

#6

Aktivace vývojových mechanismů
u dospělého organismu

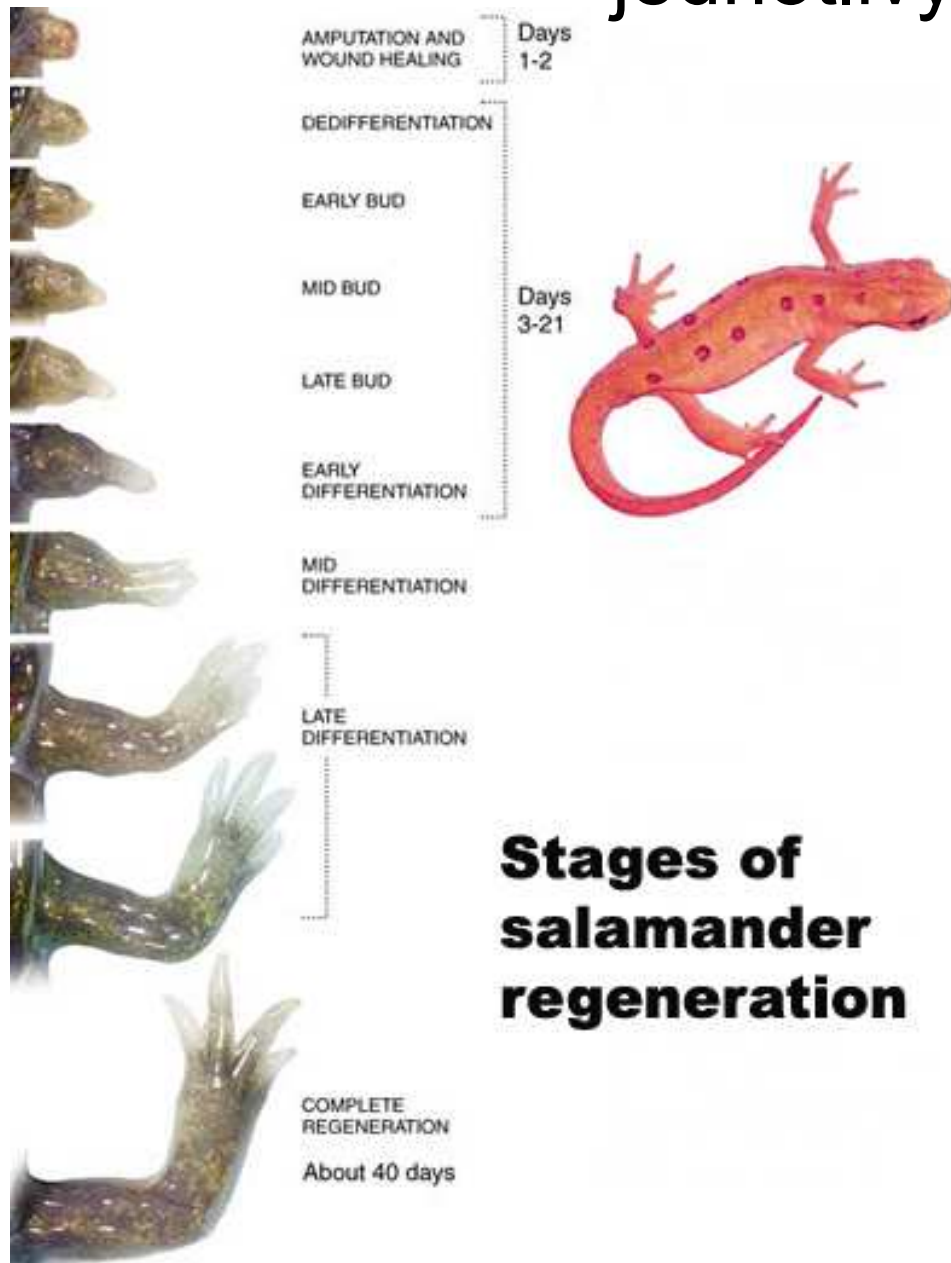
Osnova:

- Kdy a proč se reaktivují procesy spjaté s embryonálním vývojem?
- Regenerace – buněčné a molekulární mechanismy
- Kmenové buňky – definice, ESCs, adult stem cells, stem cell niche
- When things go wrong – tvorba nádorů
- Cancer stem cell concept, markery, signální dráhy (...a jsme zpět u velké pětky)

Kdy a proč se reaktivují procesy spjaté s embryonálním vývojem?

- odpověď je logická: tehdy, když je potřeba znovu vytvářet struktury poškozené či zničené tj. když je potřeba regenerovat
- v lidském organismu běžně regenerují celé tkáně – např. vlasové kořínky (doba „života“ 3-4 roky), krypty v tlustém střevě, epitel plic, krevní buňky nebo játra

přesto se Schopnosti regenerace se liší mezi jednotlivými organismy



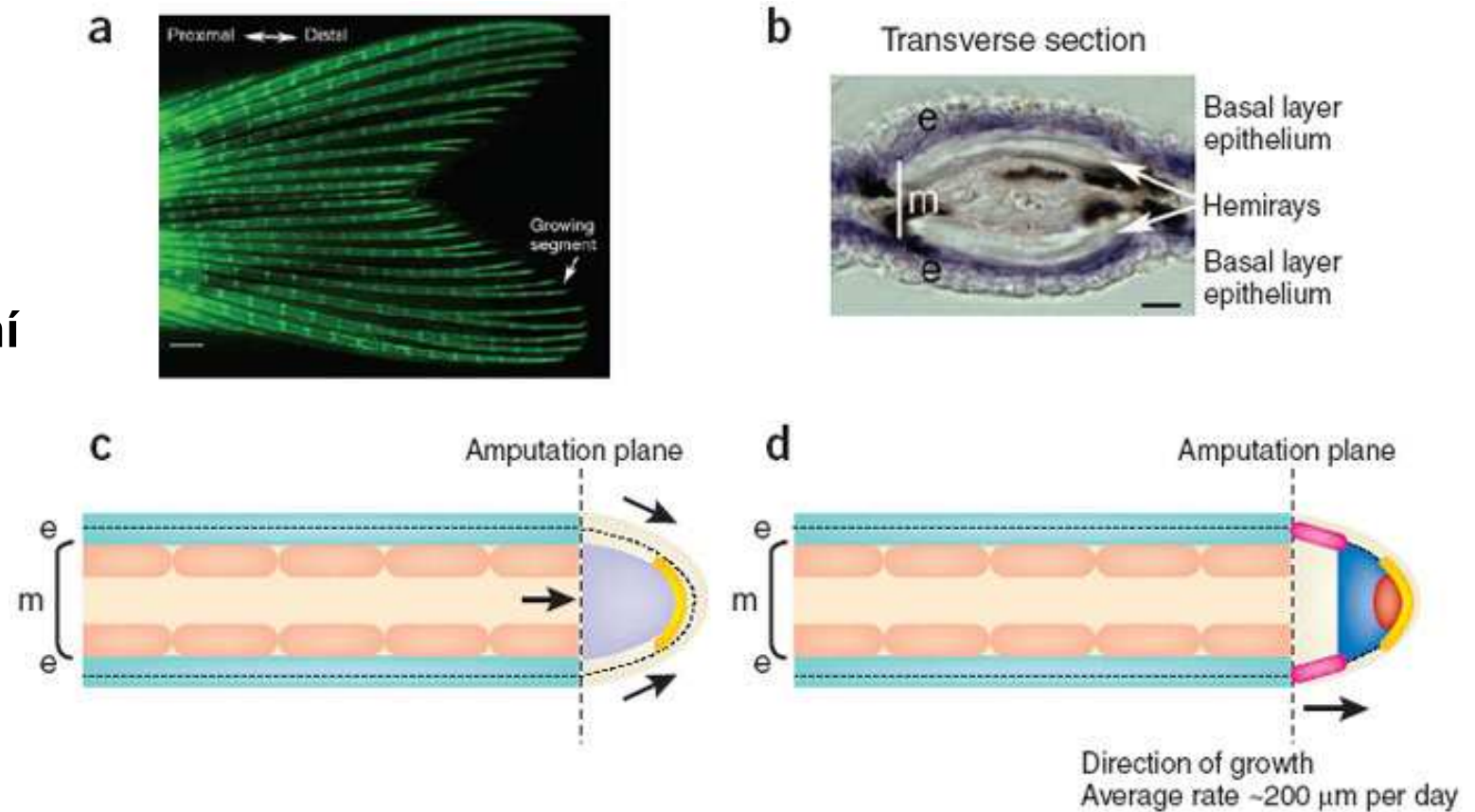
po amputaci



po 10 letech

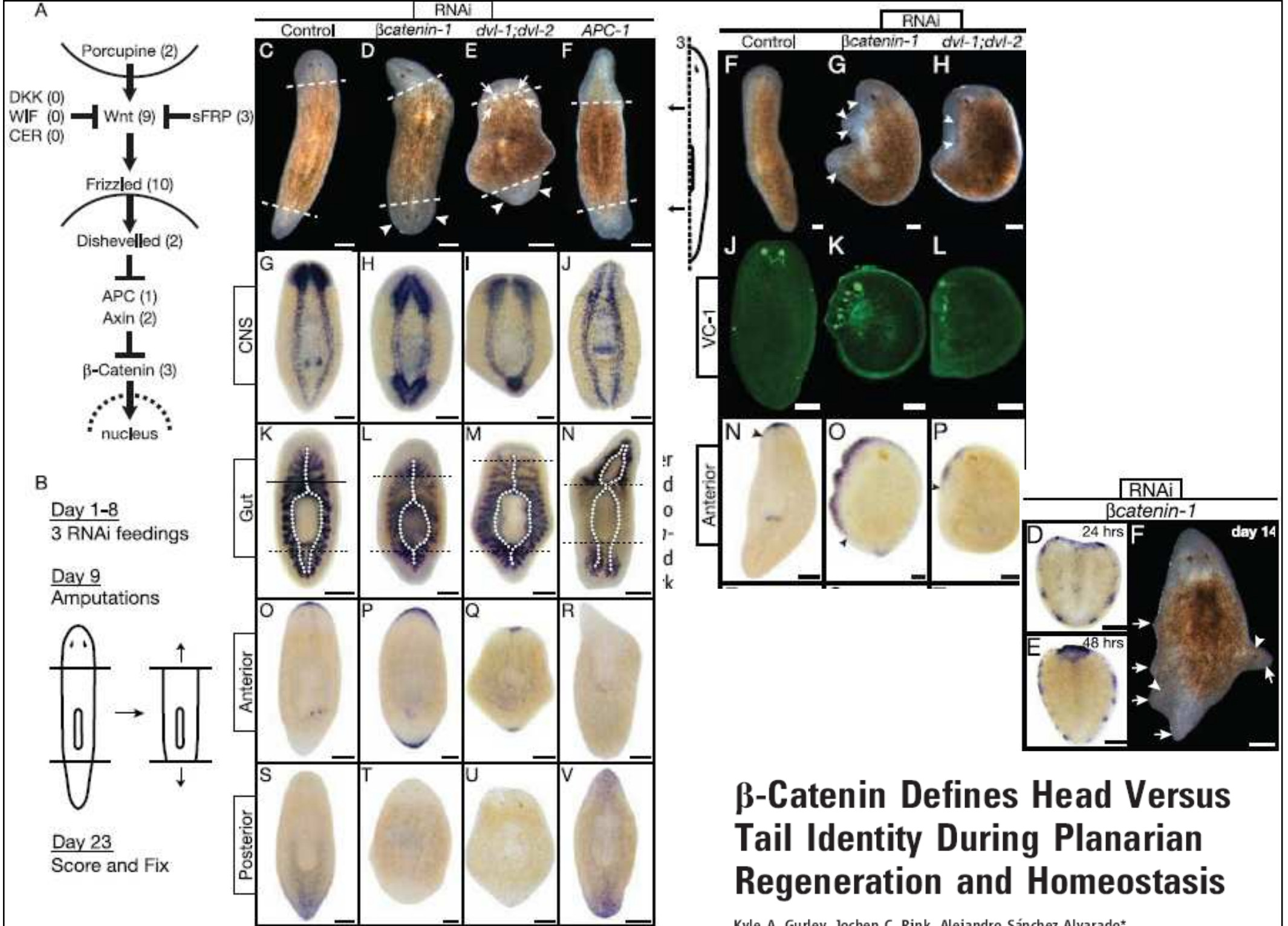


Blastéma – masa nediferencovaných pluripotentních buněk, která se vytváří po poškození a dává vznik nové struktuře



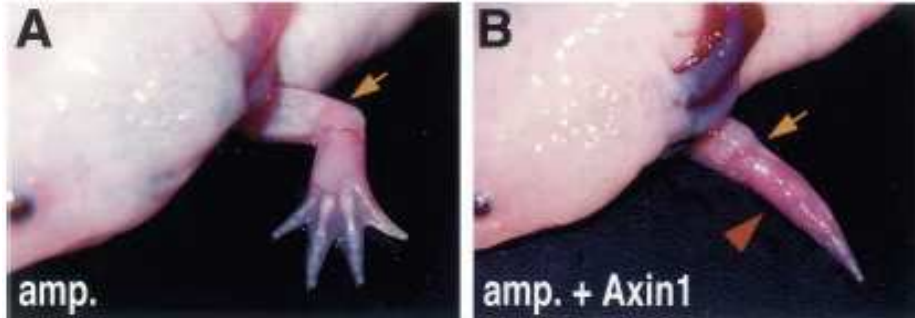
(a) Zebrafish caudal fin stained with calcein so that bony fin rays are visible. Scale bar, 200 μm. (b) Transverse section through a single fin ray. Hemirays (white arrows) are visible as crescents of bone surrounding mesenchymal tissue (fibroblasts, undifferentiated cells, artery, nerve, melanocytes). The bony rays are surrounded by an epithelium, and the basal layer of the epidermis has been labeled (purple stain and arrowheads). Scale bar, 15 μm. (c) Cartoon of a longitudinal section of a single amputated fin ray after formation of the immature blastema. The basal layer of the epithelium is indicated by the dotted line. Cells from the epithelium (light blue) cover the wound (side arrows), forming the wound epidermis and apical epidermal cap (AEC). Cells from the stump mesenchyme migrate distally (center arrow), forming the immature blastema (purple). (d) Cartoon of a longitudinal section of a single fin ray following blastemal reorganization. The basal layer of the epithelium is indicated by the dotted line, the AEC in yellow. The distal blastema (red) is distal to the proximal blastema (dark blue). Proliferating cells from the proximal blastema will migrate laterally, toward the basal epidermal layer (pink), and differentiate as bone forming cells. e, epidermis; m, mesenchyme.

During limb growth in vertebrates, an essential first step is the establishment of 'signaling centers', localized areas in the limb bud that contribute to the mechanisms regulating outgrowth and pattern formation by secreting signaling molecules such as Fgfs, Wnts, transforming growth factor (TGF)-s or bone morphogenetic proteins (BMPs), and hedgehogs (Hh). In the zebrafish, gene profiling studies have shown that the expression levels of these and other signaling molecules are regulated during the course of regeneration.

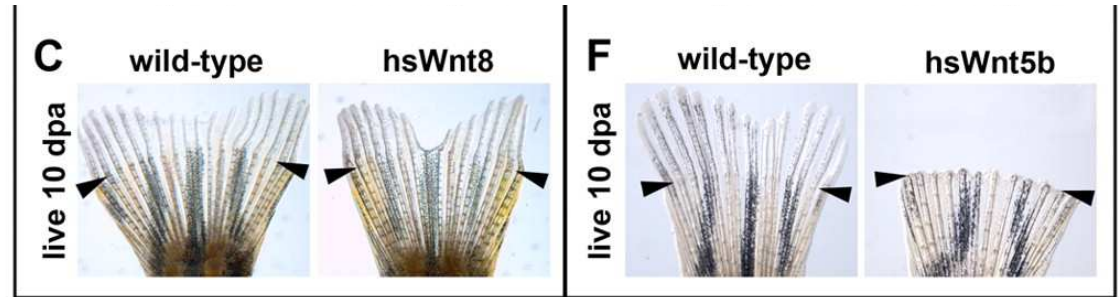


Morphogenetic pathways (canonical Wnt signalling, Hedgehog, TGF, Notch) are required for regeneration in multiple organisms

