

Image processing workflows for Tomography

Winter School on Structural Cell Biology

Julio Ortiz

Wolfgang Baumeister

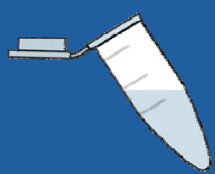
MPI of Biochemistry

Dept. of Structural Molecular Biology

Martinsried, Germany

Summary/Outlook

Sample Preparation



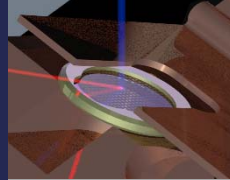
In vitro



In situ



Plunge freezing
-180 °C



FIB

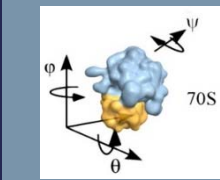
Electron Cryotomography



Single-axis tilt

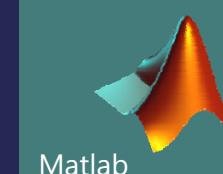


Dual-axis tilt



TM

Software



Matlab
toolbox



PyTom



Amira



Chimera

Instrumentation



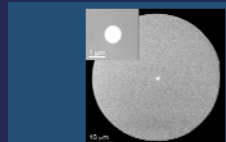
FEI Quanta 3D



FEI Titan Krios



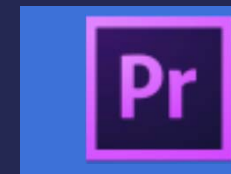
Direct Electron
Detectors



Phase Plate



3ds Max



Adobe CS6



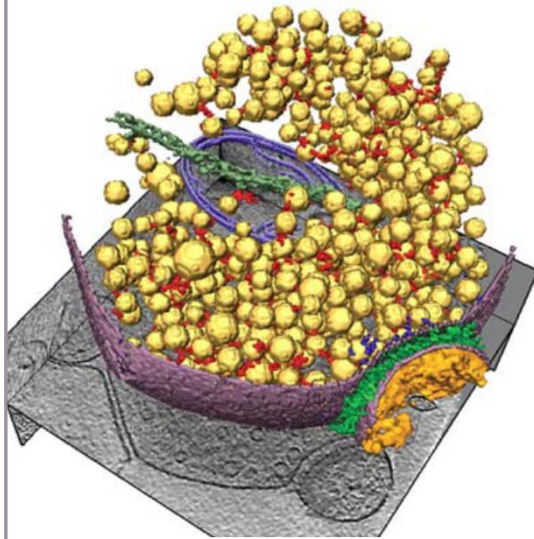
CET combines the power of 3-D imaging with a close-to-life preservation of cellular structures and **allows to study macromolecular structures *in situ*.**

Cryo-electron tomography (CET)



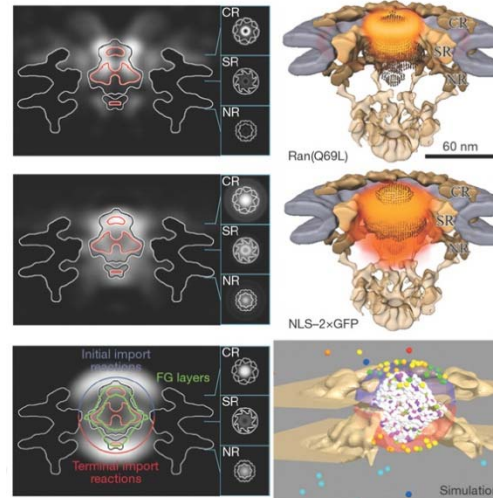
Low-level Techniques for Data Analysis in Tomography

Segmentation



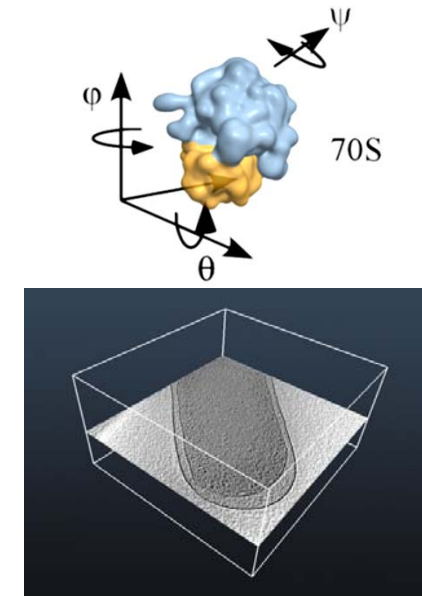
Presynaptic vesicles
Fernández-Busnadiego, R. et al.,
J. Cell. Biol. (2010)

Electron-dense labeling



NPC substrate localization
Beck, M., et al., *Nature* (2007)

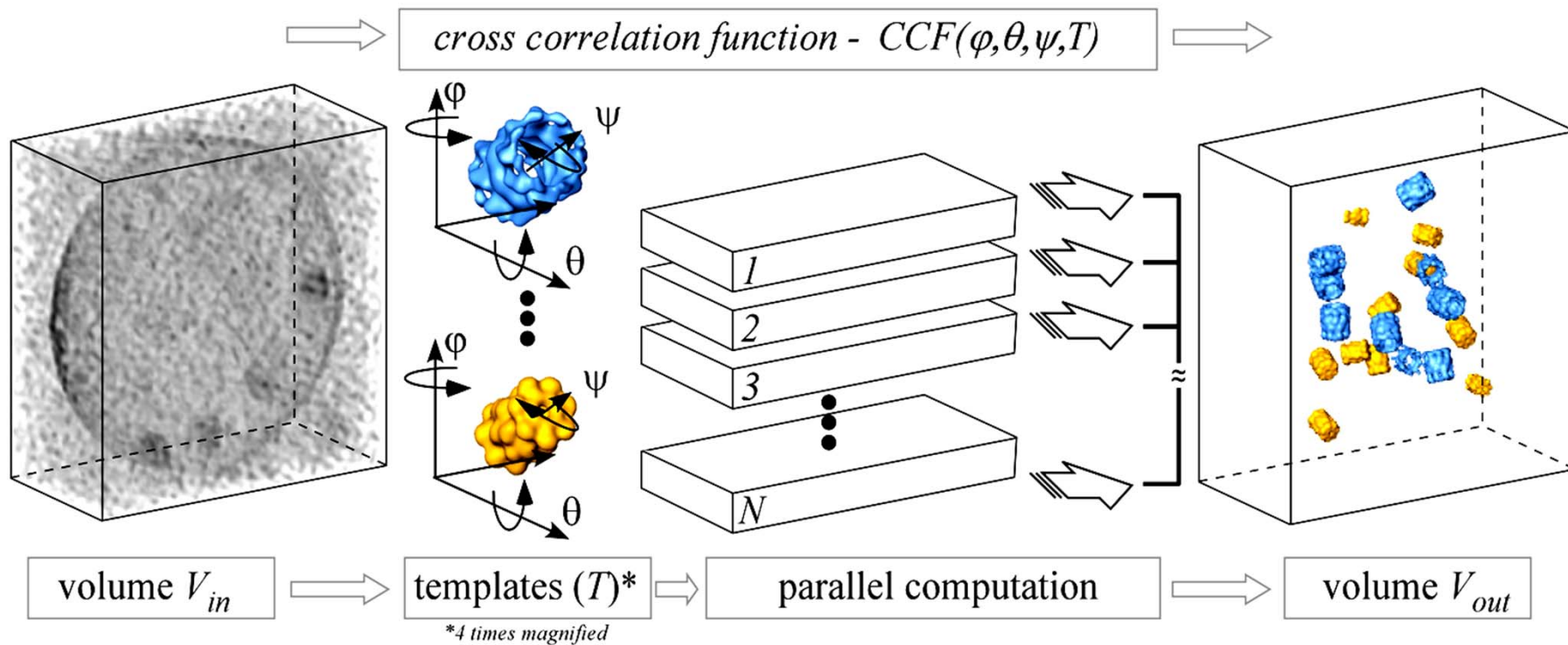
Template Matching



Boehm, J. et al. *Proc Natl Acad Sci U S A.* (2000)



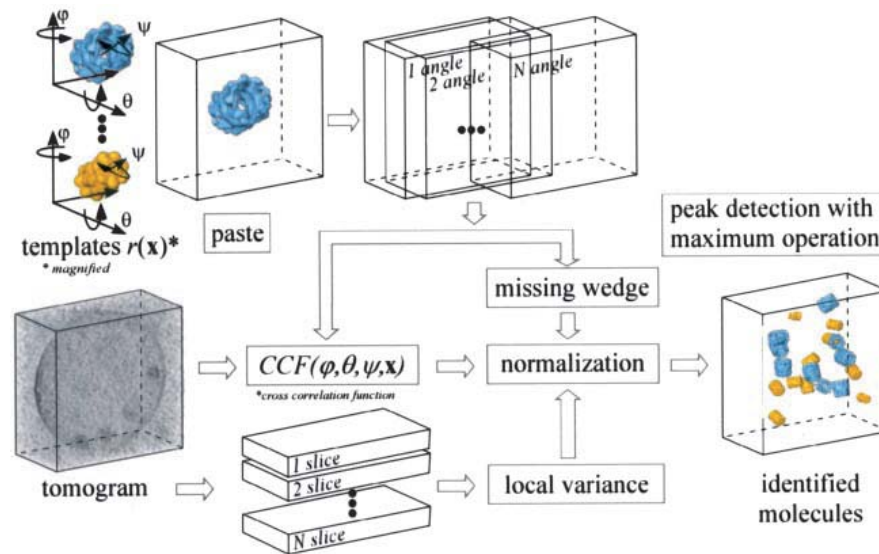
Detection and identification strategy



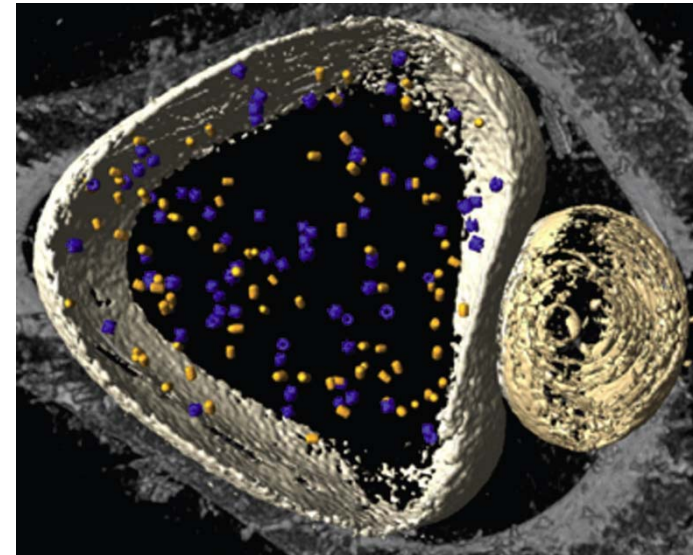
A.S. Frangakis, J. Böhm, F. Förster, S. Nickell, D. Nicastro, D. Typke, R. Hegerl, W. Baumeister:
Proc Natl Acad Sci USA 99 (2002) 14153-14158



Identification of macromolecular complexes in cryoelectron tomograms of phantom cells



Schematic flow diagram showing the detection and identification strategy

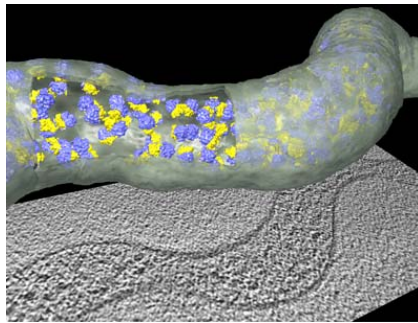


Thermosomes and 20S proteasomes encapsulated in liposomes

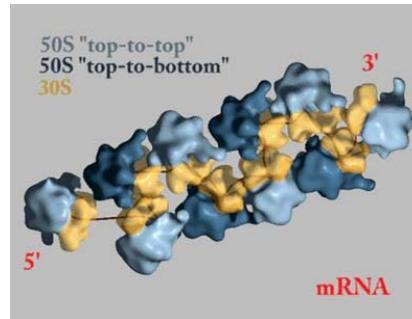
Frangakis, A. et al., *PNAS* (2002)



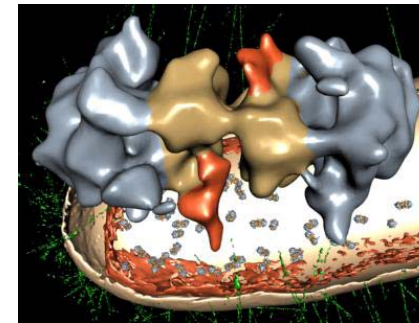
2002-2012: Template Matching Achievements



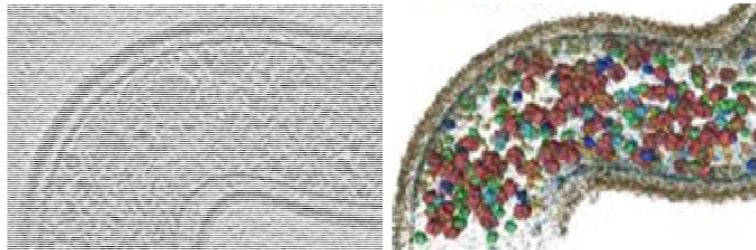
Spiroplasma
Ortiz, J., et al. *J. Struct. Biol.* (2006)



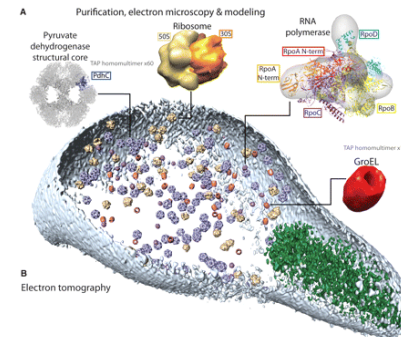
Polysomes *in vitro*
Brandt, F., et al. *Cell.* (2009)



Hibernating ribosomes (*in vitro* & *in vivo*)
Ortiz, J., et al. *J. Cell. Biol.* (2010)



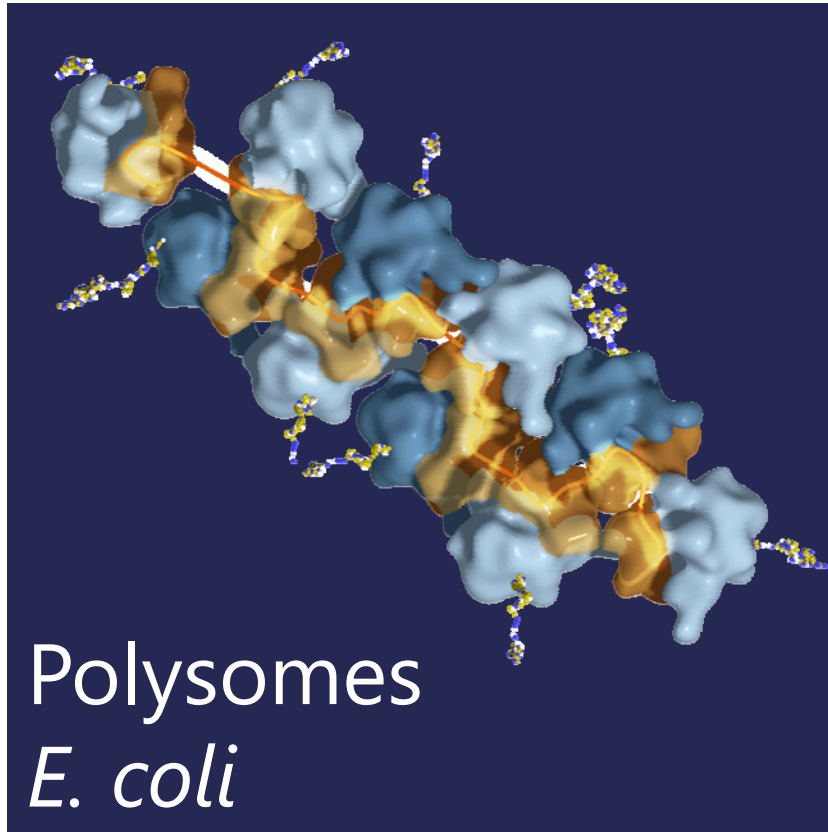
Visual Proteomics - *Leptospira*
Beck, M., et al. *Nat. Methods* (2009)



