Lab class no. 4 – Evaluation of disc diffusion test and MIC, determination of MBC, determination of concentration of bacteria by cultivation

(in pairs)

Aims:

- Evaluation of disc diffusion test
- Evaluation of Escherichia coli sensitivity to chloramphenicol (MIC)
- Determination of minimal bactericidal concentration of chloramphenicol (MBC)
- Determination of concentration of Escherichia coli by cultivation

Material

Paper ruler, transluminator, Nutrient broth – solid in Petri dish and liquid, bacteriological loop, microbiological tubes, tube with calibration, vortex, automatic pipette

Working procedure

1. Evaluation of disc diffusion test

• *E. coli* growth was inhibited by chloramphenicol and around discs there are inhibition zones - measure their diameter using paper ruler and evaluate sensitivity or resistance according to EUCAST - write down to protocol no. 3.

2. Evaluation of Escherichia coli sensitivity to chloramphenicol (MIC)

- MIC is lowest concentration of antibiotics still able to inhibit growth of bacteria
- evaluate MIC of chloramphenicol from turbidity in wells
- compare with EUCAST and wrote to protocol no. 3, if the bacteria are sensitive or resistant

3. Determination of minimal bactericidal concentration of chloramphenicol (MBC)

- Petri dish with Nutrient broth agar divide into quadrants (on the bottom side) number of quadrants correspond to number of wells in microtitration plates without turbidity
- using bacteriological loop inoculate all 4 wells of corresponding concentration into the quadrant
- in the same manner inoculate the other concentrations without turbidity (e.g. 8 ug/ml, 16 ug/ml, 32 ug/ml) - always with new loop!

4. Determination of microbiological quality of a medicinal product (determination of number of microbes)

- in pairs determine concentration of bacteria in liquid sample of *Micrococcus luteus*
- prepare set of 5 microtubes, add to each 900 μl of Nutrient broth
- mix the sample of bacteria by vortex and take out 100 μ l of sample to the first tube; mix well, take out 100 μ l again and transfer to second tube and so on
- mix all 5 tubes again, take out 100 μl from each and transfer to agar plate using tip (divided in 5 segments on bottom by marker)

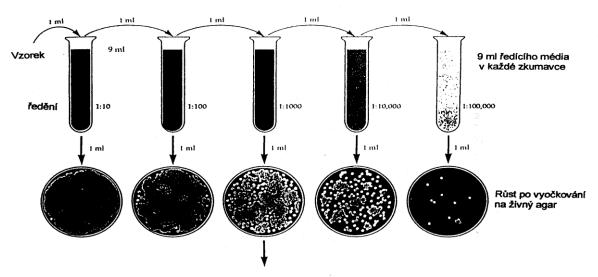
Cultivate of all plates at 37°C for 24 hours. Then the plates will be kept in fridge.

Protocol no. 4 will contain:

- definition of MIC and MBC; in last lab class you are going to fill in MBC value to your protocol - evaluate chloramphenicol as bacteriostatic or bactericidal
- in last lab class determine concentration of *E. coli* in unknown sample; draw the scheme of dilution into your protocol.

Miscellaneous agents	MIC breakpoint (mg/L)		Disk content (µg)	Zone diameter breakpoint (mm)		Notes Numbered notes relate to general comments and/or MIC breakpoints. Lettered notes relate to the disk diffusion method.
	Ss	R>		Sz	R<	
Chloramphenicol	8	- 8	30	17	17	1. Quality control of collistin must be performed with both a susceptible QC strain (E. coll ATCC 25922 or P. aeruginosa ATC
Colletin ¹	2	2		Note*	Note ^A	 Agar dilution is the reference method for fosformycin. MICs must be determined in the presence of glucose-6-phosphate (25 mg/L in the medium). Follow the manufacturers' instructions for commercial systems. Trimethoprim:sufamethoxazole in the ratio 1:19. Breakpoints are expressed as the trimethoprim concentration.
Daptomycin					- 5.	
Fosfomycin Iv	322	32 ²	200 ^B	24 ^{0.0}	240.0	
Fosfomycin oral (uncomplicated UTI only)	322	32 ²	200 ^B	24 ^{0,0}	24 ^{0,0}	
Fusidic acid	- 3	19		-	- 1	
Metronidazole	-	5.0		50 (- 7	
						C. Zone diameter breakpoints apply to E. coll only. For other Enterobacteriaceae, use an MiC method.
Nitrofurantoin (uncomplicated UTI only), E. coll	64	64	100	11	- 11	D. Ignore isolated colonies within the inhibition zone (see pictures below).
Nitroxoline (uncomplicated UTI only), E. coll	16	16	30	15	15	
Rifampicin		-		•		
Spectinomycin	-	- 9		- 9	- 8	
Trimethoprim (uncomplicated UTI only)	2	4	5	18	15	
Trimethoprim-sulfamethoxazole ¹	2	4	1.25-23.75	14	11	

Sériové ředění vzorku a přímé stanovení počtu živých bakterií (kolonie tvořících jednotek – KTJ; colony forming units – CFU)



Sériové ředění vzorku

Výpočet: počet kolonií na misce x ředění vzorku = KTJ/ml