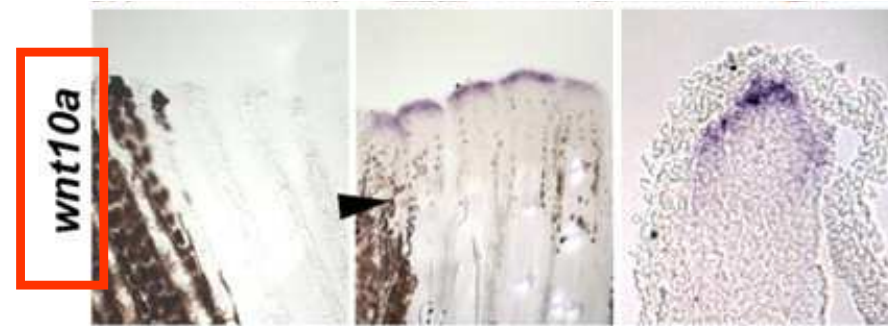
axolotl



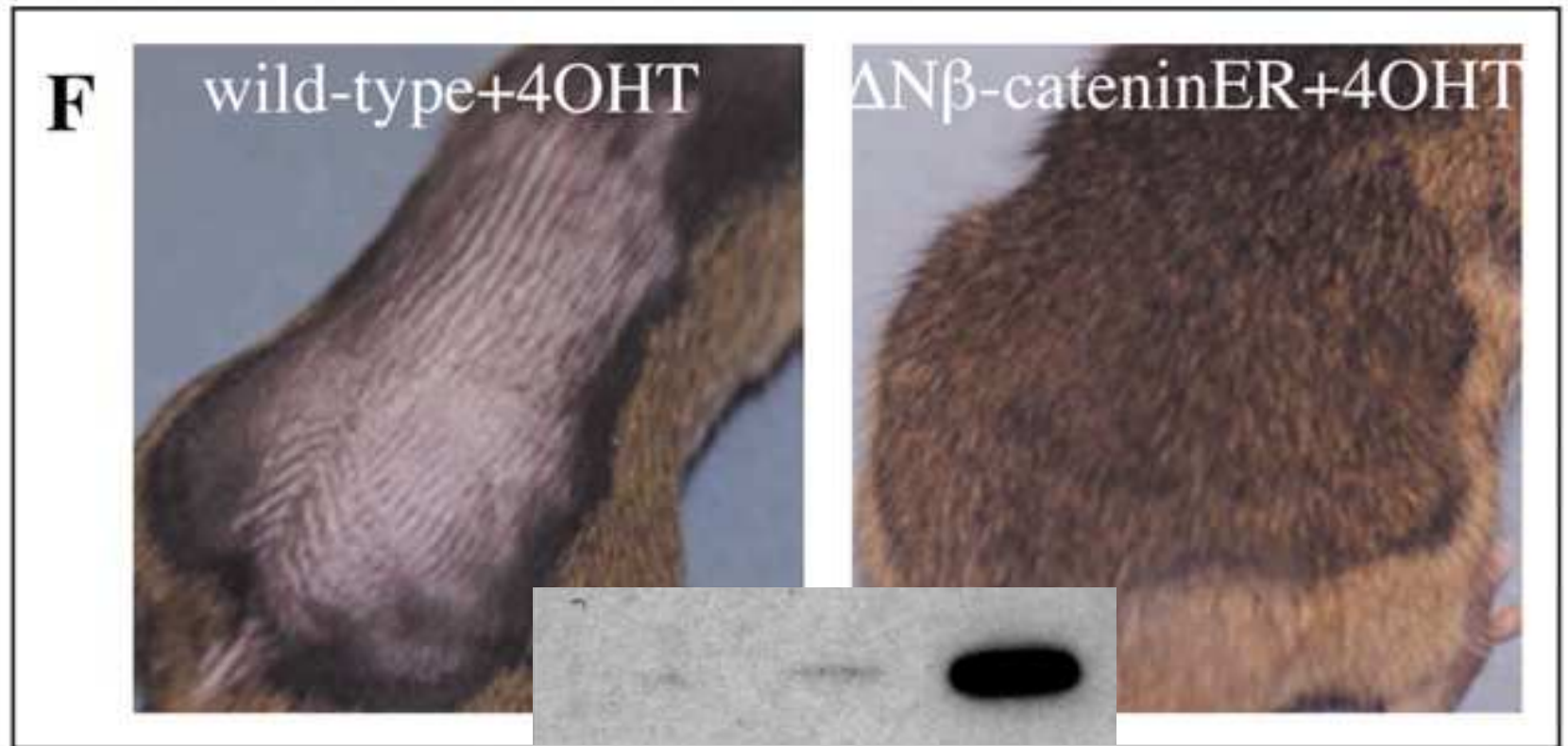
zebrafish



chick



Stejné mechanismy aktivují regeneraci i u savců a lidí.



Lo Celso, C. L. et al. Development 2004;131:1787-1799

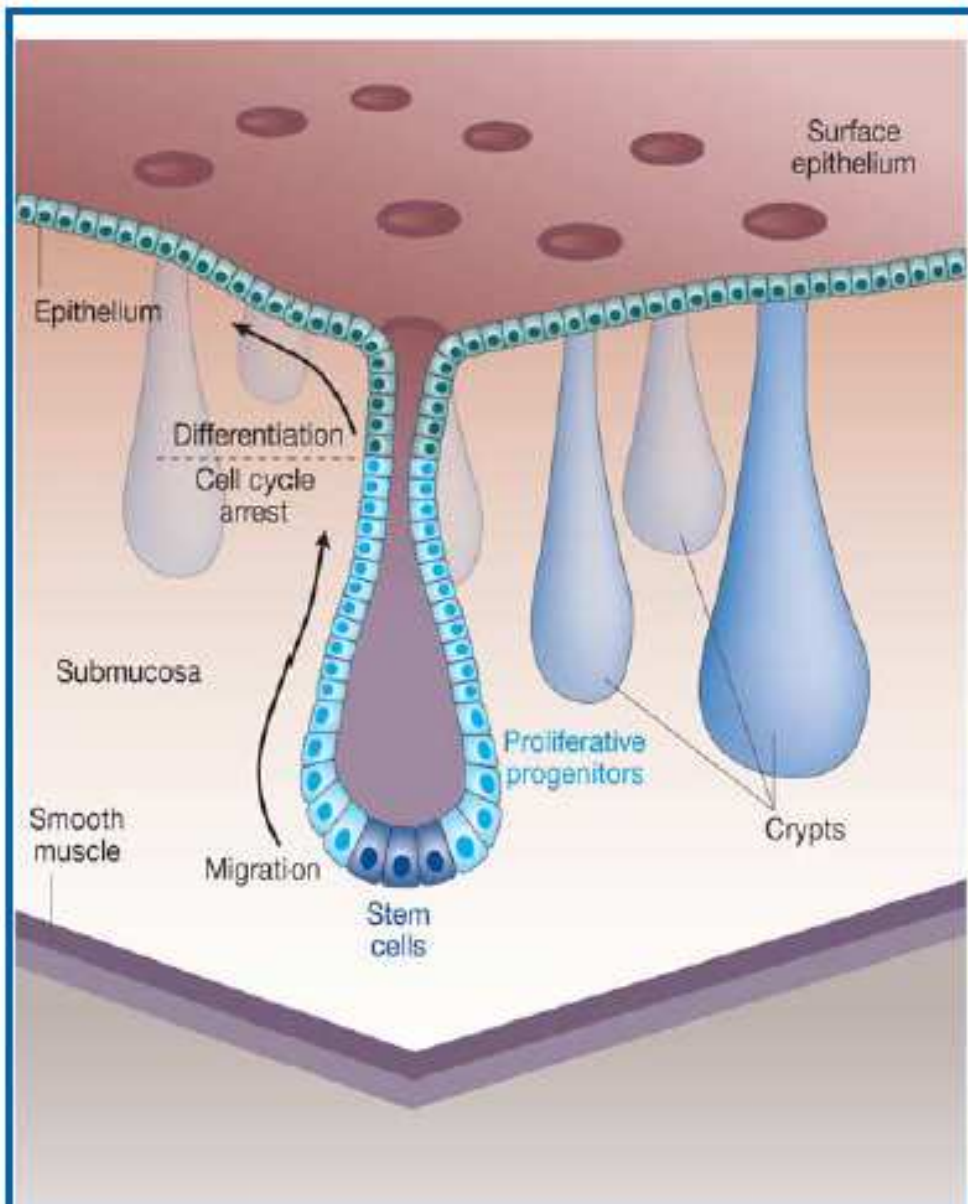


Figure 3 Tissue anatomy of the colonic epithelium. Putative stem cells (dark blue) reside at the crypt bottom. Proliferating progenitor cells occupy two-thirds of the crypt. Differentiated cells (green) populate the remainder of the crypt and the flat surface epithelium. (Adapted from ref. 89.)

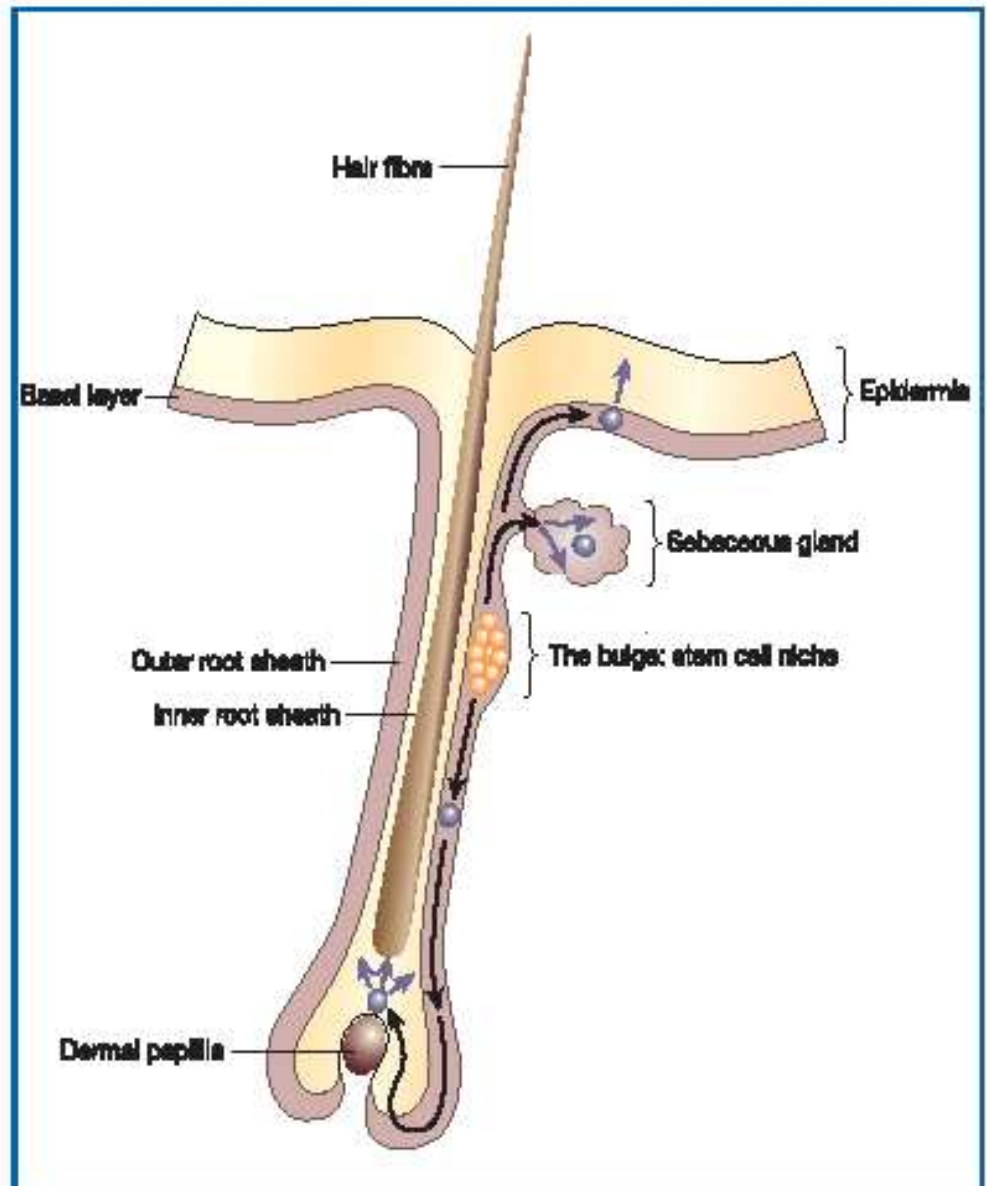


Figure 4 The hair follicle. Stem cells reside in the bulge niche. Cells can migrate upwards from here to populate the sebaceous gland and the interfollicular epidermis. Cells that migrate downwards enter the matrix where they rapidly proliferate and then differentiate to form the hair. (Adapted from ref. 90.)

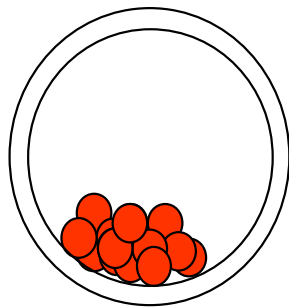
Kmenové buňky

a) sebeobnova (selfrenewal)

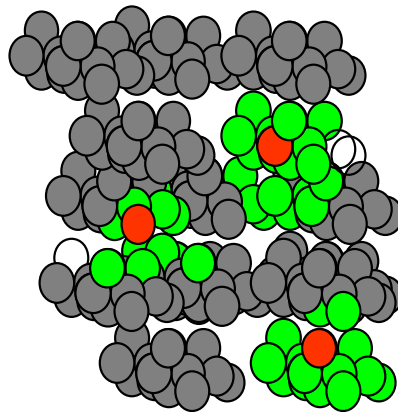
b) multipotence

- embryonální kmenové (ES)
- tkáňově specifické

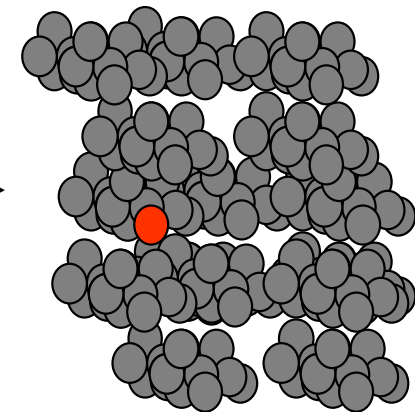
blastocysta



vyvíjející se tkáň (orgán)

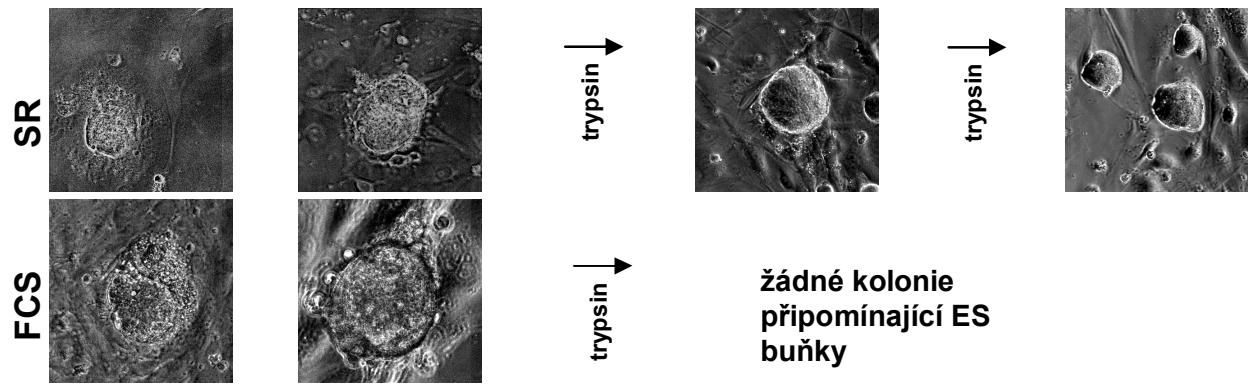
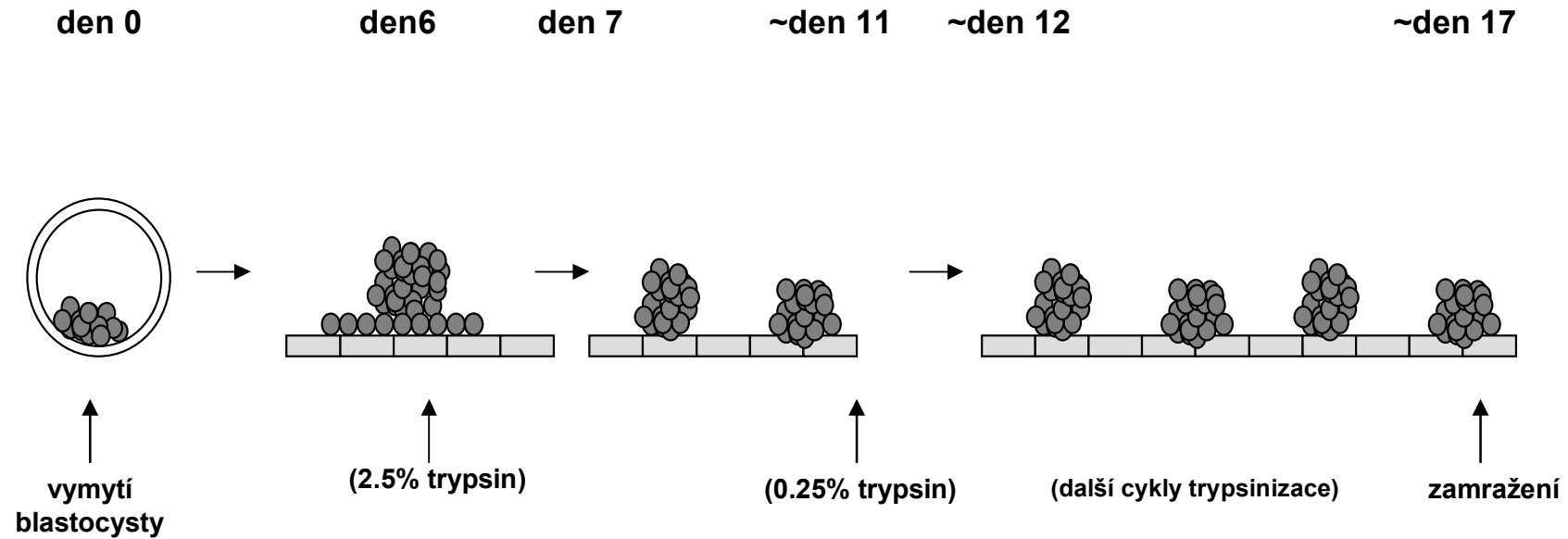