Visual Proteomics - *Mycoplasma*
Kuerner, S., et al. *Science* (2009)

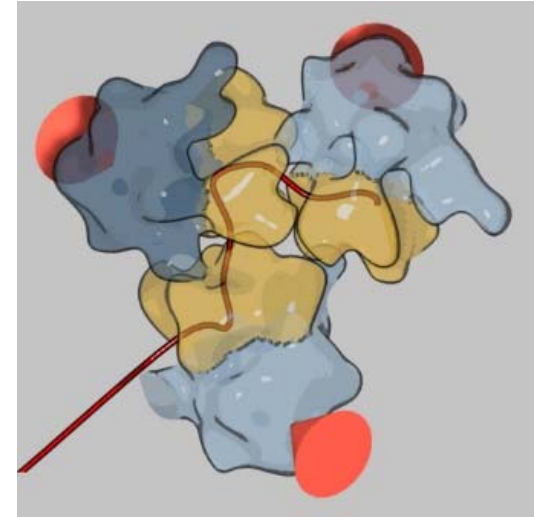


Polysomes *in situ*



Brandt, F. et al. *Cell*. 2009 **23**:261-71

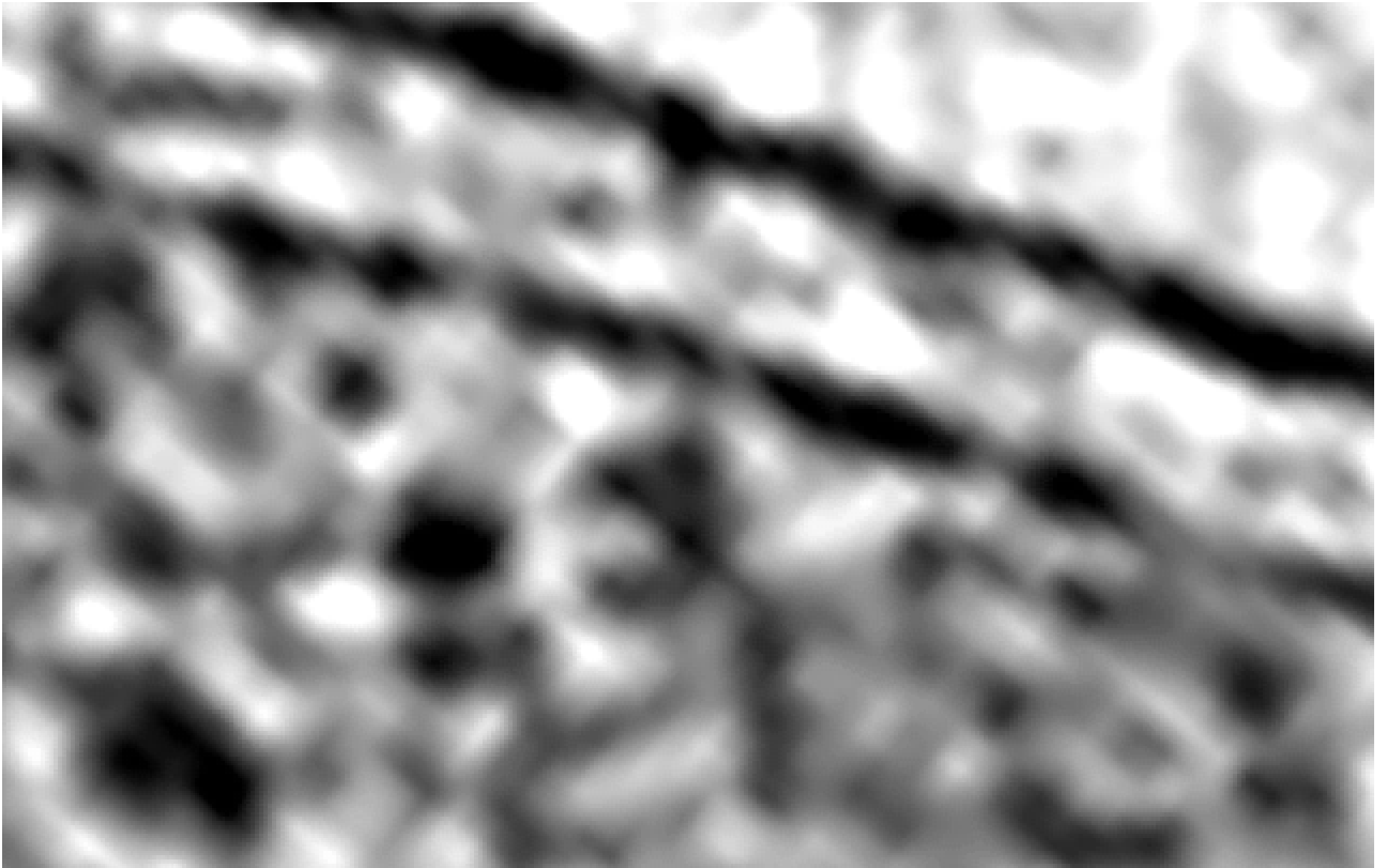
E. coli



Aim:

To detect ribosomal supramolecular arrangements in the cytoplasm of fast growing *E. coli* cells.



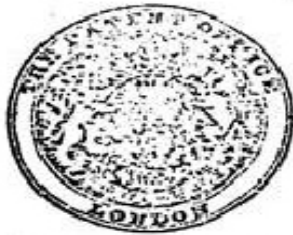


Tomogram of thin whole *E. coli* cell
(slow growing cell; CCD- camera)

Cryo-ultramicrotomy- Diamond Knife

PATENT SPECIFICATION

799,498



Date of Application and filing Complete Specification: Sept. 2, 1955.
No. 10682/57.
Application made in Sweden on Sept. 4, 1954.
(Divided out of No. 799,497).
Complete Specification Published: Aug. 6, 1958.

Index at acceptance:—Class 60, D1(D4X: H17), D2(A15: A20: K).
International Classification:—B24b.

COMPLETE SPECIFICATION

Improvements in or relating to a method of Polishing a Cutting Edge of a Diamond for a Cutting Tool

I, HUMBERTO FERNÁNDEZ-MORÁN, of Instituto Venezolano de Neurología e Investigaciones Cerebrales, Ministerio de Sanidad, Caracas, Venezuela, a National of Venezuela, do hereby declare the invention, for which I pray that a patent may be granted to me, and the method by which it is to be performed, to be particularly described in and by the following statement:—

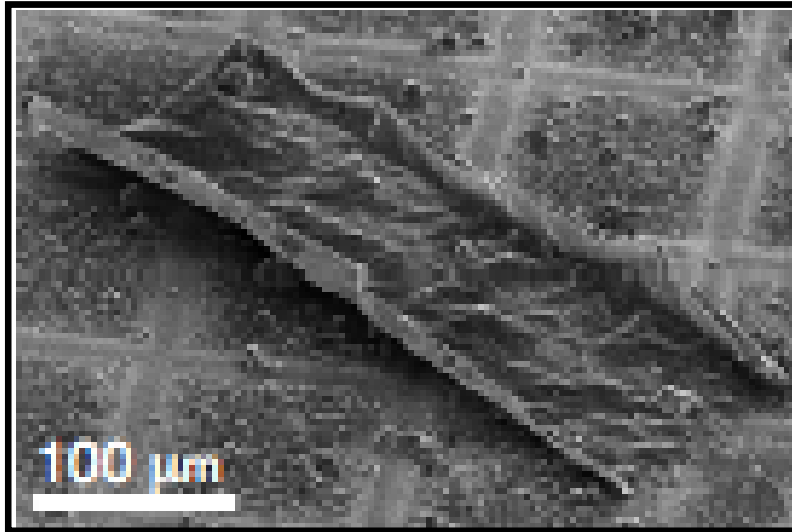
The advantageous effect of the grain size chosen has been found in practice and may perhaps be explained as follows:—

By using a paste comprising diamond powder of a grain size between 0.002 and 0.005 micron, the frictional effects on the diamond being polished are such as to produce a high temperature having a burning-off effect on surface irregularities, rather than a wearing-

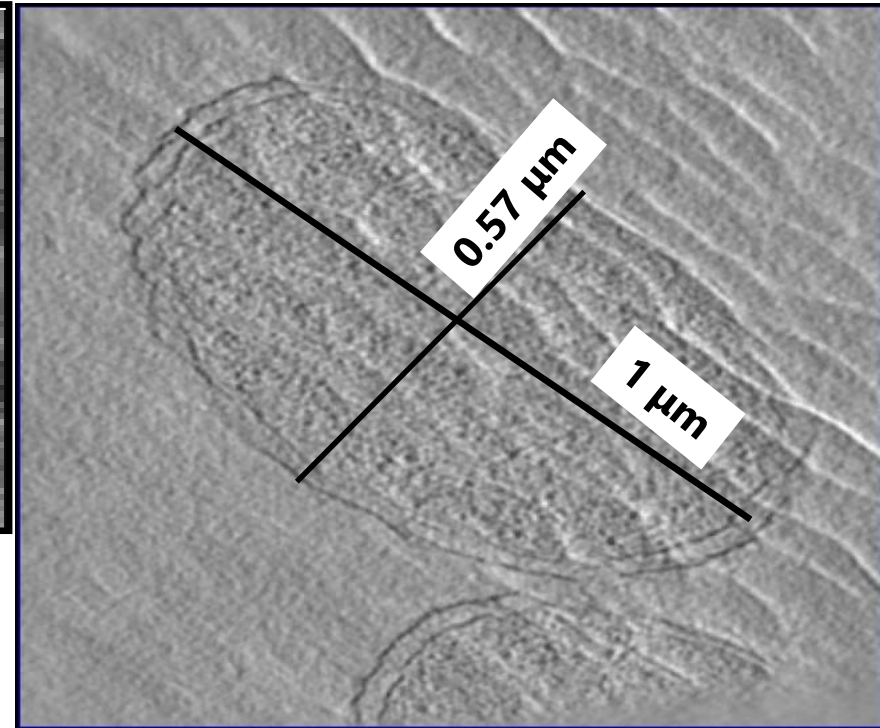


Cryo-ultramicrotomy

Cryo-section



E. coli cell

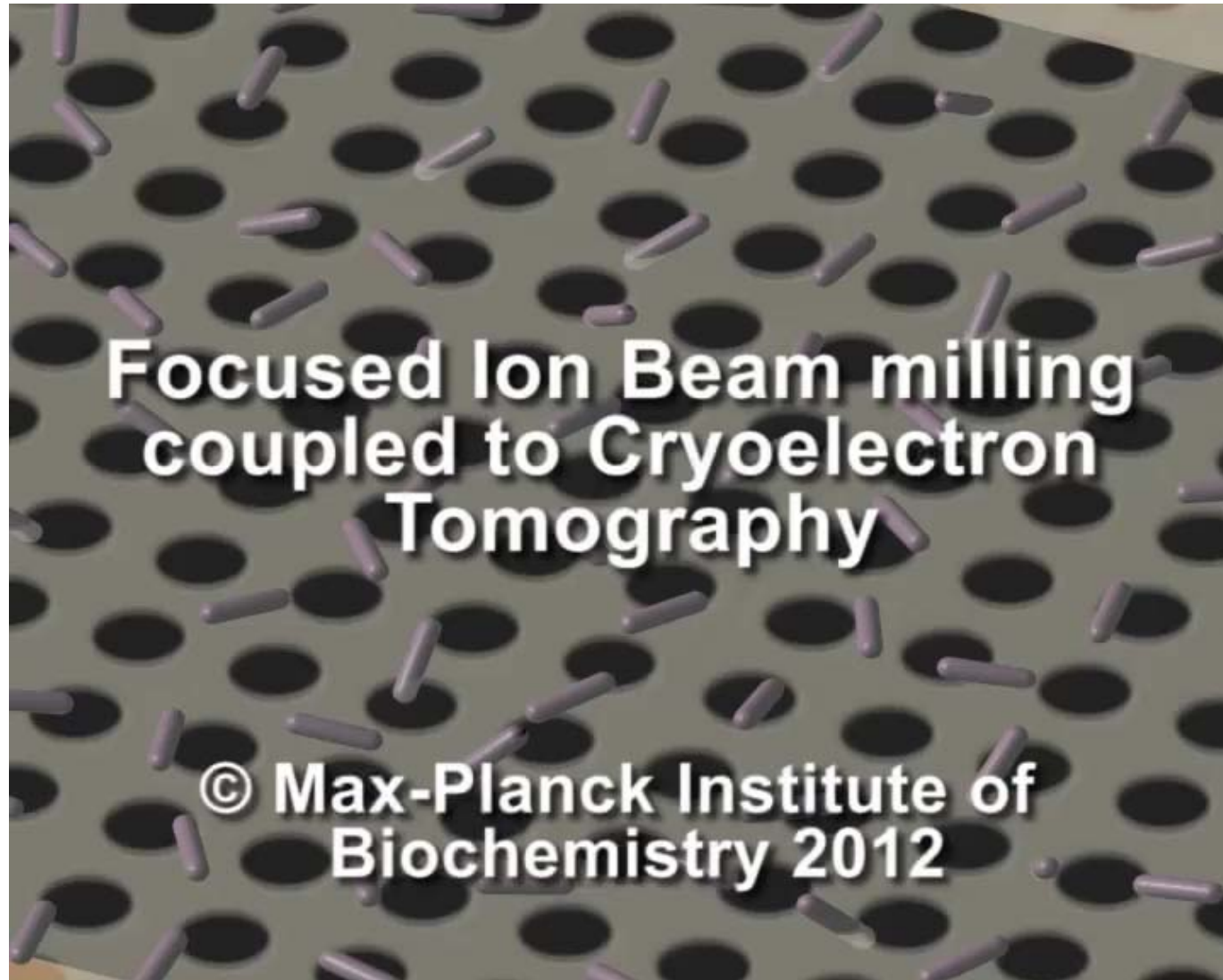


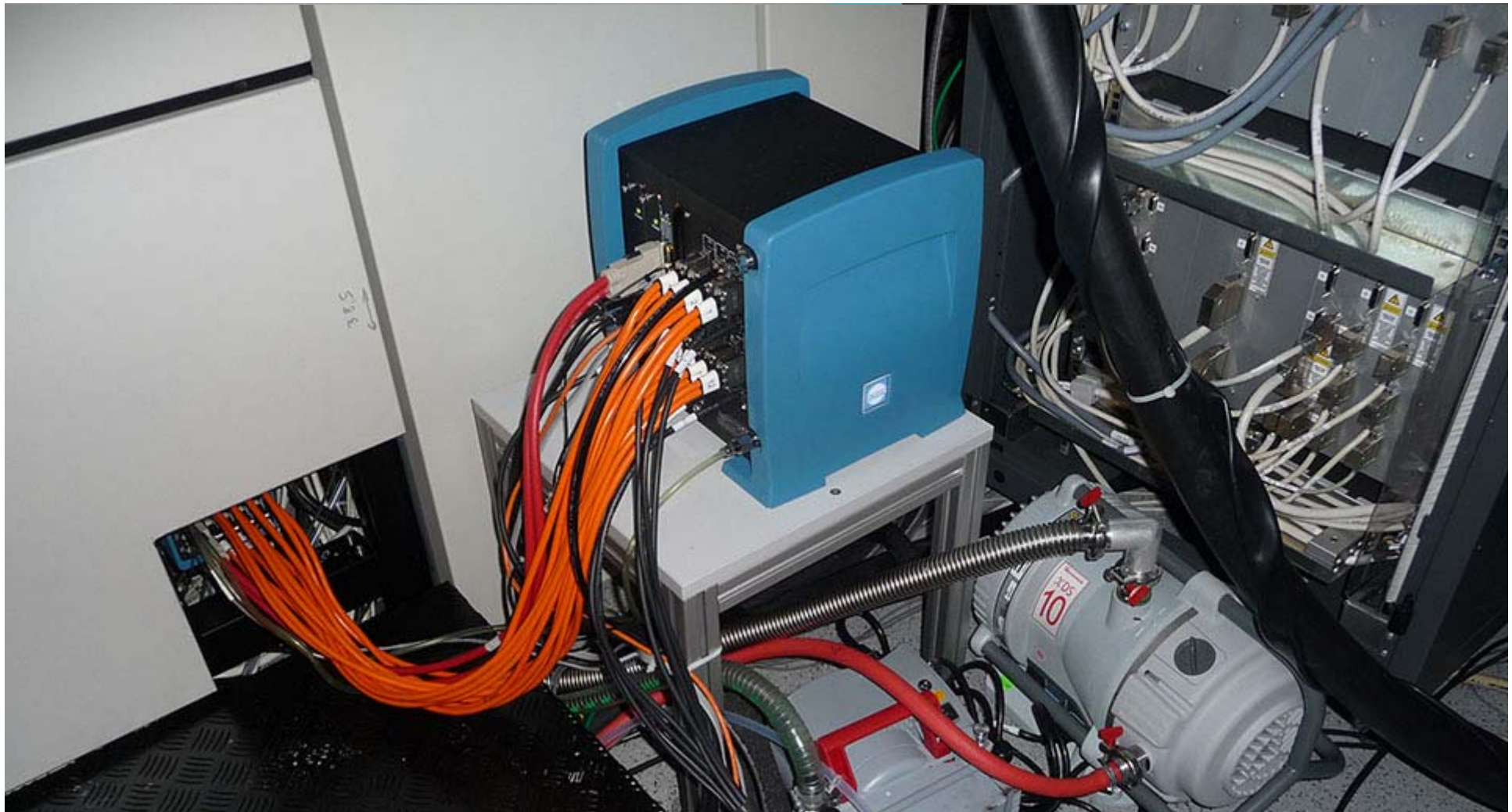
NAD filtered slice from tomogram
~ 30 % compression

