

Phytochemistry

Backgrounds of Separation Methods

CHROMATOGRAPHY

Principles of Chromatography

- Distribution of compounds between two phases:
 - Mobile phase – motile, moving
 - Stationary phase – stable, not-moving
 - Separation based on the way, which distributes the compounds between phases.
 - Distribution coefficient: $D = \frac{c_{stat. phases}}{c_{mob. phases}}$
 - Compounds in constant and dynamic balance between both phases.
 - For a target compound is valid: a position in system is generated by an interaction with stationary phase and a competition of interaction with mobile phase.

- **Stationary phases:**

- Solid
- Liquid

- **Mobile phases:**

- Liquid
- Gassy

Classification of chromatographic methods

- **According to the alignment:**

- Column chromatography – scale from capillary analytical methods to large preparative columns
- Planar chromatography – different forms of TLC

- **According to the modus of separation:**

- Adsorption
- Partitioning
- According to the charge:
 - Ion exchange
 - Ion pair
- Based on size exclusion
- Bio-affinity

- Dynamic balance of compounds distribution in stationary and mobile phase – different molecules spend different time both in mobile and stationary phase.

- When compounds distributed in mobile phase only, they move in the same speed and time.

- Stationary phase makes the retention.
 - The rate of retention could be described by:
 - Retention time.
 - Retention volume.
 - Capacity factor.

- Retention time t_R :
 - Time between injection of sample into column and detected elution of compound.
 - Depends directly on flow rate of mobile phase in column.
 - Sum of time which compound spends on stationary phase and time spent in mobile phase t_M
 - Corrected retention time: $t'_R = t_R - t_M$
- Retention volume: $V_R = F \cdot t_R$
F... flow rate
- Capacity factor k' : $k' = \frac{t_R - t_0}{t_0}$
t₀... retention time of non-retained compound
- Retention time of bounded compound: $t_R = L/v \times (1 + k')$

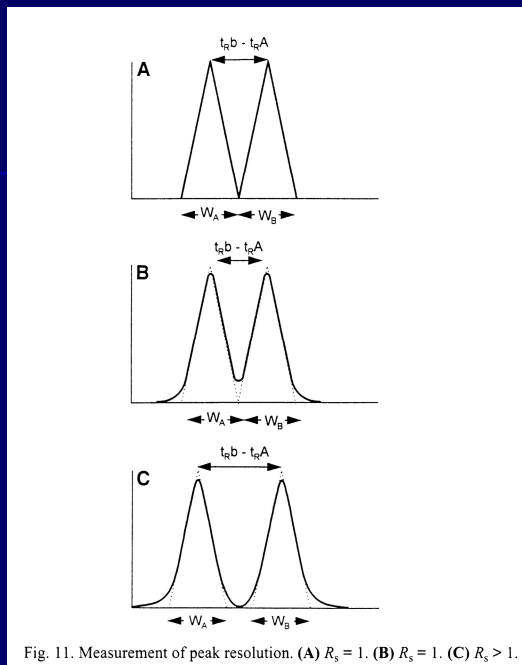
- Column Efficiency:
 - Repeated formation of balanced state.
 - Not in steps, but dynamic process.
 - Number of theoretical plates of column N :
 - How many times the balance is established.
 - Higher number \longrightarrow better separation
 - Could be used for description of separation efficiency.
 - Can describe how broad will be zones of separated compounds.
 - Depends on the column length. Longer column \longrightarrow better separation.

