In vivo biomarkers of effects / response

Do we know the agent ? Do we expect the effect ? : specific biomarkers / non-specific changes

Behavioural and Clinical biomarkers Pathology Clinical chemistry and hematology Enzymatic changes Protein synthesis biomarkers Oxidative stress markers Behavioural and clinical biomarkers

Behavioural and clinical biomarkers

Parameters evaluated

- body weight
- food consumption
- fitness & welness

Interpretation

- : are these ? biomarkers ? (effects already demonstrated in vivo)
- biomarkers of existing serious stress / intoxication

Behavioural and clinical biomarkers

Table 7.4 Effect of some agricultural chemicals on behavioural parameters of the rainbow trout

Chemical	LD ₅₀ Swimming (96hr) capacity		Swimming activity	Strike frequency	Daphnia consumed	% consuming daphnia	% survival from predation	
Carbaryl	1.95	0.1-1	0.1–1	>1	0.1–1	0.1–1	<0.01	
Chlordane	0.042	>0.02	0.002-0.02	0.002-0.02	0.002-0.02	0.0020.02	0.002-0.02	
DEF	0.66	0.05-0.1	0.005-0.05	0.005-0.05	< 0.005	0.005-0.05	0.005-0.05	
2,4-DMA	100	5-50	5-50	5-50	5 -50	0.5-5	5-50	
Methyl parathion	3.7	>0.1	< 0.01	0.01-0.1	<0.1	0.01-0.1	0.01-0.1	
Pentachlorophenol	0.052	>0.02	0.002-0.02	0.002-0.02	0.0002-0.002	>0.02	0.002-0.02	

DEF: tributyl phosphorotrithioate

2,4-DMA: 2,4-dichlorophenoxyacetic acid

After Little et al. (1990).

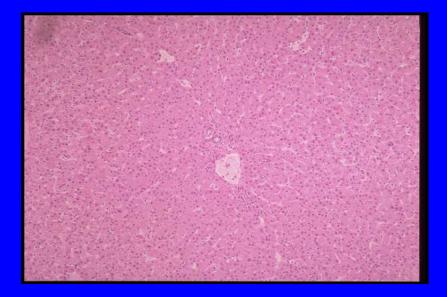
(Histo)pathology biomarkers

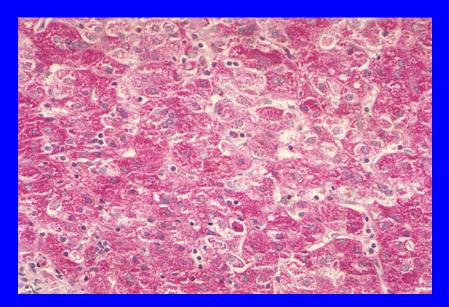
Pathology

(-) Destructive methods, Time consuming, Professional requirements(+) High relevance – organ/tissue changes

1) microscopy of internal organs

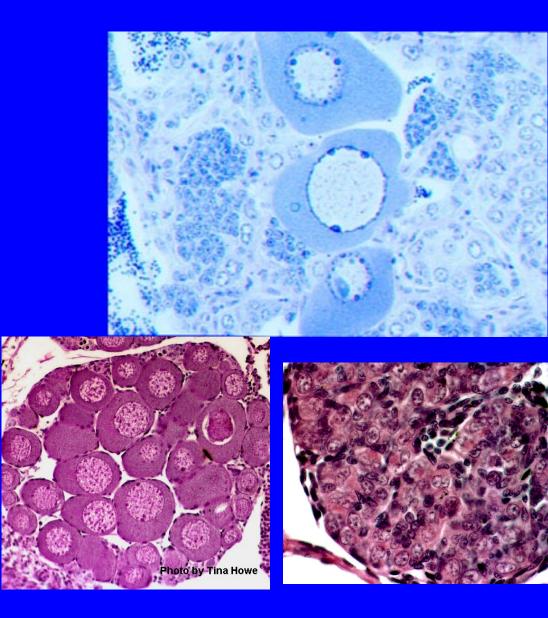
- : non-specific changes in internal organs
- : specific changes in liver (dioxin-like POPs, cyanobacterial toxins ..)
- : intersex / imposex formation (xenoestrogenicity)





Example: Liver damage by cyanobacterial toxins microcystins

Endocrine disruption: Intersex microscopy



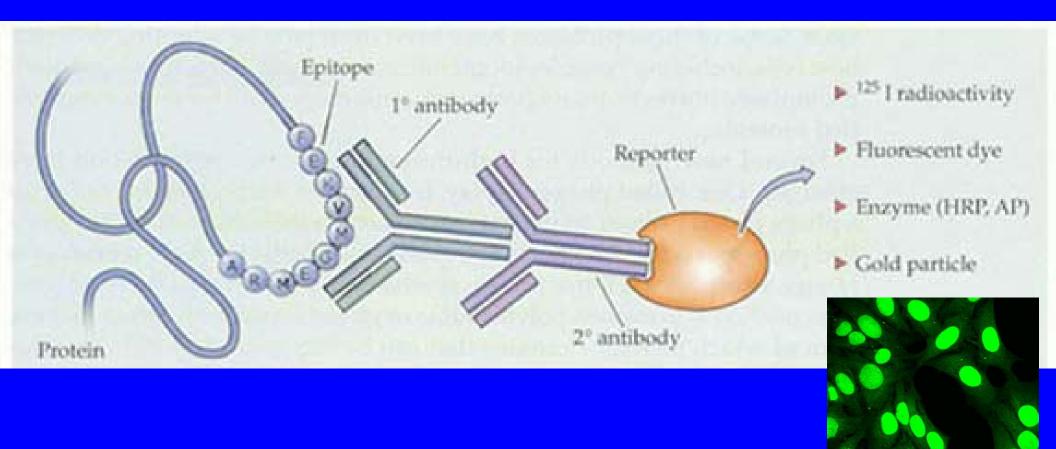
Oocytes in testicular tissue

Pathology

2) immunohistochemistry & microscopy

- : determination of specific changes
- : Fluorescein (FITC) labeled antibodies (Ab) applications
 - toxicant induced autoimmunity:

anti-nuclear Ab, ANA



Pathology

3) Nuclear DNA characterization

- micronuclei evaluation
- chromosomal abnormalities :
 - karyotype biomarkers (human genetic disorders)
- : non-destructive (blood samples; plant tissues)