Polara 300 KV, Mag: 55 KX, Pixel size: 0.55 nm/pixel, Def: -14 nm, Thickness: ~100 nm



Focused Ion Beam milling coupled to CET



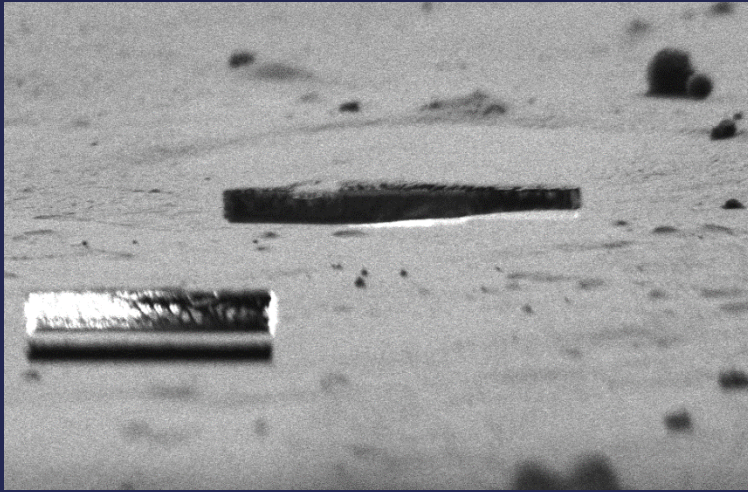


Advanced Transmission Electron Microscopy

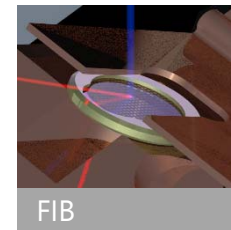
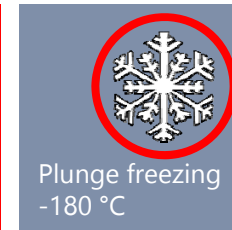
Dual-tilt axis CET with K2 camera and Energy filter



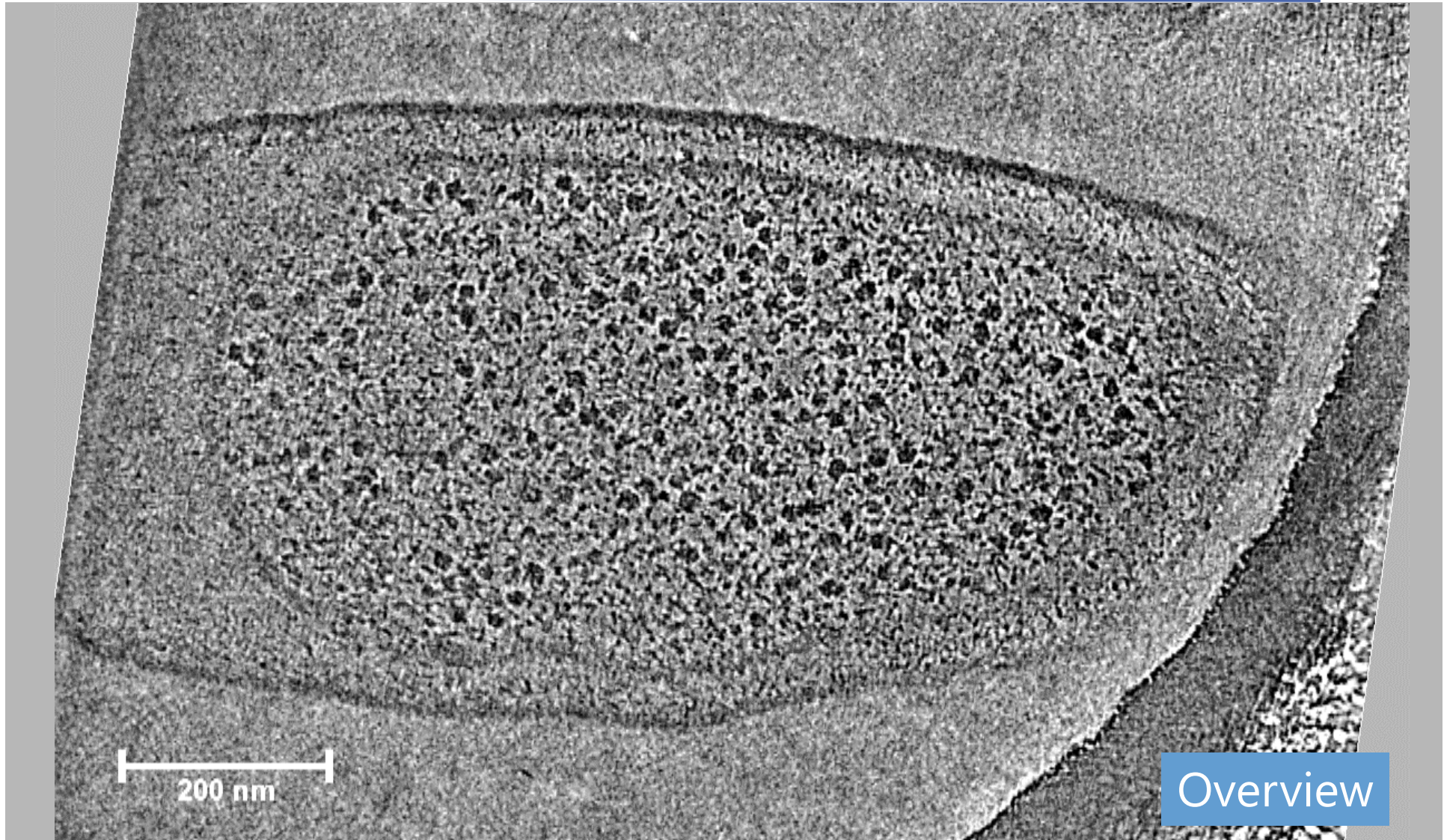
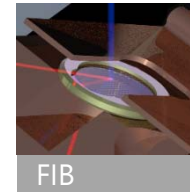
Polysomes *in situ*



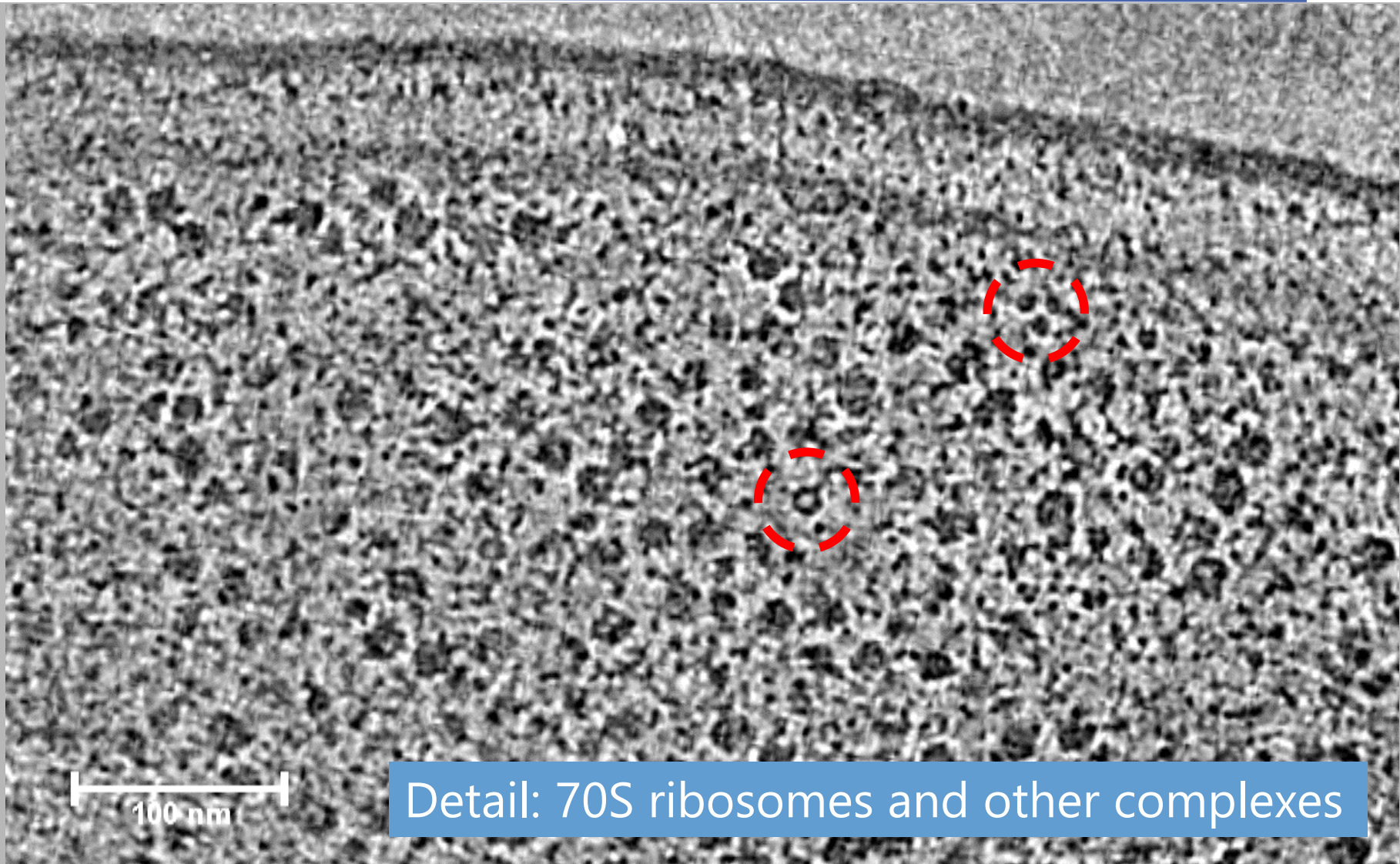
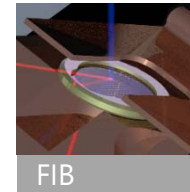
Polysomes
E. coli



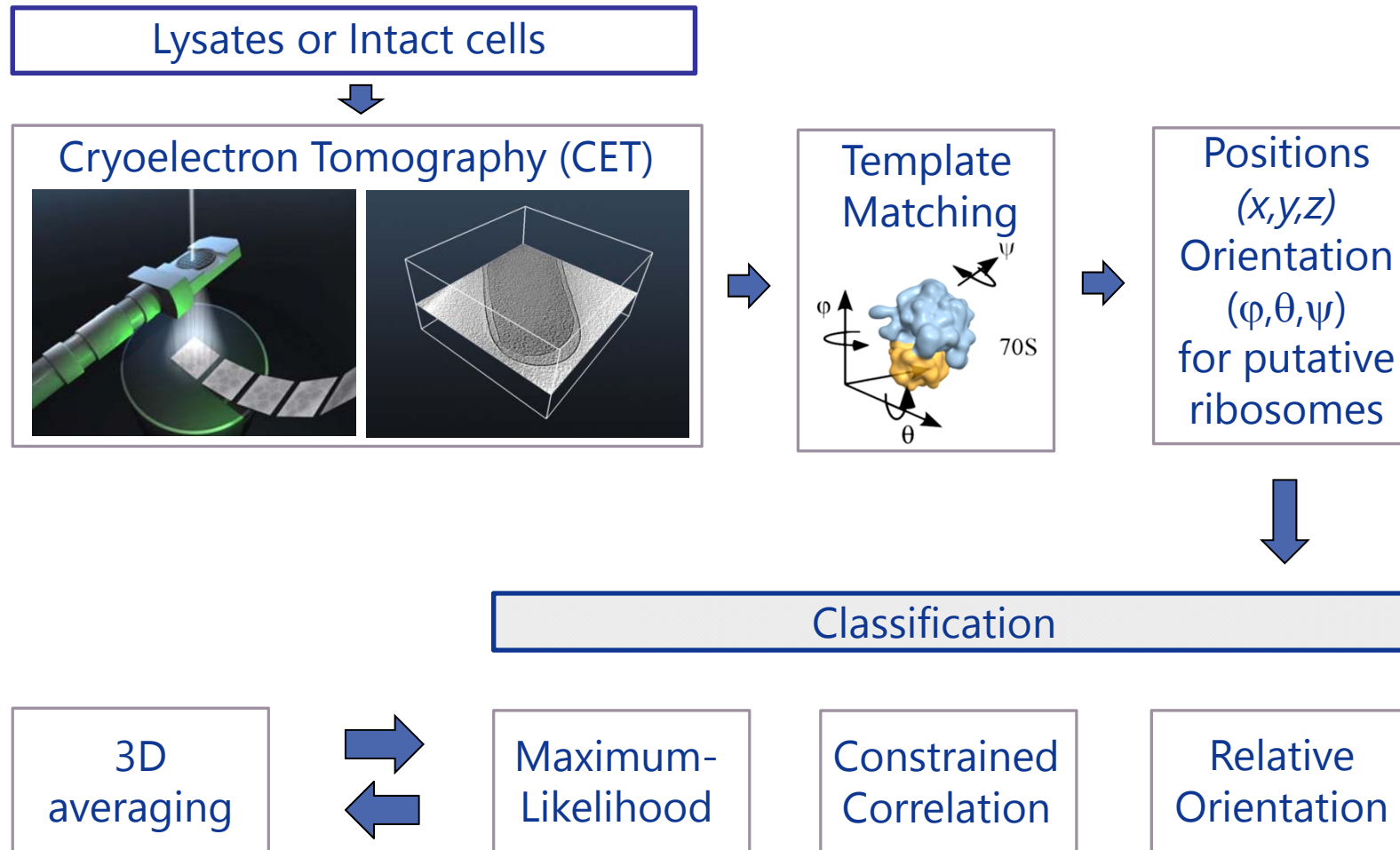
E. coli BL21 MtlA³⁸⁵-SecM (non-stalling conditions)