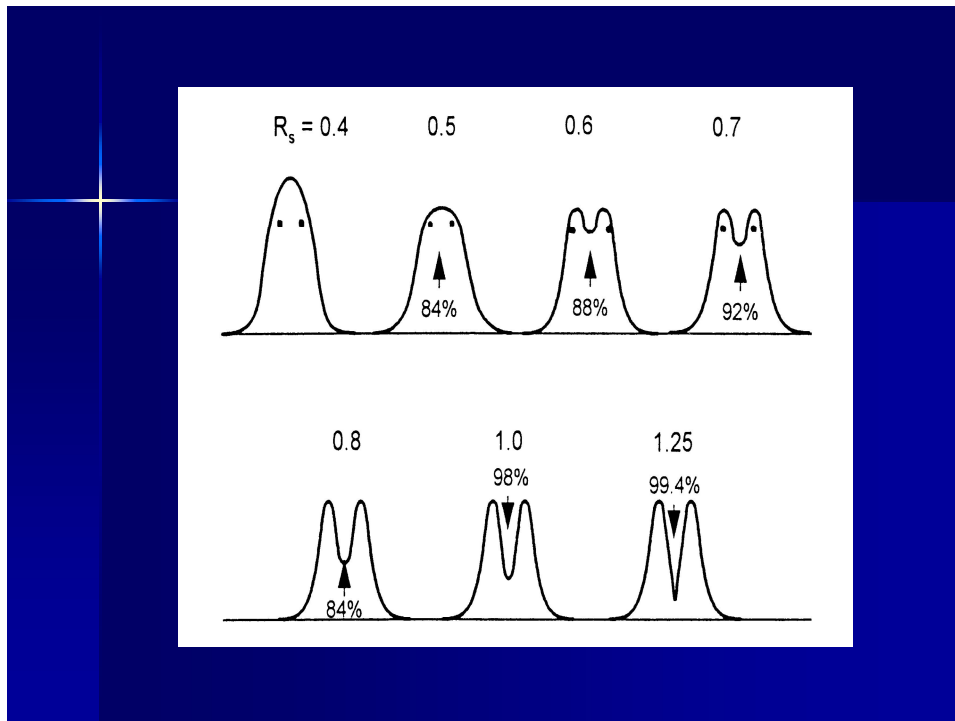
■ Separation and selectivity α :

- Different compounds → different distribution coefficients. $\alpha = K_B / K_A = k'_B / k'_A = t_{RB} / t_{RA}$
- Compounds retained selectively → different retention times.

■ Resolution R_s :

- Two parameters for calculation:
 - Distance of peak apexes.
 - Width of peaks at base.
- $$R_s = \frac{t_{RB} - t_{RA}}{1/2 \times (w_A + w_B)}$$





How to read a chromatogram to get an information.

1. Identification (quality)

- Retention time- each compound displays the characteristic retention time under stable and defined conditions.
- The same retention time t_R for two samples \rightarrow probably the same compound.
- Method of simultaneous injection.
- Verification with help of detection method.

2. Physico-chemical character of compound

- Non-polar compounds \rightarrow slow elution on reversed phase
 \rightarrow fast elution on normal phase
- Polar compounds \rightarrow fast elution on reversed phase
 \rightarrow slow elution on normal phase

3. Amount (quantification)

- For target compound is the AUC (Area Under Curve) direct proportional to amount of compound.
- Standard with known concentration \rightarrow calibration curve.
- Estimation of quantity according to the peak height.

■ **Quantitative analysis**

■ **Method of internal normalization**

- Basic condition – all components of sample must be eluted.
- Detector must give linear response depending on concentration of all analyzed compounds
- Percentual composition of mixture is derived from AUC of all peaks.
- Method is simple. Problems when detector response different or not linear for all analyzed compounds.

■ **Method of absolute calibration**

- Based on injection of known amounts of sample and standard under the same experimental conditions.
- Content is evaluated from calibration curve or from direct comparison of AUC of standard and sample peaks.
- At least two injections on column (sample and standard).

■ **Quantitative analysis**

■ **Method of internal standard**

- The mixture of different weight ratio is prepared from standard and pure analyte, the analysis is then carried out.
- Calibration curve is based on ration $AUC(\text{sample})/AUC(\text{standard})$ and $\text{weight}(\text{sample})/\text{weight}(\text{standard})$
- When a sample with unknown concentration is analyzed, this sample is added to standard and from the ration of AUC is possible to calculate the amount of target compound.
- Advantage – it is not necessary to know the exact amount of injected sample.
- Disadvantage – difficult to find good standard.

■ **Method of standard addition**

- Based on addition of defined amount of analyzed compound to a sample.
- Two analyzes: 1. without addition, 2. with addition.
- Increase of AUC of compound peak is directly proportional to an amount of added analyte.

How to improve a chromatographic separation?

- Capacity factor
- Theoretical plate number
- Selectivity

Capacity factor k'

- $k' = 0$ compound not retained
- $k' > 20$ t_R not effective (too long)

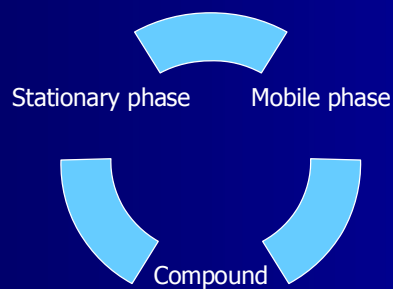
How to affect:

1. Good choice of mobile phase.
2. Using of gradient elution.

Efficacy

- Increase of N affects the peak width and quality of resolution, but did not affect the principle of separation. Relative t_R stays unchanged.
- Affect: stationary phase particle size.