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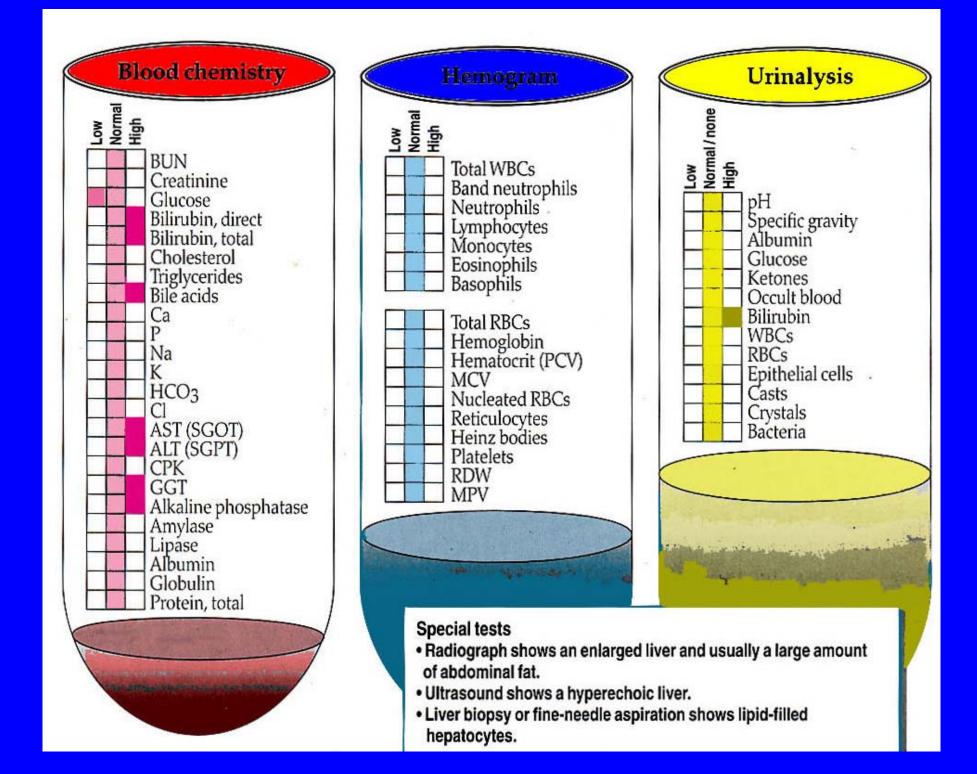
Clinical chemistry & hematology biomarkers

Non-destructive (BLOOD, URINE sampling)

Multipe parameters can be measured

responses to various types of stresses (including toxic stress)

 "normal" value ranges known for humans, rats and few other species (*limited use as biomarkers in other organisms*)



Methods:

- automatic biochemical and hematological analyzers
- different "analytes" various principles of methods

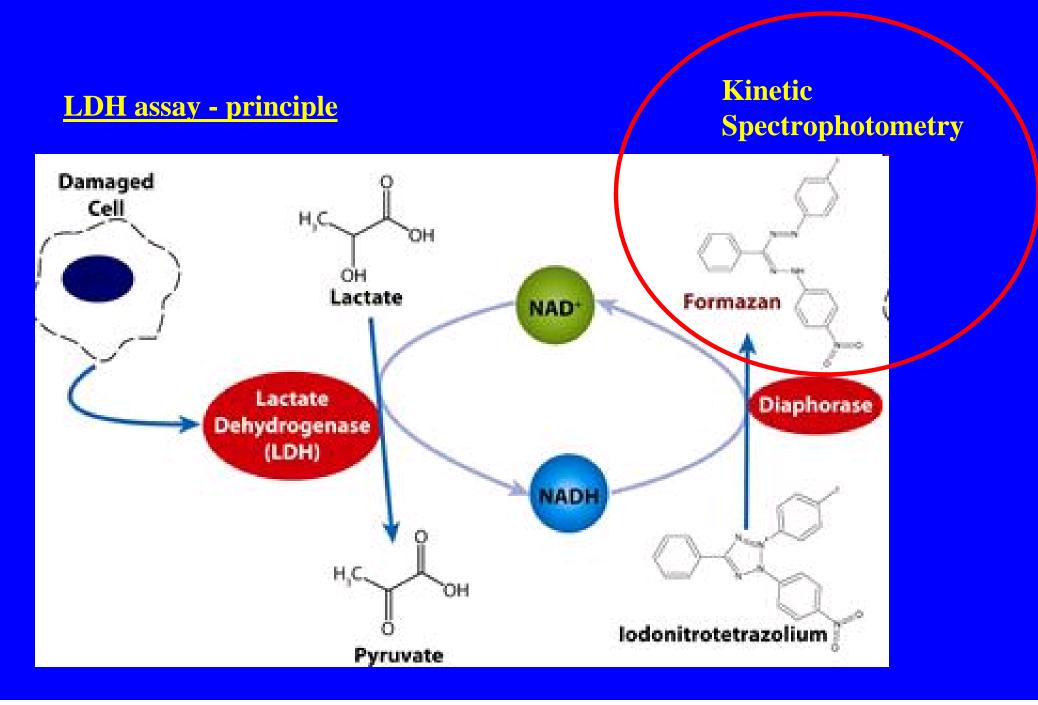


Often with specific interpretation:

- determination of enzymatic activities in blood
- tissue/organ-specific damage damage

Examples (toxicological studies)

- <u>liver damage</u> <u>AST</u> (Aspartate aminotransferase),
 <u>ALT</u> (Alanine aminotransferase) in blood...
 : cyanotoxins, dioxin-like POPs
- lactate dehydrogenase (LDH) general cell damage
- muscle damage: creatine kinase in serum
 - : isozymes tissue specific (brain, muscle, heart);



Example – changes in rat serum enzymes after CCL4 exposure

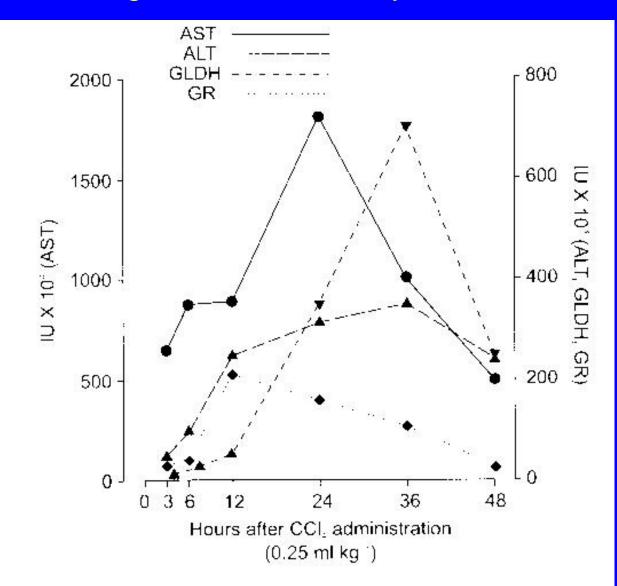


Figure 3 Serum enzyme levels in rats following dosing with carbon tetrachloride (CCI₄, 0.25 ml kg⁻¹). Redrawn from Zimmerman (1978).