dospělá tkáň

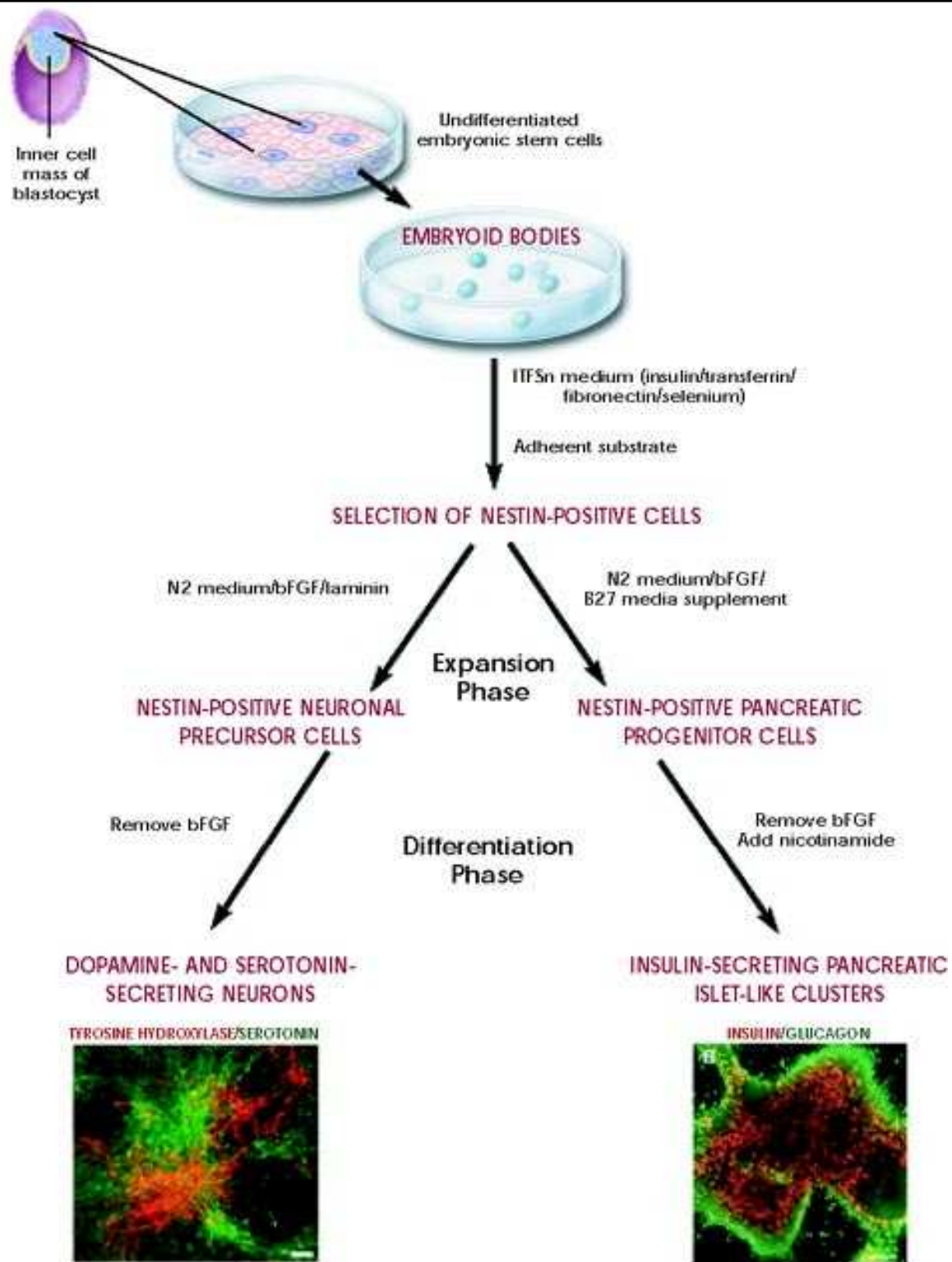


Příprava embryonálních kmenových buněk:

Bryja V. et al.: An efficient method for the derivation of mouse embryonic stem cells. Stem Cells, April 2006)





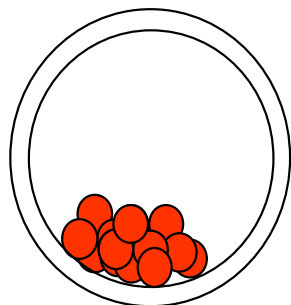


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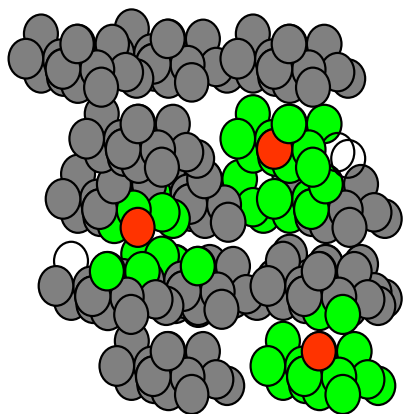
embryonální vývoj

In vivo

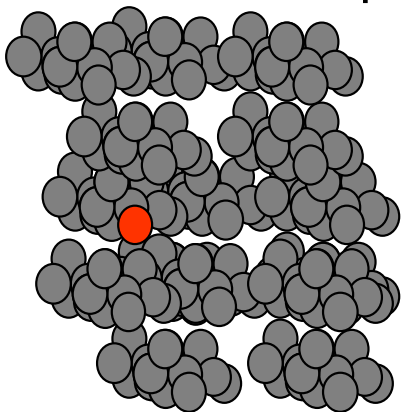


blastocysta

vyvíjející se tkáň (orgán)



dospělá tkáň



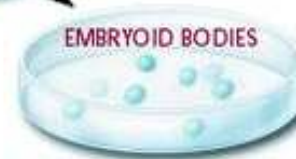
In vitro



Inner cell mass of blastocyst



Undifferentiated embryonic stem cells



EMBRYOID BODIES

IIFSn medium (insulin/transferrin/fibronectin/selenium)

Adherent substrate

SELECTION OF NESTIN-POSITIVE CELLS

N2 medium/bFGF/taminin

N2 medium/bFGF/B27 media supplement

Expansion Phase

NESTIN-POSITIVE NEURONAL PRECURSOR CELLS

NESTIN-POSITIVE PANCREATIC PROGENITOR CELLS

Remove bFGF

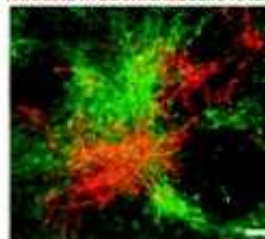
Differentiation Phase

Remove bFGF Add nicotinamide

DOPAMINE- AND SEROTONIN-SECRETING NEURONS

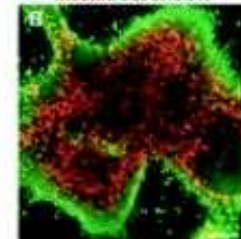
INSULIN-SECRETING PANCREATIC ISLET-LIKE CLUSTERS

TYROSINE HYDROXYLASE/SEROTONIN

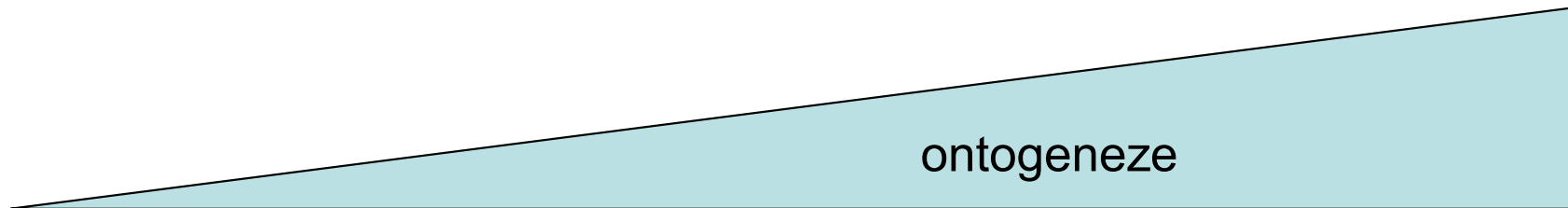


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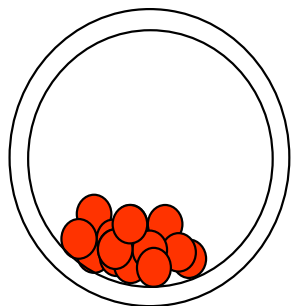
INSULIN/GLICAGON



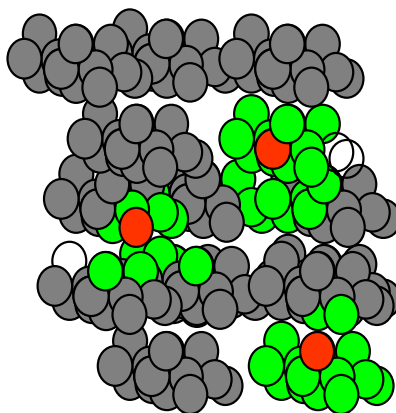
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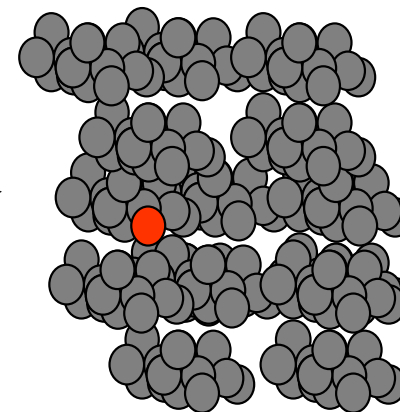
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vyvíjející se tkáň (orgán)

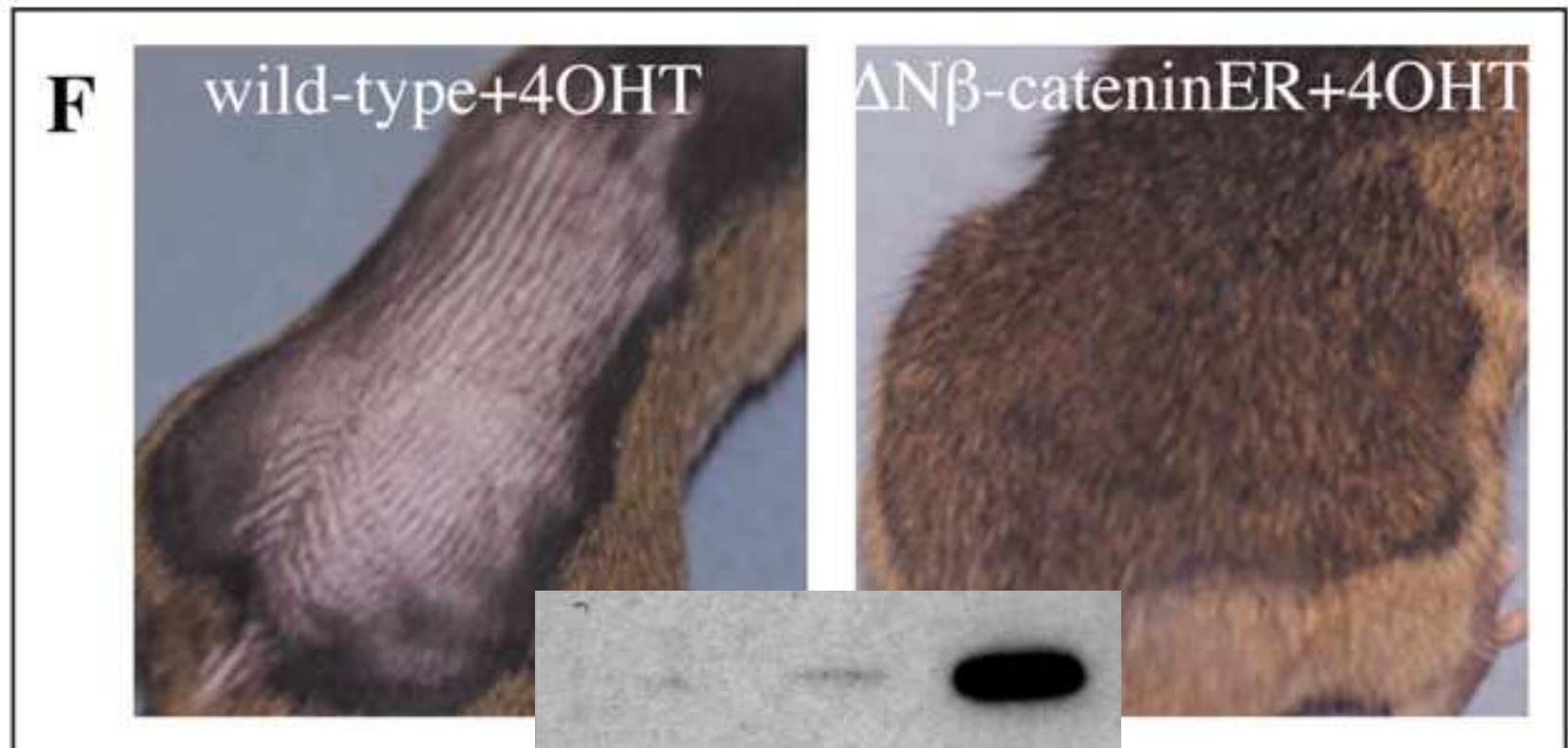


dospělá tkáň





Klíčové regulátory (Velká Pětka): Wnt, Shh, BMP/TGF, RTK (FGF), Notch



**Kde jsou adult stem cells a jak
vypadají?**

Prostředí kmenových buněk (stem cell niche)

vlasový kořínek

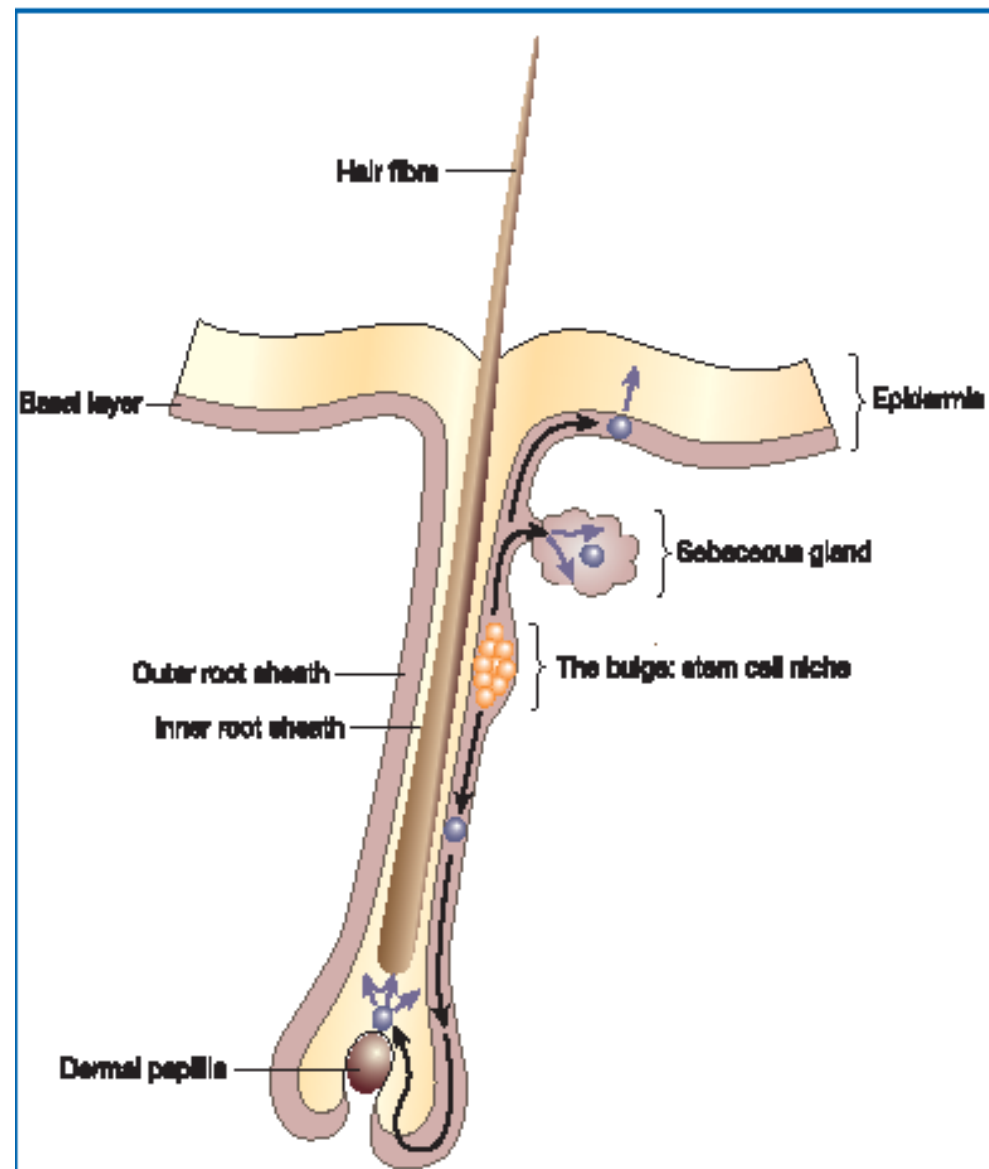


Figure 4 The hair follicle. Stem cells reside in the bulge niche. Cells can migrate upwards from here to populate the sebaceous gland and the interfollicular epidermis. Cells that migrate downwards enter the matrix where they rapidly proliferate and then differentiate to form the hair. (Adapted from ref. 90.)

