



instruct
Integrating
Biology

I2PC

Instruct Image Processing Centre

JM CARAZO

Image Processing in SPA: Principles and Workflows

José María Carazo (carazo@cnb.csic.es)

Spanish National Center for Biotechnology

Instruct Image Processing Center

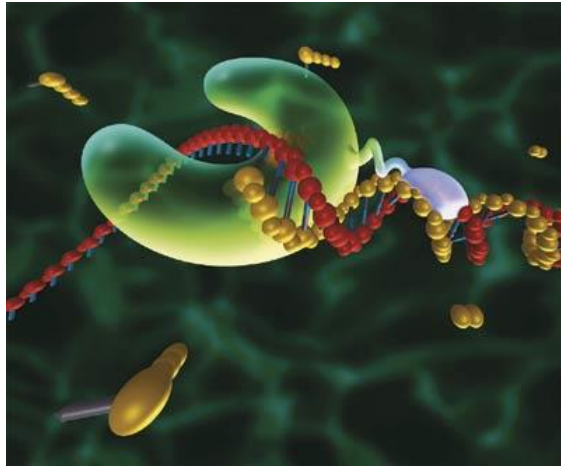


Cryo-EM Course Lectures Outline

Monday	
13:00-13:45	JO: General introduction to three-dimensional cryo microscopy
13:45-14:30	SdC: Sample preparation and vitrification
15:00-16:30	MW: Modelling/Fitting
Tuesday	
09:00-9:45	JMC: Single Particle Analysis (SPA): The basics
09:45-10:30	JMC: Image processing workflows for SPA
Thursday	
09:00-09:45	JO: Image processing workflows for Tomography
09:45-10:30	SdC: New EM technologies



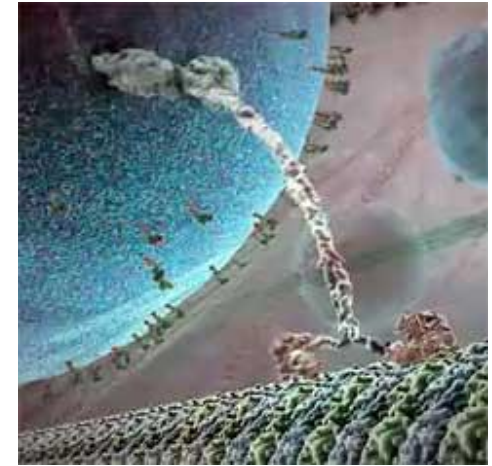
Life is based on macromolecular machines



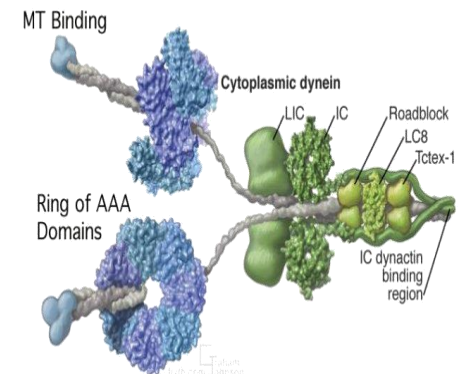
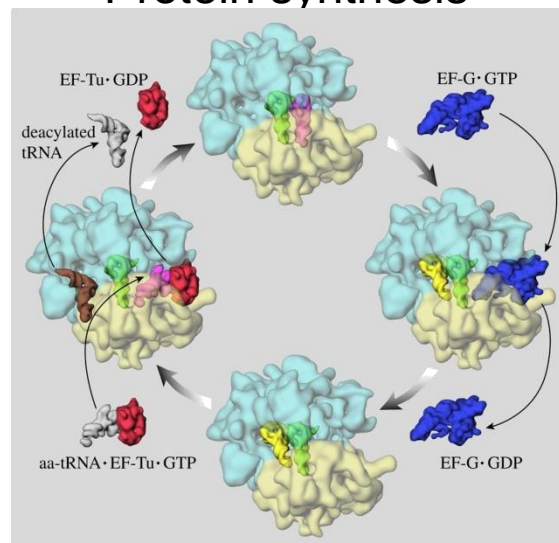
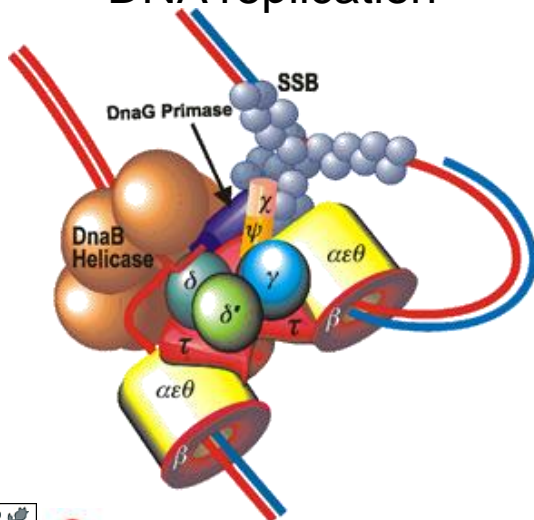
DNA replication



Protein synthesis



Dynein motion



This is our objective!

ARTICLE

doi:10.1038/nature12822

Structure of the TRPV1 ion channel determined by electron cryo-microscopy

Maofu Liao^{1*}, Erhu Cao^{2*}, David Julius² & Yifan Cheng¹

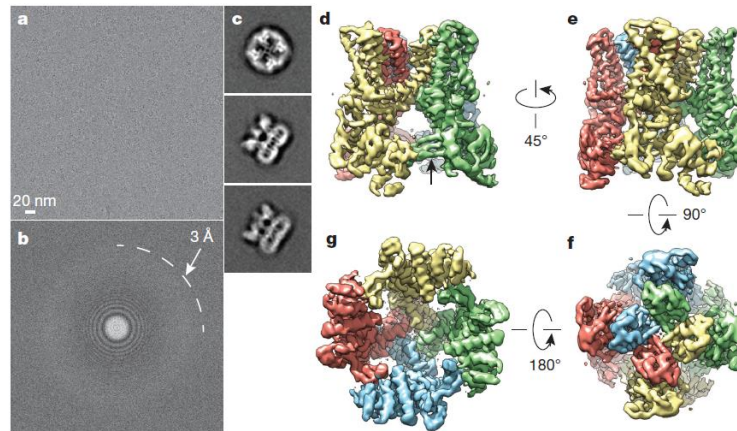
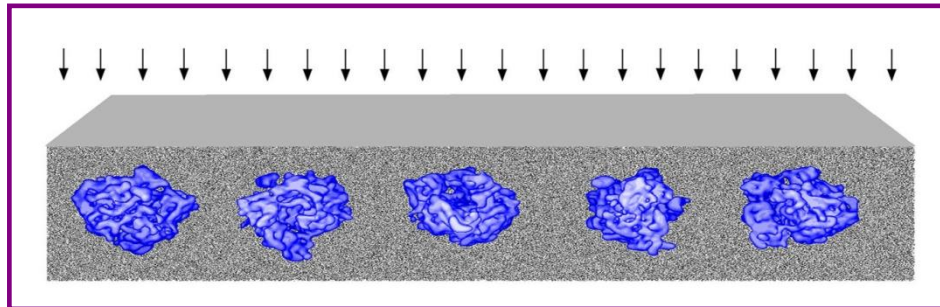


Figure 1 | 3D reconstruction of TRPV1 determined by single-particle cryo-EM. **a**, Representative electron micrograph of TRPV1 protein embedded in a thin layer of vitreous ice recorded at a defocus of 1.7 μm . **b**, Fourier transform of micrograph shown in **a**, with Thon rings extending to nearly 3 \AA . **c**, Enlarged views of three representative 2D class averages show fine features of

tetrameric channel complex. **d-g**, 3D density map of TRPV1 channel filtered to a resolution of 3.4 \AA (scaled to atomic structure) with each subunit colour-coded. Four different views of the channel are shown, from side (**d, e**), top (**f**) and bottom (**g**). The arrow in panel **d** indicates β -sheet structure in the cytosolic domain of TRPV1.

The cryo-EM SPA pledge

- In 3D Electron Microscopy *individual macromolecules* are visualized down to *atomic resolution*.
- Trapped in ice, these molecules are free to expose their internal flexibility/plasticity.



The cryo-EM SPA pledge

- In 3D Electron Microscopy *individual macromolecules* are visualized down to *atomic resolution*
 - *Attention to every detail of the image formation process*
 - *Very precise image processing*
- Trapped in ice, these molecules are free to expose their internal flexibility/plasticity
 - *Need to classify individual images in 3D*



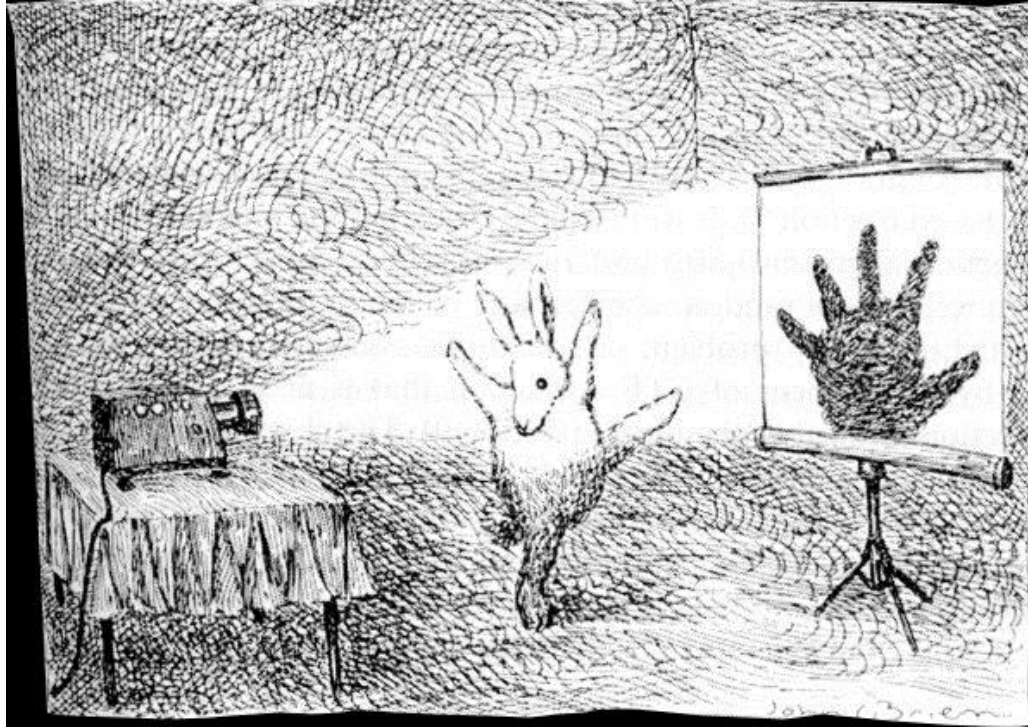
The cryo-EM SPA pledge

Attention to every detail!