Selectivity 3 groups of molecular interactions

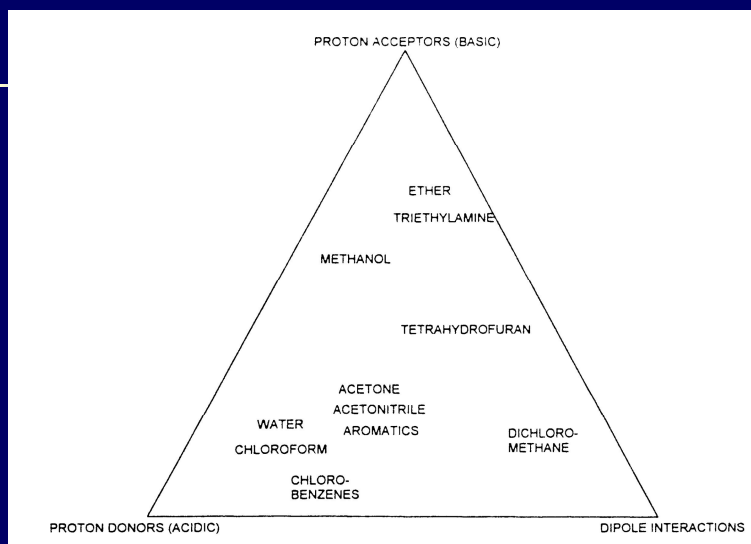


Eluotropic series

Eluotropic Series of Solvents

Solvent	$E^0(\text{Al}_2\text{O}_3)$	Boiling pt., °C	Viscosity, $\text{mN}\cdot\text{s}\cdot\text{m}^{-2}(20^\circ\text{C})$	UV Cutoff, nm
Pentane	0	36	0.24	210
Cyclohexane	0.04	69	0.98	210
CCl_4	0.18	77	0.97	265
Toluene	0.29	111	0.59	286
Diethyl ether	0.38	35	0.25	218
Chloroform	0.40	62	0.57	245
Dichloromethane	0.42	40	0.44	235
Tetrahydrofuran	0.45	66	0.55	220
2-Butanone	0.51	80	0.32	330
Acetone	0.56	56	0.32	330
1,4-Dioxane	0.56	107	1.44	215
Ethyl acetate	0.58	77	0.45	255
Diethylamine	0.63	115	0.33	275
Acetonitrile	0.65	82	0.37	190
2-Propanol	0.82	82	2.50	210
Ethanol	0.88	78	1.20	210
Methanol	0.95	64	0.59	210
Water	1.00	100	1.0	—

Snyder triangle



Liquid chromatography

- The most universal and most often used method in natural compounds separation

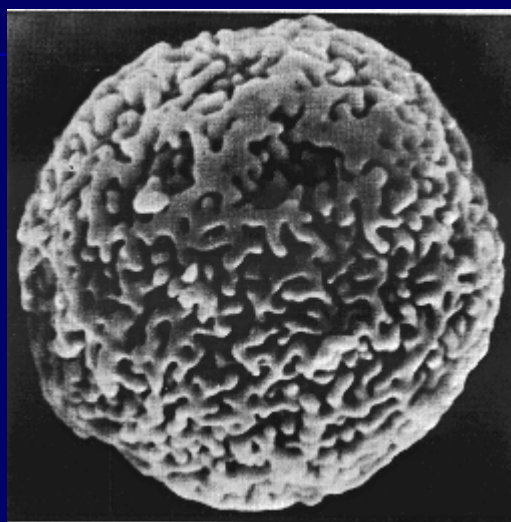
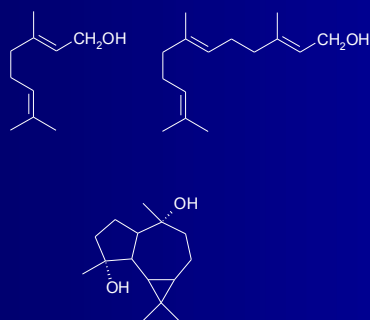
Mechanisms of Separation	Stationary phase example
Adsorption	Silica, aluminium oxid, polyamides
Rozdělování	RP- materials (C2, C8, C18), cellulose
Ion pair	RP- materials (C18)
Ion exchange	Ionex, catex
Complexation	catex with Ag ⁺
Chiral separation	Chiral phases or mobile phases with chiral modifier
Gel filtration, size exclusion	Gels, Sephadex LH-20