PHAHs		
DDE	+ Quail	Dieter (1974)
	+ Starling	Dieter (1975)
DDT	= Redstart	Karlsson et al. (1974)
PCBs	= Redstart	
	+ Quail	Dieter (1974)
	+ Starling	Dieter (1975)
Endrin	– Fish	Sharma <i>et al.</i> (1979)
	(Ophiocephalus)	
Photomirex	+ Rat	Chu et al. (1981)
OPs		
Malathion	+ Rat	Dragomirescu et al. (1975)
	+ Quail	Dieter (1974)
	+ Starling	Dieter (1975)
	– Carp	Dragomirescu et al. (1975)
Methylparathion	+ Chicken	Somlyay et al. (1989)
Phosmethylan	+ Chicken	2000 yuy et ul. (1909)
Methidathion	+ Carp	Asztalos et al. (1990)
Metals		
Cadmium chloride	= Brook trout	Christensen et al. (1977)
Copper sulphate	+ Carp	Dragomirescu et al. (1975)
Lead nitrate	= Brook trout	Christensen et al. (1977)
Mercuric chloride	+ Quail	Dieter (1974)
	= Brook trout	Christensen et al. (1977)
	+ Fish	Verma and Chand (1986)
	(Notopterus)	(
Methylmercury	+ Starling	Dieter (1975)
Ithers		
Oil	= Striped mullet	Chambers et al. (1979)
Paraquat	+ Carp	Asztalos et al. (1990)

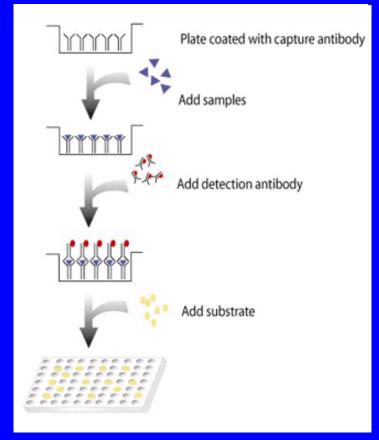
+ Human: E

Excretory products in urine Tumor genes and tumor markers

- cancer genes ras, myc,
- α -fetoprotein (AFP)
- suppressor genes p53, Rb

Methods of determination:

- ELISA (enzyme linked immunosorbent assays)



Changes in enzyme activities

Enzymatic changes

Toxicity mechanisms related to "enzyme changes":

Inhibitions of AcChE (organo-phosphates) d-Aminolevulinic Acid Dehydratase (ALAD) (lead - Pb) Proteinphosphatases (microcystins)

Inductions of detoxication & oxidative stress enzymes (hepatopancreas / liver / blood)

> MFO [CYP classes - **<u>EROD</u> / MROD / BROD] <u>Phase II enzymes</u> (GSTs) Glutathion metabolism enzymes (GPx, GRs)**

(+) Rapid enzymatic assays, specific responses(-) Some ~ EXPOSURE biomarkers

AcChE inhibition mechanism &

effects in birds

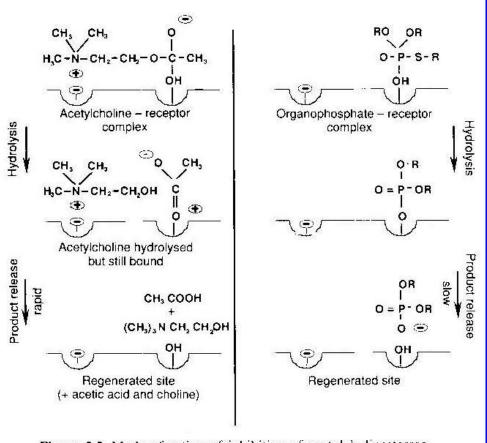
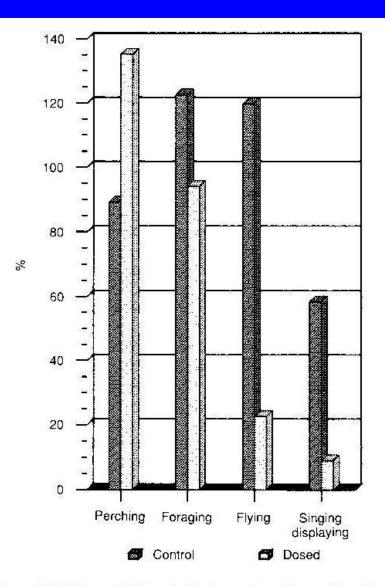
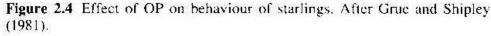


Figure 2.2 Mode of action of inhibition of acetylcholinesterase.

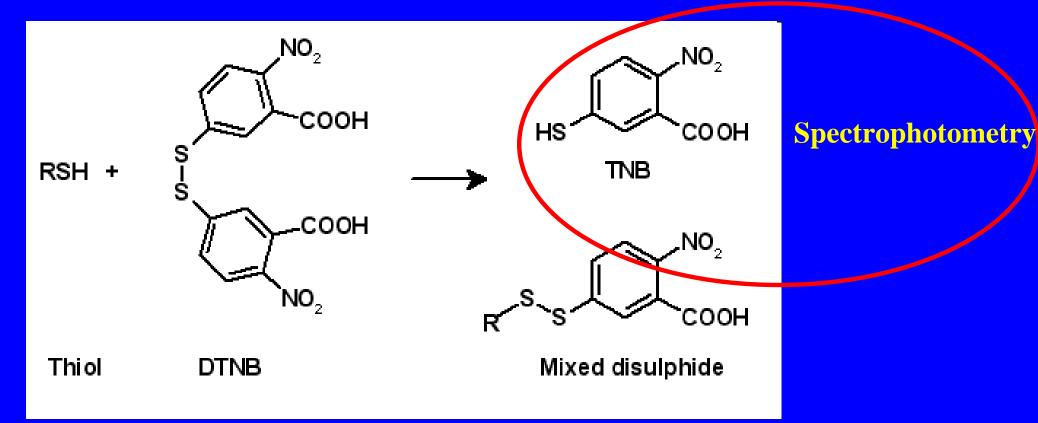




AcChE inhibition assay

Model Substrate (butyryl-thio-choline, acetyl-thio-choline)

- cleaved by <u>AcChE</u> -> formation of free –SH groups
- SH: thiol reactive probes: Ellman's reagent (DTNB)
- DTNB-S-choline: yellow colour (spectrophotometry A420)



