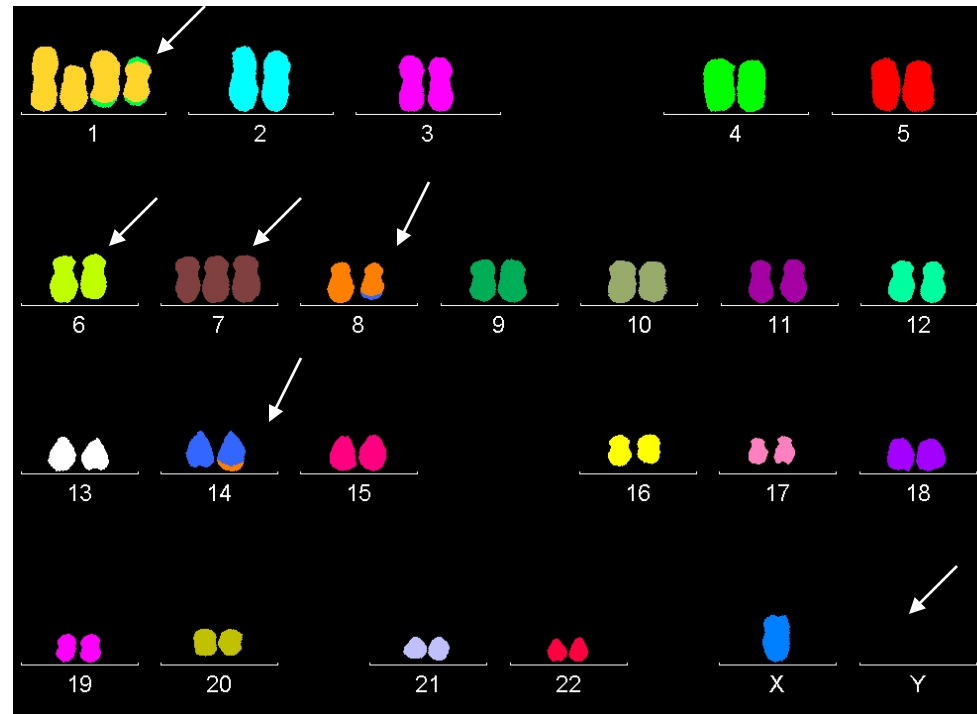
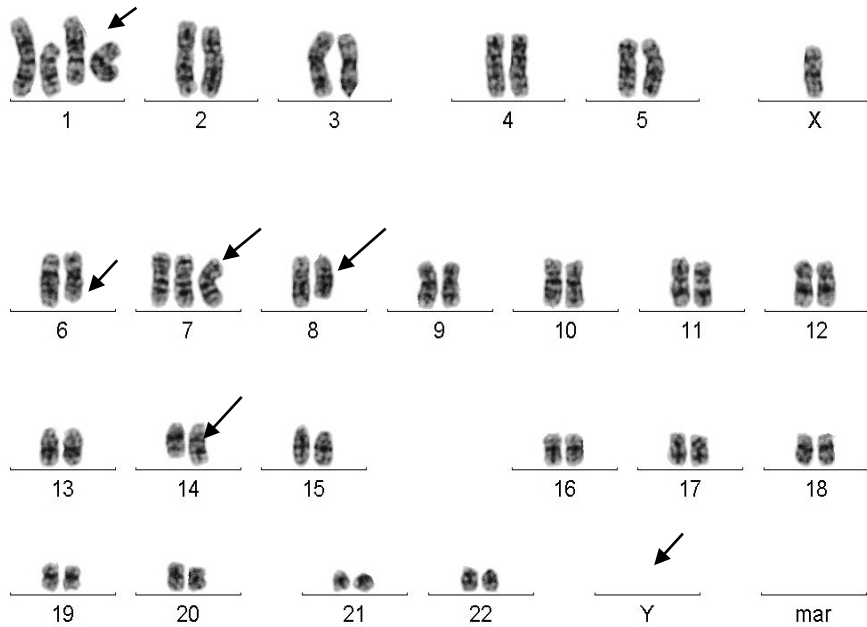
NON-HODGKIN LYMPHOMAS IN CHILDREN

- 4-7% of tumors in children and young adults
- Incidence increases with changes
- WHO classification 2008
- frequency of histological subtypes different from adults