Reya & Clevers 2005, Nature

Prostředí kmenových buněk (stem cell niche)

kostní dřeň

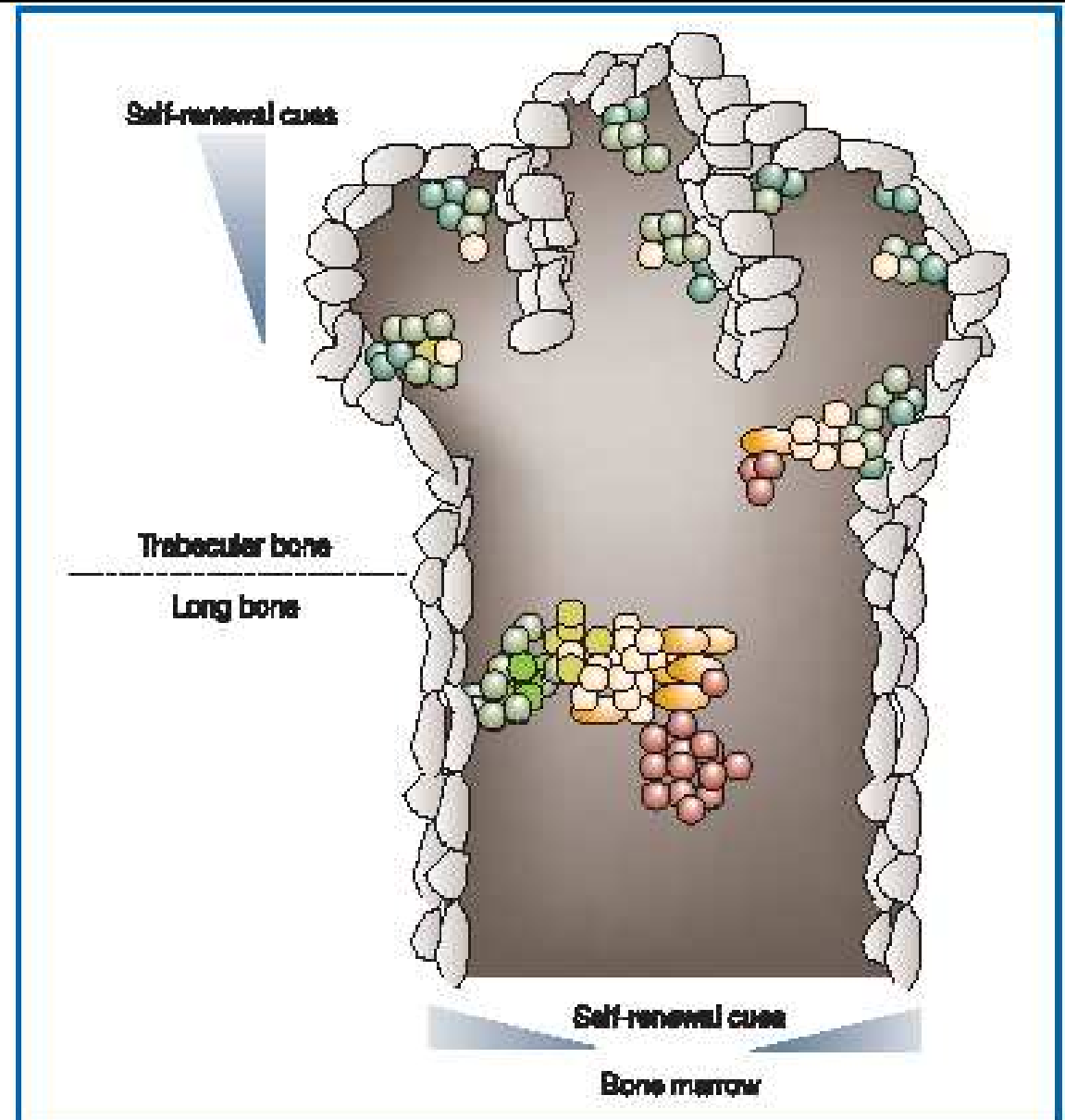
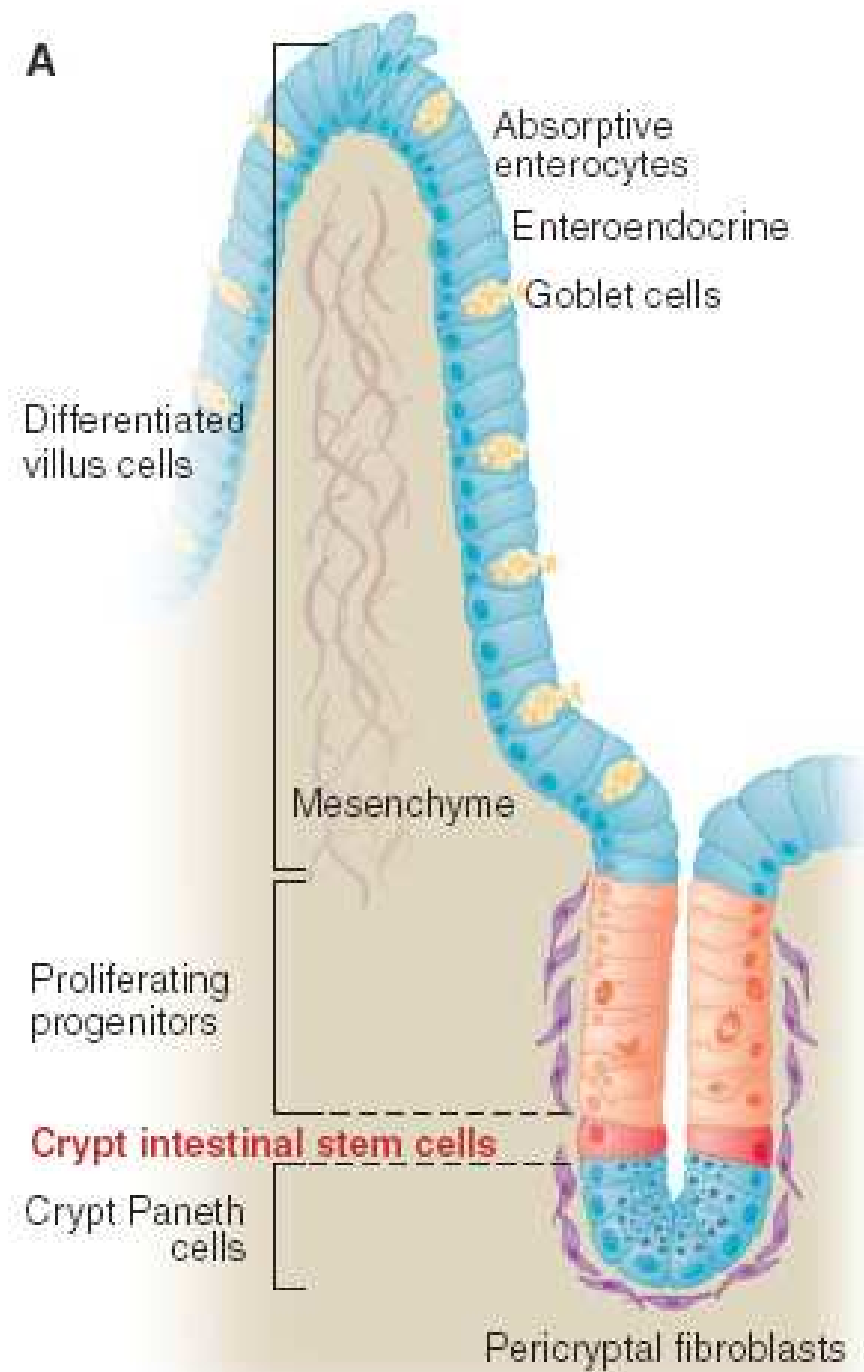


Figure 5 Proposed model of HSC development in the niche. HSCs are shown (dark green) at the endosteal marrow adjacent to the bone's surface mainly at the trabecular bone, and are postulated to migrate inward in the central marrow as they differentiate (precursors in light green; differentiated cells in yellow, orange and red) away from a possible gradient of self-renewal cues. (Adapted from ref. 44.)

Reya & Clevers 2005, Nature

Prostředí kmenových buněk (stem cell niche)

střevní epitel



Prostředí kmenových buněk (stem cell niche)

střevní epitel – latest developments
aneb jak opravdu na to (Barker et al.,
Nature, October 2007)

A. Příprava transgenní myši č. 1 za
účelem zjistit, kde je nový potenciální
stem cell marker exprimován (in vivo
expression profiling). *Lgr5* je
exprimován specificky v buňkách ve
spodní části krypty.

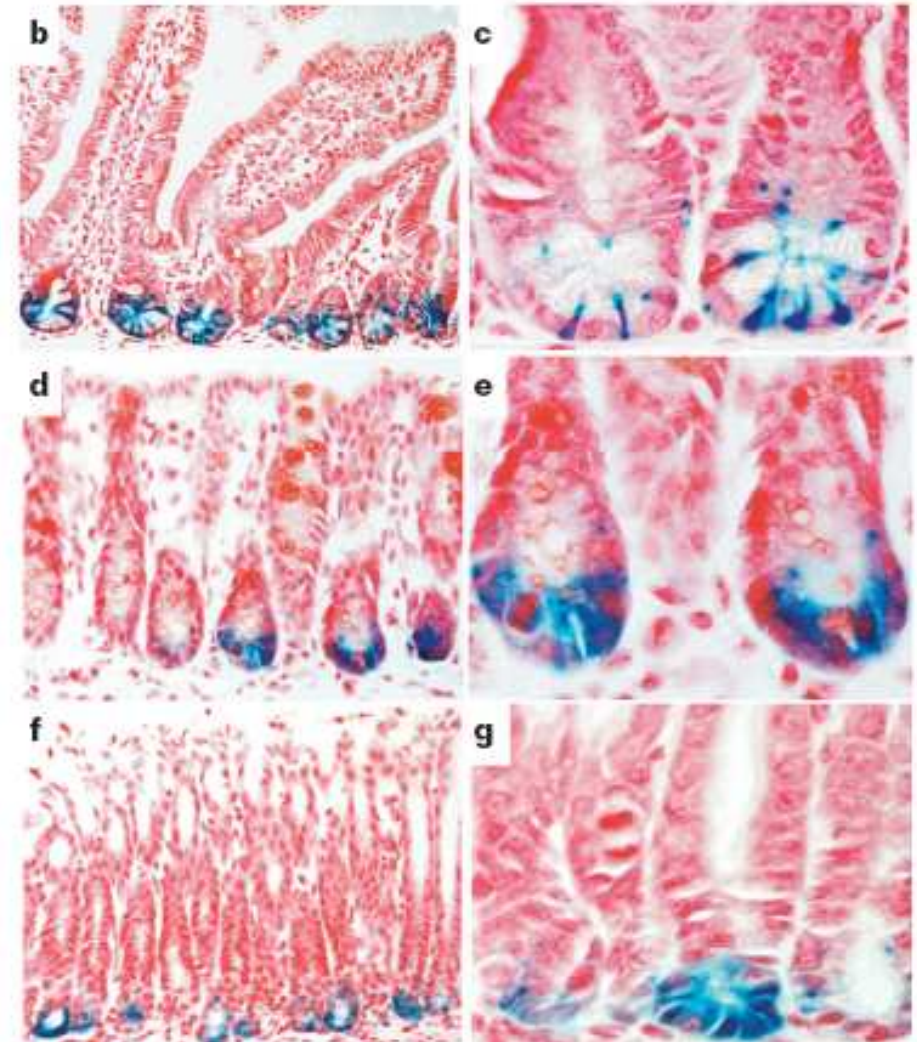
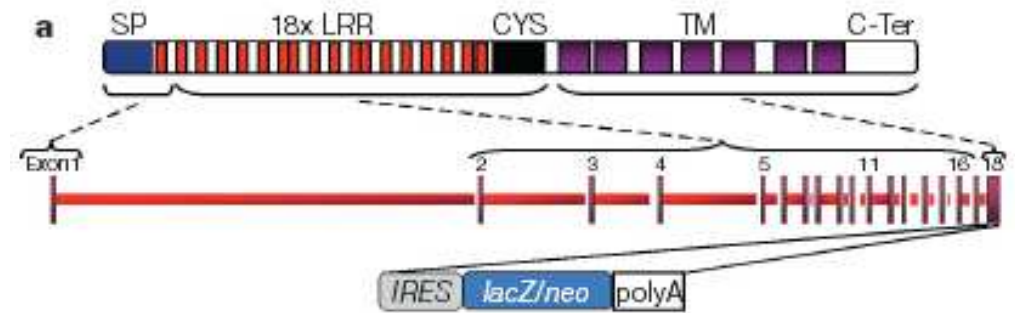


Figure 3 | Restricted expression of an *Lgr5-lacZ* reporter gene in adult mice. **a**, Generation of mice carrying *lacZ* integrated into the last exon of the *Lgr5* gene, removing all transmembrane (TM) regions of the encoded *Lgr5* protein. Neo, neomycin resistance cassette; SP, signal peptide; LRR, leucine-rich repeat region; C-Ter is carboxy terminus. **b–h**, Expression of *Lgr5-LacZ* (blue) in selected adult mouse tissues. **b, c**, In the small intestine, expression is restricted to six to eight slender cells intermingled with the Paneth cells at the crypt base. **d, e**, In the colon, expression is confined to a few cells located at the crypt base. **f, g**, Expression in the stomach is limited to the base of the glands.

Prostředí kmenových buněk (stem cell niche)

střevní epitel – latest developments
aneb jak opravdu na to (Barker et al.,
Nature, October 2007)

B. Příprava transgenní myši 2, 3 a 4
za účelem zjistit, co všechno vzniká
z Lgr5-pozitivních buněk (Lgr5+
lineage tracing). Lgr5 pozitivní buňky
dávají vzniknout všem částem
buněčného epitelu.

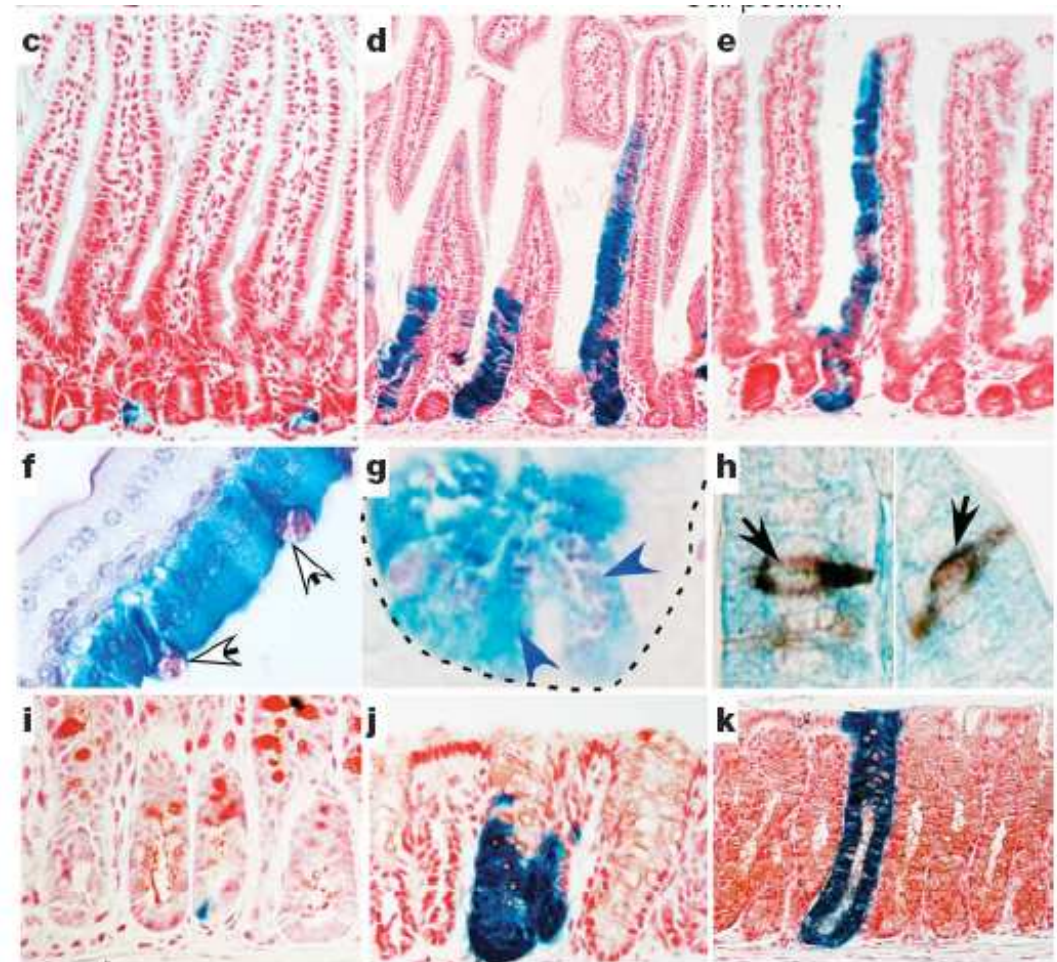
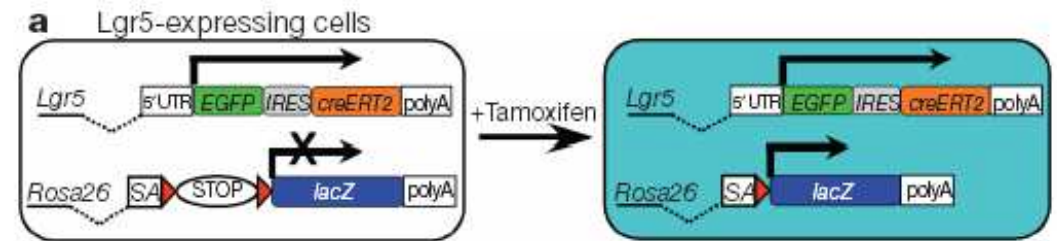


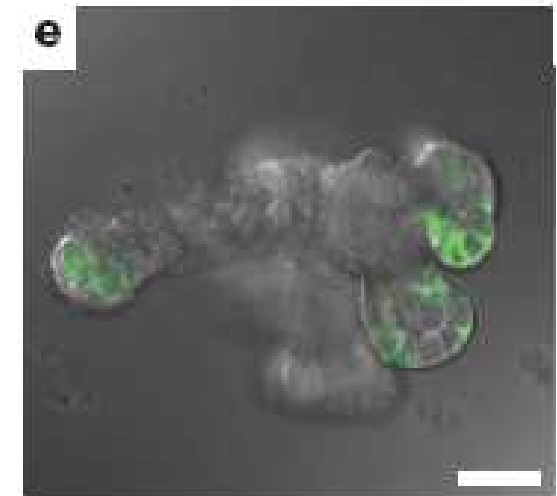
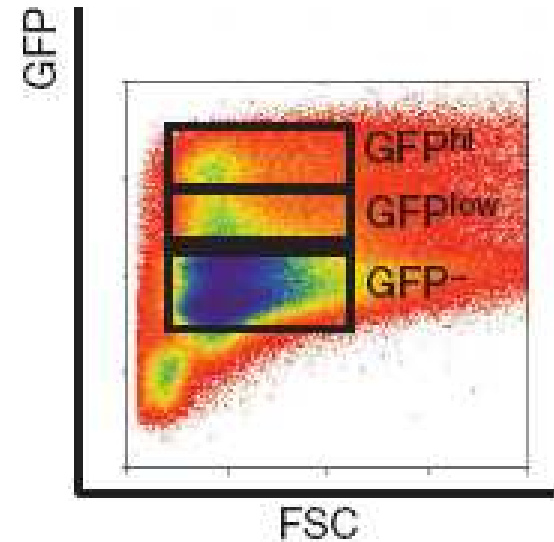
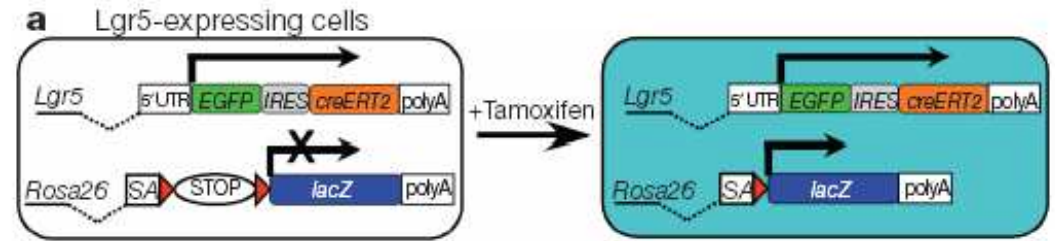
Figure 5 | Lineage tracing in the small intestine and colon. a, *Lgr5-EGFP-IRES-creERT2* knock-in mouse crossed with *Rosa26-lacZ* reporter mice 12 h after tamoxifen injection. b, Frequency at which the blue cells appeared at

carrying activated Cre. c–e, Histological analysis of LacZ activity in small intestine 1 day after induction (c), 5 days after induction (d) and 60 days after induction (e). f–h, Double-labelling of LacZ-stained intestine using PAS demonstrates the presence of goblet cells (f, white arrows) and Paneth cells (g, blue arrows) in induced blue clones. Double-labelling with synaptophysin demonstrates the presence of enteroendocrine cells within the induced blue clones (h, black arrows). i–k, Histological analysis of LacZ activity in colon 1 day after induction (i), 5 days after induction (j) and 60 days after induction (k).