AcChE inhibition mechanism &

effects in birds

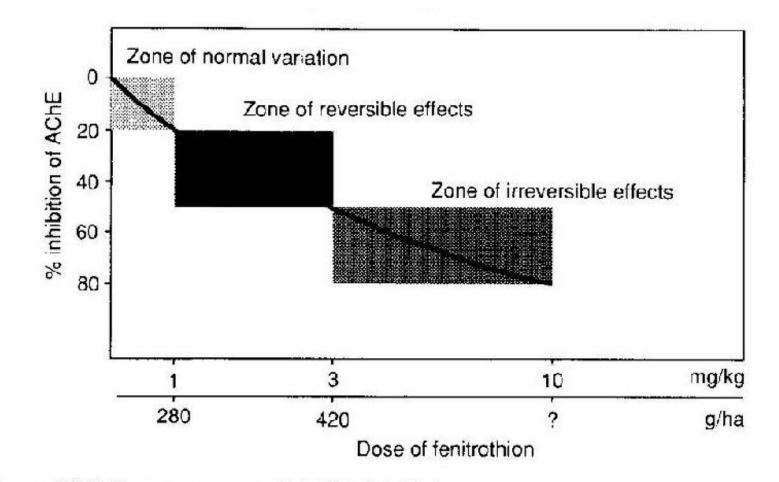
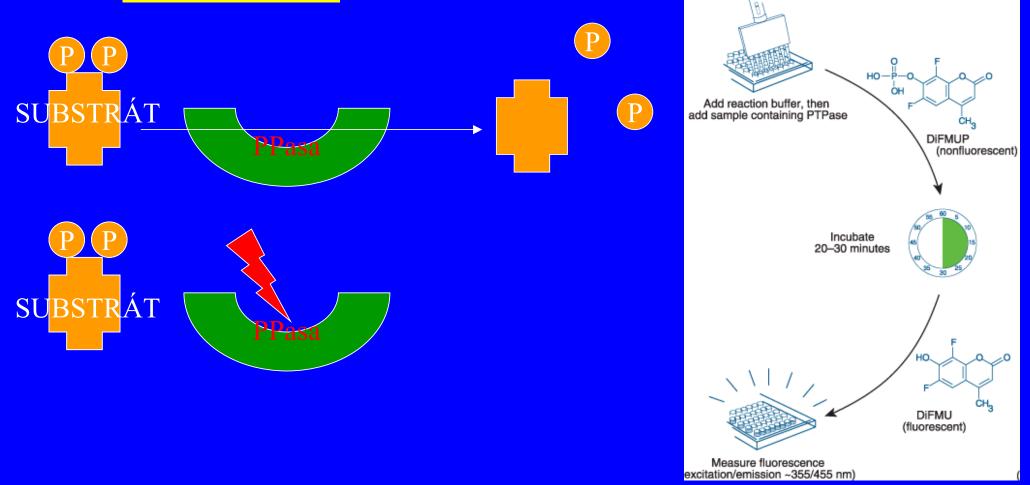


Figure 10.2 Dose response of AChE inhibition.

Proteinphosphatase inhibition assay

<u>Model substrates cleaved by PPase</u> ³²P-labelled protein -> free ³²P radioactivity 6,8-difluoro-4-methylumbelliferyl phosphate -> <u>fluorescence</u>



MFO (CYP) activities

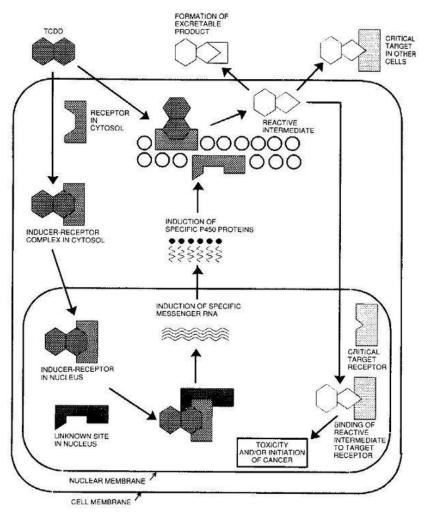


Figure 5.1 Diagram of MFO system. Nebert and Gonzalez (1987).

Table 5.1 Classification of P450s

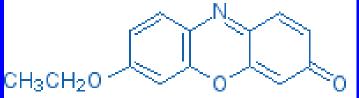
Nomenclature	Induced by/specificity			
P4501	Polycyclic aromatic, TCDD			
P4501I	Phenobarbital-inducible family*			
P450IIA	Specific for testosterone hydroxylase			
P450IIB	PB inducible			
P450IIC	PB inducible			
P450IID	Specific for debrisoquine 4-hydroxylase			
P450IIE	Ethanol inducible			
P450III	Steroid inducible			
P450IV	Specific to lauric acid w-hydroxylation			
P450XI	Located in mitochondrion			
P450XIA				
P450XIB				
P450XVII	Formation of steroid 17-hydroxylases			
P450XIX	Involved in synthesis of oestrogens			
P450XX1	Formation of steroid 21-hydroxylases			
P450LI	Plant/yeast			
P450CI	Prokaryote			

* PB-inducible genes largely confined to P450IIB and C. After Nebert and Gonzalez (1987).

MFO (CYP) activities

EROD assay

Determination of CYP450 activity



substrate: Ethoxyresorufin
 -> Oxidation by CYP1A1 -> Fluorescence
 EthoxyResorufin-O-Deethylase activity EROD
 (other substrates: CYP isozymes:
 BROD - butoxy..., MROD, PROD ...)

Biomarker of organic pollution (exposure & effects)

- : AhR-activating compounds (PCDD/Fs, PCBs, PAHs)
- : often used in environmental studies

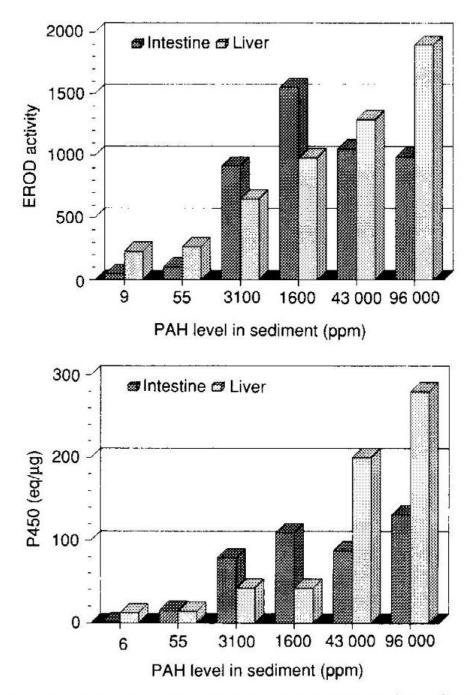


Figure 5.6 Relationship of sediment concentration of PAHs to EROD activity in liver and intestine of spot. After Van Veld *et al.* (1990).