- Characterization of each image
- A posteriori characterization of the Projection Geometry
- 3D reconstruction process
- 3D classification
- Validation



As we were saying yesterday



2D projections : lack of information

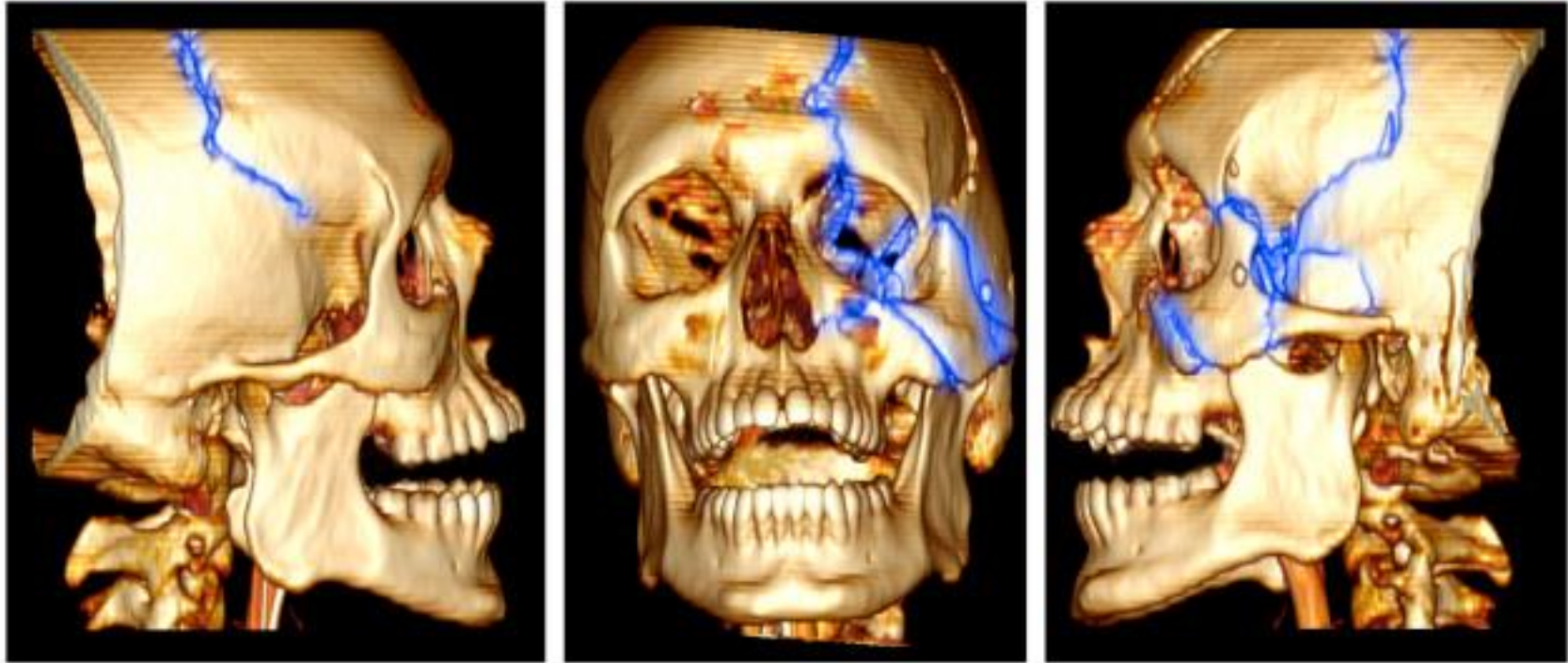


Limits the comprehension
of complex objects

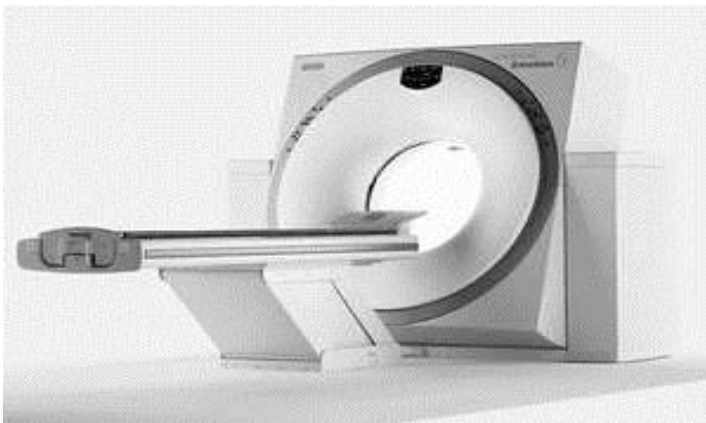
The value of a «radiography»