Prostředí kmenových buněk (stem cell niche)

střevní epitel – latest developments
aneb jak opravdu na to (Barker et al.,
Nature & Sato, Nature 2009)

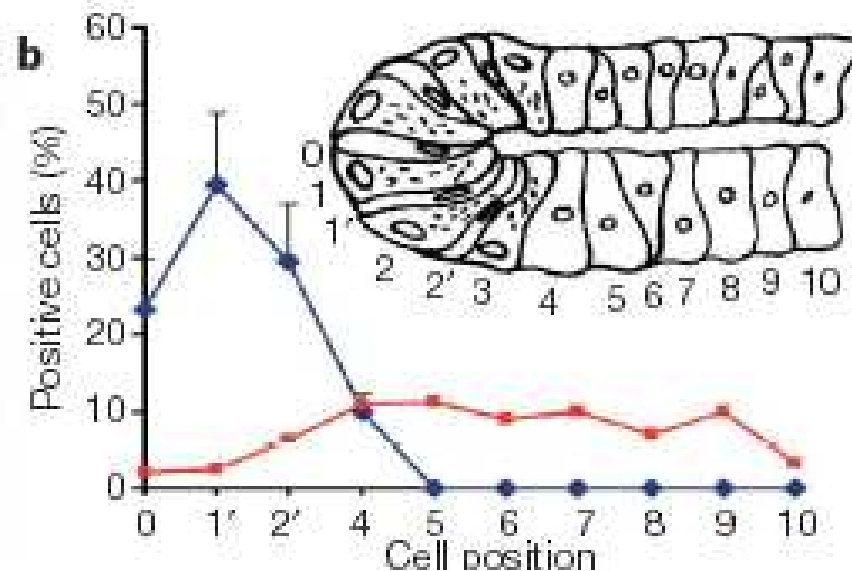
C. Lgr5 pozitivní buňky in vitro dávají
vzniknout kompletní villus-crypt
strukturu in vitro (Doposud se to s
žádnými jinými buňkami nepodařilo)



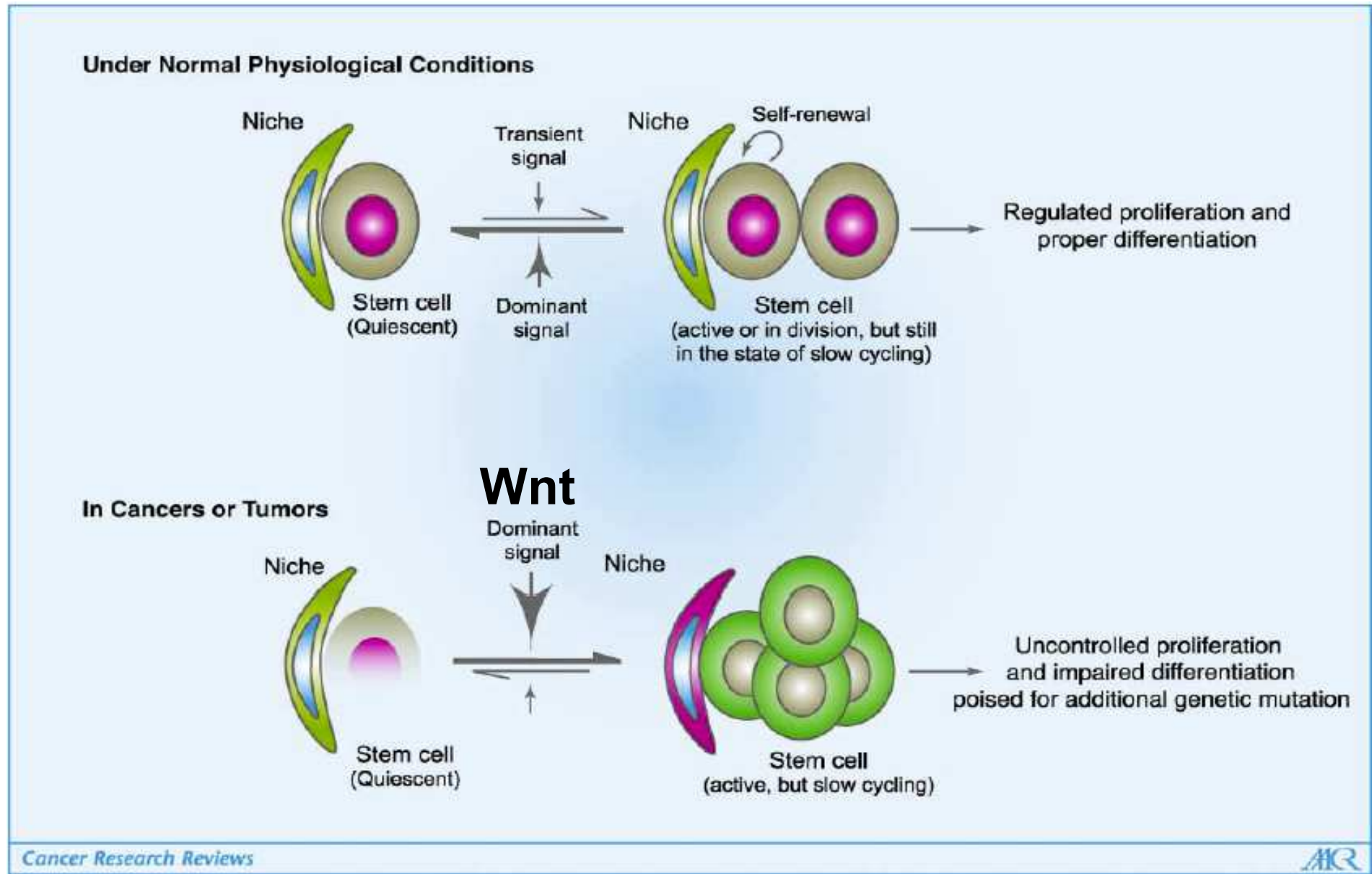
Prostředí kmenových buněk (stem cell niche)

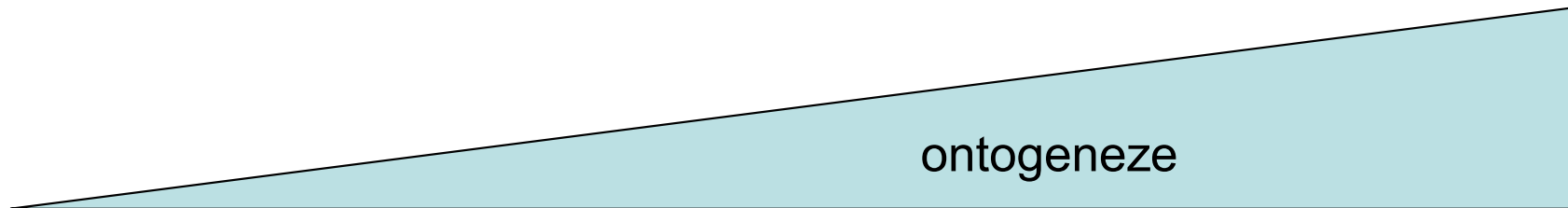
**střevní epitel – latest developments
aneb jak opravdu na to (Barker et al.,
Nature, October 2007)**

**D. Závěr: Kmenové buňky epitelu
tlustého i tenkého střeva jsou
protáhlé, dříve nepovšimnuté buňky,
v relativní pozici 1', 2' a 3' od spodu
krypty. Všechno co se říkalo předtím
byly jen žvásty 😊**

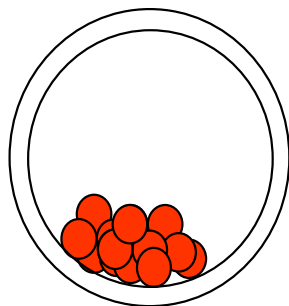


Prostředí kmenových buněk (stem cell niche)

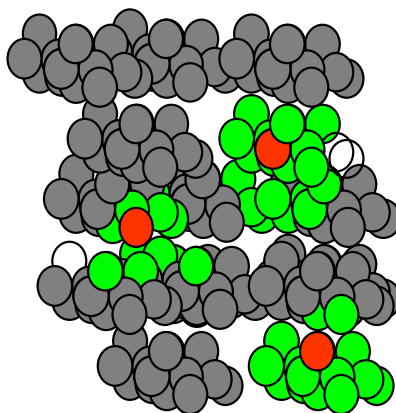




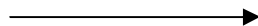
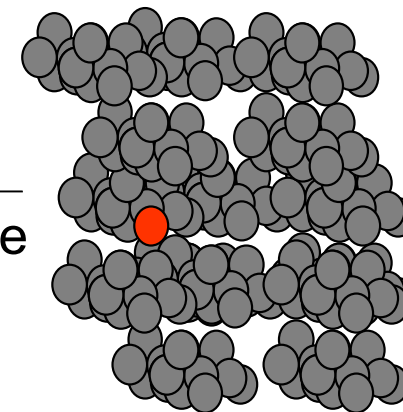
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vyvíjející se tkáň (orgán)

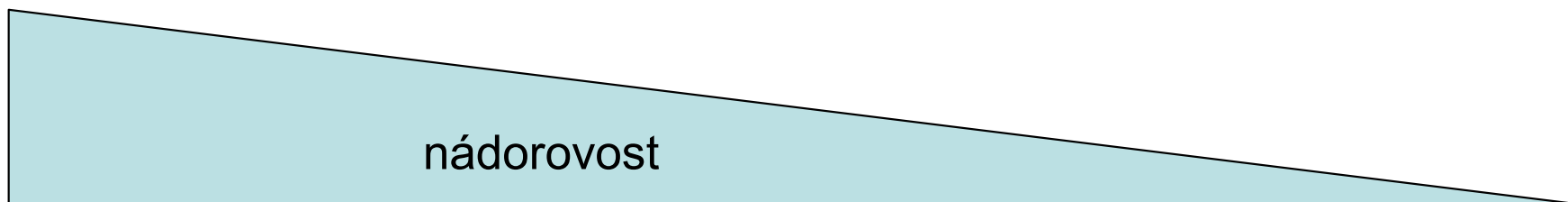


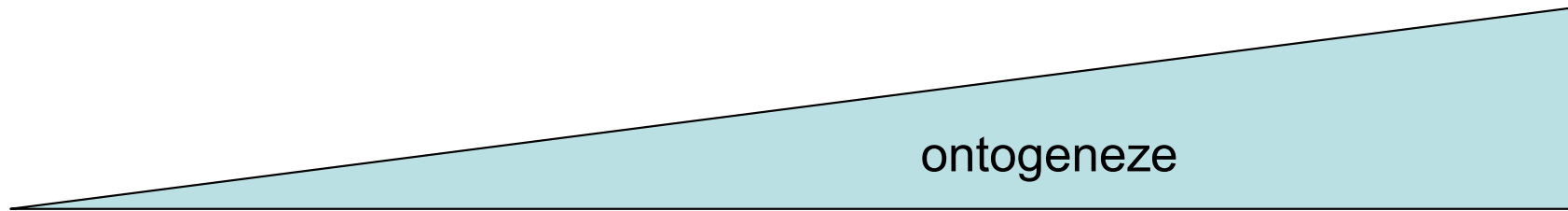
dospělá tkáň



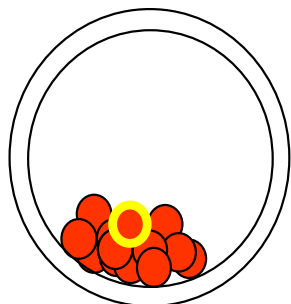
← regenerace

← tvorba nádoru

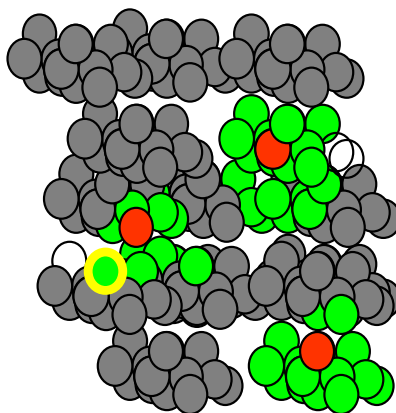




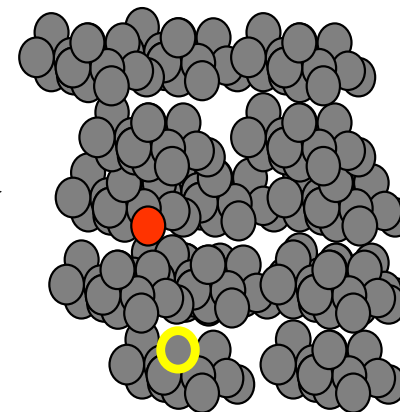
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vyvíjející se tkáň (orgán)

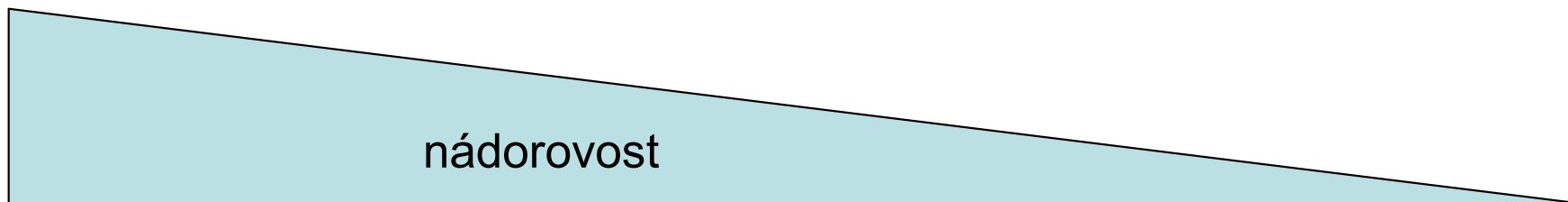


dospělá tkáň

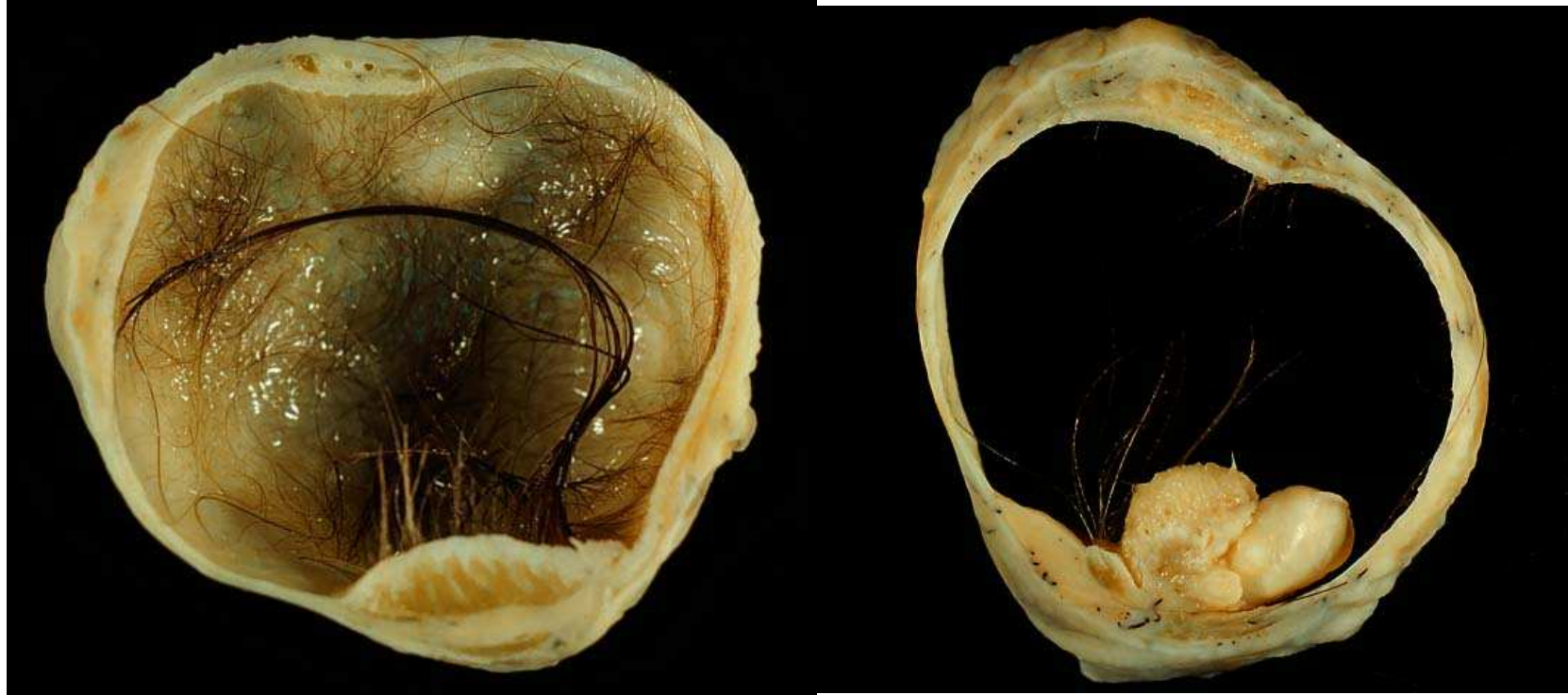


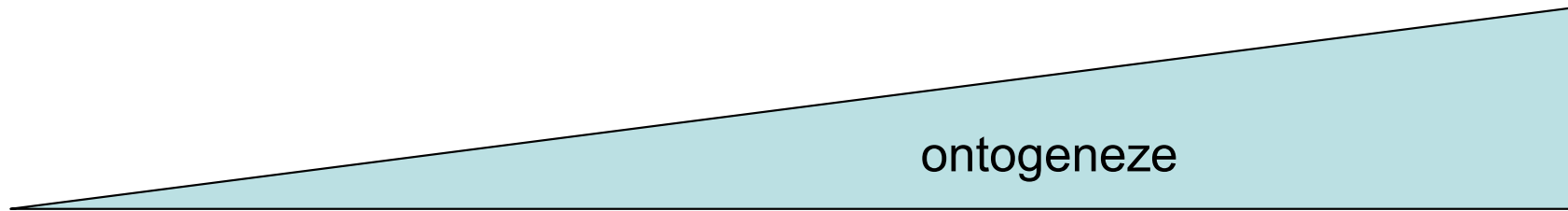
**Počet mutací nutných
pro vznik nádoru:**