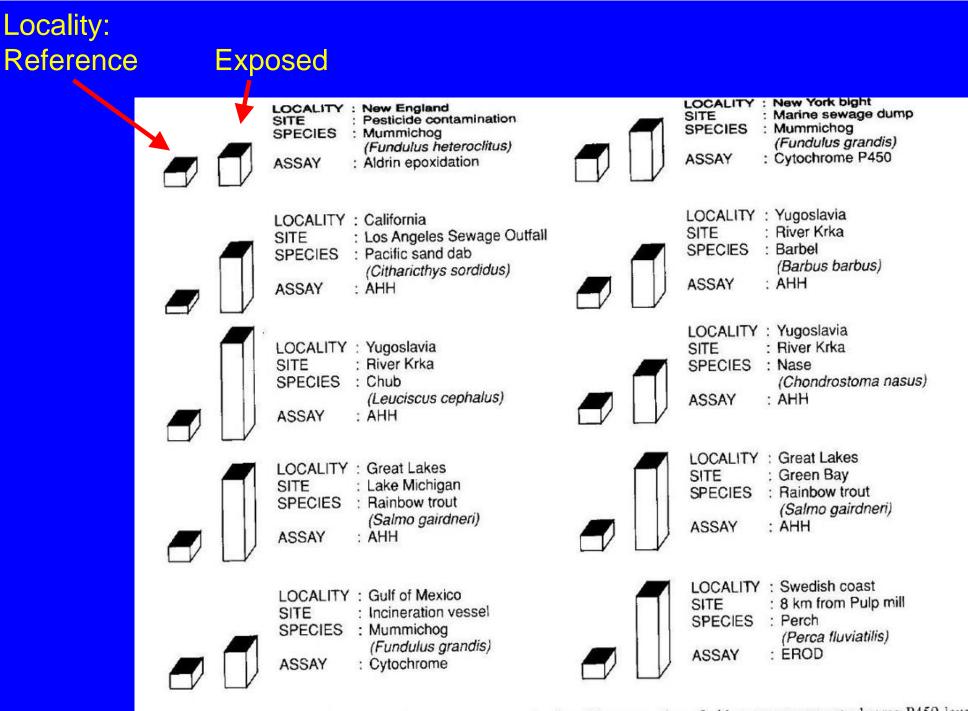
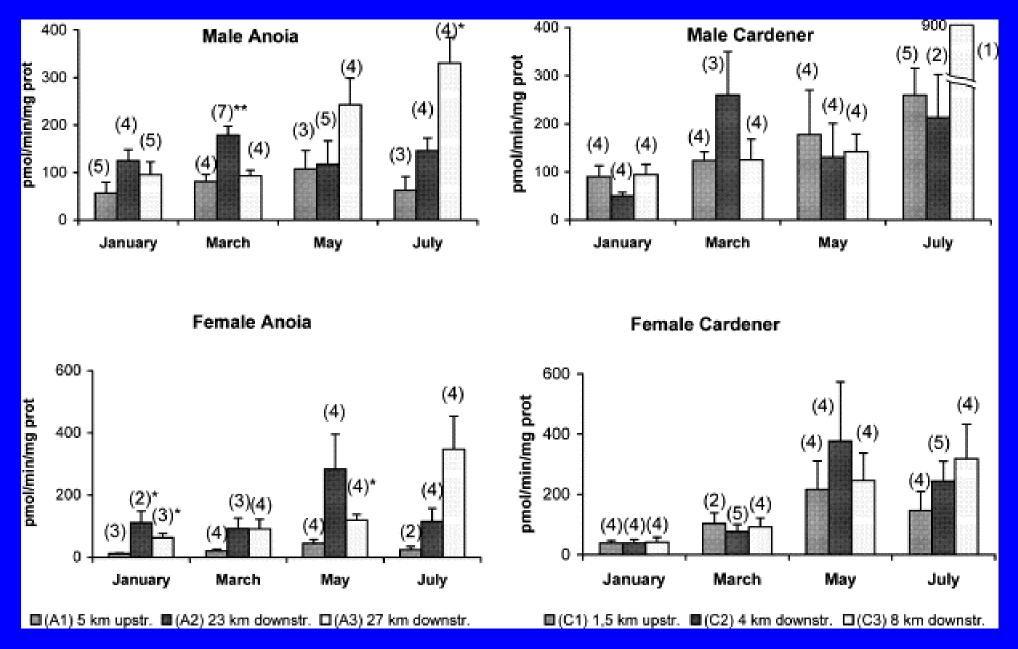


Figure 5.5 MFO changes in fish exposed to organic contamination. The proportion of either enzyme or cytochrome P450 levels letected at reference (short towers) and experimental sites (long towers) is presented in schematic form. All differences between reference and experimental sites were statistically significant (P < 0.05 or better). Payne *et al.* (1987).



EROD variation on male and female carp from the Anoia and Cardener tributaries – *seasonal variability & response at contaminated localities*

MFO-responses are SPECIES – SPECIFIC & not always related to clinical signs

Table 3.3 Comparison of the effects of PCB congeners on the reproduction of mink and rats

Clinically normal to change in cytochrome P450	Clinically normal No change in cytochrome
Io change in cytochrome P450	No change in cytochrome
	P450
lo induction of MFO	Some induction of MFO
enzymes	enzymes
evere anorexia and	Clinically normal
diarrhoea	
ncrease of cytochrome P450	Increase in cytochrome P450
lo induction of MFO	Induction of MFO enzymes
enzymes	
ŧ	enzymes evere anorexia and diarrhoea crease of cytochrome P450 o induction of MFO

After Gillette et al. (1987a).

MFO-responses are SPECIES – SPECIFIC & relative activity decreases with body size

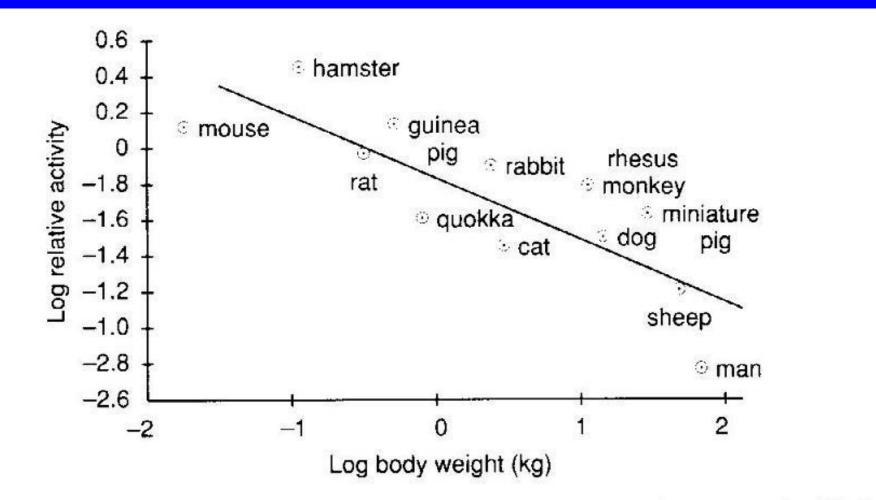


Figure 5.3 Relationship of body weight to MFO activity in mammals. Walker (1978 and 1980).