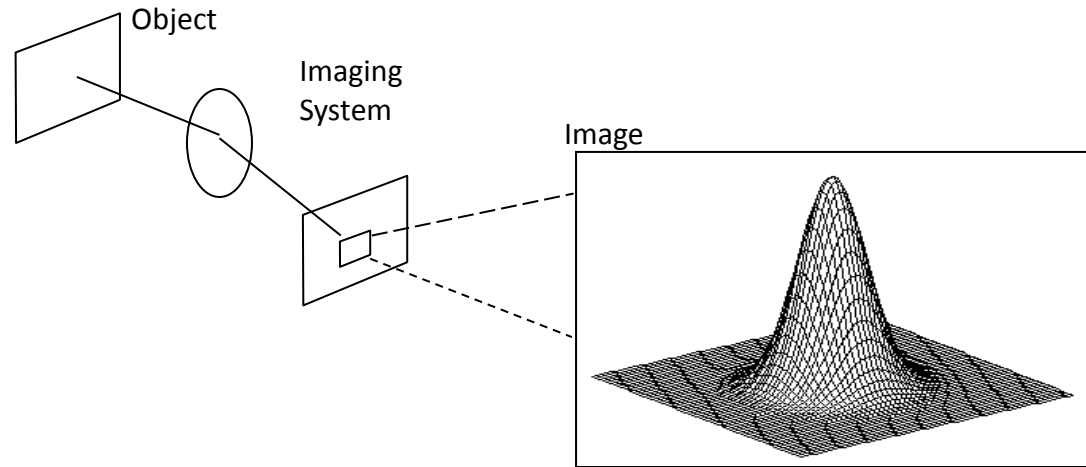
Compared to full 3D CT



Compared to full 3D CT

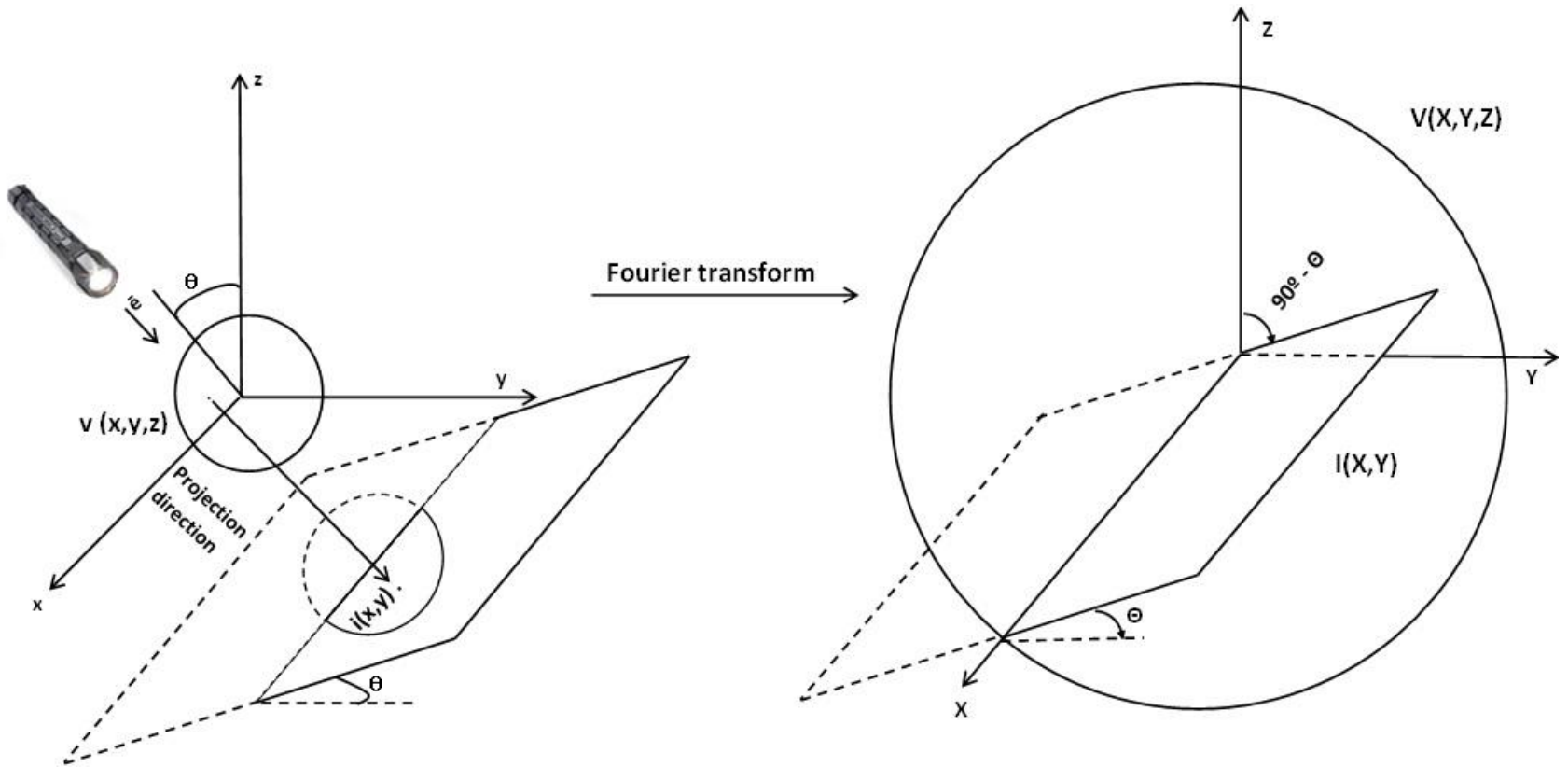


Realistic Image Formation Model: CTF characterization

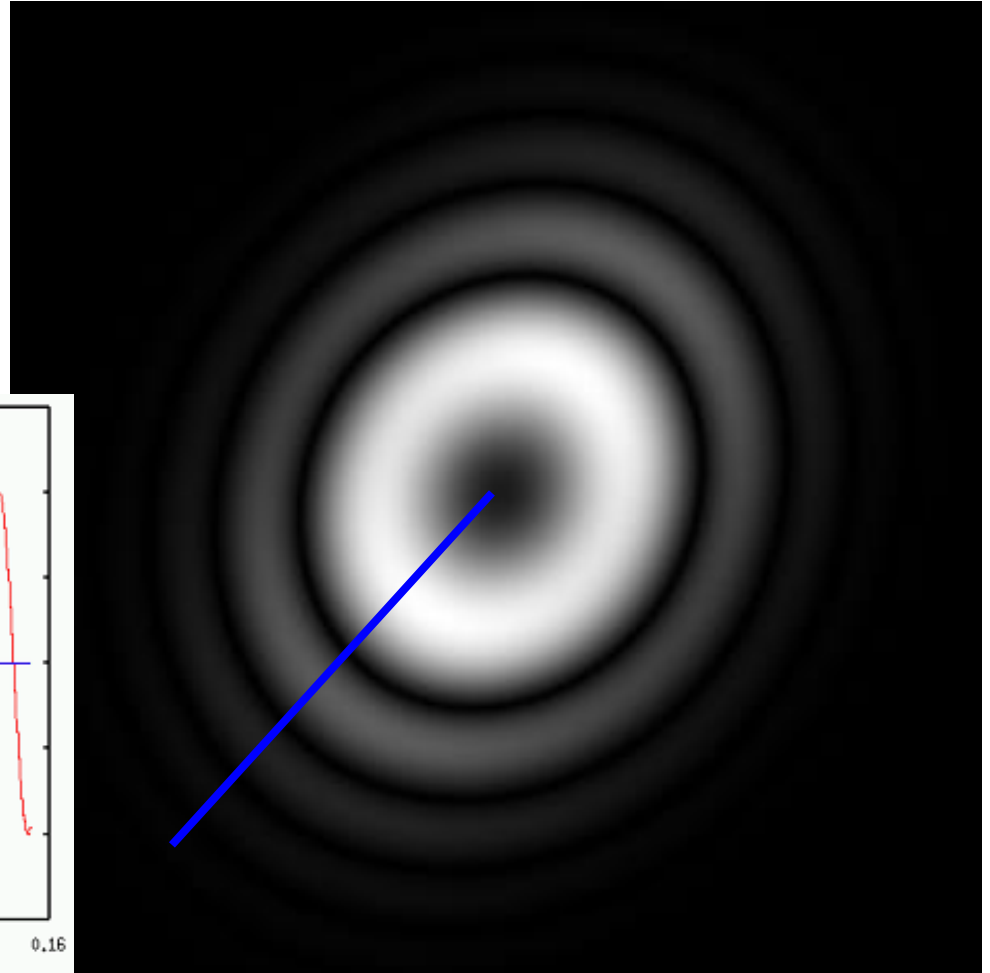
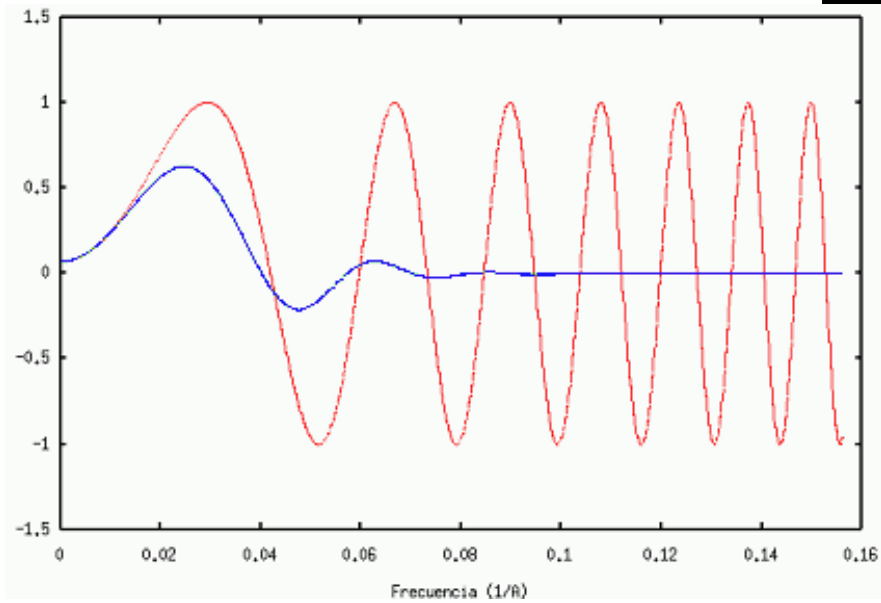


PSF:
Point Spread Function

Real space and Fourier Space



CTF Profile in Fourier Space



CTF Challenge

1 CTF Challenge: Result Summary

2 Roberto Marabini⁽¹⁾, Bridget Carragher⁽²⁾, Shaoxia Chen⁽⁹⁾,
James Chen⁽¹³⁾, Anchi Cheng⁽²⁾, Kenneth H. Downing⁽⁴⁾, Joachim Frank⁽⁵⁾,
Robert A. Grassucci⁽⁵⁾, Bernard Heymann⁽¹²⁾, Wen Jiang⁽⁶⁾,
Slavica Jonic⁽¹⁰⁾, Hstau Y. Liao⁽⁵⁾, Steven J. Ludtke⁽³⁾, Shail Patwari⁽¹¹⁾,
Angela L. Piotrowski⁽¹¹⁾, Adrian Quintana⁽⁷⁾, Carlos O.S. Sorzano⁽⁷⁾, Henning Stahlberg⁽⁸⁾,
Javier Vargas⁽⁷⁾, Neil R. Voss⁽¹¹⁾, Wah Chiu⁽³⁾, Jose M. Carazo⁽⁷⁾

3 August 4, 2014

4 ⁽¹⁾Escuela Politécnica Superior, Universidad Autónoma de Madrid, 28049
5 Cantoblanco, Madrid, Spain.

6 ⁽²⁾ The National Resource for Automated Molecular Microscopy, The Scripps
7 Research Institute, La Jolla, CA 92037, USA

8 ⁽³⁾ Baylor College of Medicine, Houston, Texas 77030, USA



CTF Challenge

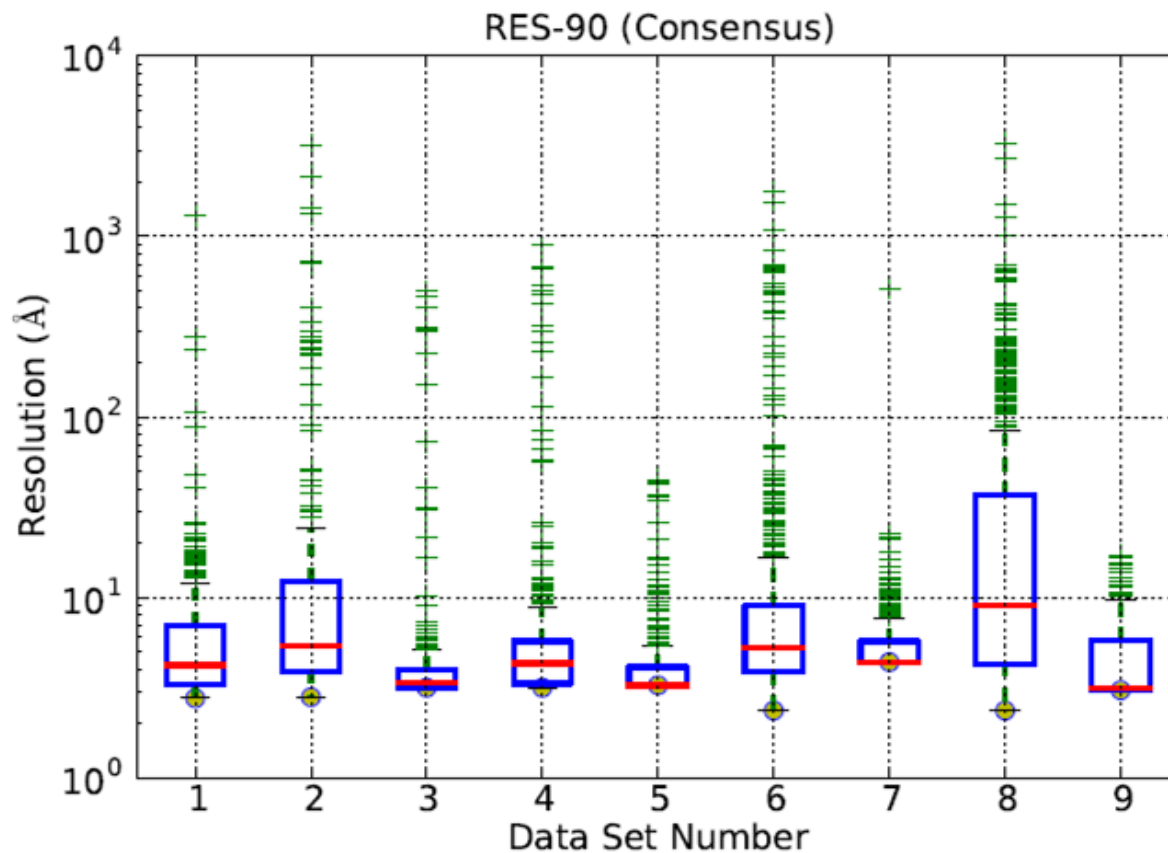


Figure 9: RES-90 analysis. X-axis ticks refer to data sets, Y-axis represents an estimation of the resolution limit imposed by the accuracy in the CTF determination. Yellow circles show the Nyquist frequency for each data set.