0

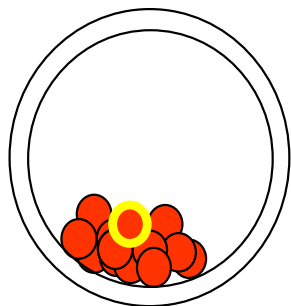


Teratoma

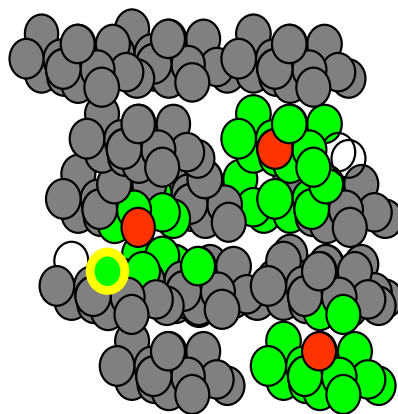




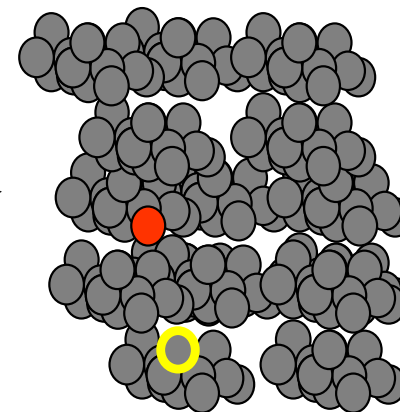
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vyvíjející se tkáň (orgán)



dospělá tkáň

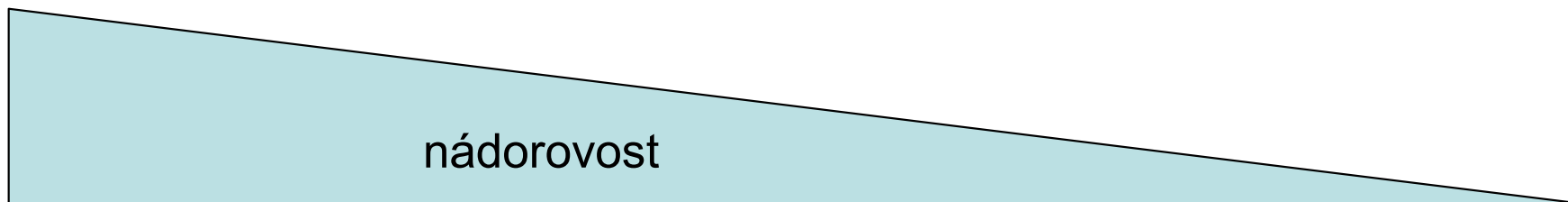


Počet mutací nutných
pro vznik nádoru:

0

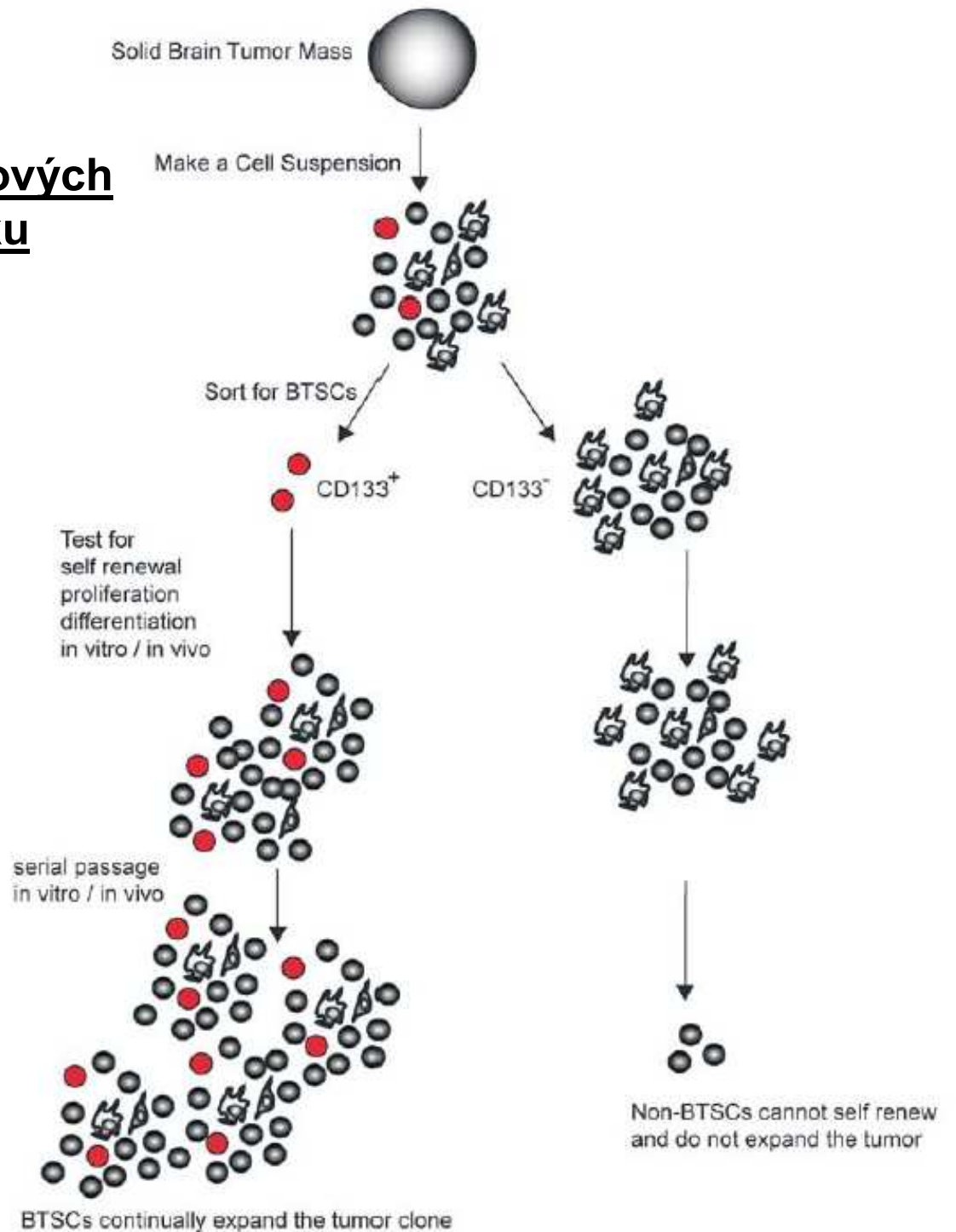
1-5

5 a více



Jak víme o nádorových kmenových
buňkách?

Izolace a identifikace nádorových kmenových buněk mozku



Aktivace nádorových kmenových buněk jako důsledek chronického poškození a regenerace

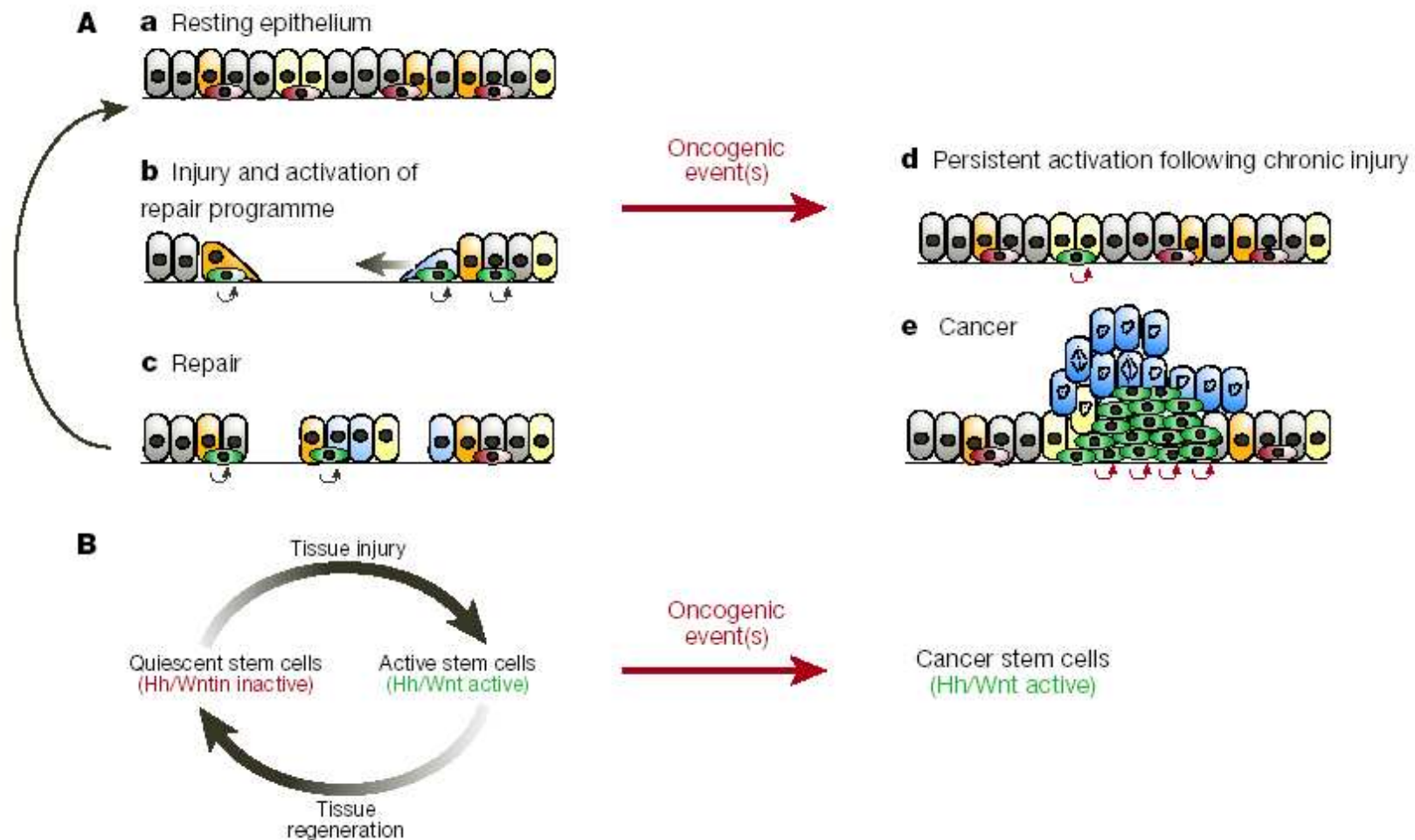


Figure 2 Model for carcinogenesis resulting from persistence of a state of injury repair.

A, Cellular events of epithelial repair. **a**, Resting epithelium with several differentiated cell phenotypes (brown, orange, and yellow) derived from tissue stem cells, now quiescent (red). Pathways such as Hh and Wnt signalling pathways that have a role in the renewal of stem cells are not active. **b**, Epithelial defect resulting from acute injury. Loss of epithelial continuity activates a repair program which is driven by Hh or Wnt signalling. This program results in the acquisition by epithelial cells of a more mesenchymal phenotype, including flattening and movement of cells (straight arrow) to cover the wound, activation (green), and expansion of stem cells through renewal divisions (curved arrows). **c**, The wound is repaired, first by rapid cell movement, and then by restoration of cell numbers resulting from the amplification of stem cells and

the differentiation of their progeny. Subsequently, either epithelial continuity and patterning is restored, Hh and Wnt signalling ceases, and the stem cell compartment returns to quiescence (**a**); or oncogenic event(s) may trap a stem cell in an activated state of continuous renewal, which is driven by autonomous Wnt or Hh signalling (**d**). Further genetic or epigenetic change in such a persistently activated stem cell (curved red arrows) might produce a cancer stem cell (green) which is capable of aggressively propagating a cancer (**e**). This may result from enhanced proliferation and production of more cancer stem cells as well as from differentiated cancer cells (blue). **B**, Stem cells cycle between quiescence and activity as a consequence of Hh/Wnt driven responses to injury. Oncogenic event(s) may trap activated stem cells in a permanent state of Hh/Wnt driven activity, resulting in cancer stem cells.

Klasické morfogenetické dráhy (Wnt, Hh, Notch a další) regulují regeneraci, tkáňové specifické kmenové buňky i nádory