Potencies to induce CYPs (AhR)

PCDD/Fs and co-planar PCBs

- induction of MFO is structure-dependent; potencies & toxicities among compounds differ
- international agreement on <u>TEF/TEQ approach</u> to characterize dioxin-toxicity in environmental samples (WHO)
- each compound (only few selected in WHO agreement) relative potency (TEF) related to 2,3,7,8-TCDD
 - 2,3,7,8-TCDDTEF = 1Several other PCDD/Fs0.1-1PCBs $10^{-5} 0.1$ (No. 77, 126)
- species-specific TEFs for humans / fish / birds
- chemical analyses of samples

 => SUMA (concentrations x TEF) = TEQ (ng TCDD / sample)
 EASY comparison of sample contamination

TEFs for selected PCDDs

CONCENED

TOVIC FOUNDAL ENCY FACTOR (TEP)

CONGENER	TOXIC EQUI	IVALENCY FAC	FACTOR (TEF)			
	HUMANS/ MAMMALS	FISH ^a	BIRDS ^a			
2,3,7,8-TCDD	1	1	1			
1,2,3,7,8-PeCDD	1	1	1 f			
1,2,3,4,7,8-HxCDD	0.1 ^a	0.5	0.05^{-f}			
1,2,3,6,7,8-HxCDD	0.1 ^a	0.01	0.01^{-f}			
1,2,3,7,8,9-HxCDD	0.1 ^a	0.01 e	0.1^{-f}			
1,2,3,4,6,7,8-HpCDD	0.01	0.001	< 0.001 f			
OCDD	0.0001 ^a	-	-			

TEFs for PCBs

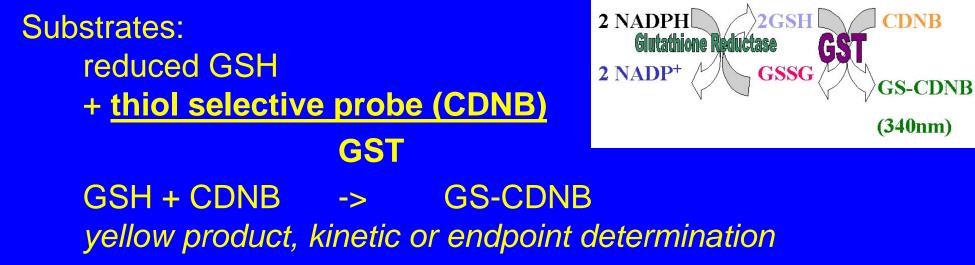
			1997 WHO TEFs(2)				
Congener Number	IUPAC Chlorobiphenyl Prefix	1994 WHO TEFs(1)	Humans/ Mammals	Fish	Birds		
PCB-77	3,3',4,4'-Tetra-	0.0005	0.0001	0.0001	0.05		
PCB-81	3,4,4',5-Tetra-		0.0001	0.0005	0.1		
PCB-105	2,3,3',4,4'-Penta-	0.0001	0.0001	<0.00005	0.0001		
PCB-114	2,3,4,4',5-Penta-	0.0005	0.0005	<0.00005	0.0001		
PCB-118	2,3',4,4',5-Penta-	0.0001	0.0001	<0.00005	0.00001		
PCB-123	2,3',4,4',5'-Penta-	0.0001	0.0001	<0.00005	0.00001		
PCB-126	3,3',4,4',5-Penta-	0.1	0.1	0.005	0.1		
PCB-156	2,3,3',4,4',5-Hexa-	0.0005	0.0005	<0.00005	0.0001		
PCB-157	2,3,3',4,4',5'-Hexa-	0.0005	0.0005	<0.00005	0.0001		
PCB-167	2,3',4,4',5,5'-Hexa-	0.00001	0.00001	<0.00005	0.00001		
PCB-169	3,3',4,4',5,5'-Hexa-	0.01	0.01	0.00005	0.001		
PCB-170	2,2',3,3',4,4',5-Hepta-	0.0001					
PCB-180	2,2',3,4,4',5,5'-Hepta-	0.00001					
PCB-189	2,3,3',4,4',5,5'-Hepta-	0.0001	0.0001	<0.00005	0.00001		

Phase II conjugation enzymes - GSTs

<u>GSTs</u>

- soluble and membrane (ER) variants
- activities in cytoplasm or microsomes

<u>Methods</u>



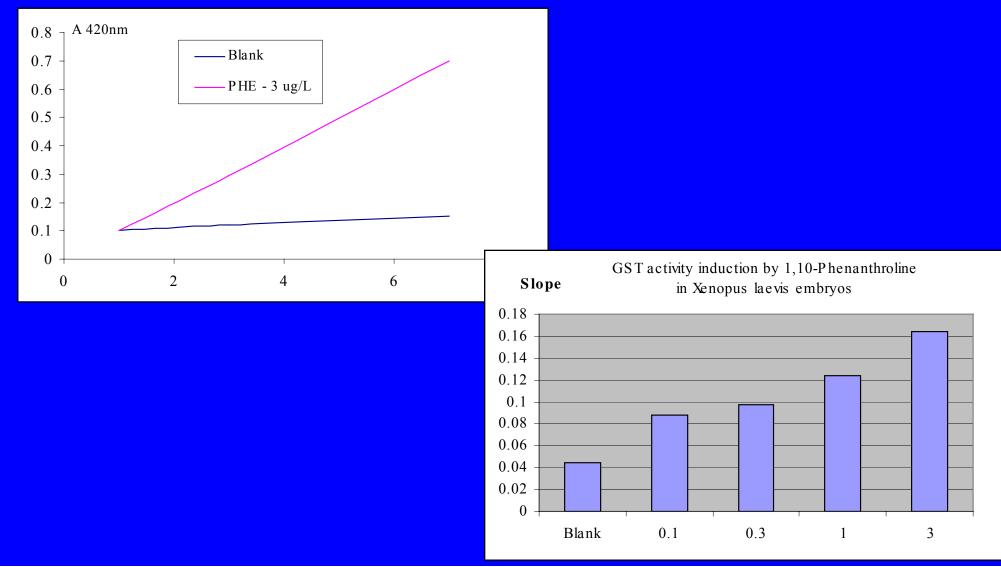
Kinetic assessment

stress -> Induction of GSTs faster reaction -> slope of kinetic increase

GST activity - example

Kinetic assessment of GSTs

stress -> Induction of GSTs faster reaction -> kinetic slope increases



Protein levels (synthesis) biomarkers

PROTEIN SYNTHESIS

Protein determination

- amount (concentration)
- activity (see enzymatic assays)

Amount quantification

- mRNA levels (in vitro assays)
- protein
 - electrophoresis and Western-(immuno)blotting
 - ELISA techniques

Examples

heat shock proteins (hsp90, hsp60, hsp 70, ubiquitin) metalothioneins

Vitellogenin(-like) Vtg proteins in male

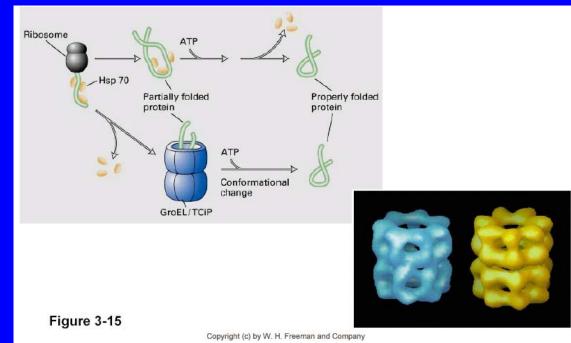
Heat Shock Proteins (hsp)