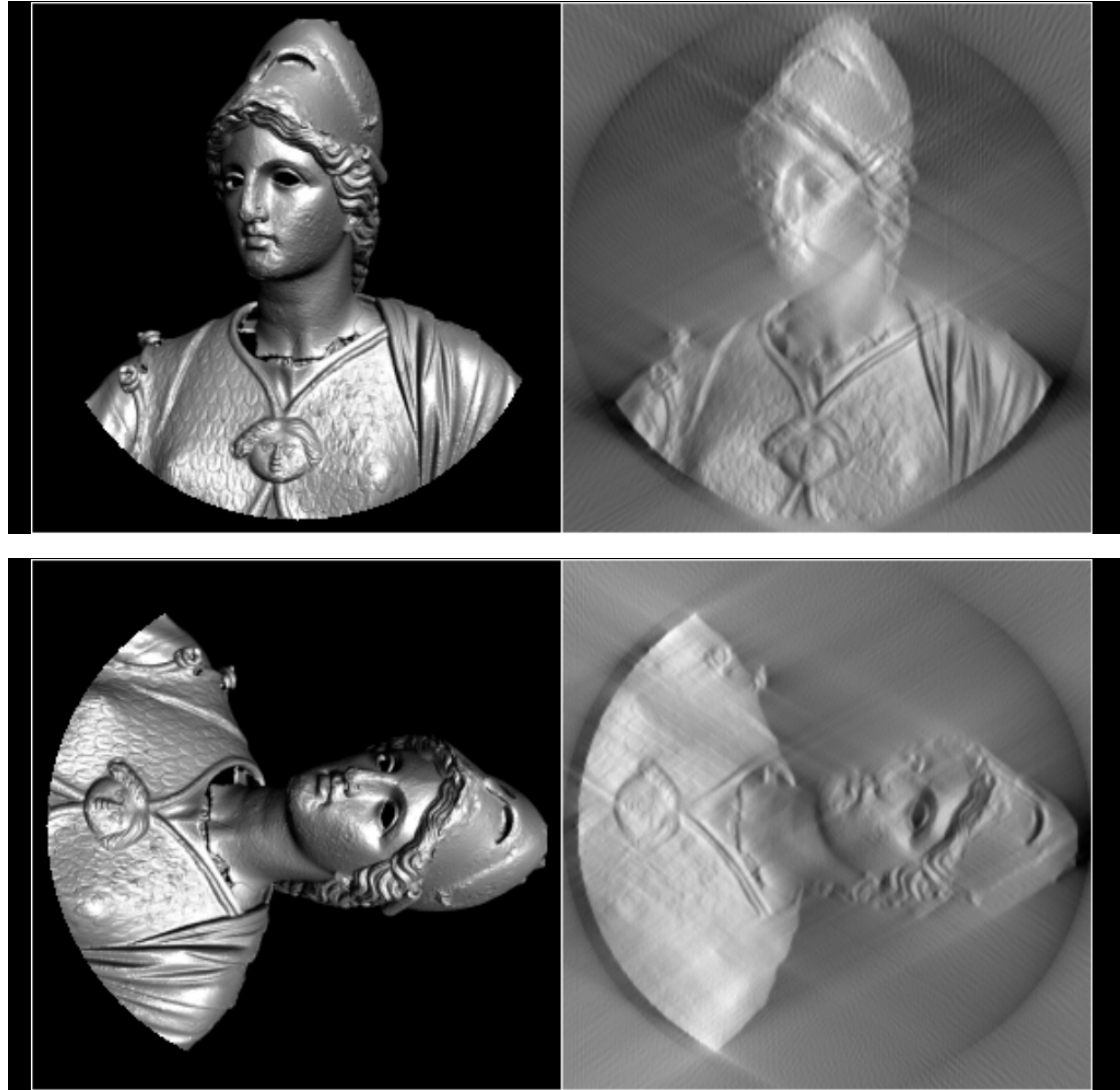
E. coli BL21 MtlA³⁸⁵-SecM (non-stalling conditions)



Methods



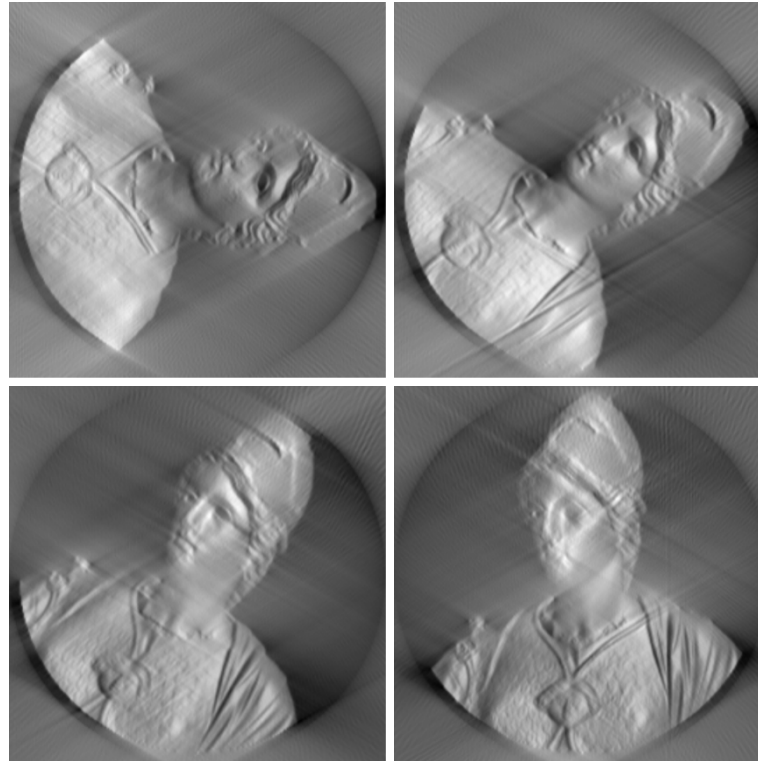
The art of averaging: Missing Wedge effect



Combining reconstruction by averaging



2D Model



Reconstructions from identical objects
in different orientations

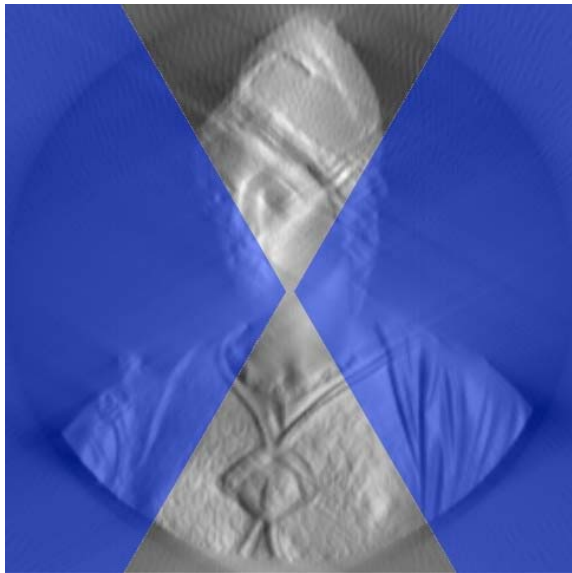


Combined reconstruction



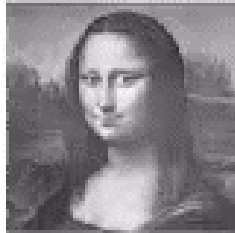
The art of averaging

Combining objects in different orientations...