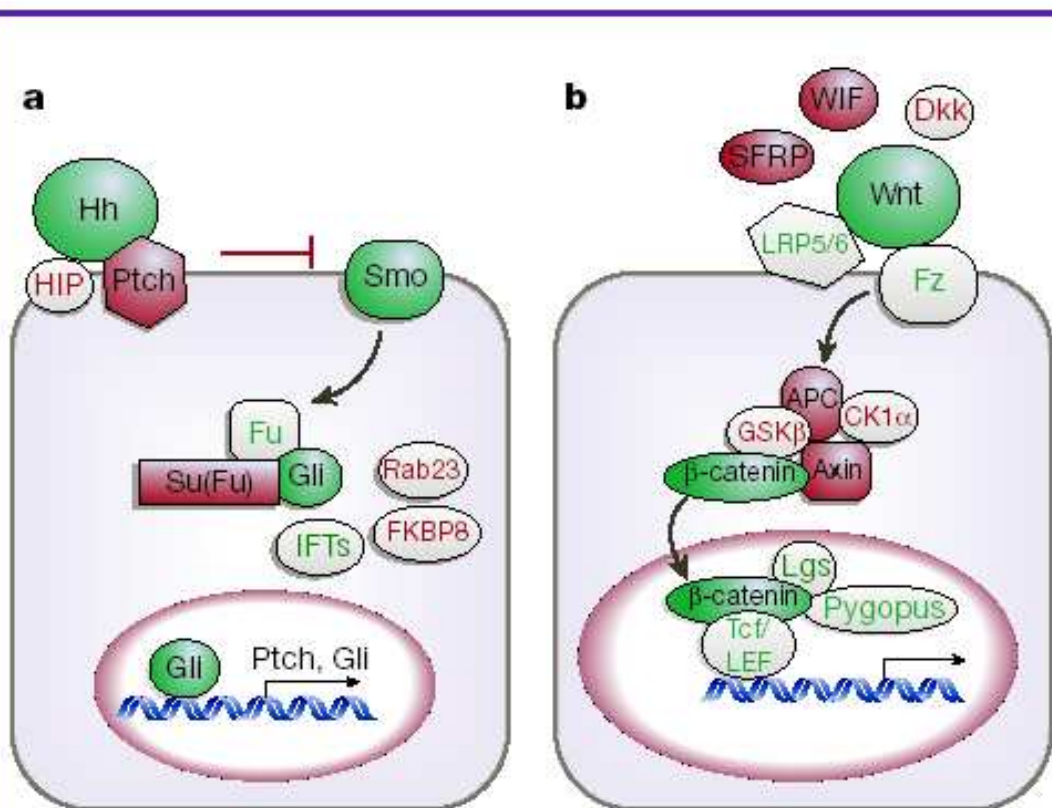


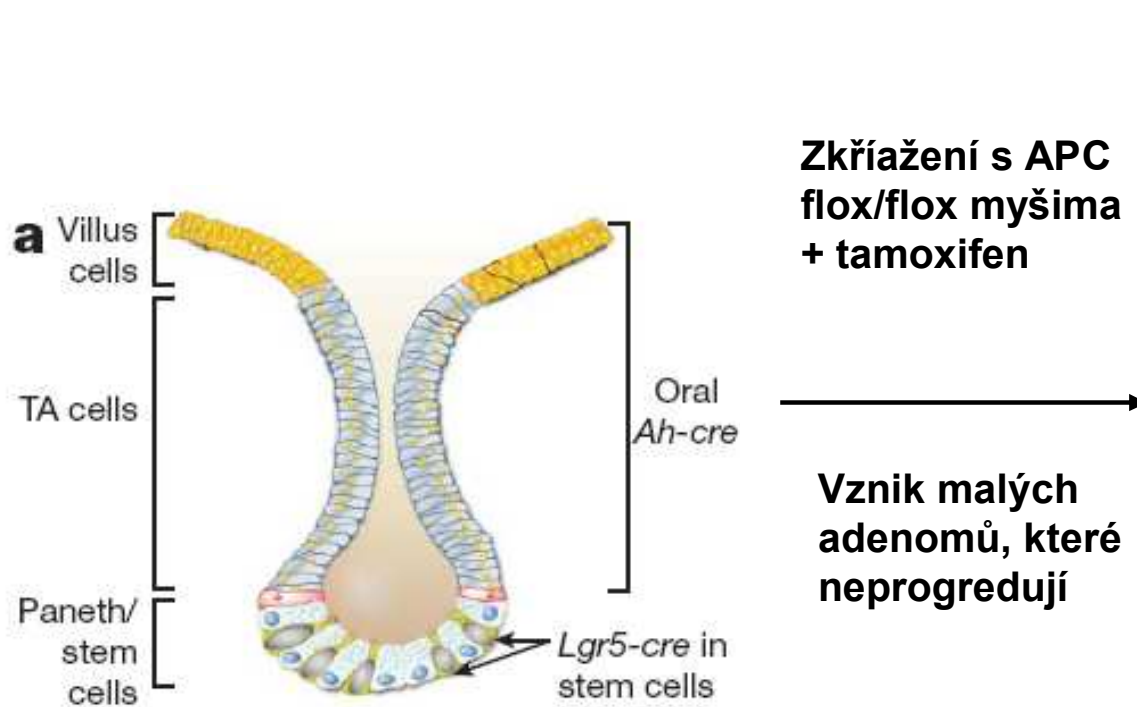
Figure 1 Hh and Wnt signalling pathways. Simplified views of the Hh and Wnt signalling pathways, with emphasis on components implicated in cancer or tissue regeneration. Green and red colours denote pathway components with primarily positive or negative roles, respectively, in pathway activation. Shaded components have been causally implicated in tumorigenesis (see Table 2 and text; more complete pathway descriptions are available in refs 32–34 for Hh and refs 17, 46 for Wnt). **a**, Activation of the Hh signalling pathway is initiated by binding of a Hh ligand to Ptch. This lifts suppression of Smo, activating a cascade that leads to the nuclear translocation of Gli and the activation of target genes. HIP is a membrane protein that antagonizes pathway activity by binding to Hh ligands, and Fu, Su(Fu), Rab23, FKBP8 and the IFTs (intraflagellar transport proteins) act downstream of Ptch and Smo to regulate Gli. The function of Rab23, FKBP8 and the IFTs outside the CNS is not established. HIP, Hh-interacting protein; Rab23, a member of the Rab family of GTPases; FKBP8, a member of the FK506-binding protein family. **b**, The Wnt signalling pathway is activated by binding of Wnt ligands to their receptors Fz and LRP5/6, leading to the release of β -catenin from the degradation complex and facilitating its entry into the nucleus, where it regulates target gene transcription through association with Tcf/LEF, Legless (Lgs) and Pygopus. SFRP, WIF and Dkk are secreted antagonists of Wnt signalling. APC, Axin, GSK3 β and CK1 α are components of the β -catenin degradation complex. WIF, Wnt inhibitory factor; Dkk, Dickkopf; GSK3 β , glycogen synthase kinase 3 β ; CK1 α , casein kinase 1a.

Table 2 **Hh and Wnt pathways in cancer**

Tissue	Tumour	Evidence of pathway involvement	References
Hh pathway			
Brain	Medulloblastoma	Tumorigenesis by inactivation of <i>PTCH</i> ; allograft and cell-line growth inhibition by cyclopamine; inhibition of autochthonous tumour growth by synthetic small molecule antagonist	37, 81; reviewed in 6
		Tumorigenesis by inactivation of <i>Su(fu)</i>	86
	Glioma	Gli amplification; growth inhibition of some cell lines by cyclopamine	87, 88
Skin	Basal cell carcinoma	Tumorigenesis by inactivation of <i>PTCH</i> ; <i>in vivo</i> tumorigenesis by expression of activating form of <i>SMO</i> or by <i>Shh</i> overexpression and <i>in vitro</i> growth inhibition by synthetic Hh pathway antagonist; inhibition of human tumour growth topical cyclopamine	82, 83; reviewed in 6
Muscle	Rhabdomyosarcoma	Tumorigenesis by inactivation of <i>PTCH</i>	reviewed in 6
Oesophagus	Adenocarcinoma	Cell-line growth inhibition by cyclopamine, Hh blocking antibody	42
Stomach	Adenocarcinoma	Cell-line growth inhibition by cyclopamine, Hh blocking antibody	42
Pancreas	Adenocarcinoma	Xenograft and cell-line growth inhibition by cyclopamine, Hh blocking antibody; tumour initiation (in mouse) by <i>Shh</i> overexpression	42, 43
Biliary tract	Adenocarcinoma	Xenograft and cell-line growth inhibition by cyclopamine, Hh blocking antibody	42
Lung	Small-cell lung cancer	Xenograft and cell-line growth inhibition by cyclopamine, Hh blocking antibody	41
Prostate	Adenocarcinoma	Xenograft and cell-line growth inhibition and suppression of metastasis by cyclopamine; increased xenograft growth by <i>Shh</i> and <i>Gli</i> overexpression	29, 89, 90
Bladder	Urothelial carcinoma	Increased tumour induction (in mouse) by alkylating agent in <i>Ptch</i> heterozygote	91
Oral cavity	Squamous cell cancer	Growth inhibition of cell lines by cyclopamine;	92
Wnt pathway			
Colon	Adenocarcinoma	Tumorigenesis by inactivation of APC, Axin; tumorigenesis by stabilization of β -catenin; epigenetic inactivation of SFRPs	47; reviewed in 45
Liver	Hepatoblastoma	Tumorigenesis (in mouse) by inactivation of APC and by stabilization of β -catenin	reviewed in 45
Blood	Multiple myeloma	Cell-growth inhibition by dominant negative TCF4; growth stimulation by Wnt ligand	93
Hair follicle	Pilomatricoma	Tumorigenesis (in mouse) by overexpression of β -catenin	reviewed in 45
Bone	Osteosarcoma	<i>Dkk3</i> and <i>LRP5</i> expression inhibits tumour cell growth <i>in vitro</i>	94, 95
Lung	Non-small-cell carcinoma	Apoptosis and cell-growth inhibition by short interfering RNA and a blocking antibody against Wnt2	96
Pleura	Mesothelioma	Apoptosis and cell-growth inhibition by transfection of SFRP	97

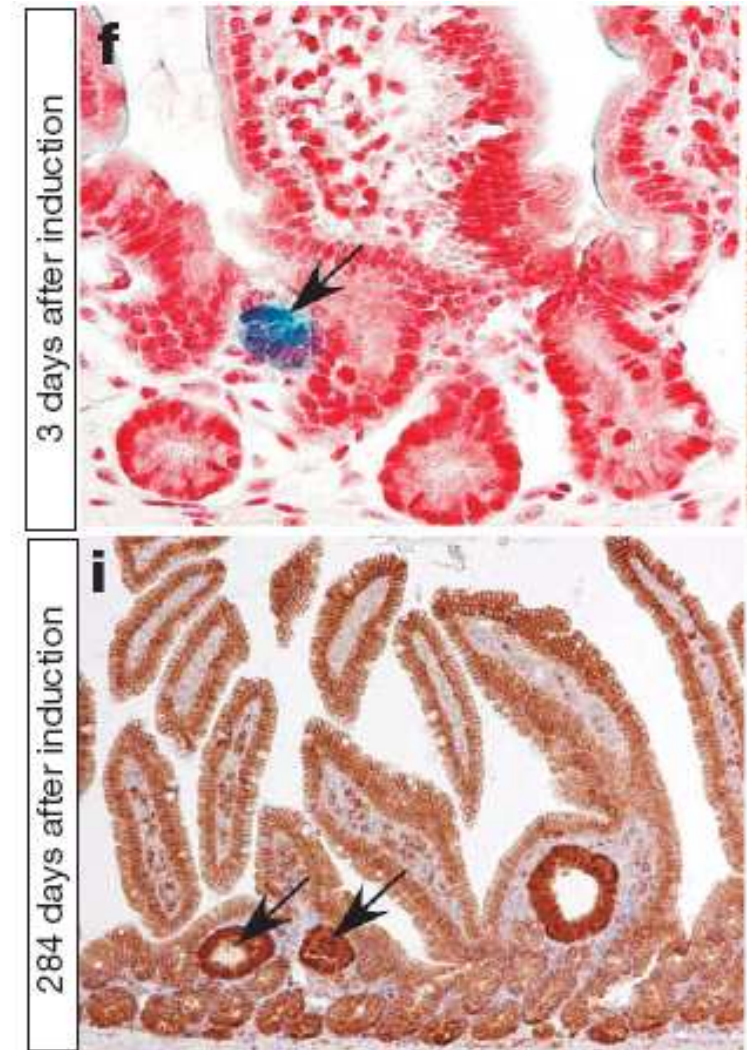
Emphasis is placed on functional data showing a requirement for pathway activation in tumour formation and/or tumour cell growth. (See Fig. 1 and text for gene abbreviations.)

Nekontrolovaná aktivace kmenových buněk má fatální následky

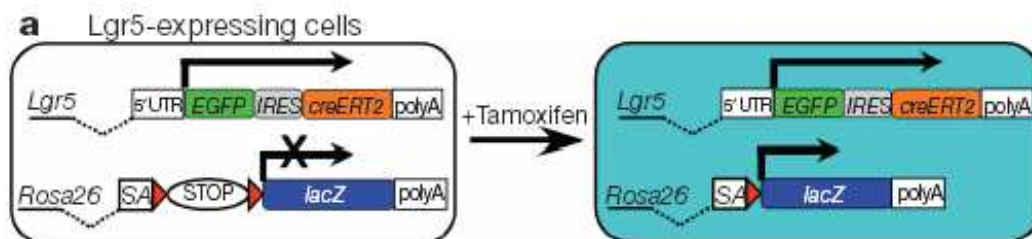


Zkřížení s APC flox/flox myšima + tamoxifen

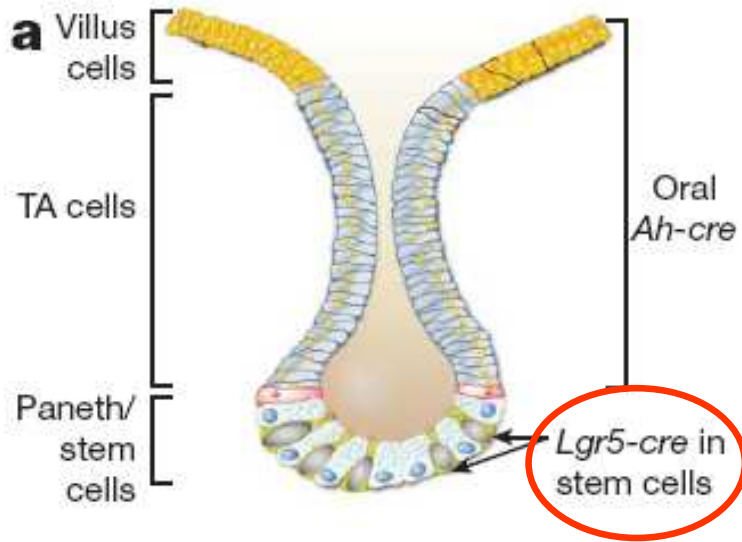
Vznik malých adenomů, které neprogredují



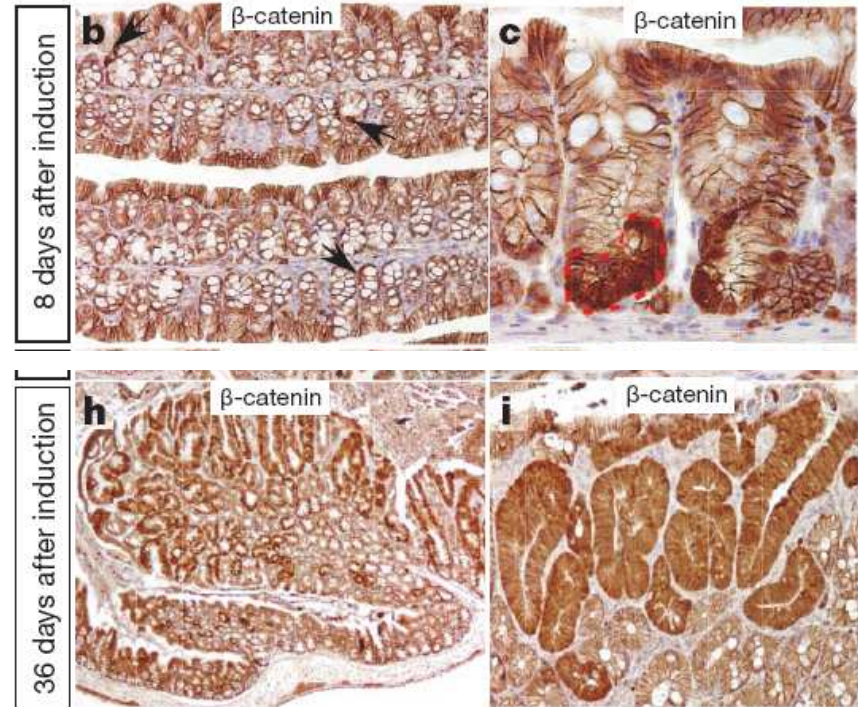
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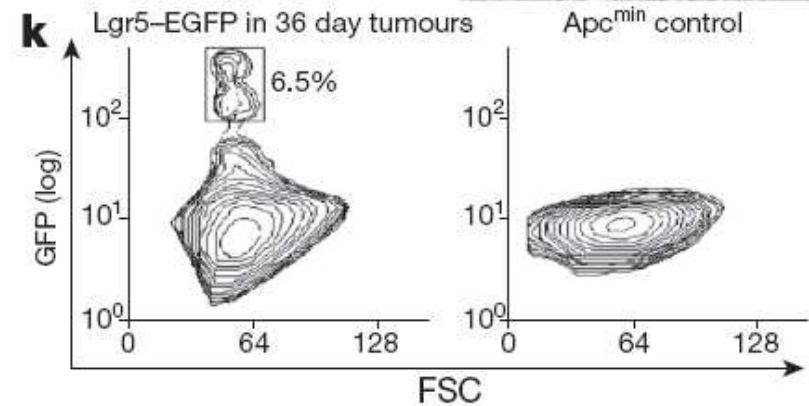
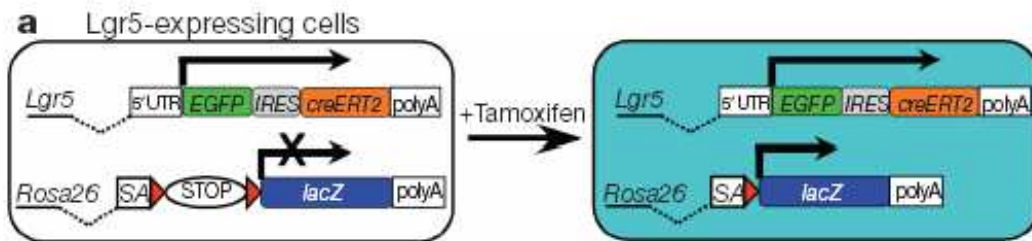
Nekontrolovaná aktivace kmenových buněk má fatální následky



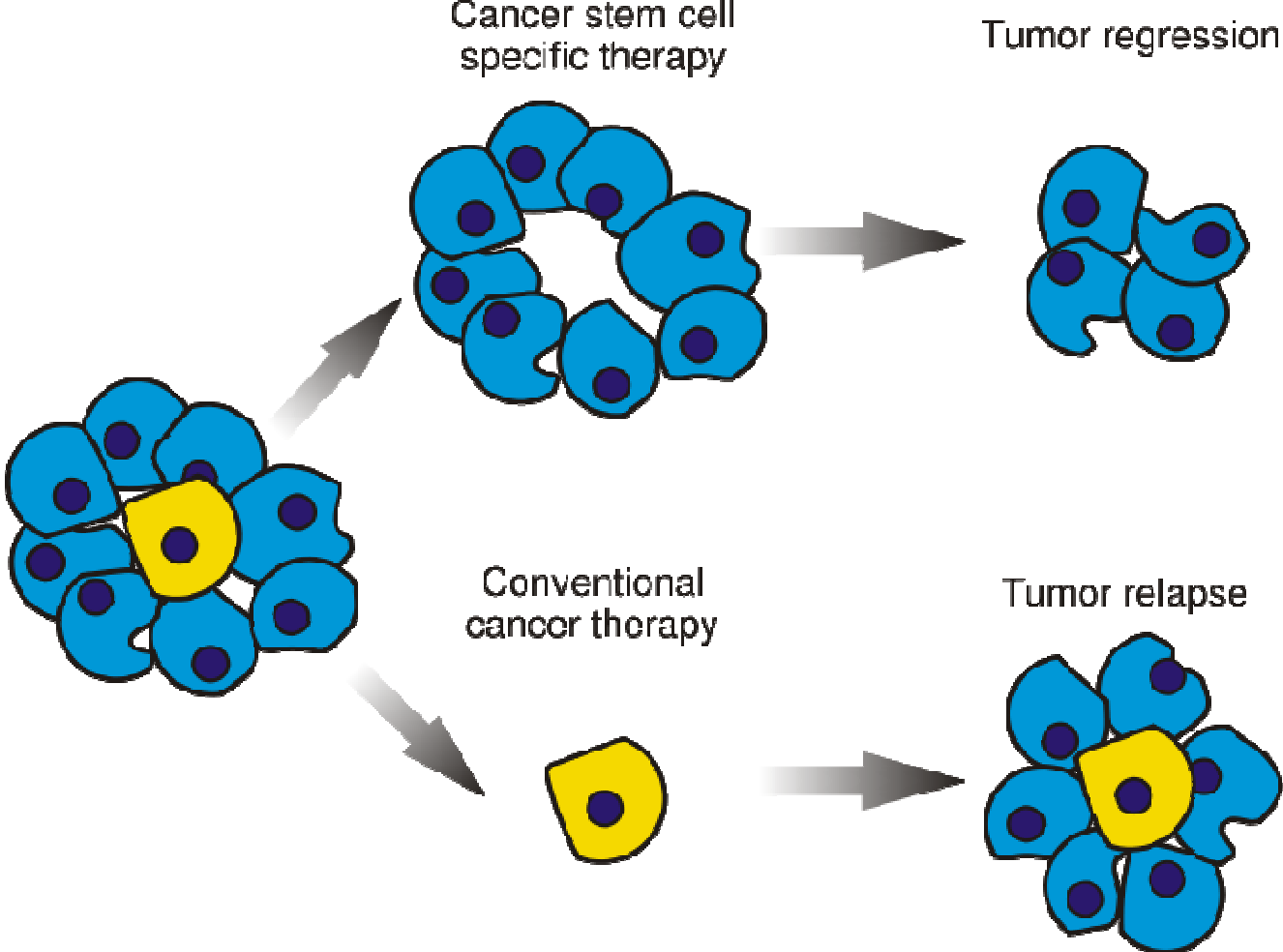
Zkřížení s APC flox/flox myšima + tamoxifen
Rychlý vznik adenomů a nádorů