Stress = synthesis of new proteins

- ~ equilibrium and homeostasis buffering
 - temperature (cold / heat) cryo-preservation
 - salinity & metals ion buffering
 - organic xenobiotics detoxication

New proteins must be folded

(3D-structure) by "CHAPERONES" - hsp90, hsp60, hsp 70 (~ 60-90 kD molecular weight kD)

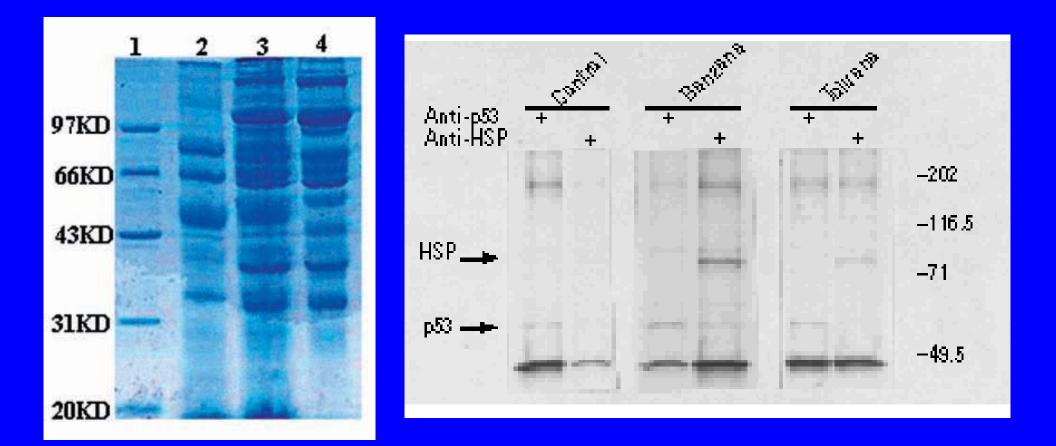


HSP determination - example

<u>HSP = GENERAL STRESS biomarker</u>, non-specific

- phylogenetically conserved (similar sequences in "all" organisms)
- structural similarity => easy determination:

electrophoresis + immunoblotting (Western blotting)



Metalothioneins (MTs, MT-like proteins)

Low MW proteins (6-10 kD) rich of Cystein (-SH)

- detected in numerous eukaryotic organisms
- induced in the presence of metals or less specific stress (low O2, T)
- long halflife (~ 25 days)
- binding of divalent metals (Zn, Cd, Hg) => exposure elimination
- natural function (?) regulation of essencial metals in cells

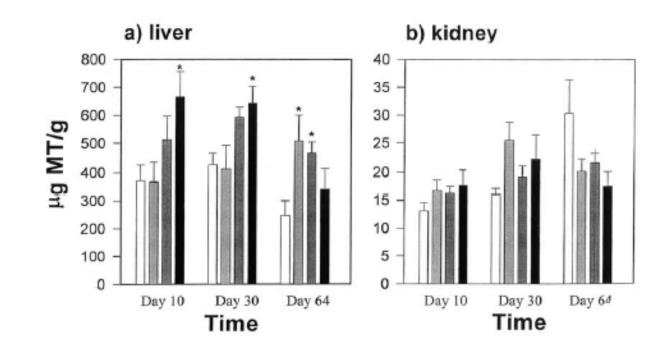


Fig. 2. Metallothionein (MT) concentrations in the (a) livers and (b) kidneys of lake whitefish fed a control diet and three As contaminated diets for 10, 30, and 64 days. Data are expressed as mean (\pm S.E.). Asterisk denotes mean is significantly different from the control at that duration (P < 0.05). See Fig. 1 for an explanation of histogram shading.

Vitellogenin

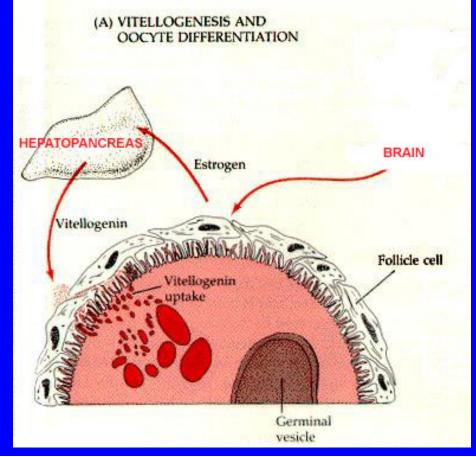
<u>Vtg</u>

precursor of yolk proteins, phospho-protein
 -> egg formations (females) at oviparous animals

synthesised in liver and distributed via blood (haemolymph)
 xenoestrogens & other endocrine disruptors

-> increased levels or early production in FEMALES

-> production in MALES



Vitellogenin

VTG Determination

1) ELISA (exposed organisms - F/M, in vitro

- in vivo exposed organisms (*biomarker in vivo*)
- in vitro production in hepatocytes exposed to effluents

(marker of estrogen-like presence

(-) specific Antibodies necessary for each species (low crossreactivity)

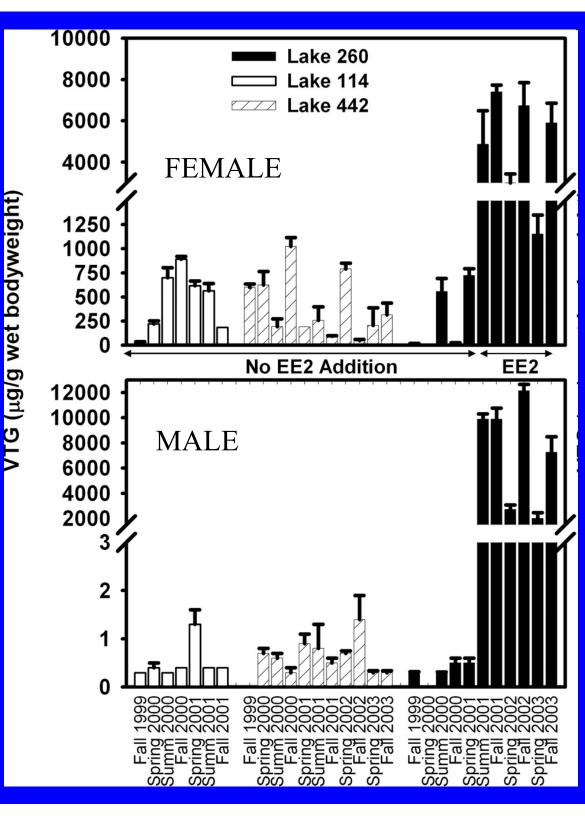
2) "Vitelin-like proteins"

- total amount of "alkali-labile" phosphate in haemolymph (mussels)
- alkaline extraction of P from sample & determination

Vitellogenin in fish

Kidd et al. (2007) PNAS

Fig. 1. Mean \pm SE (n = 4-7) VTG concentrations in whole-body homogenates of male (*Lower*) and female (*Upper*) fathead minnow captured in 1999–2003 from reference Lakes 114 and 442 and from Lake 260 before and during additions of 5–6 ng·L⁻¹ of EE2 (low catches of fish in Lake 260 in 2004 and 2005 did not allow for these analyses in the latter 2 years of the study).