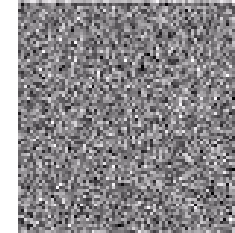


The art of averaging: The "Mona Lisa" effect – Bias caused by reference

template



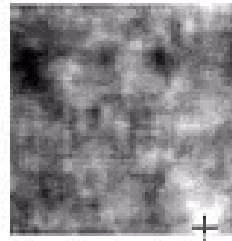
data



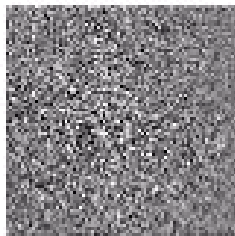
data aligned



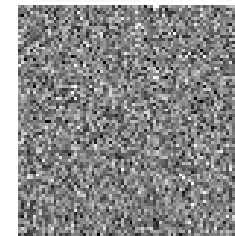
cross correlation



aligned average

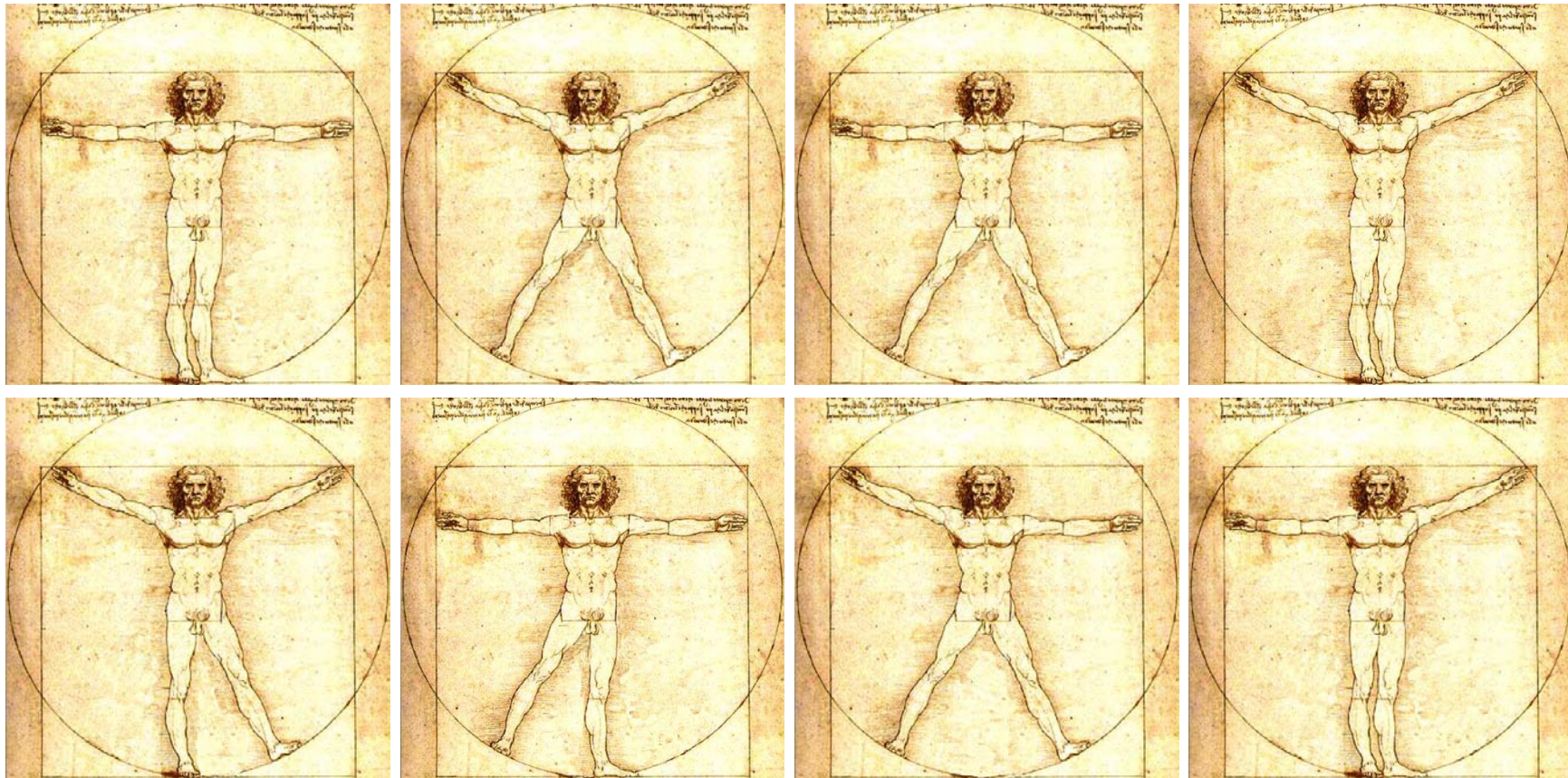


average



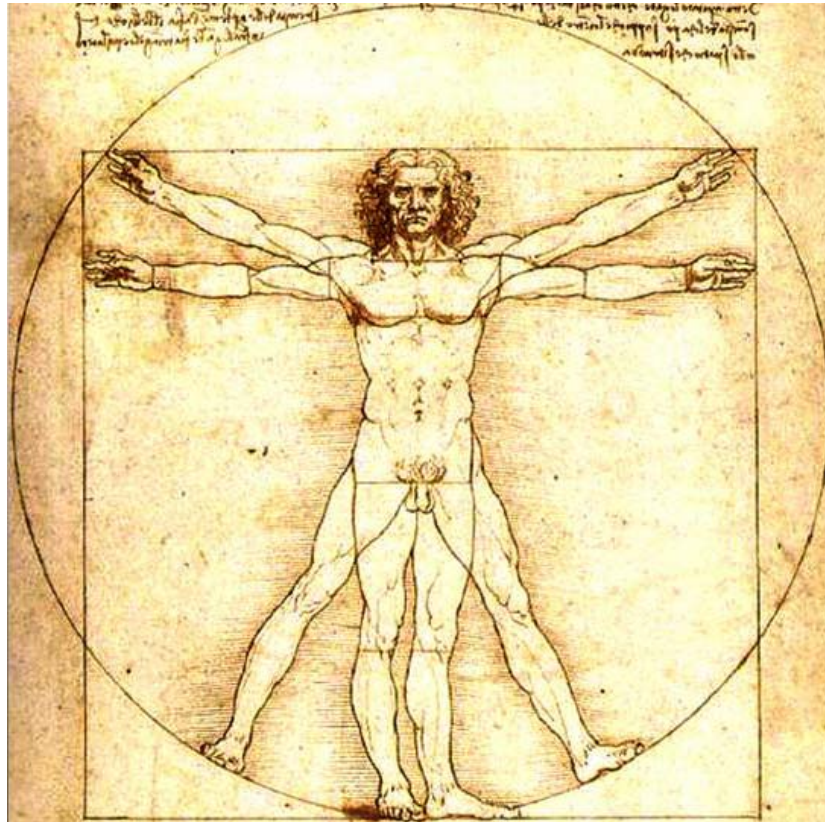
The art of averaging

The “Vitruvian man” effect – Flexibility and occupancy problems

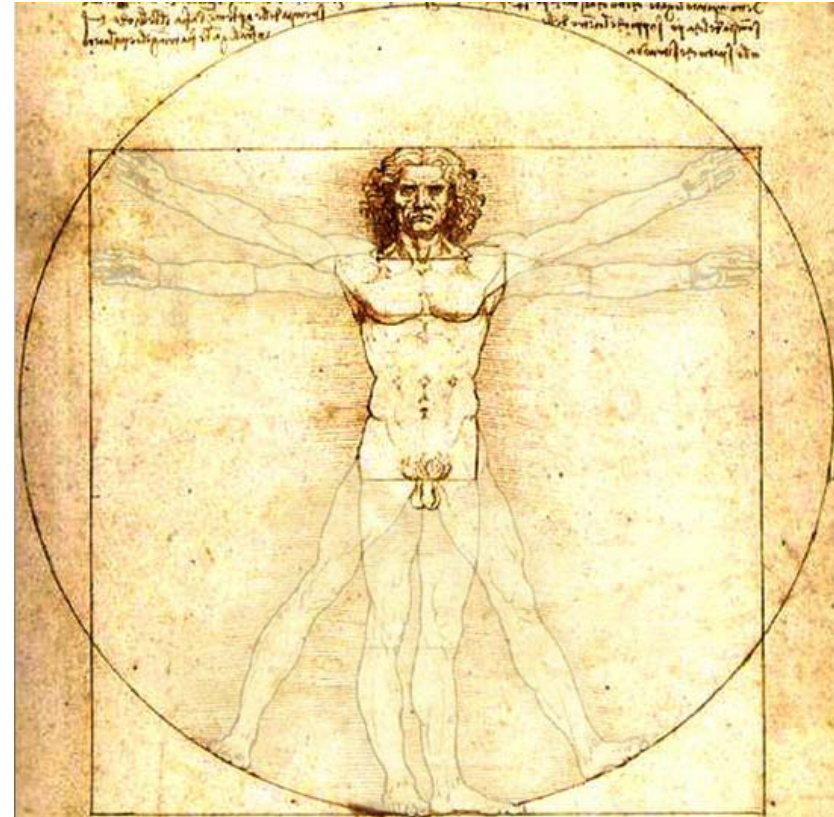


The art of averaging

The “Vitruvian man” effect – Flexibility and occupancy problems



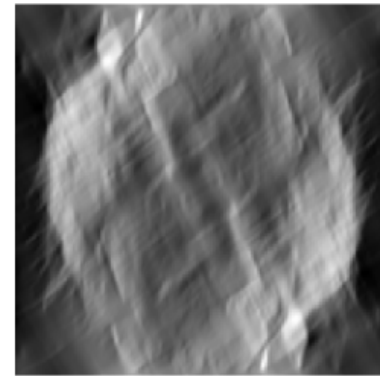
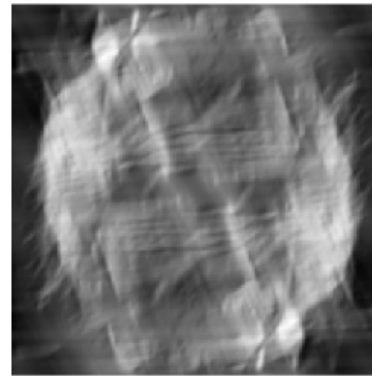
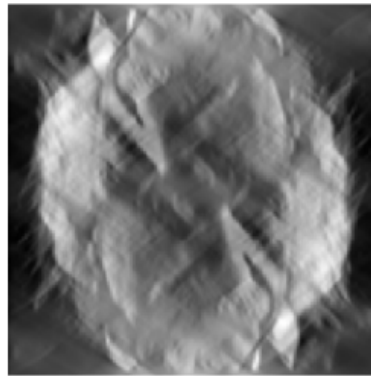
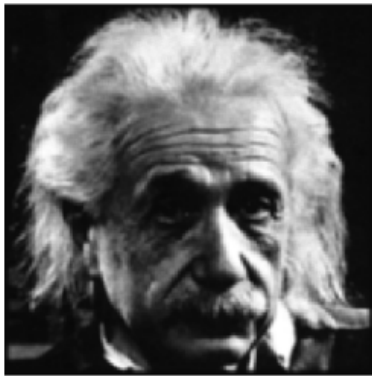
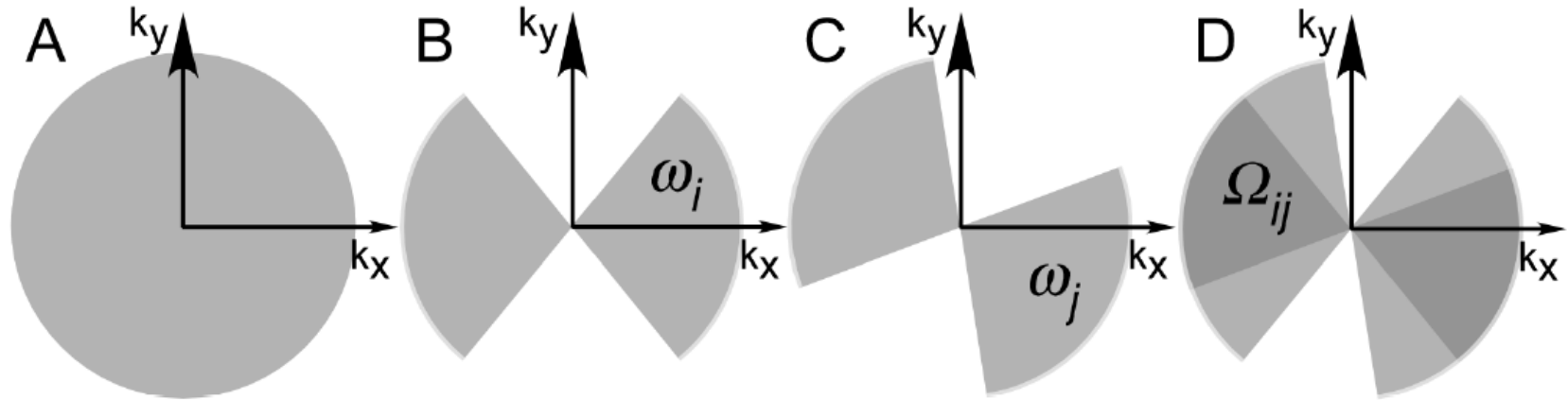
“Vitruvian man” – Leonardo Da Vinci



Average of 8 different images



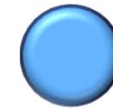
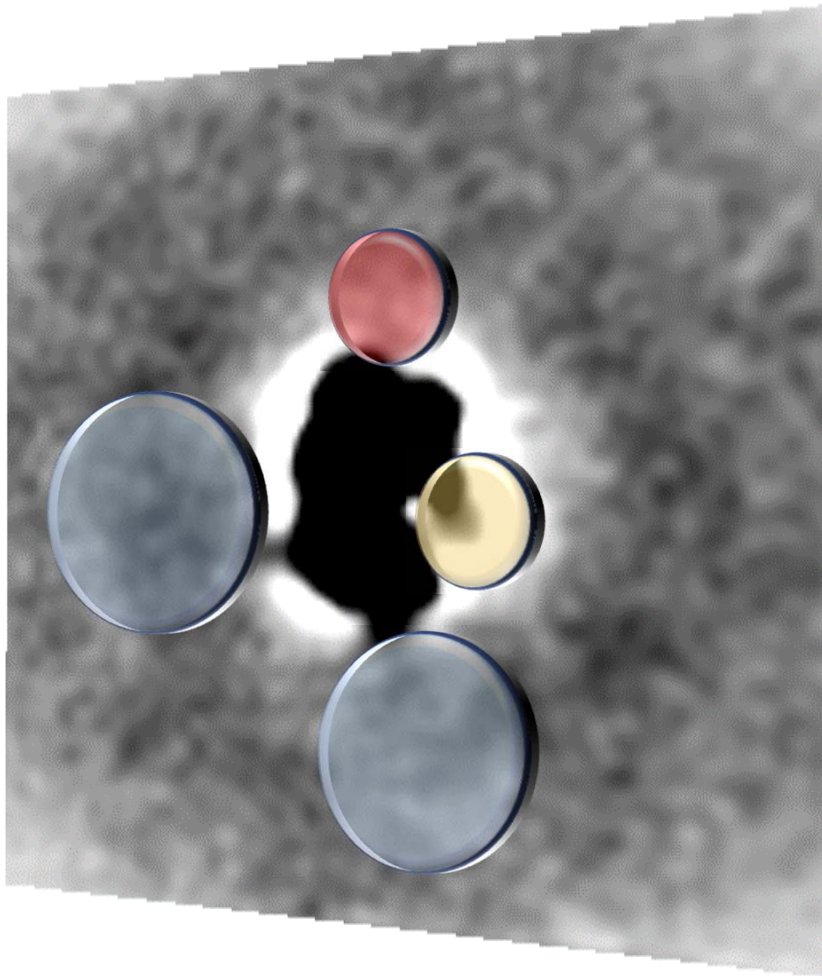
Constrained Correlation Classification



Förster F., et al. (2007) J. Struct. Biol.



Constrained Correlation Classification (3D-average selected ribosomes)



70S ribosome

Average of ~ 6000 part.
selected by
30S subunit similarity
resolution: ~2.6 nm

Areas with associated densities:



mRNA entry and
exit site



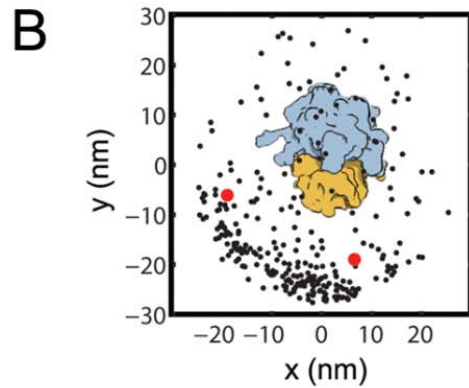
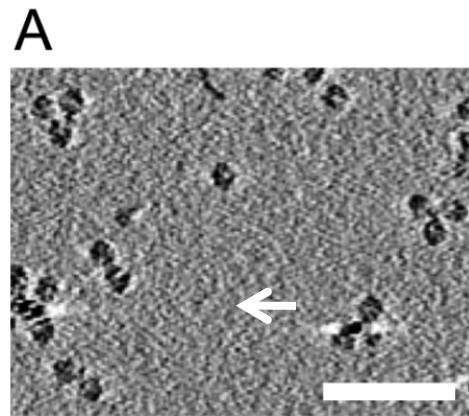
Elongation factor
binding sites



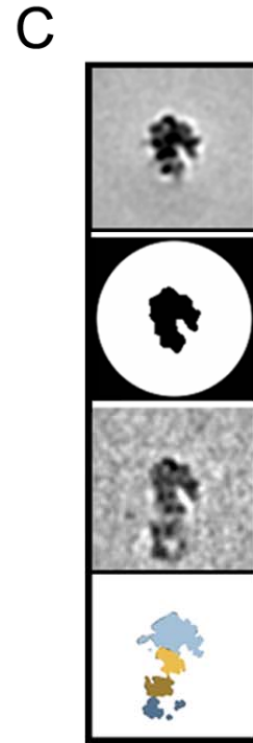
Nascent chain
exit site



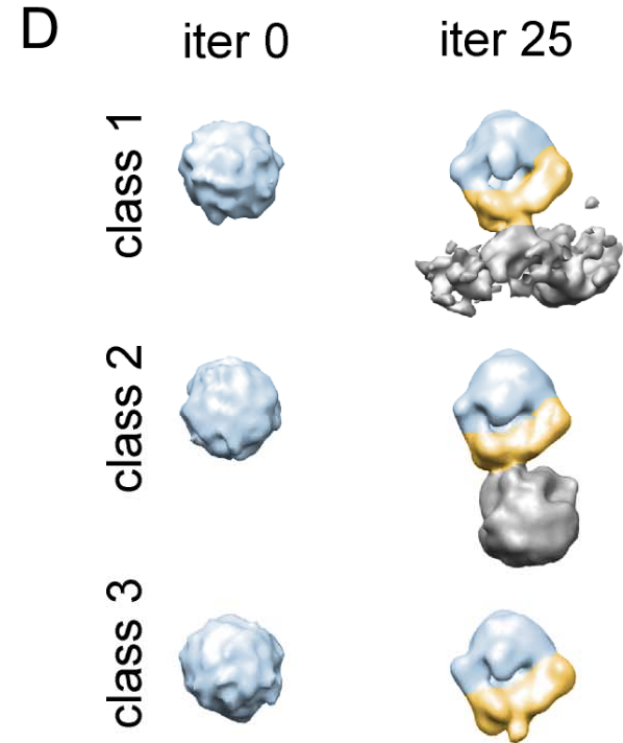
Hibernating Ribosomes in Lysates from Starved *E. coli* Cells



Relative Orientation



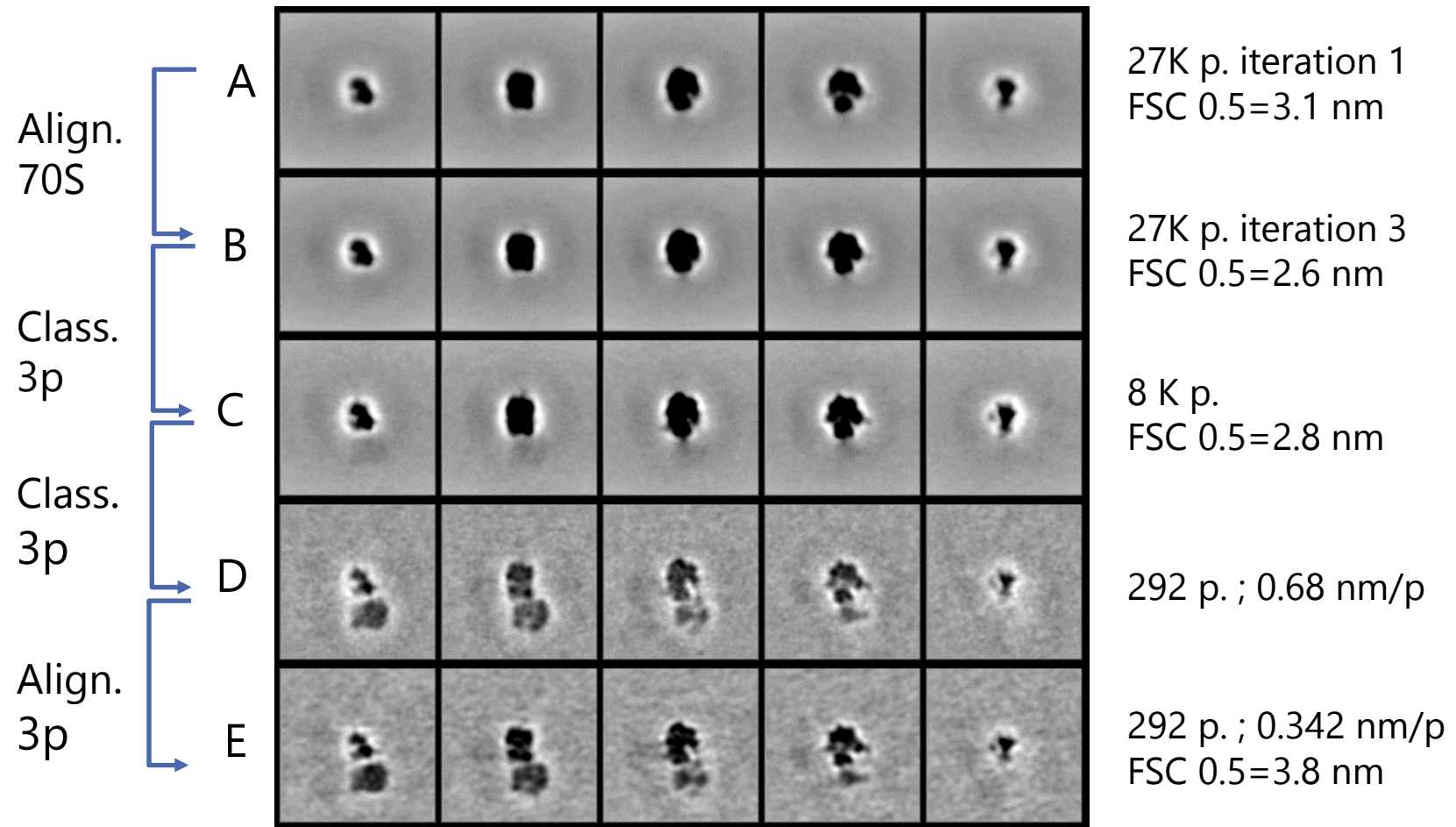
Constrained Correlation



Maximum-Likelihood



3D Alignment and Averaging







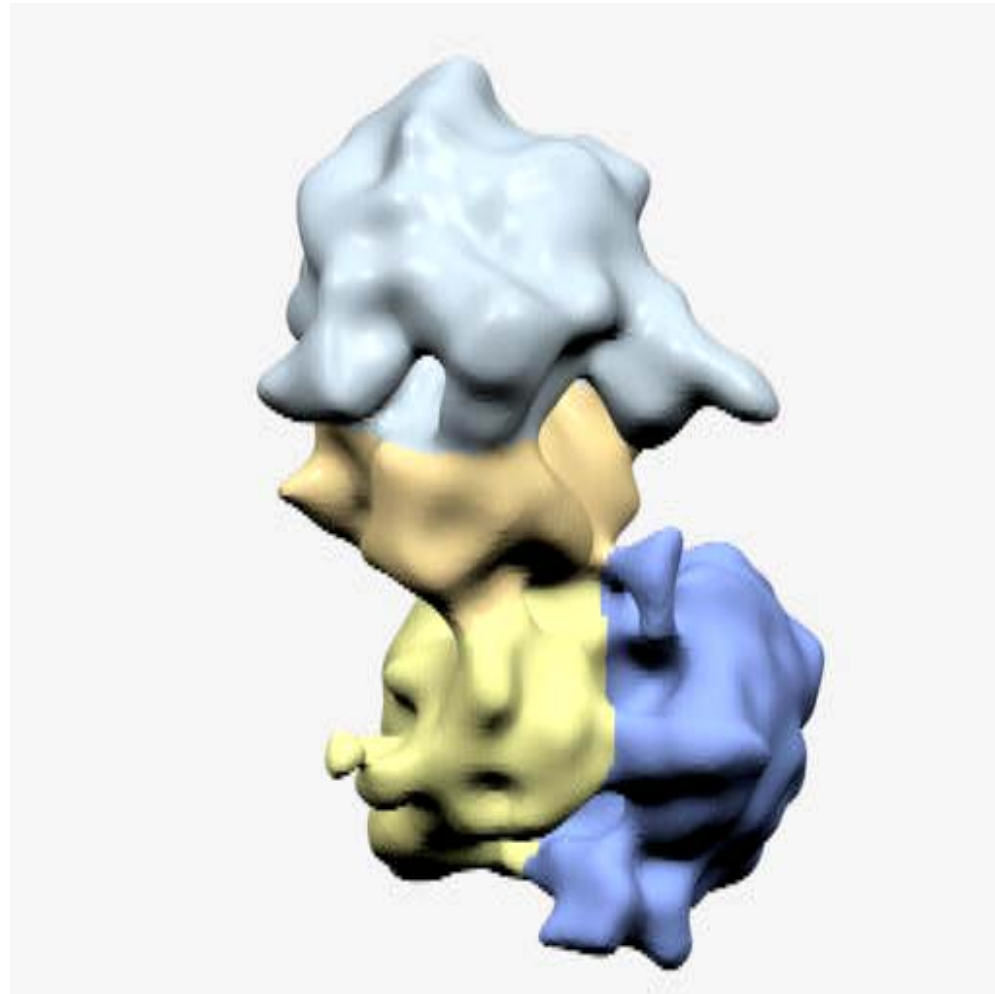
3'-neighbor ribosome attached in ca. 1% of analyzed particles