How precise should we be?

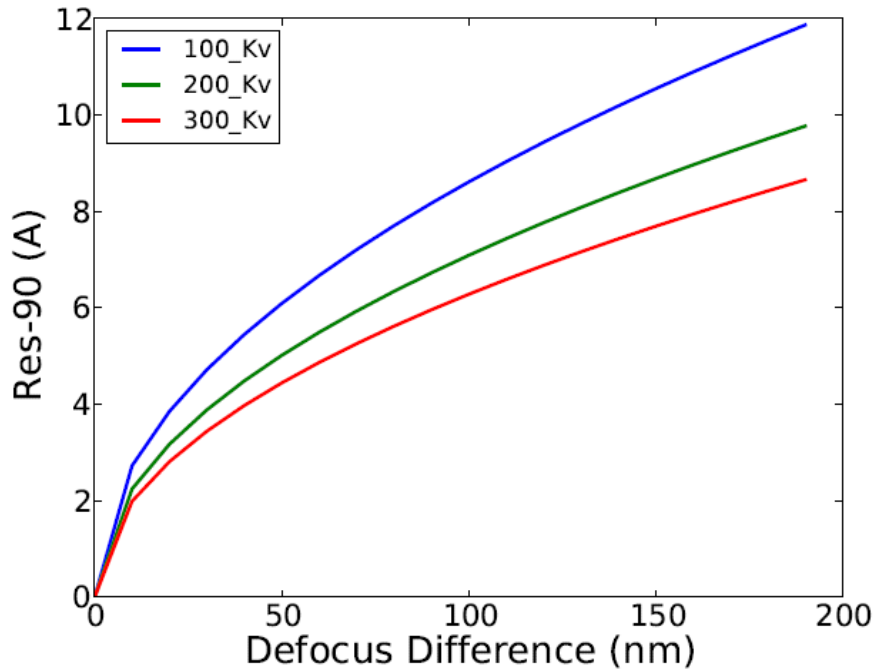


Figure 8: Resolution at which the wave aberration shift introduced by a given defocus error is 90° . Note that this magnitude depends only on the defocus error and not on the actual amount of defocus. Additionally, note that the plot would be the same if instead of considering two non-astigmatic CTF estimations we would consider the defocus difference between the two astigmatic axis, assuming no errors in astigmatic angle estimation.

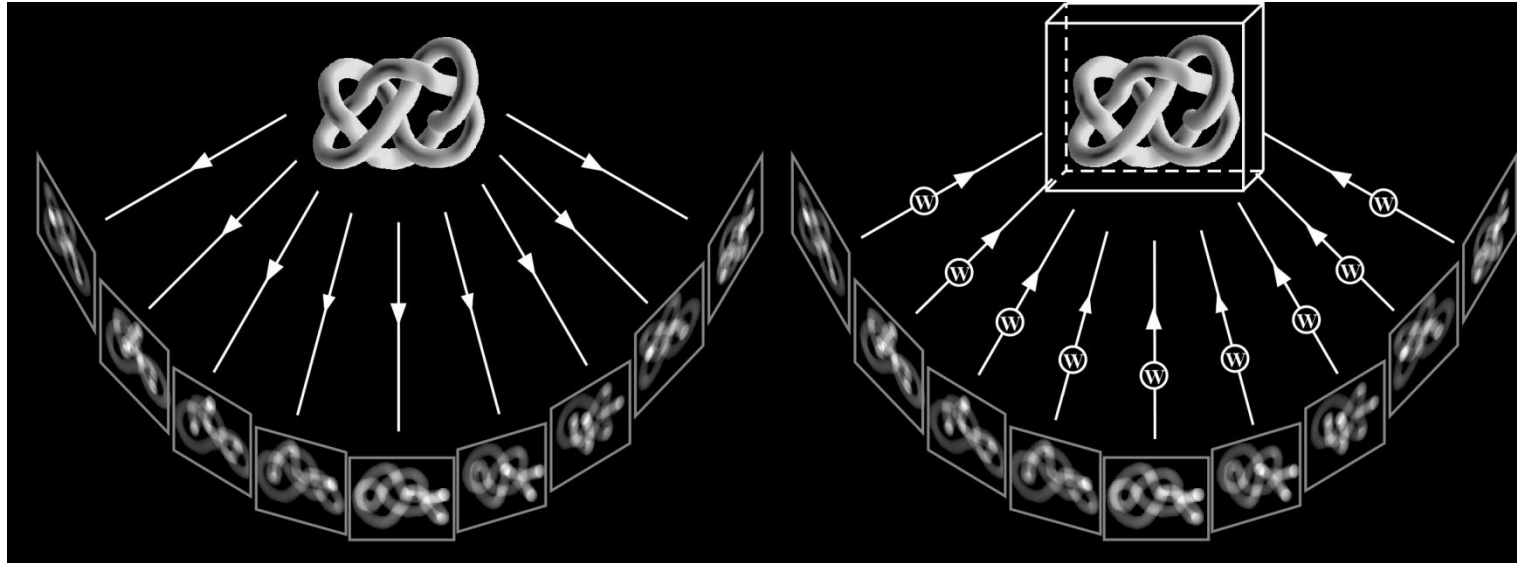
The cryo-EM SPA pledge

Attention to every detail!

- Characterization of each image
- A posteriori characterization of the Projection Geometry
- 3D reconstruction process
- 3D classification
- Validation



Tomography Principle



Acquisition of
tilted image series

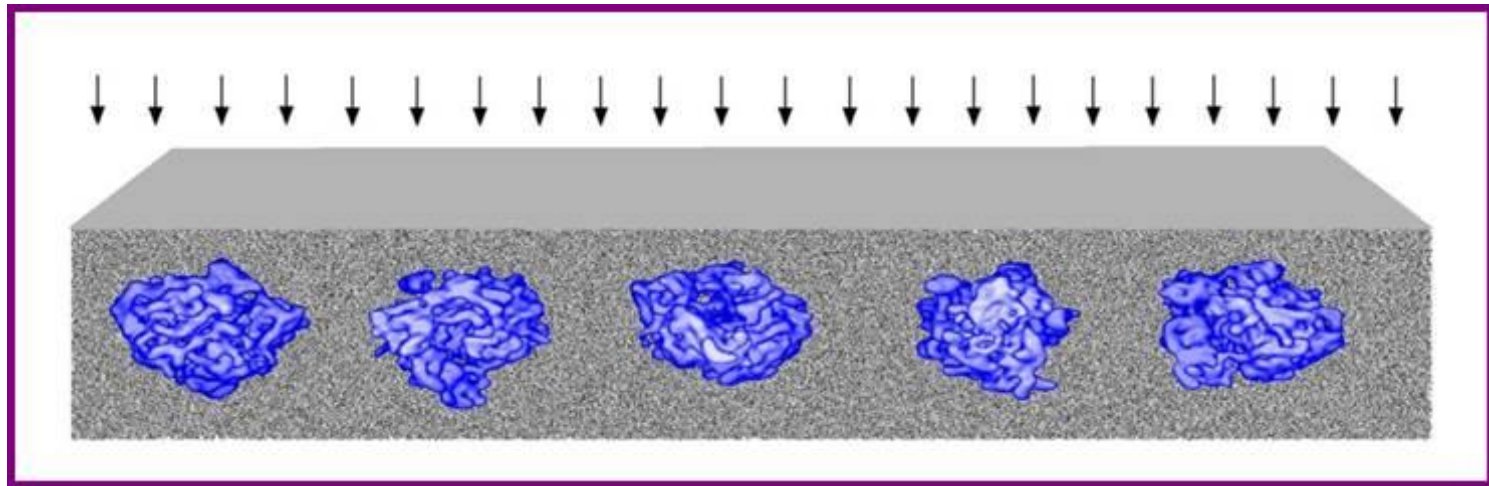


Correction of
microscope
default
(mechanical drift,
CTF...)



Reconstruction

That does not apply to SPA



Parameter space

- For each particle we need to determine 3 angles and 2 shifts. FIVE parameters.
- If we have 100.000 particle images.
- We then have a space of 500.000 parameters!