Adsorption liquid chromatography

- Interaction of analytes with stationary phase is based on adsorption
- Distribution constant based on series of parameters (volume of monomolecular layer of solvent, activity of adsorbent, elution power of solvent)
- The interaction should be physical, not chemical.
- The separation should be carried out in linear part of adsorption isotherm. In non-linear part of isotherm we will get „tailing“ peaks.
- Most common adsorbents used:
 - Silikagel or aluminium oxid (alumina)
 - Carbon organic adsorbents
- Most common mobile phases:
 - Non-polar carbohydrate + polar organic modifier

■ Silikagel

- Gel of silic acid
- Adsorption chromatography
- Normal phase
- Polar characteristics – presence of –OH functional groups.
- Silikagel is weakly acidic (used at pH = 2 to 8)
- Under normal circumstances – separation of compounds which differ in polar functionalities. Differences in non-polar parts – no efficient separation.



Aluminium oxid

- Aluminium oxid suitable for separation of basic compounds
 - Acidic
 - Neutral
 - Basic

Chromatography on bonded phases

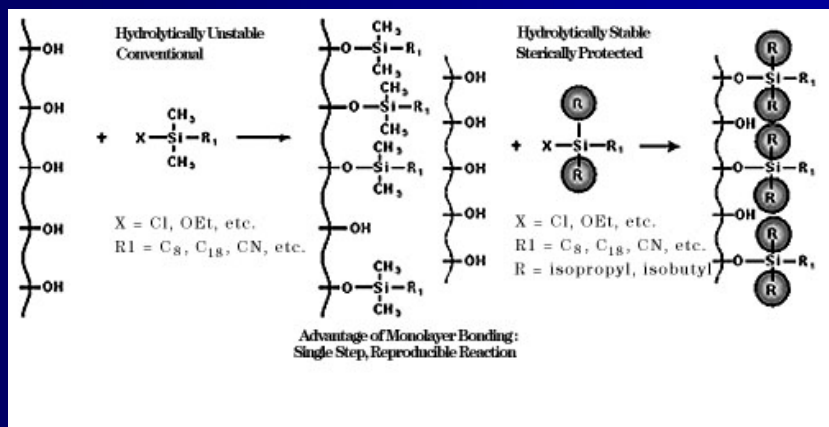
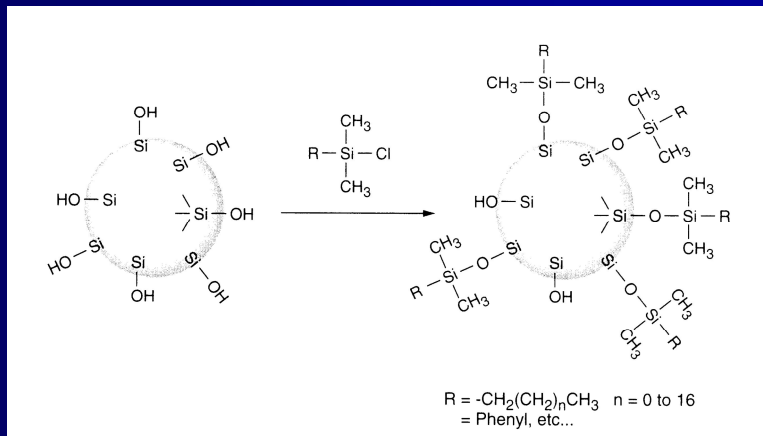
- Different functional groups are bonded on carrier (usually silikagel)
- The most common – chemically bonded phases of siloxane type (prepared by reaction of silikagel with chlorsilanes)
- Disadvantage of this sorbent type is limited pH range (2 to 8, different pH can cause the „bleeding“ of stationary phase)
- Typical example of chemically bonded phase: C8, C18, CN-, phenyl, chiral compound
- Non-reacted (free) –OH groups of silikagel should be deactivated
- According to the bonded functional group can be separated different compounds
 - For example: bonded chiral phases can separate optical izomers
- Exact description of separation process is difficult
 - Process is basic on solvophobic (hydrophobic) effect
 - Important is dissolution of compound in mobile and stationary phase and competition

Chromatography on bonded phases

- Stationary phases used are of non-polar character
- Mobile phase : water with addition of polar organic solvents (alcohols, acetonitril, dioxan, tetrahydrofuran, acetone)
- Selectivity is strongly affected by mobile phase composition
- Elution power of mobile phase rises with decreasing polarity
- Chromatography on reversed phase is suitable of separation of homologous compounds
- RP HPLC rules modern separation
- Nomenclature "reversed phases" is used from historical reasons

■ Modified silica

- Silikagel + carbohydrate side-chain
- C2, C8, C18
- Distribution chromatography
- Non-polar character
- Reversed phase
- Changes of non-polar part of analyzed molecule will affect separation.
- Limited pH range.