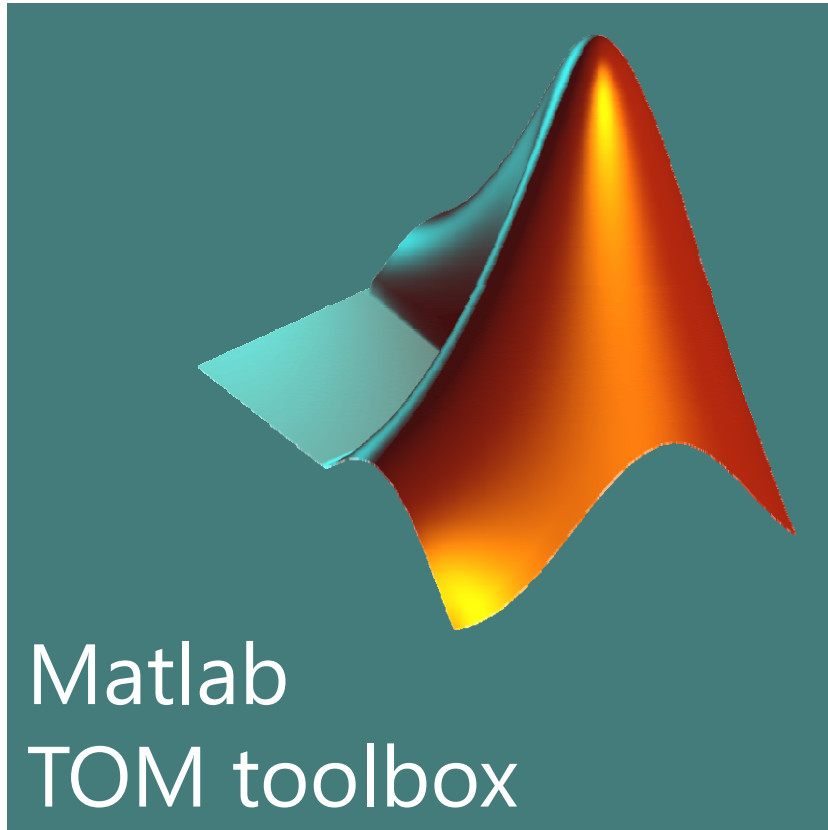
Polysomal organization *in situ*: 3D-Average “top-to-top”

- 103 particles
- $(256)^3$ p, 0,342 nm/p
- Average filtered at 3nm res.
- Particles present in all analyzed tomograms

-  50S - central ribosome
-  30S - central ribosome
-  50S – neighbor ribosome
-  30S – neighbor ribosome



Methods: Software



Matlab TOM toolbox

The toolbox supports a wide range of functions for tomography that extend the capability of the MATLAB® numeric computing environment.

Aim:

Customable procedures for tomogram reconstruction and analysis.

Nickell. S. et al. (2005) *J. Struct. Biol.* **149**:227-34



Methods: Software



PyTom

PyTom

Open-source platform that unifies standard tomogram processing steps in a single python-based toolbox.

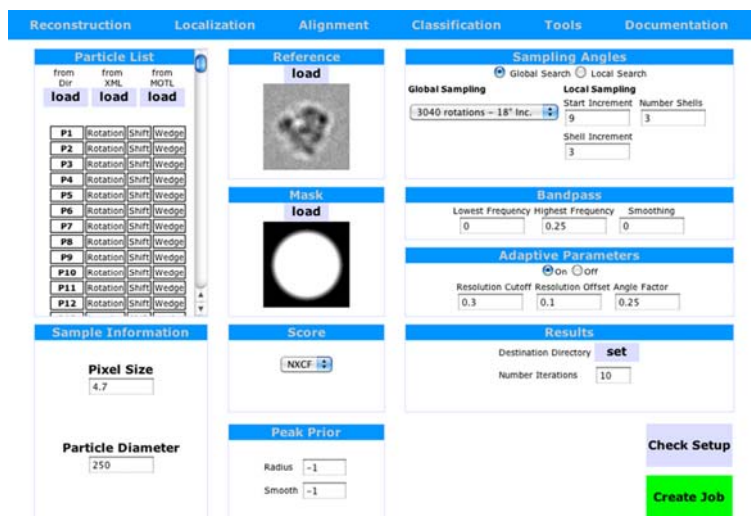
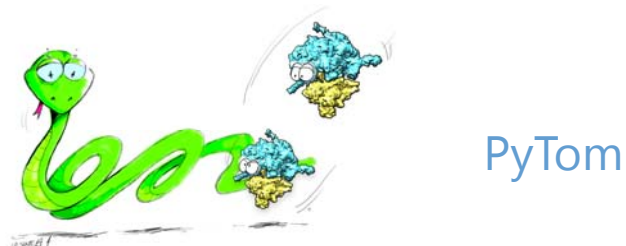
Aim:

3D alignment, averaging and classification of subtomograms. Implementation of new algorithms for image processing in ECT.

Hrabe, T., et al. (2012) JSB 178: 177-188



Recent Improvements in Data Processing Workflow



Grabe, T., et al. *J. Struct. Biol.* 2012

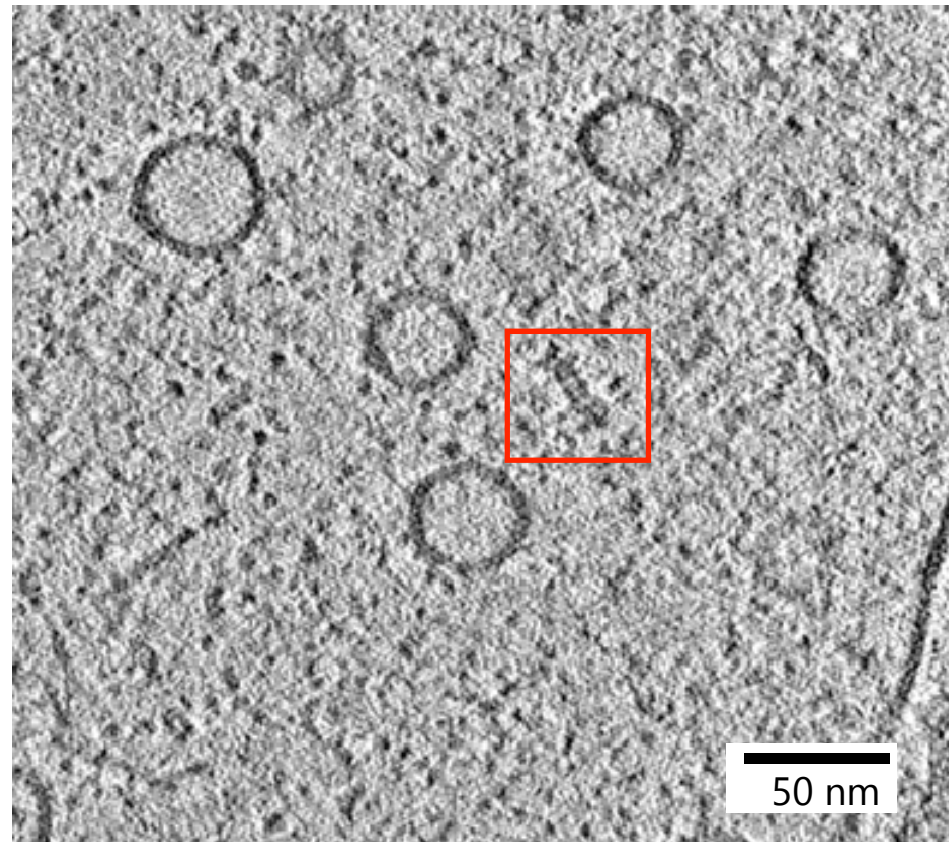
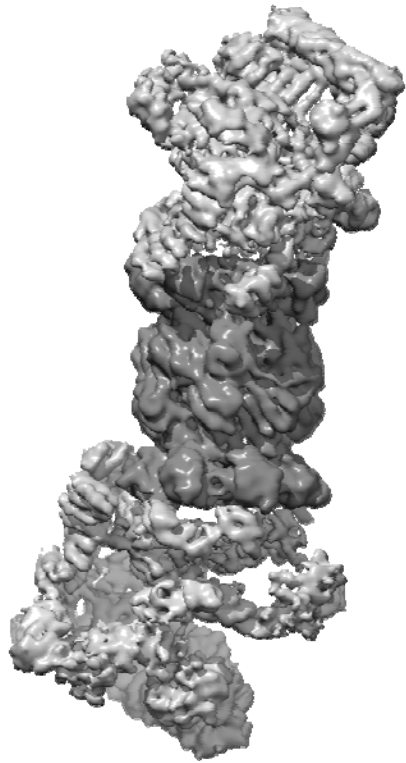
1. Reconstruction
 - Phase Flipping ('CTF correction')
 - Projection Alignment with Tilt-Specific Image Rotation + Magnification
2. Localization
 - Template Matching (+Support Vector Machines)
 - Parallel processing on LINUX cluster
 - Better angular sampling
3. Subtomogram Alignment
 - Parallel processing on LINUX cluster
 - Resolution-dependent lowpass: low bias
 - Resolution-dependent sampling
 - Direct reconstruction from projections
4. Classification
 - Multi-reference correlation with simulated annealing



CET: Direct detection and Phase Plates

Asano, S. *et al.* (2014) *Science* 23:347

Sample: Hippocampal primary cultured neuronal cell



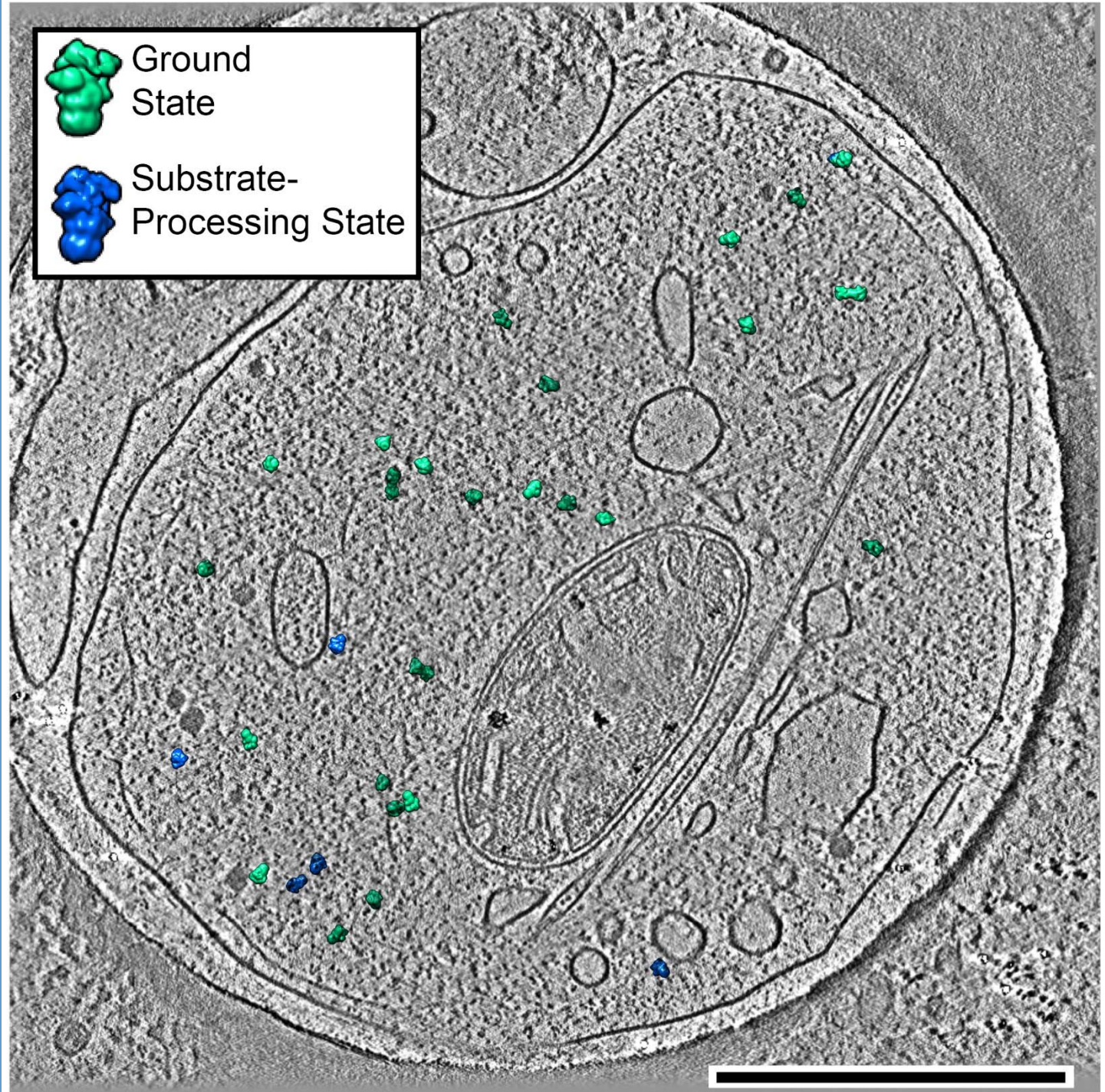
26S Proteasome *in situ*!