Vitelin-like proteins in mussels

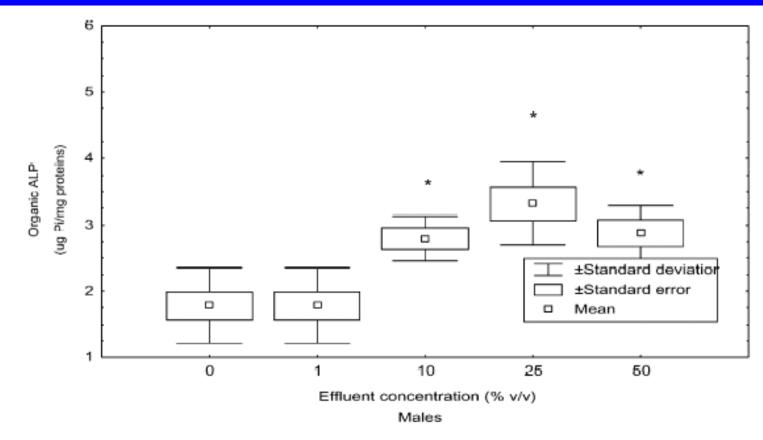


Fig. 4. Induction of Vg by exposure to a municipal effluent. Mussels were exposed for 96 h to a municipal effluent at 15°C. They were then collected for Vg and sex determinations. The asterisk (*) indicates significant difference at P < 0.05.

Biomarkers of oxidative stress

Oxidative stress markers

Several parameters respond to oxidative stress

: enzymes (GPx, GR, GSTs) - enzymatic activities (see elsewhere)

: antioxidants (GSH, vit E)

: markers of oxidative damage

- <u>MDA</u>,

- 80H-dG (see DNA damage)

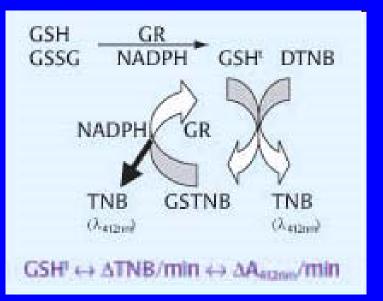
Oxidative stress markers

GSH determination

- antioxidant (scavenger of ROS) & reactive molecules
- conjugation molecules for detoxication
- probable intracellular regulatory molecule (? apoptosis ?)

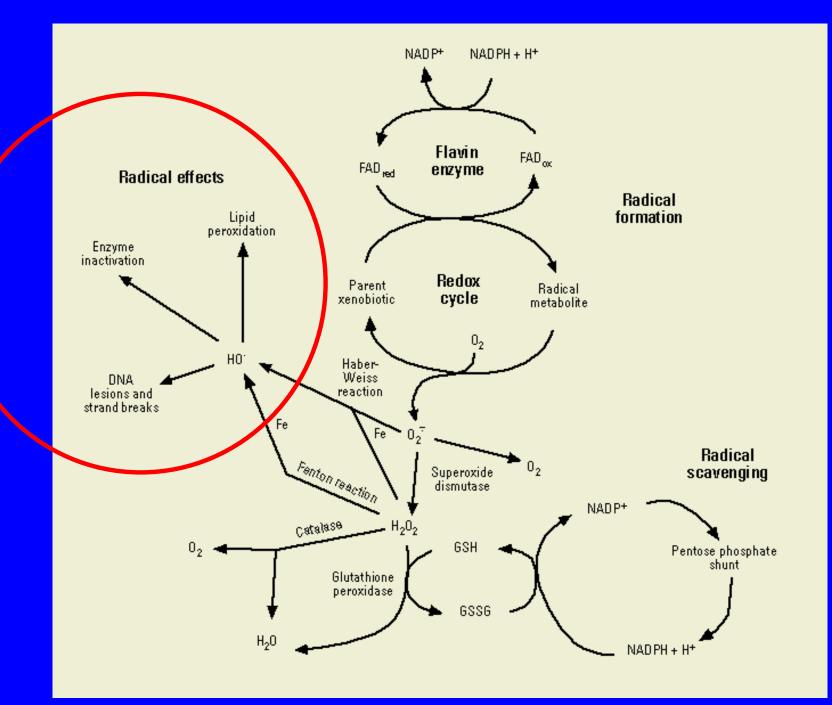
Total glutathione = reduced GSH + oxidized GSSG

GSH + <u>Ellman's reagent (DTNB)</u> -> Reduced GSH GSH + Glut.Reductase + <u>DTNB</u> -> Total GSH



Total – Reduced = Oxidized

Markers of oxidative DAMAGE



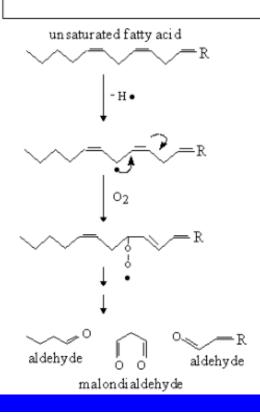
Lipid peroxidation -> Malonyldialdehyde (MDA)

<u> MDA – malondialdehyde</u>

product of Lipid peroxidation

STEPS OF LIPID PEROXIDATION

(INITIATION) LIPID + R/OH \rightarrow LIPID (PROPAGATION) LIPID + 0 \rightarrow LIPID-00 LIPID-00 + LIPID \rightarrow LIPID-00H + LIPID (TERMINATION) LIPID + LIPID \rightarrow LIPID-LIPID LIPID-00 + LIPID \rightarrow LIPID-00-LIPID (SCAVENGING) LIPID + VIT E \rightarrow LIPID + VIT E



Lipid peroxidation -> Malonyldialdehyde (MDA)

MDA – formed from oxidized membrane phospholipids

- : determination:
 - HPLC
 - TBARS method

TBARS – ThioBarbituric Acid Reactive Species

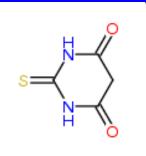
- : less specific than HPLC (+/- aldehydes)
- : easy determination (spectrophotometry)

Method:

- 1) sample extract (with MDA)
- 2) add TBA
- 3) boil (cca 30' / 90°C)



4) determination by spectrophotometry (A 540 nm)



MDA modulation - examples