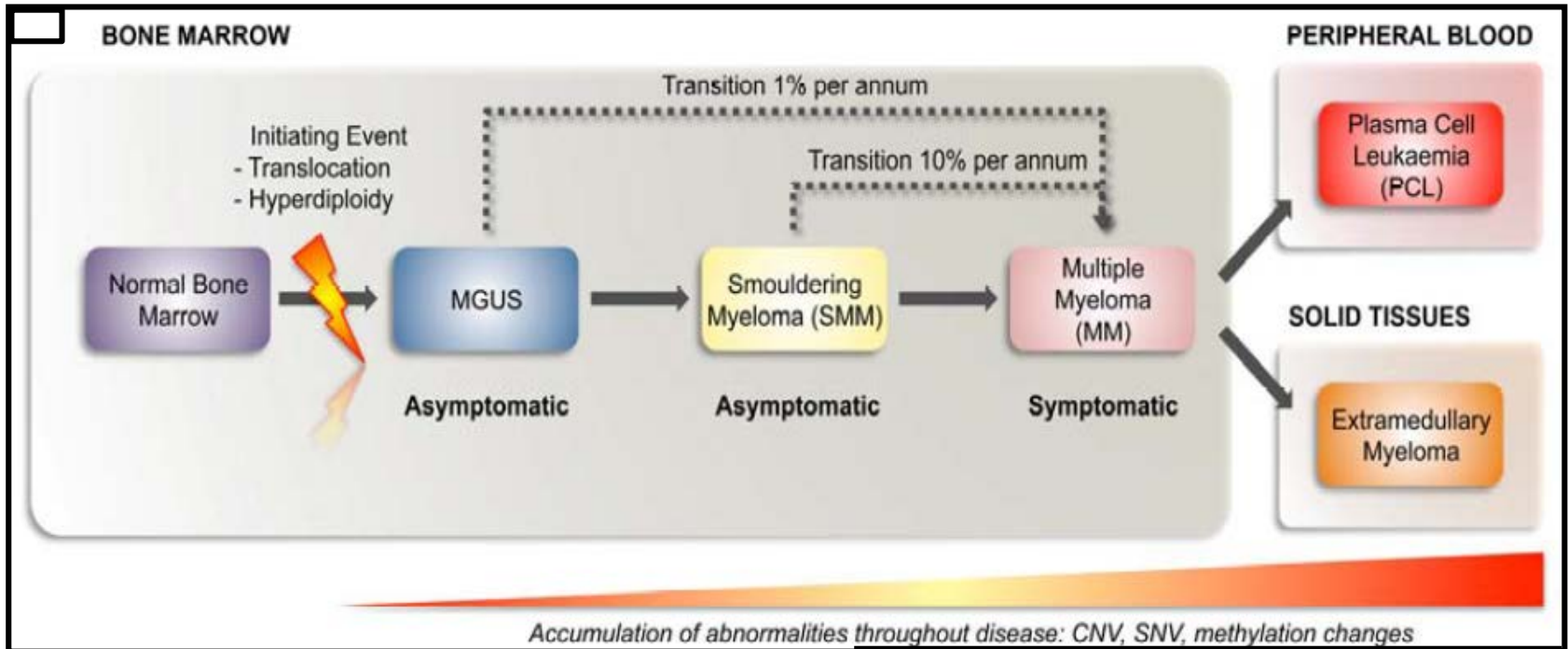
BURKITT LYMPHOMA

48,X,-Y,del(1)(p13pter),+der(1)del(1)(q?24q?ter)t(1;4)(q23;?q?),
+ider(1)(q11)del(1)(q?24q?ter)t(1;4)(q23;?q?),del(6)(q?15),+7,t(8;14)(q24;q32)(1.klon-56%)



11/2011

MULTIPLE MYELOMA



Walker B, Wardell CP, Melchor L et al., *Submitted*



SUMMARY - CYTOGENETICS

- Diagnostics and prognostic stratifications of hematologic malignancies
- Can analyze the entire genome in one run
- Allows for clarification of diagnosis by specific chromosomal aberration
- Recurrent nonrandom changes determine prognosis of disease
- Aberration classification allows monitoring of treatment efficacy

Thanks for your attention