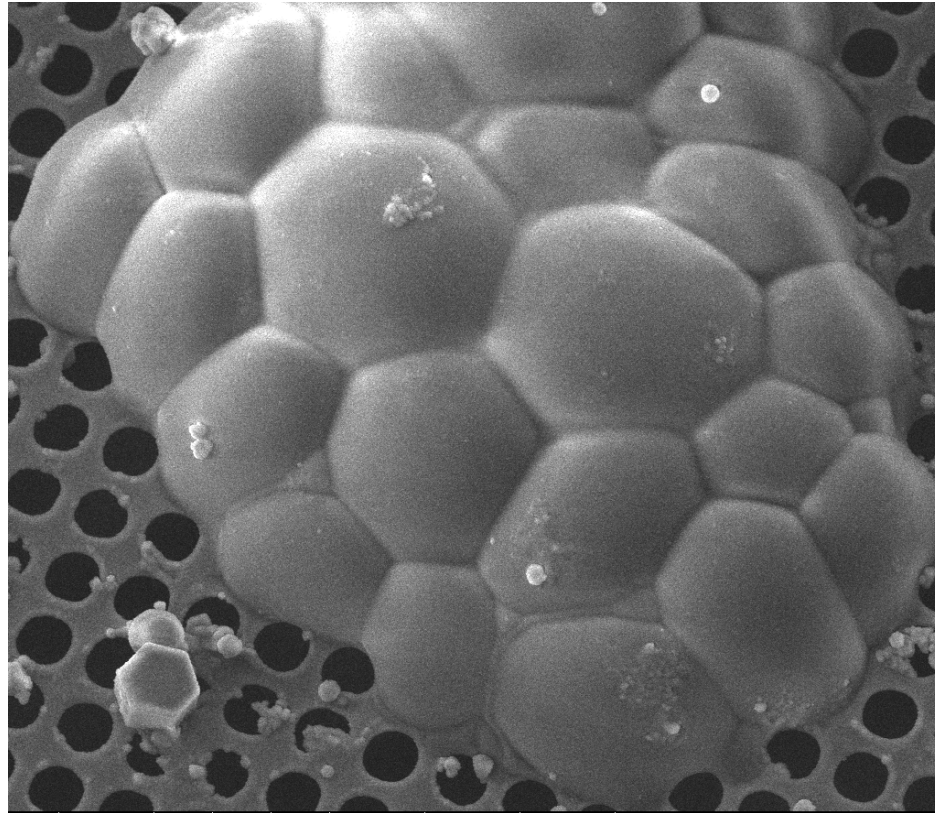
Proteasomes in neurons cytoplasm

Asano, S. *et al.* (2015)
Science 23:347

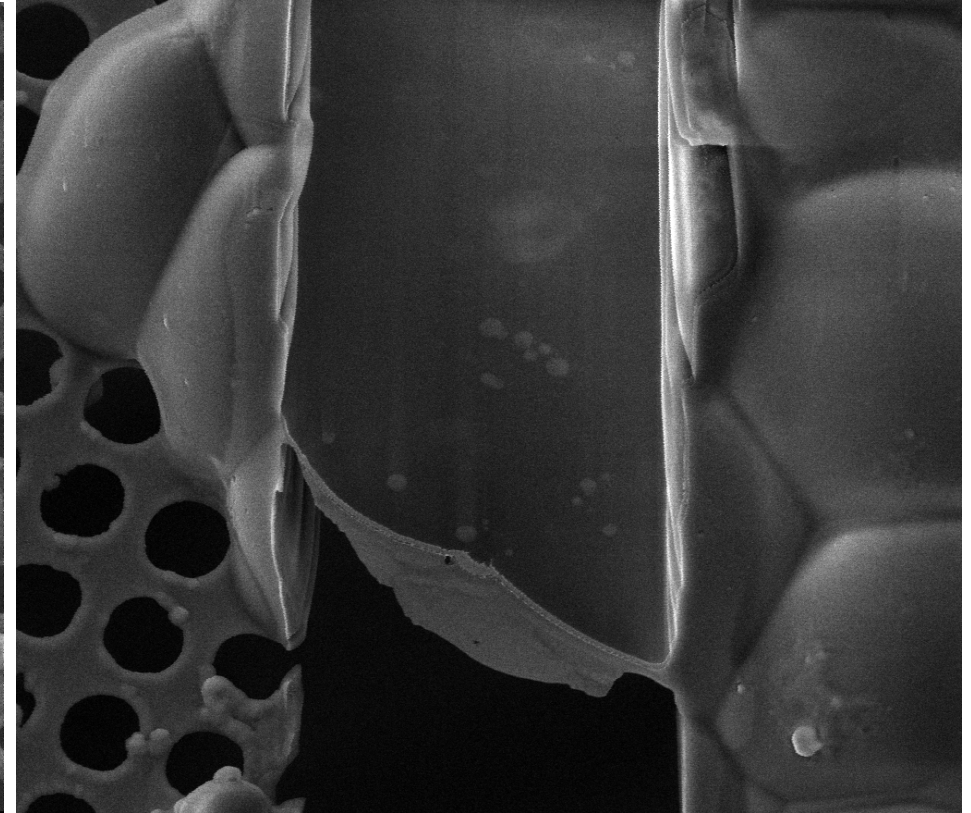


Cryo-FIB: *Chlamydomonas reinhardtii*

40 μm



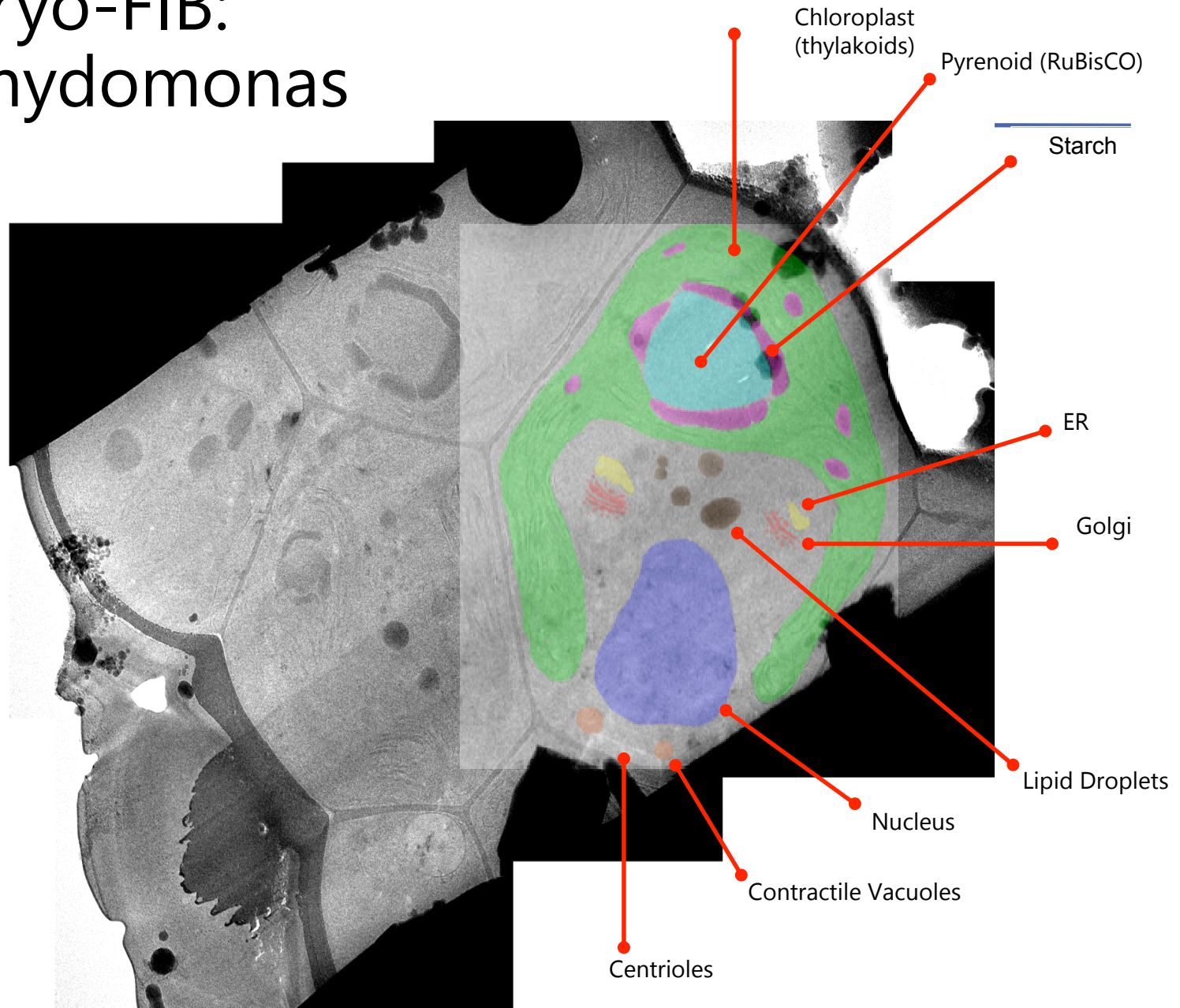
10 μm



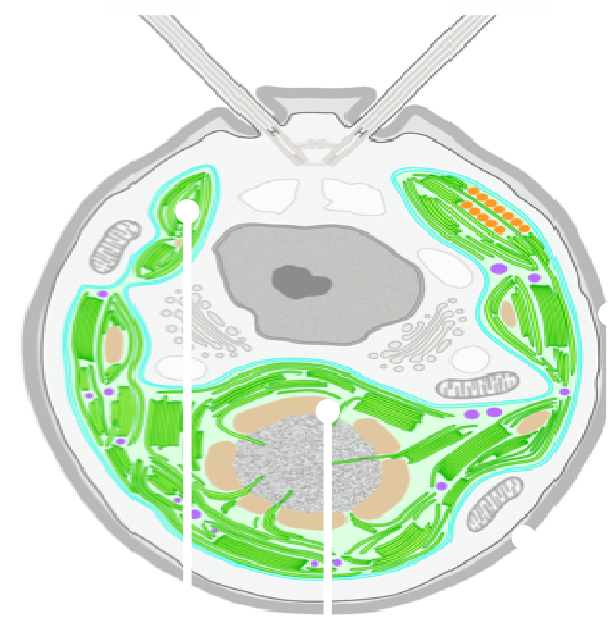
Engel, BD. *et al.* (2015) *Elife* 13;4



Cryo-FIB: Chlamydomonas



Cellular CET: 'fibbed' Chlamydomonas



Starch

Globules

Pyrenoid

Thylakoids

Engel, BD. *et al.* (2015) *Elife* 13;4



Cryo-FIB: *Chlamydomonas reinhardtii*

Engel, BD. *et al.* (2015) *Elife* 13;4

RuBisCO



CCD

Pyrenoid

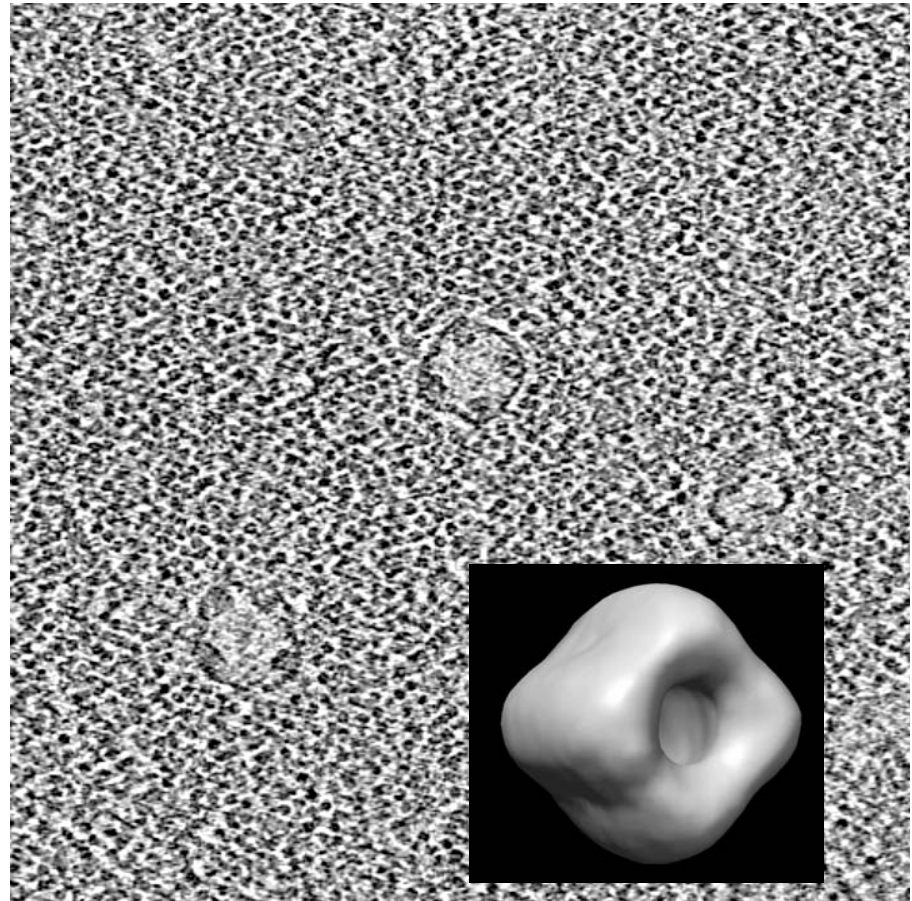


CRYO-FIB: *Chlamydomonas reinhardtii*

RuBisCO



CCD



Pyrenoid

K2 Summit

pixel size: 0.424 nm; 10,000 particles out of ~300,000 from ONE tomogram



Robust membrane segmentation based on tensor voting

The screenshot shows the memlabel2 software interface. The main window displays a 2D tomogram with segmented membranes in red and green. The interface includes several control panels:

- View:** Radio buttons for Original, Filter, Threshold, Label, Dir. filt., Cleft, and Material (selected).
- Set Material:** Display Cur... button, Label: 2.
- 2D Size Th.:** Update button, XY: 0, XZ: 0, YZ: 0.
- Results:** Save Labels, Save Cleft, Save Mate... buttons, Format: .mrc (selected), .em.
- Thresholding:** 10.26 Thresh., 48 Z slice, Crop Update button, X: 131 464, Y: 1 464, Z: 1 120.
- Size Threshold:** Update Lab... button, Display Curs... button, S. Th. 10.
- Cleft:** Distance, Cleft dist., Min: 0, Max: 0.
- Orientation:** Get Mb. Pt., Get PSR Pt., Get Orient. buttons.
- Dir. Filter:** Update Filtr., Dist.: 1, Sens: 0.
- Morph. Op.:** Erode, Dilate buttons.

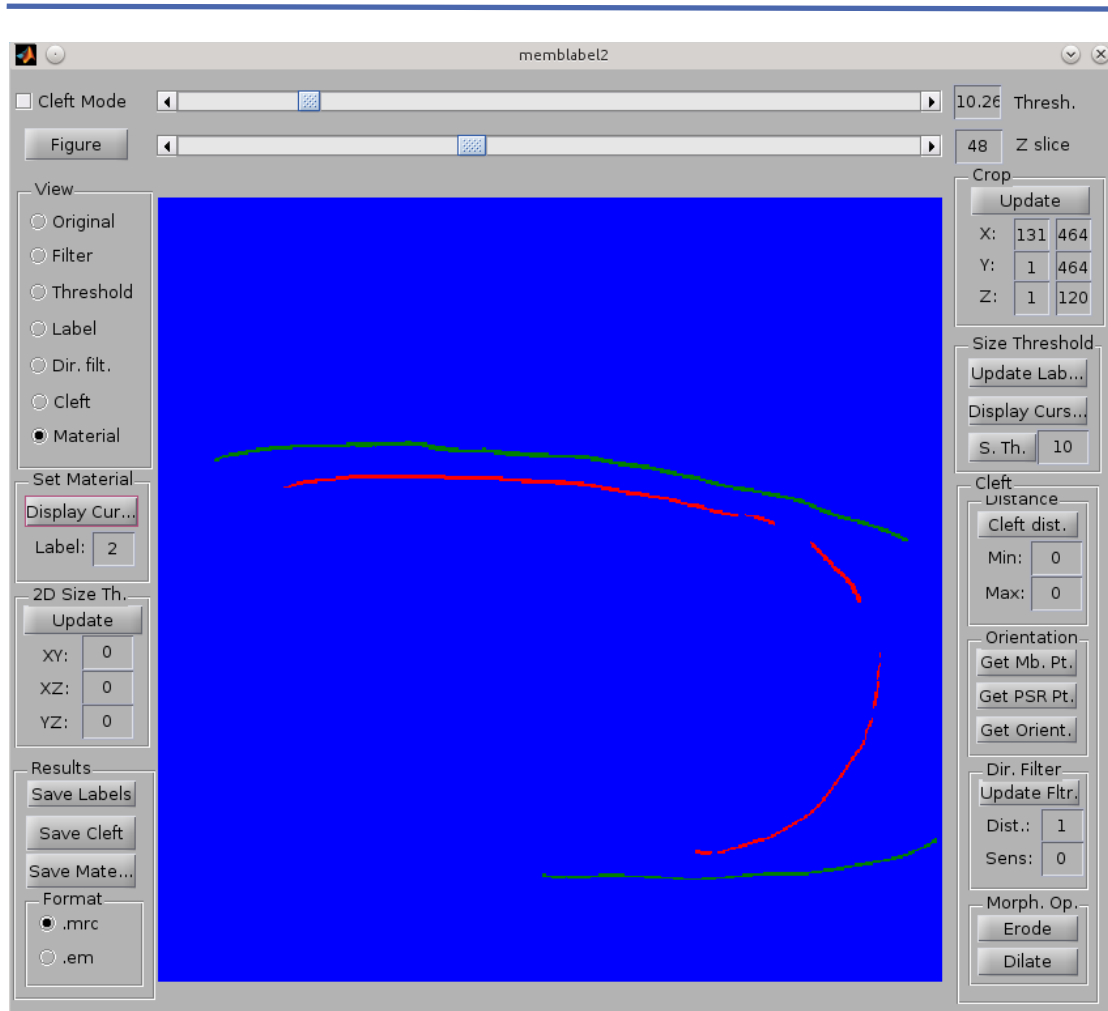
The diagram on the right illustrates the workflow:

- Original tomogram
- Tensor voting
- Filtered tomogram
- Segmentation by thresholding
- Volume-based labeling
- Materials

Martinez-Sanchez, A., et al., (2014) *J. Struct. Biol.* in press



Robust membrane segmentation based on tensor voting



The screenshot shows the memlabel2 software interface. The central window displays a blue tomogram with segmented membranes in red and green. The interface includes several control panels:

- View:** Radio buttons for Original, Filter, Threshold, Label, Dir. filt., Cleft, and Material (selected).
- Set Material:** A "Display Cur..." button and a "Label: 2" input field.
- 2D Size Th.:** "Update" button and input fields for XY: 0, XZ: 0, and YZ: 0.
- Results:** "Save Labels", "Save Cleft", "Save Mate...", and "Format" (radio buttons for .mrc and .em).
- Thresholding:** "10.26 Thresh.", "48 Z slice", "Crop" (Update), and "Size Threshold" (Update Lab..., Display Curs..., S. Th. 10).
- Cleft:** "Distance" (Cleft dist., Min: 0, Max: 0).
- Orientation:** "Get Mb. Pt.", "Get PSR Pt.", "Get Orient.".
- Dir. Filter:** "Update Fltr.", "Dist.: 1", "Sens: 0".
- Morph. Op.:** "Erode", "Dilate".