Chromatography of ion pairs

- Often used for ion compounds, combination with chemically bonded phases (C18)
- Mobile phase contains ion pair reagent with opposite charge, than is the charge of analyzed compound
 - For anion analysis tetraalkylammonium salts are often used
 - For cation analysis dodecyl sulphate is often used
- Incurred ion pair displays properties of polar organic molecule and pass from mobile phase to stationary phase.

Ion exchange chromatography

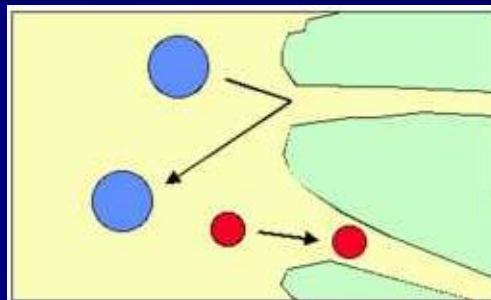
- Technique suitable for separation of ionic nature based on electrostatic interaction
- Analyte during the separation migrates in the direction of ion exchanger, than diffuses through the exchanger where ion exchange takes place.
- Proper choice of ion exchange can affect parameters of separation.
 - Different types of catexes and anexes
 - Parameters of separation are based on size of molecules, size of and polarity of charge, on ionic power of mobile phase, degree of solvation...
- Affinity of compound separated rises with increasing charge of ionex
- When parameters of charge and size of different molecules separated are the same, superiority is given to compound in higher concentration

Size exclusion - gel permeation chromatography

- Separation based on size and shape of compound particles
- Mobile phase just carry the separated compounds through the column, it does not participate on separation process directly.
- Huge molecules (macromolecules) do not permeate into pores of gel and elute from the column as first
- Small molecules permeate into pores of gel, their movement through the column slows.
- Exclusion limit = size of particles which will not permeate into pores and therefore will not be separated.
 - Can be used for determination of molecular weight
- Material
 - agarose and dextran gels
 - organic polymers (co-polymer styrene + divinylbenzene)
 - Use of porous, rigid glasses and silicagel
- Mobile phase must dissolve analytes, but should not react with analytes, must wet surface of stationary phase and must be compatible with a mode of detection.
- Often used: water and water buffers, isopropanol.
- **Analyses of macromolecules, proteins, peptides, allergens**

■ Sephadex LH-20

- Hydroxypropyl-dextrane gel
 - According to conditions:
 - Adsorption
 - Partitioning
 - Gel filtration
 - Combination