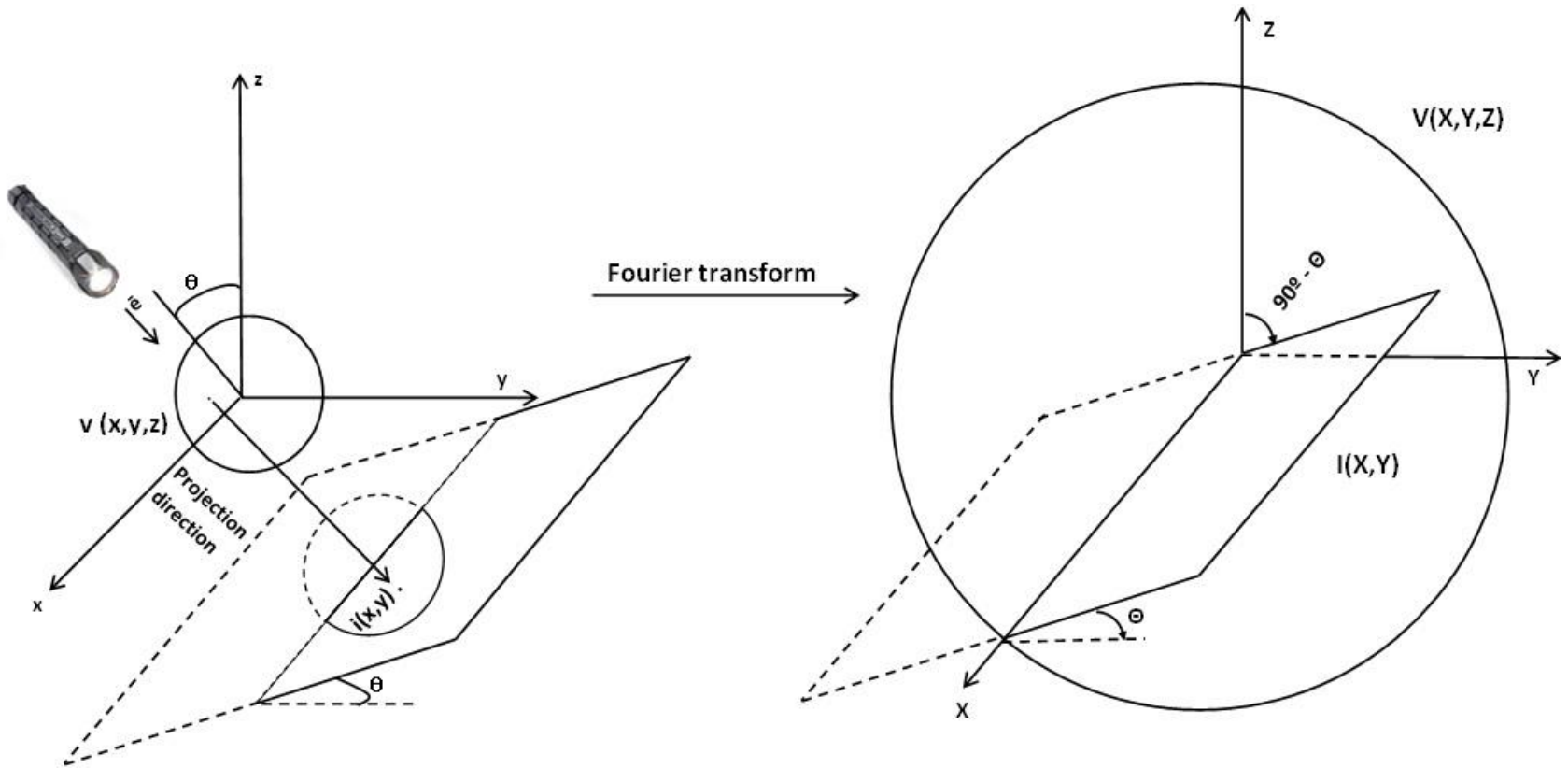
The cryo-EM SPA pledge

Attention to every detail!

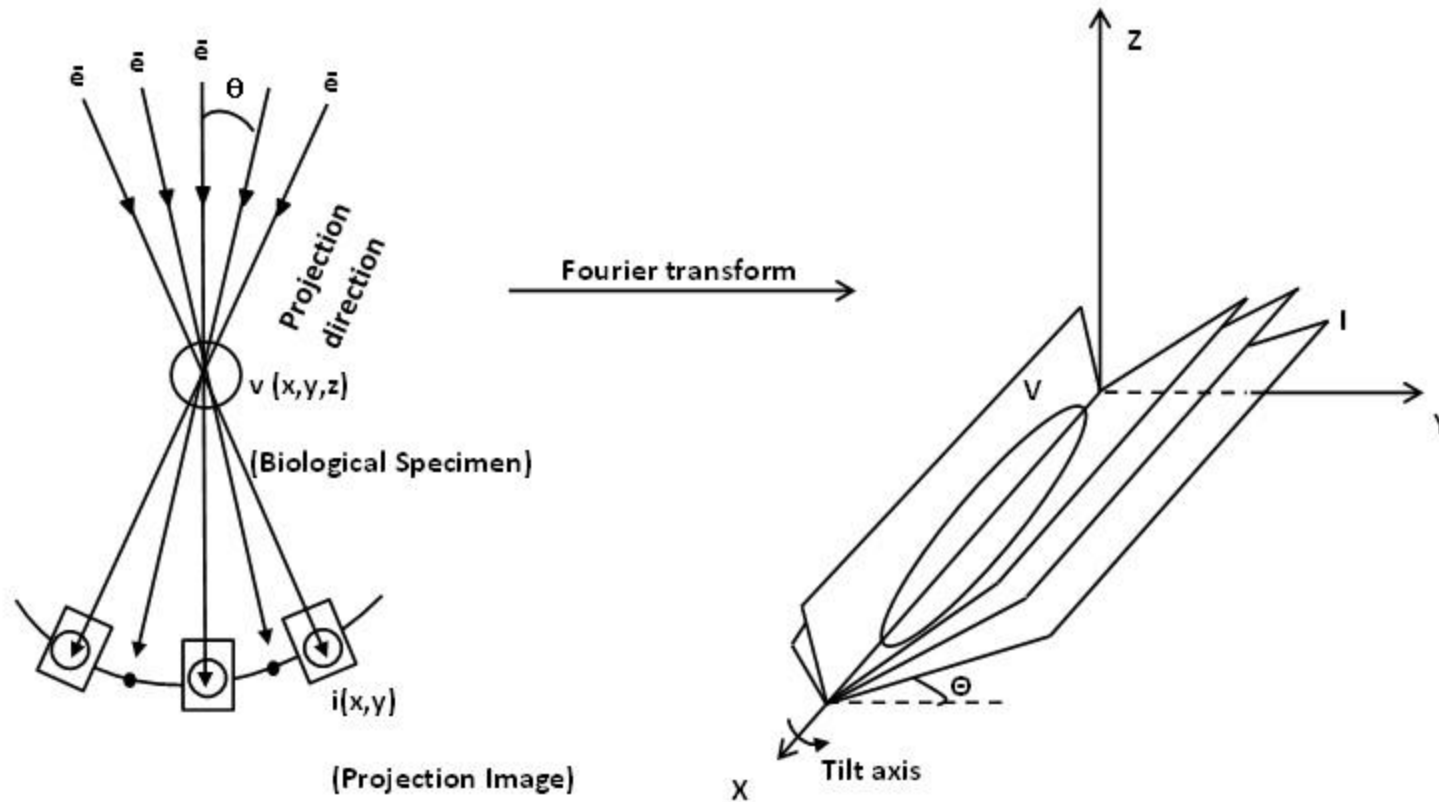
- Characterization of each image
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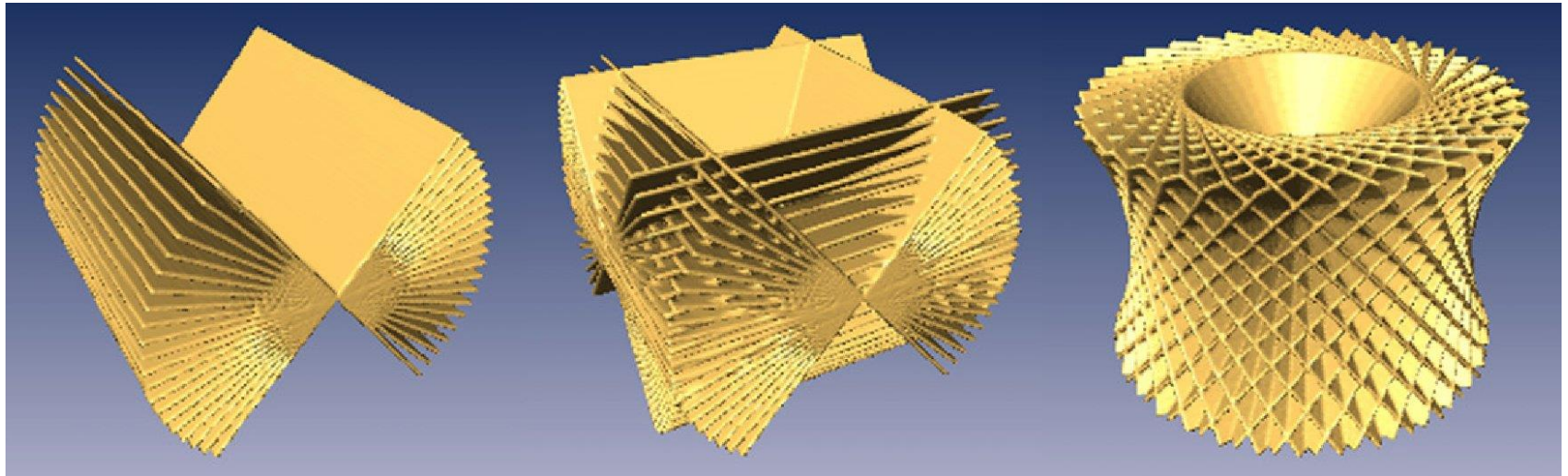
Real space and Fourier Space