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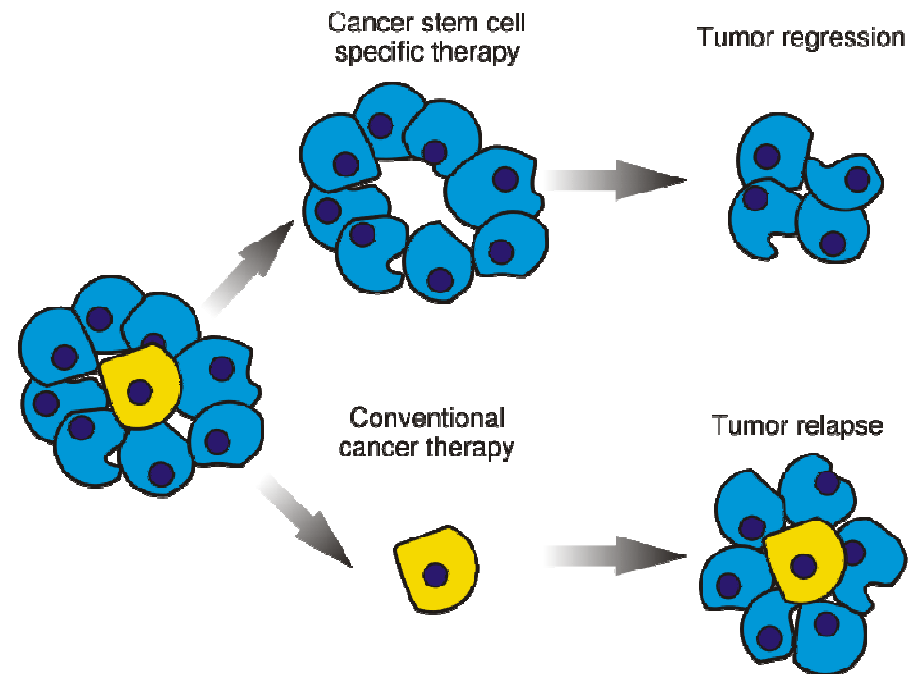


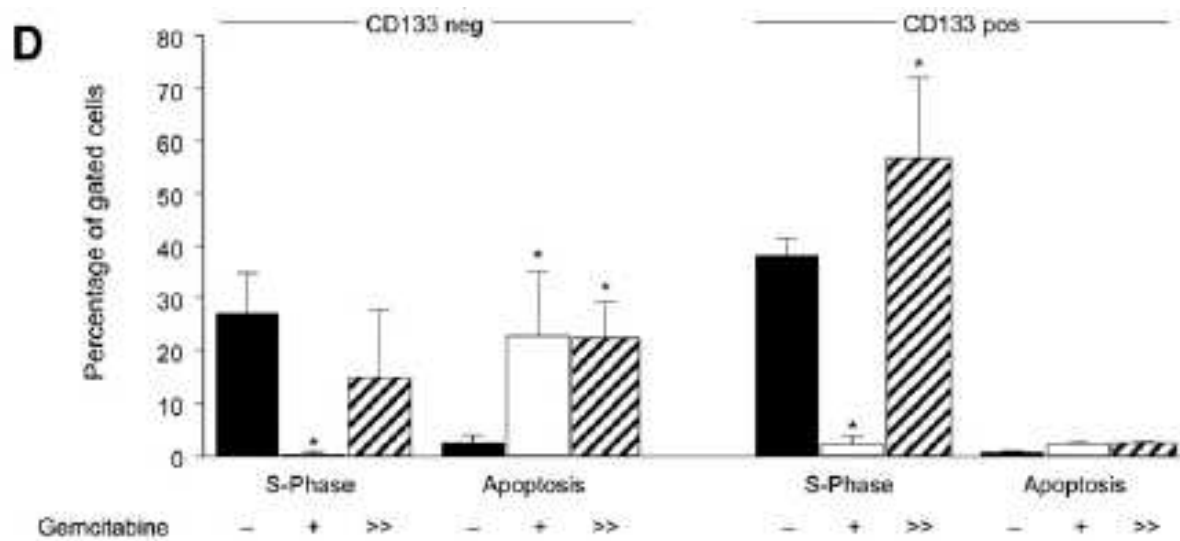
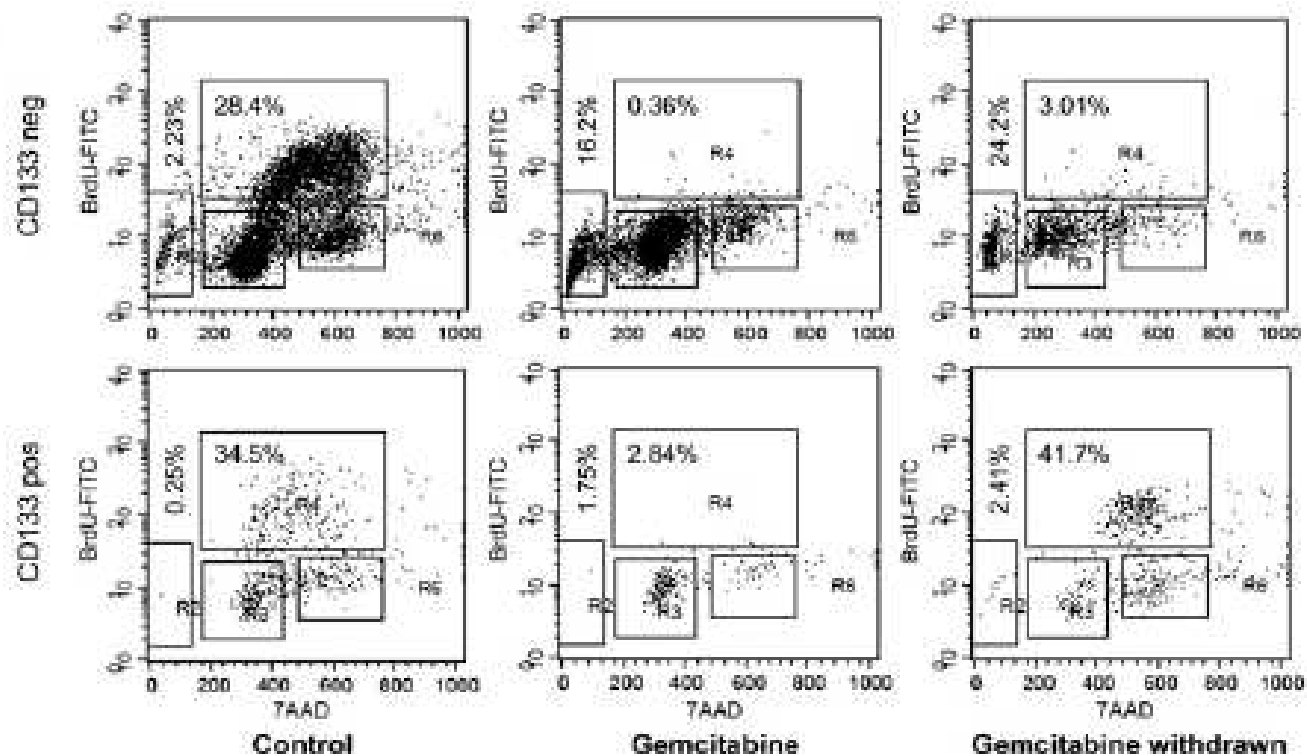
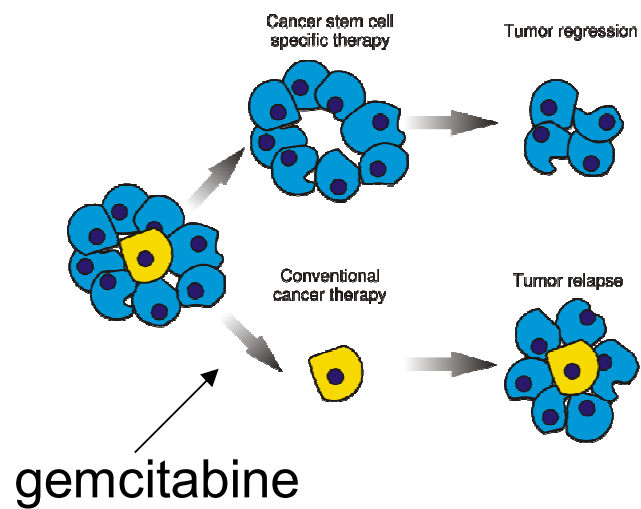
Cancer stem cell based therapy



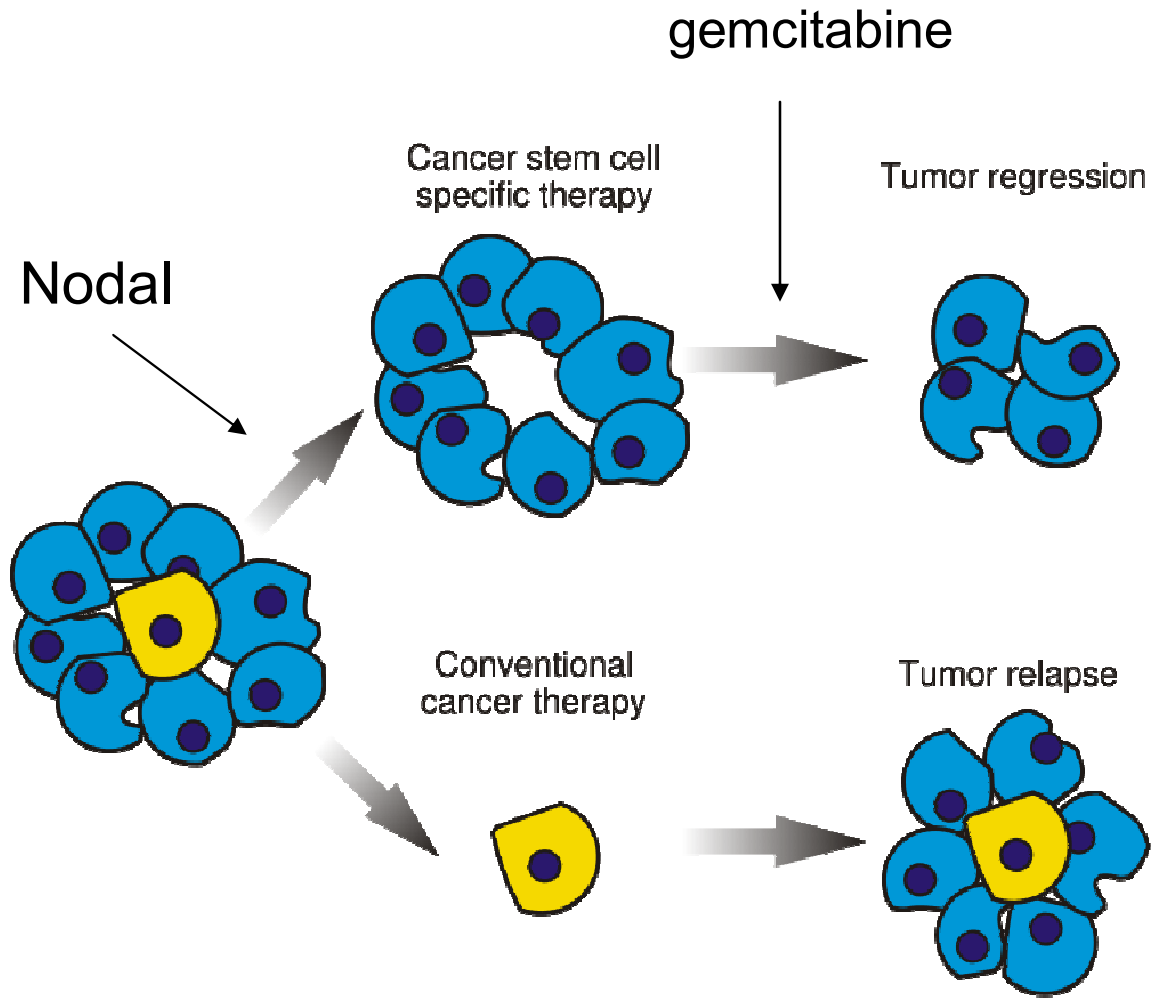
Cancer stem cell based therapy - naděje pro nemocné s nádorem slinivky?

- Pancreatic adenocarcinoma - čtvrtý nejčastější důvod úmrtí u pacientů nádorových onemocnění
- Median survival - 4-6 months
- 5-year survival - 1%
- Treated with gemcitabine - does not work

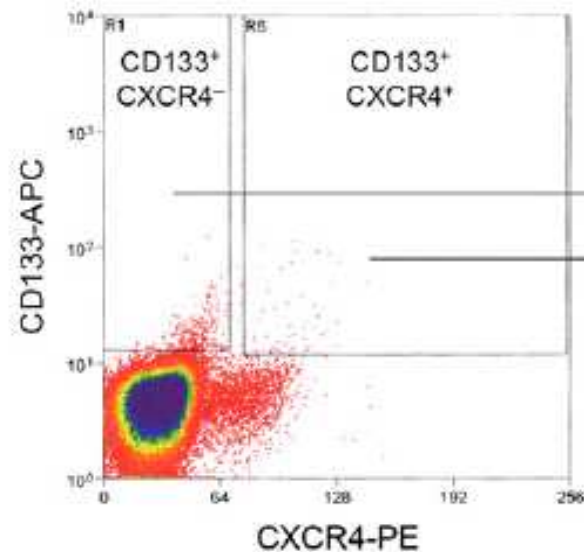




Terapie budoucnosti?



Metastasis driving cancer stem cells



Isolation of

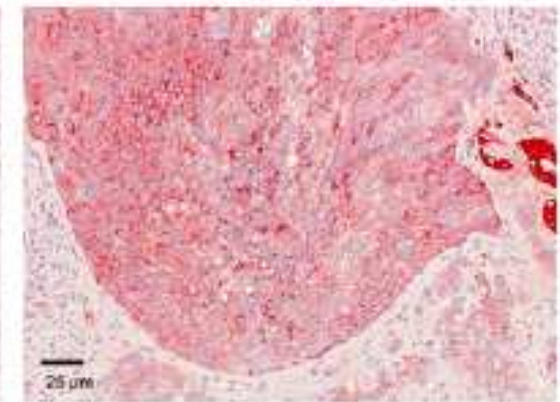
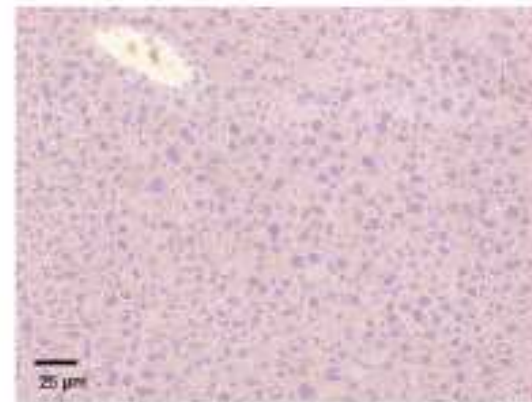
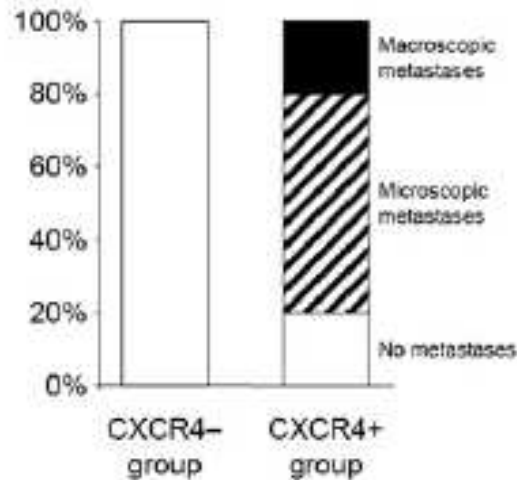
Noninvasive CD133⁺ CXCR4⁻

Highly invasive CD133⁺ CXCR4⁺

CXCR4⁻ group
(10³ cells)

CXCR4⁺ group
(each population 5x10² cells)

D



CXCR4⁻ group

CXCR4⁺ group

SDF1/CXCL12 - chemokin, ligand exprimovaný v plicích, játrech, kostní dřeni a lymfatických uzlinách; tj. hlavních cílech metastáz