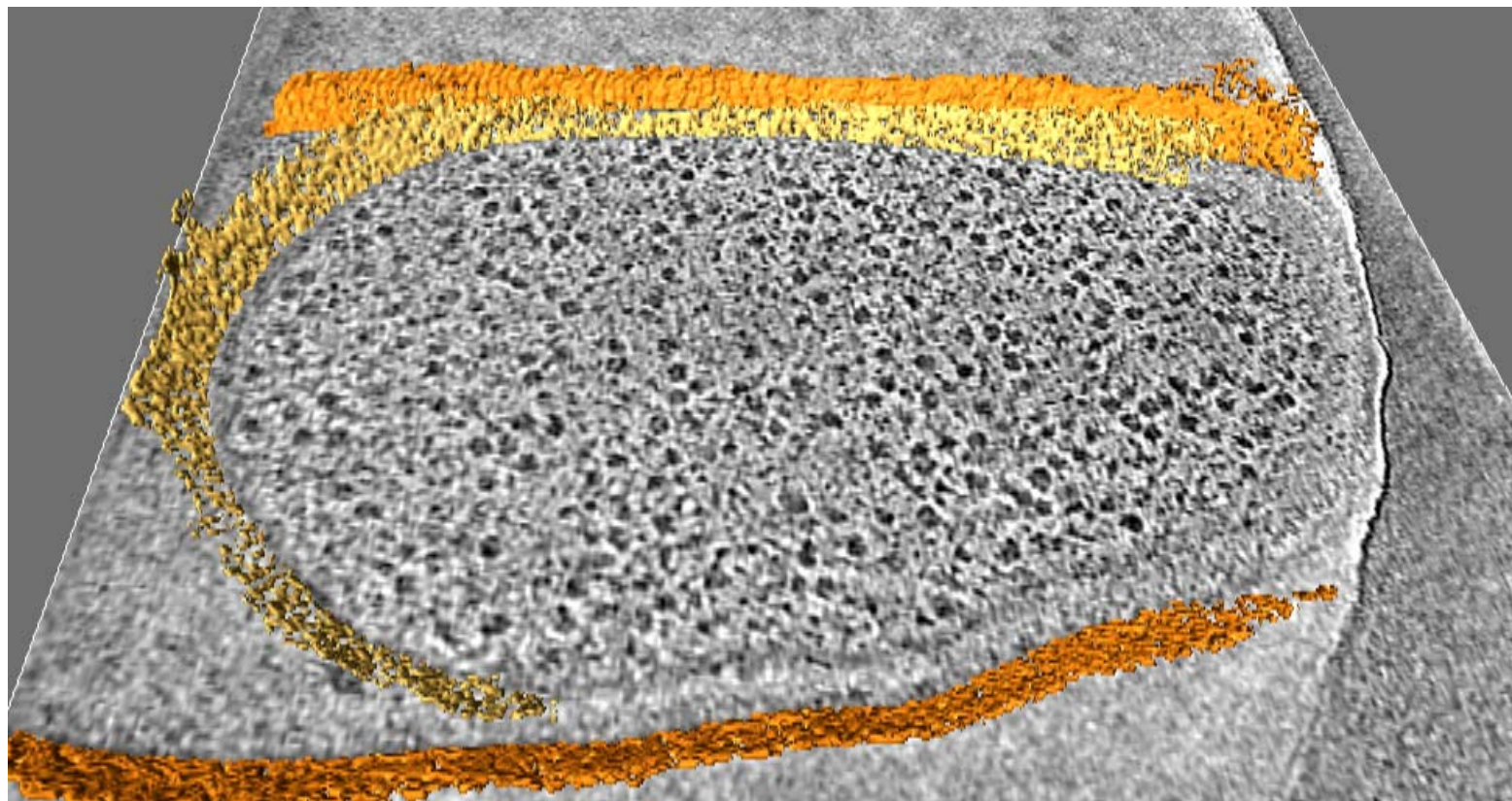
Flowchart illustrating the process:

- Original tomogram
- Tensor voting
- Filtered tomogram
- Segmentation by thresholding
- Volume-based labeling
- Materials


Martinez-Sanchez, A., et al., (2014) *J. Struct. Biol.* in press



Robust membrane segmentation based on tensor voting

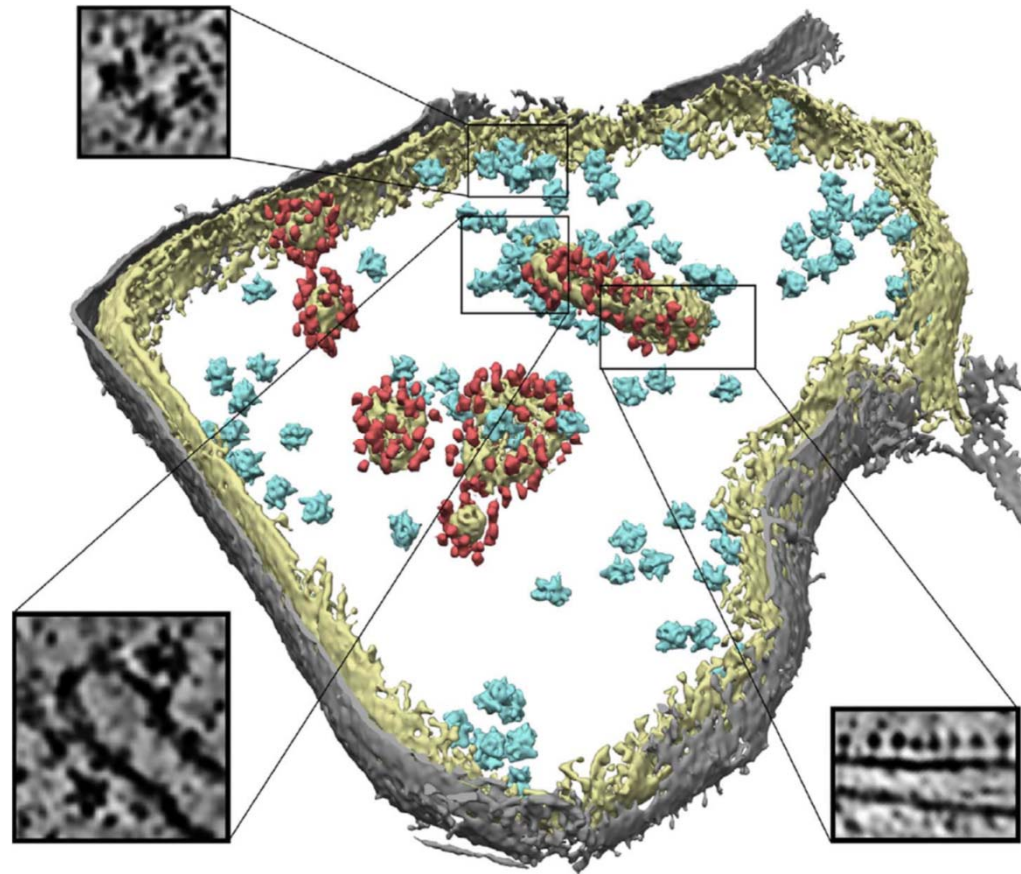


 segmented outer membrane

 segmented inner membrane



Organization of the mitochondrial translation machinery studied *in situ* by cryoelectron tomography

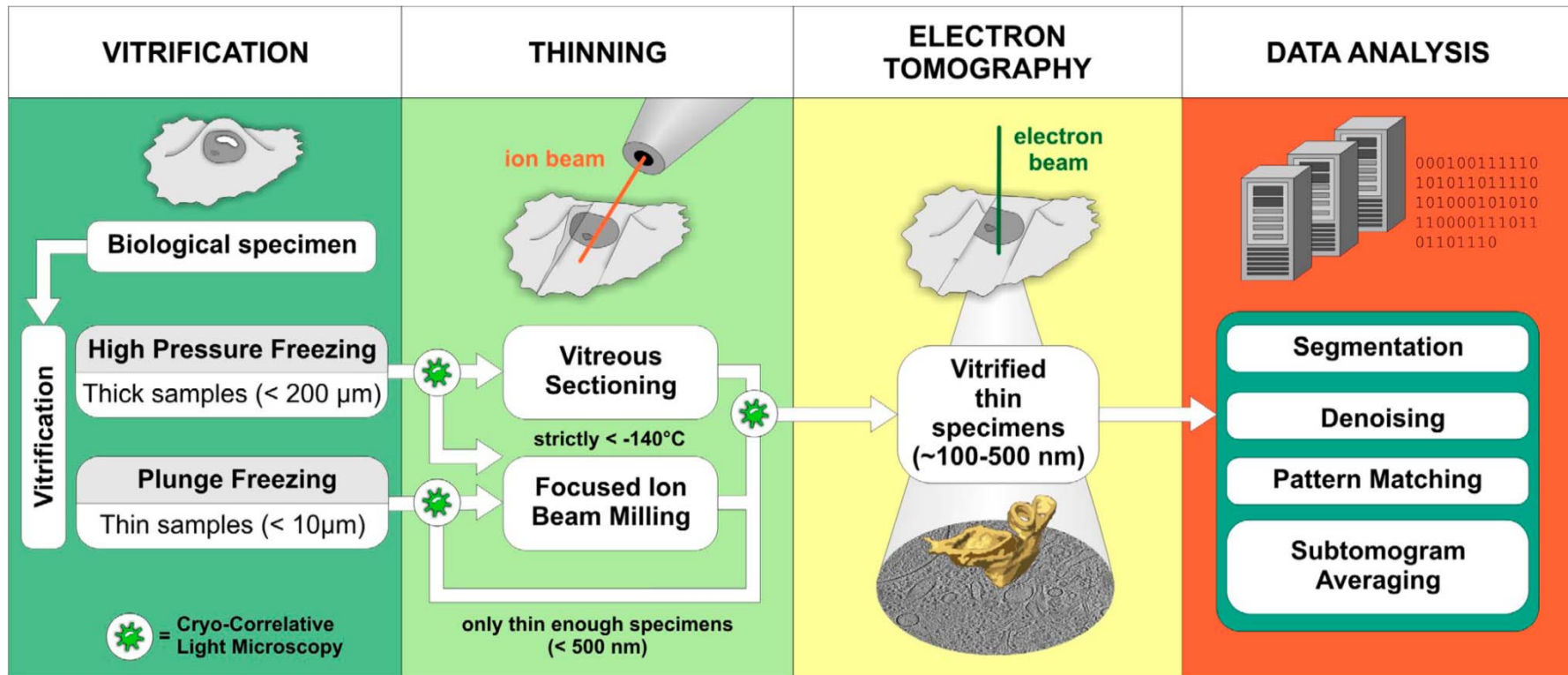


Distribution of mitoribosomes in translation-competent yeast mitochondria.

Pfeffer, S. et al., (2015)
Nature Communications



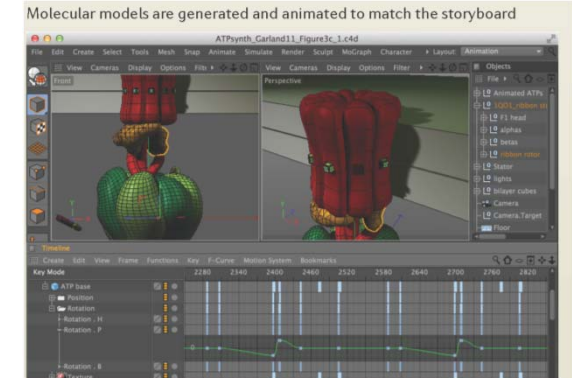
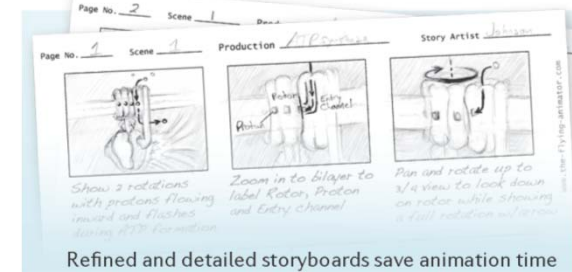
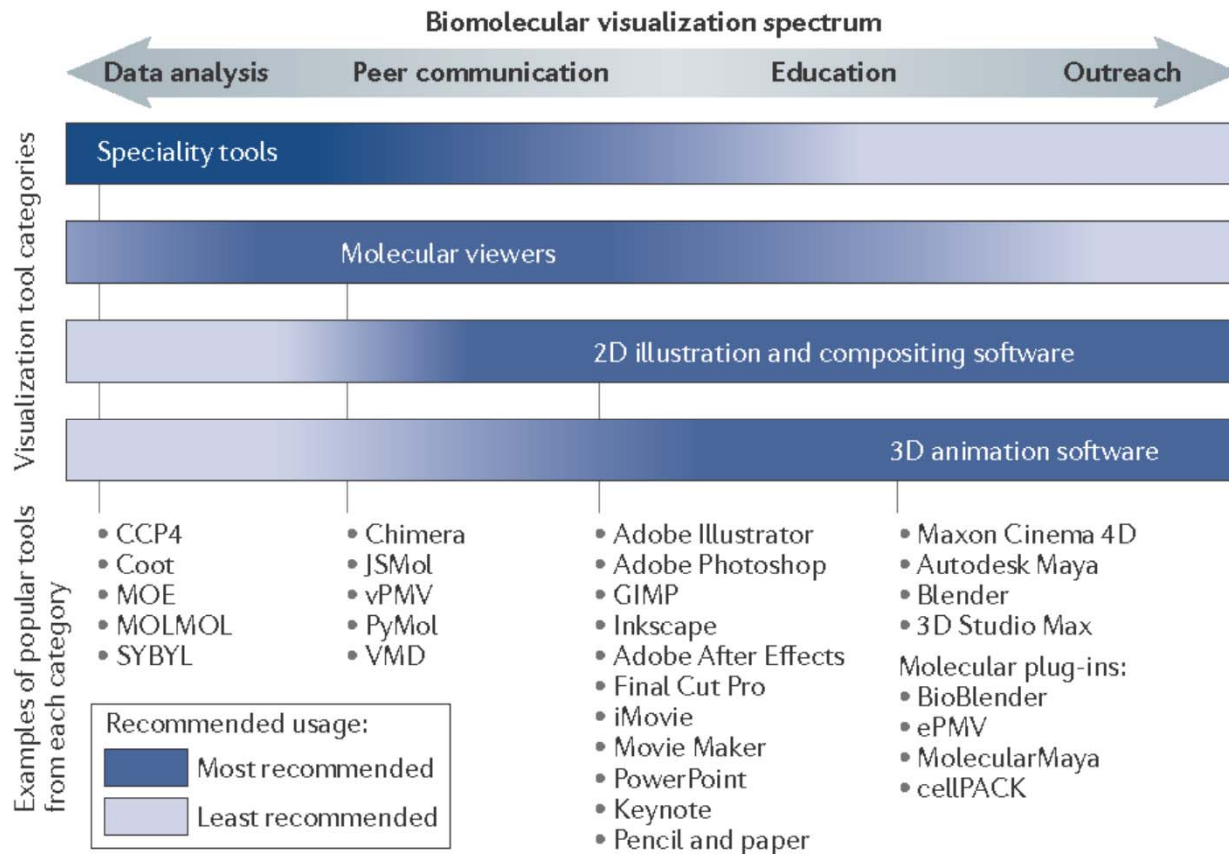
Schematic representation of the cryo-ET workflow



Lučić, V. et al. *JCB* (2014)



Biomolecular Visualization Tools



Johnson, G and Hertig, S. *Nat Rev Mol Cell Biol.* (2014) 10:690-8.





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