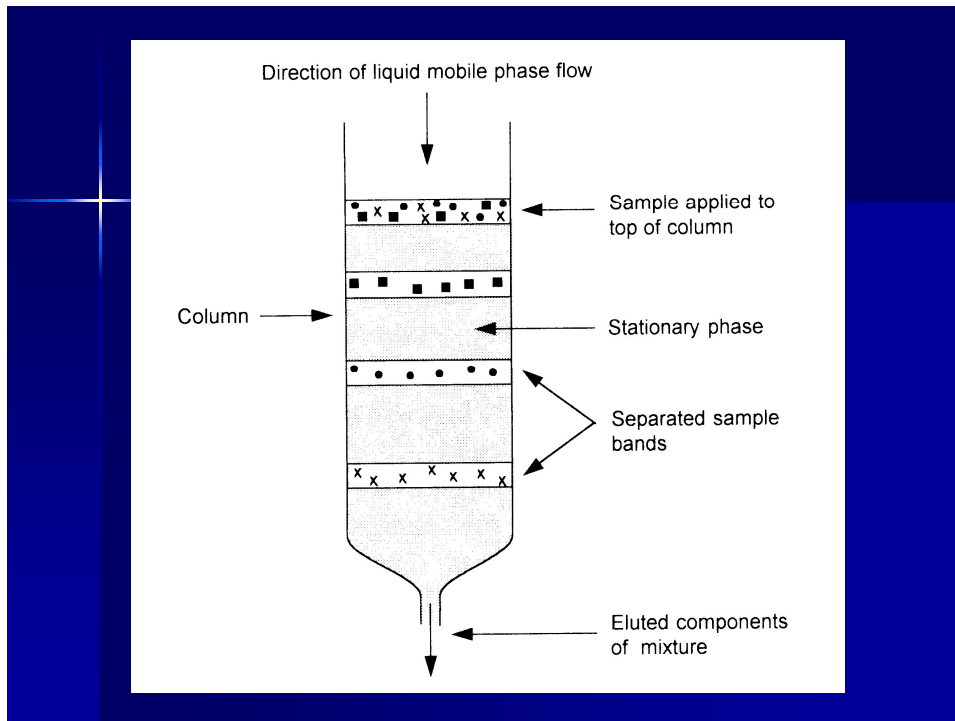


Bio-affinity chromatography

- It is based on specific interaction between biological active compounds and their „anti-compounds“
- Examples of such as specific interaction:
 - *Antibody- Antigen*
 - *enzyme – substrate*
 - *lecithine – glycoprotein*
- Bio-affinity chromatography is used for separation, isolation, and purification of sample
- Stationary phase must be first at first bonded on suitable inert carrier.
- Ligand (stationary phase) must have high affinity for determined compound
- Bond of ligand and analyte can be disturbed by change of experimental conditions or using a deforming buffer.

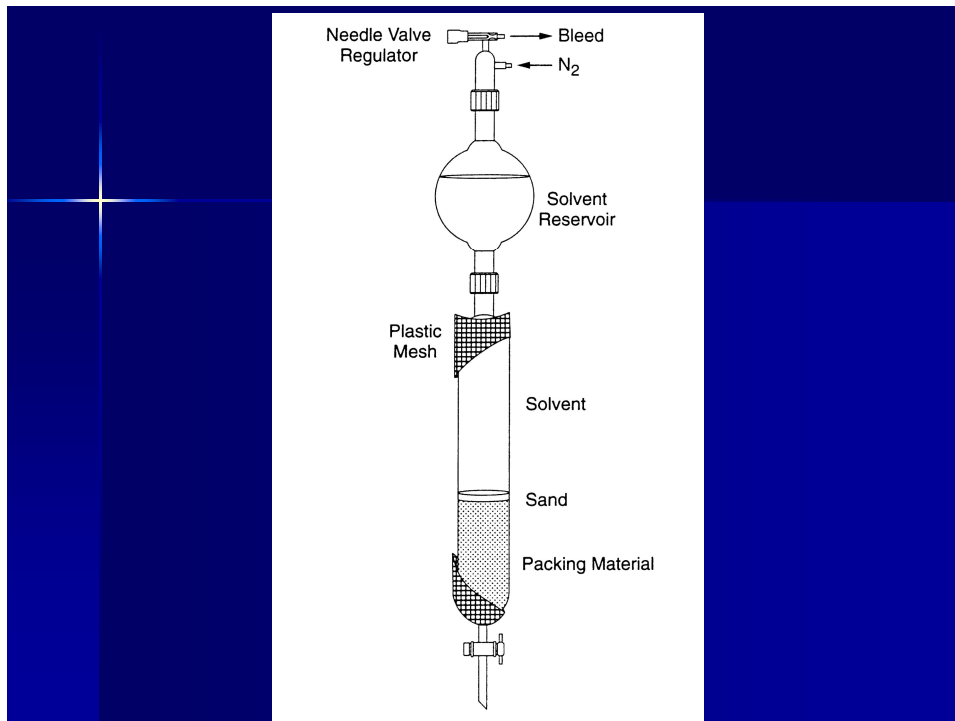
Types of liquid chromatography

- **Classical column chromatography**
 - Oldest, simplest.
 - Commonly normal phase (silikagel, aluminiumoxid).
 - Large particle size (60-200 µm)
 - First step of separation.
 - Big sample amounts.
 - Sample application:
 - Liquid sample (good solubility in mobile phase)
 - Solid sample (bad solubility, adsorption on silikagel 1:1)



■ Flash chromatography

- The same principles as classical column.
- Overpressure upon the mobile phase (air, N₂).
- Higher pressure → higher velocity of mobile phase → lower sorbent particle size 40-60 μm.
- Faster and/or better separation



- Low-pressure and medium pressure chromatography (LPLC a MPLC)

- LPLC

- Hybrid of flash chromatography and HPLC.
- Mobile phase is pressurized to 1-6 atm.
- Simple pump.
- Particle size similar to flash chromatography.
- Different column size, different types of sorbents.

- MPLC

- Sophisticated apparatus.
- Smaller particles 10-40 μm .
- Pressure 3-50 atm.