Principles of Fourier Reconstruction Method



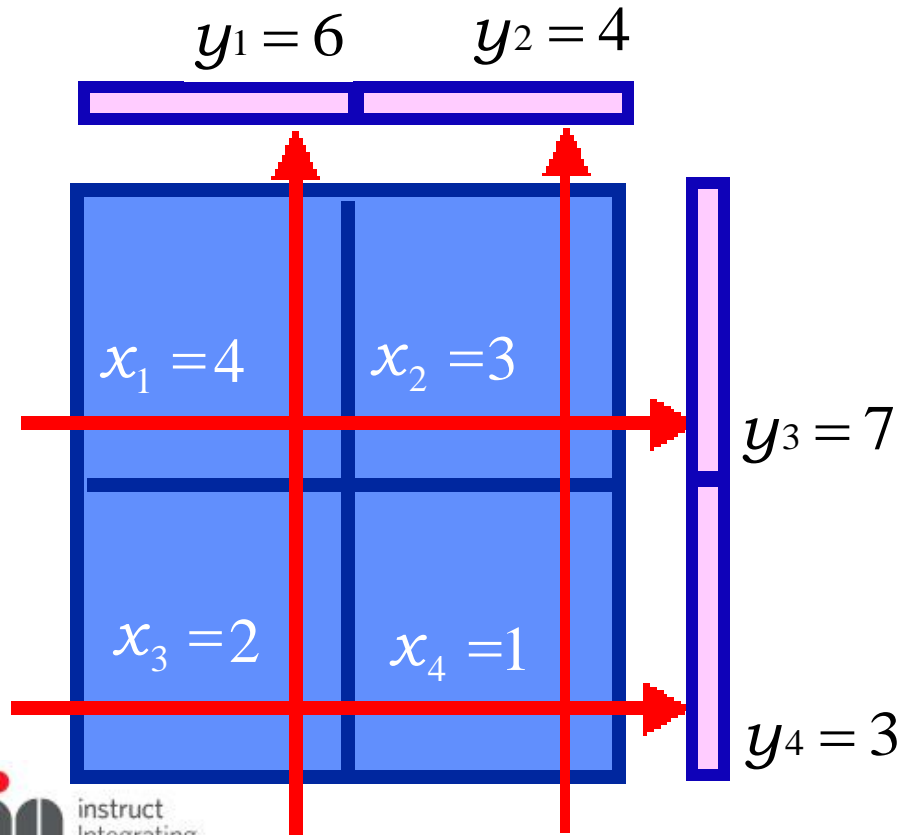
More complex geometries in Fourier space



$$f(\mathbf{r}) \approx \sum_{j=1}^J x_j b_j(\mathbf{r})$$

Reconstruction as a linear set of equations

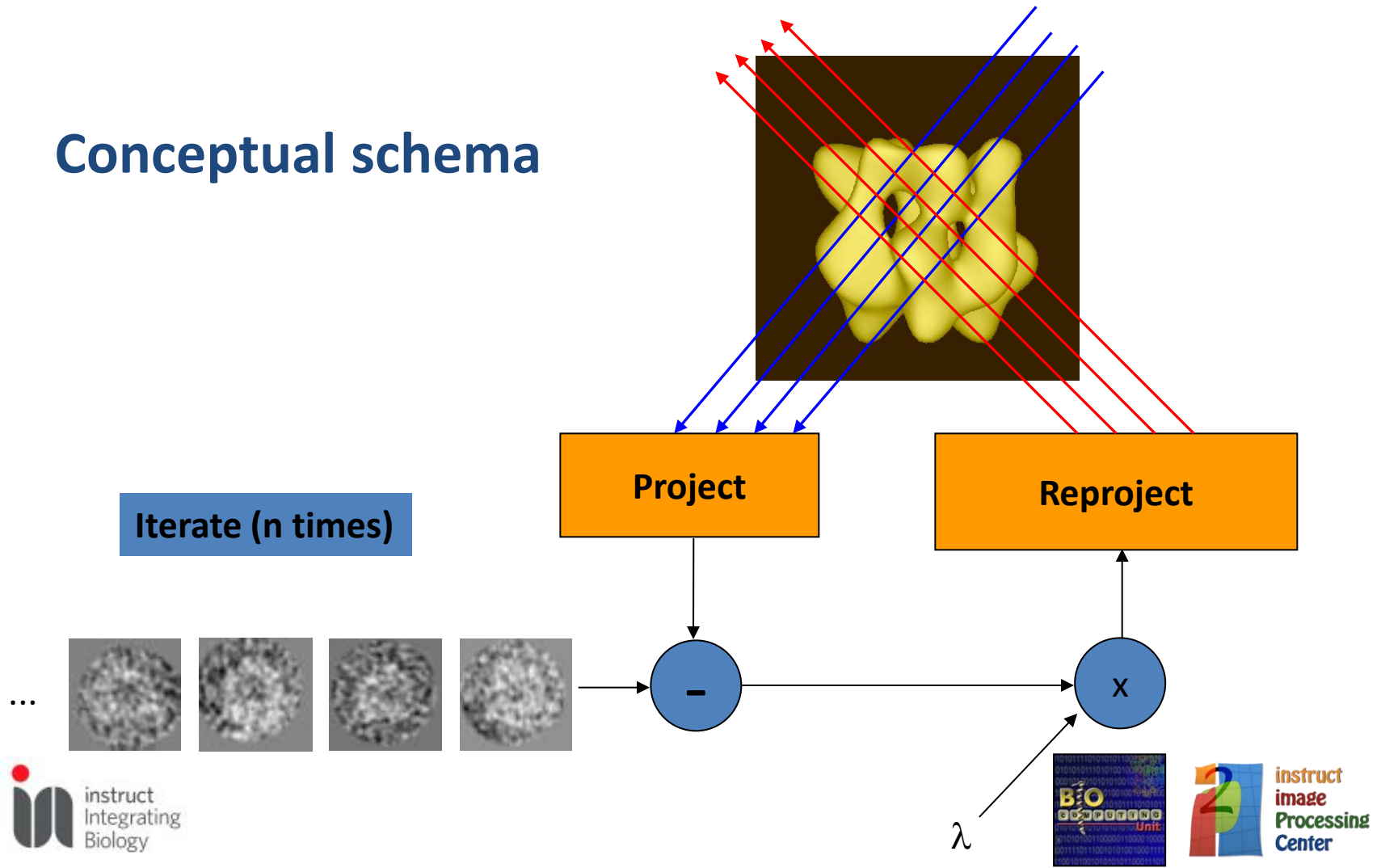
$$y_i \approx \sum_{j=1}^J l_{i,j} x_j \quad l_{i,j} = 1,0$$



$$\left\{ \begin{array}{l} x_1 + x_3 = 6 \\ x_2 + x_4 = 4 \\ x_1 + x_2 = 7 \\ x_3 + x_4 = 3 \end{array} \right.$$

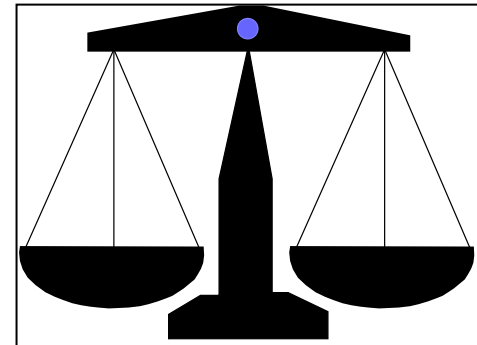
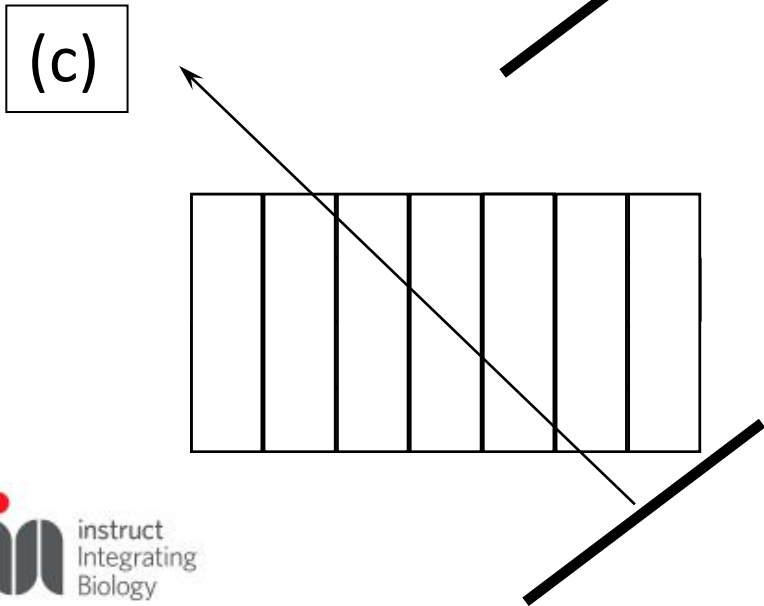
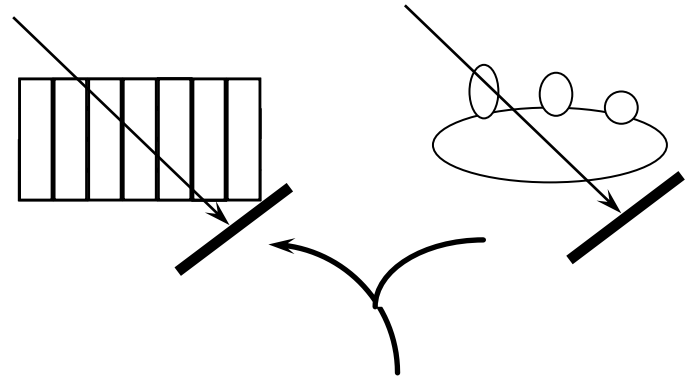
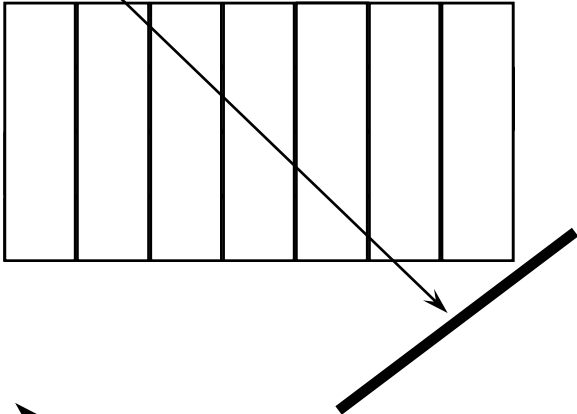
ART, the “basics”

Conceptual schema



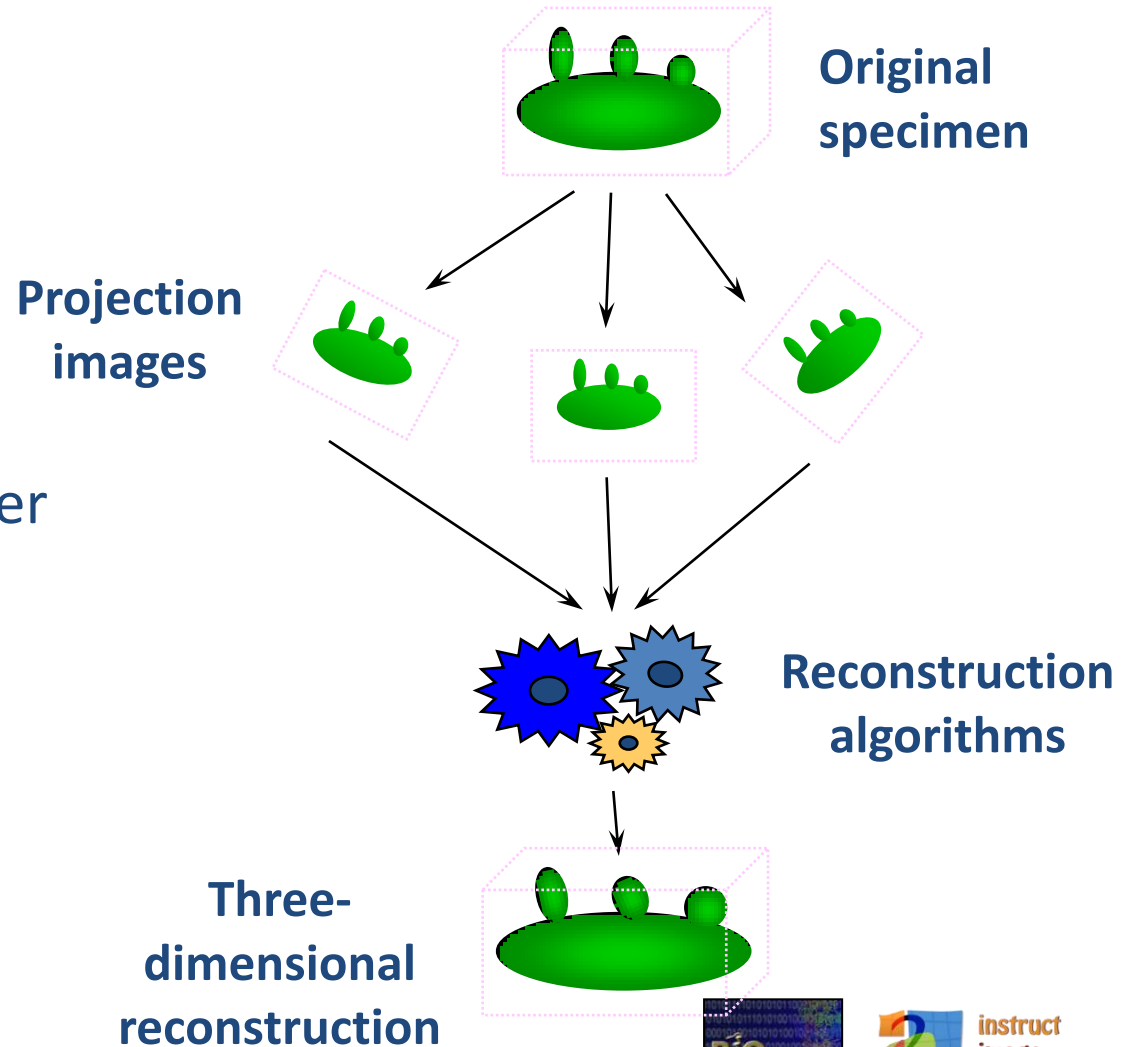
ART, the “basics”

(a) (b)



3D reconstruction approach:

- Limited number of projections
- Image noise
- Partial lack of control over particle homogeneity
- Particle lack of control over data collection geometry



The cryo-EM SPA pledge

Attention to every detail!

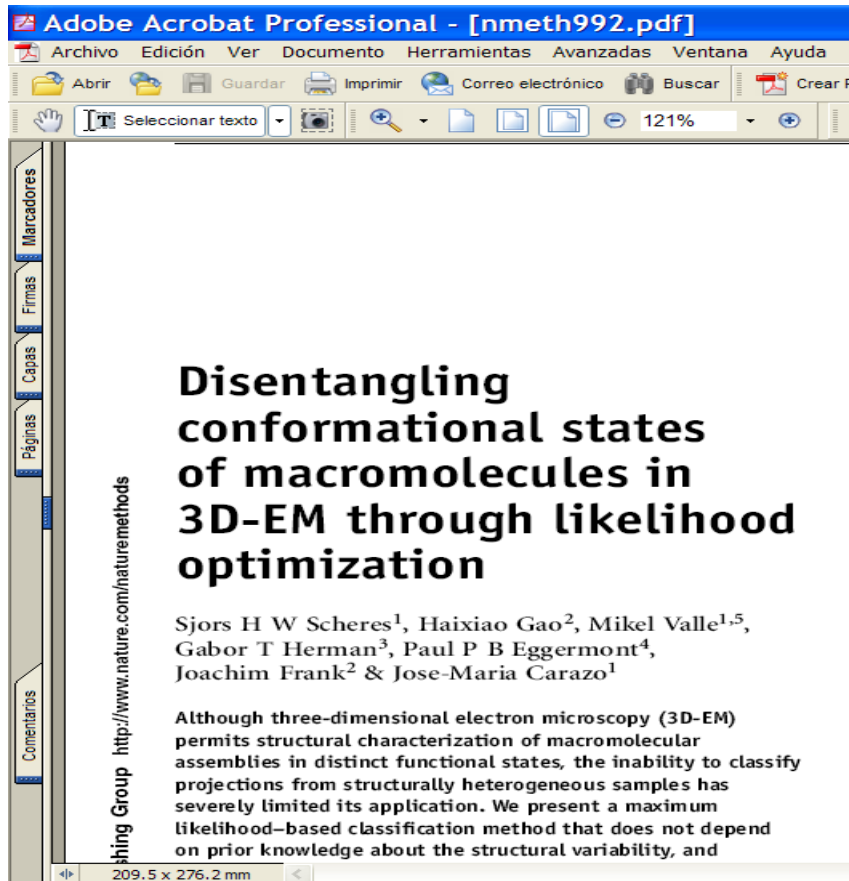
- Characterization of each image
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The 3D flexibility challenge



The 3D flexibility challenge



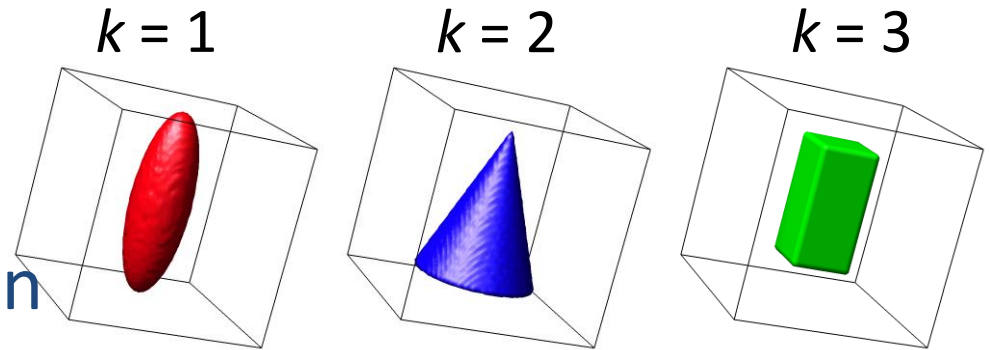
position in each image. The computational effort, using some 4,000 CPU hours on a computer cluster, is perhaps the most audacious application of the expectation-maximization algorithm ever performed. It also showcases an extremely powerful new tool for structural biology.

NOW in RELION



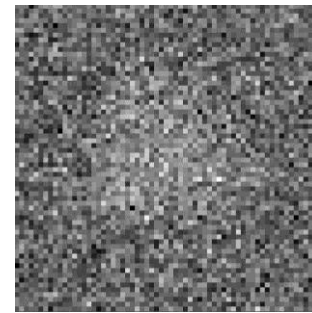
Statistical model

Each image is a projection of one of K underlying 3D objects k



with addition of noise

Unknowns: the 3D objects k , orientations

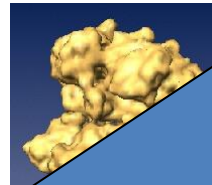
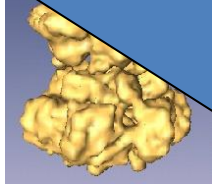
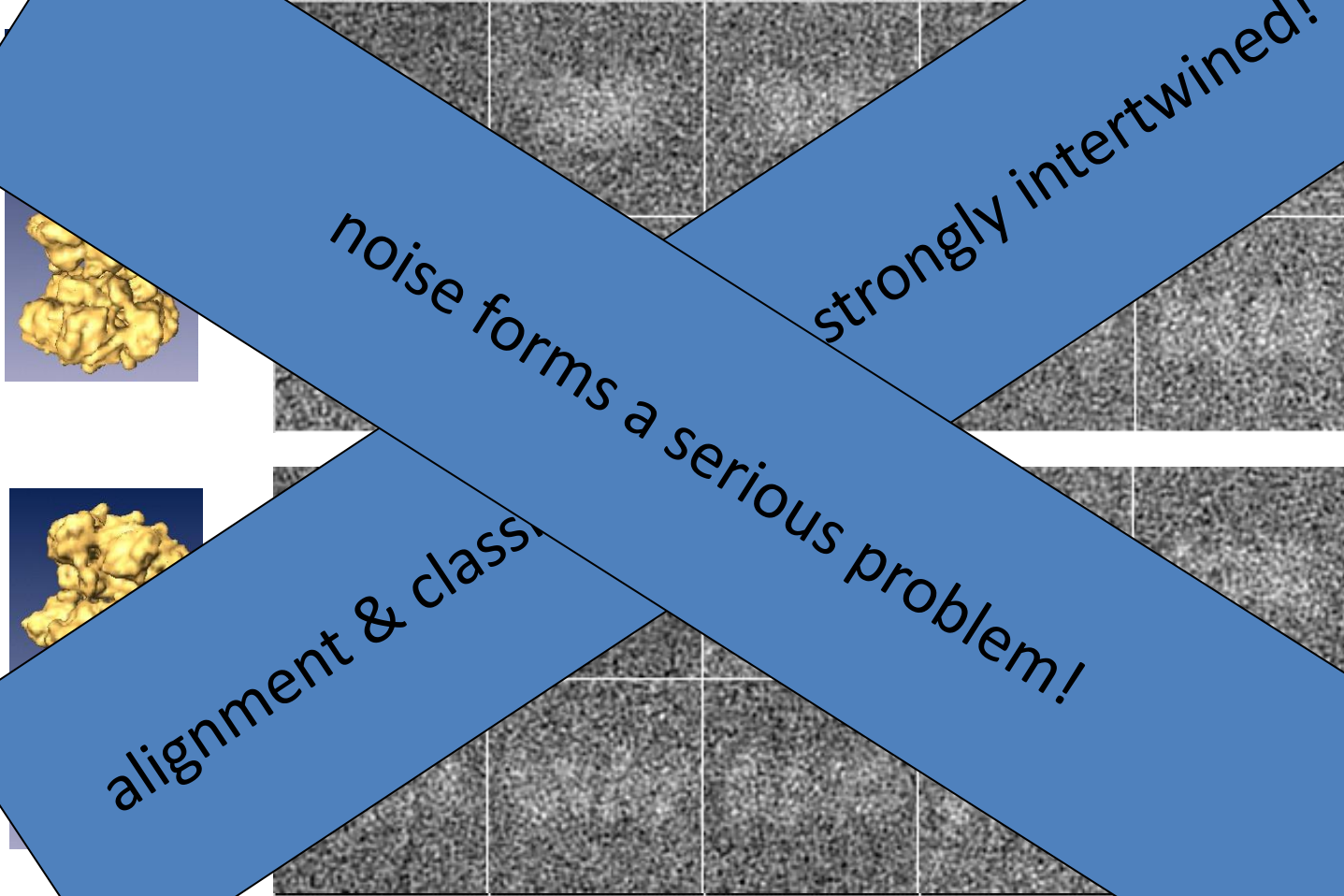


Parameter space of cryo EM SPA

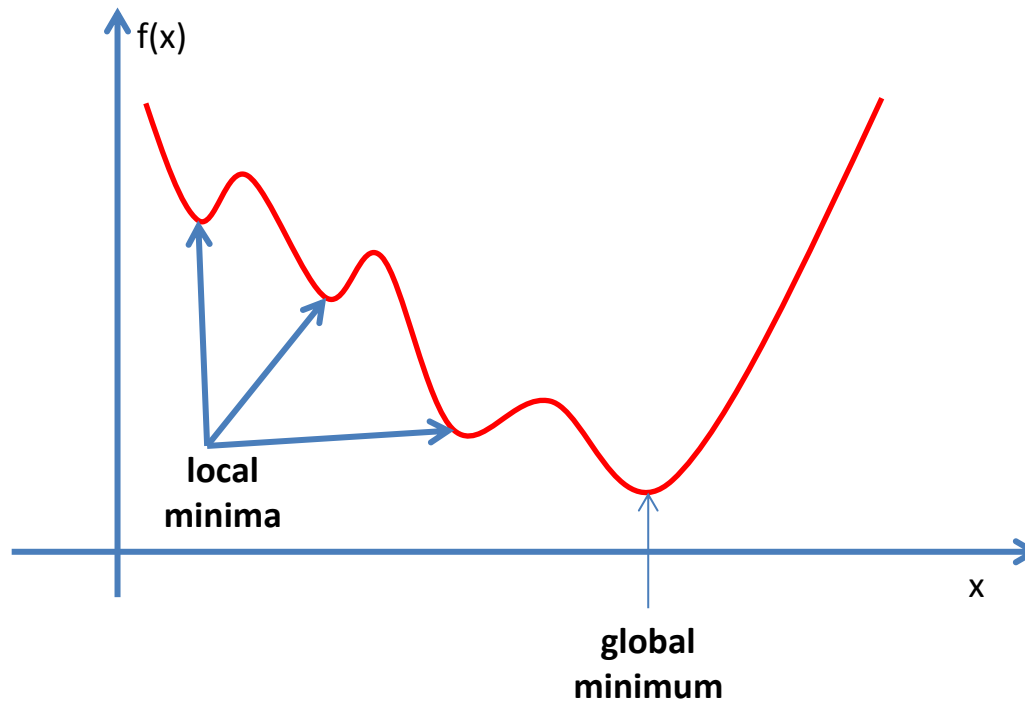
- Target (the X's): A volume of (for example) 100 x 100 x 100 voxels = 10^{**6} variables
 - (Plus 500.000 = $5 \times 10^{**5}$ geometry variables)
 - (plus 100.000 x k (classes))
- Measurements (the Y's): 100.000 particle images of 100 x 100 pixels = 10^{**9}
- But we have noise!: $2 + 2 = 5$ (or 3, or 6 ...)



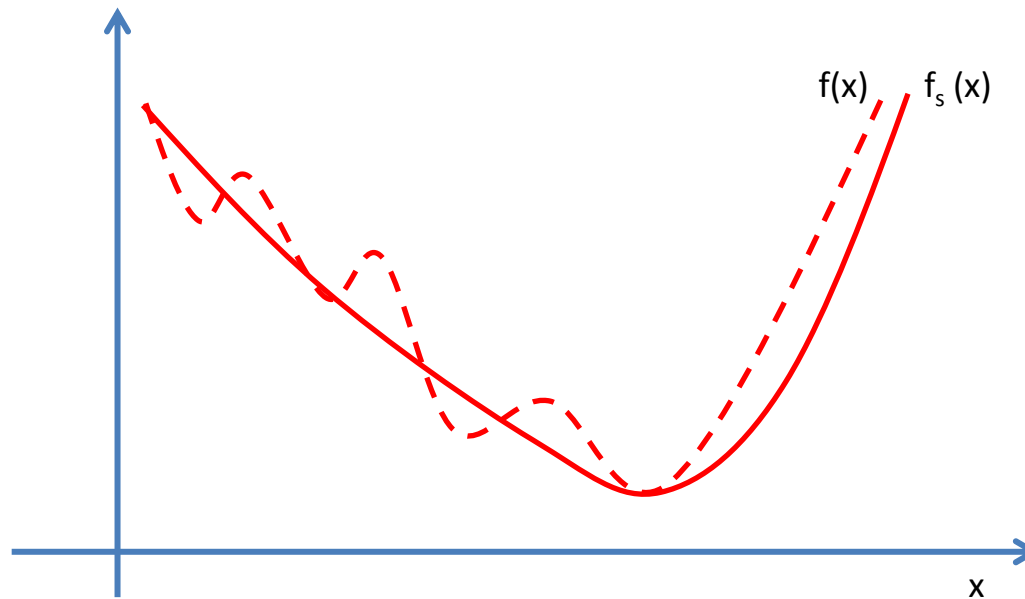
Everything is mixed!!!



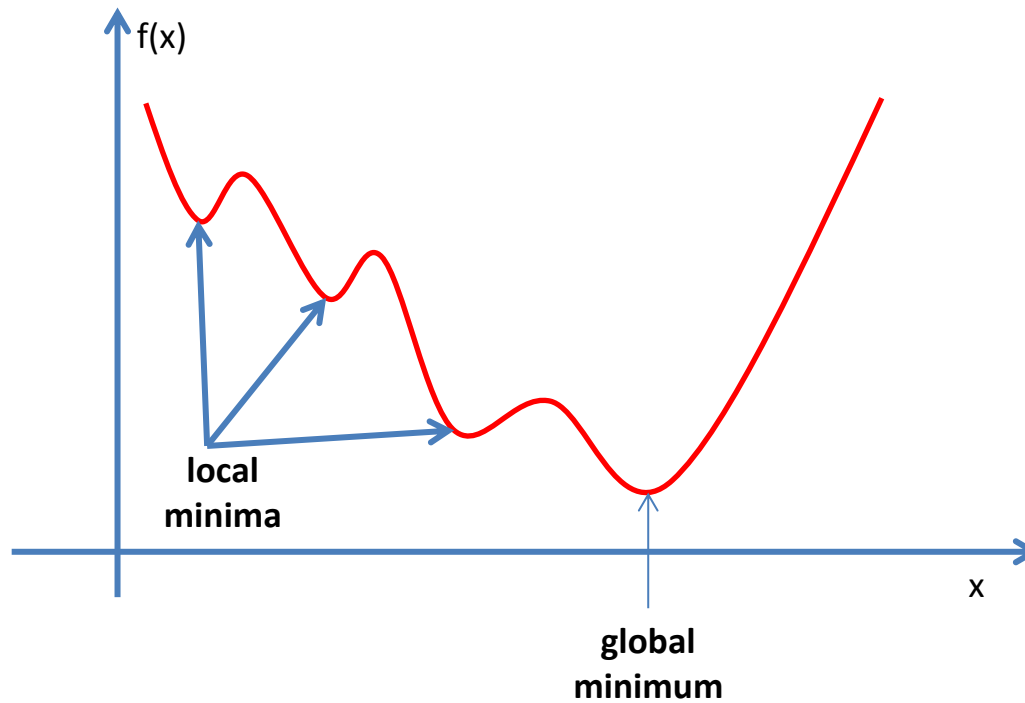
Parameter space of cryo EM SPA



Parameter space of cryo EM SPA



The Initial Volume Problem in SPA



The cryo-EM SPA pledge

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- Characterization of each image
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Validation

doi:10.1016/j.jmb.2011.09.008

J. Mol. Biol. (2011) 413, 1028–1046



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journal homepage: <http://ees.elsevier.com/jmb>



Tilt-Pair Analysis of Images from a Range of Different Specimens in Single-Particle Electron Cryomicroscopy

Richard Henderson^{1*}, Shaoxia Chen¹, James Z. Chen²,
Nikolaus Grigorieff², Lori A. Passmore¹, Luciano Ciccarelli³,
John L. Rubinstein⁴, R. Anthony Crowther¹, Phoebe L. Stewart⁵
and Peter B. Rosenthal⁶

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²Rosenstiel Basic Medical Science Research Center, Howard Hughes Medical Institute, Brandeis University, Waltham, MA 02154, USA

³Max Planck Institute for Biophysics, D-60438 Frankfurt, Germany

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⁵Department of Molecular Physiology and Biophysics, Vanderbilt University Medical Center, Nashville, TN 37232, USA

⁶Division of Physical Biochemistry, MRC National Institute for Medical Research, London NW7 1AA, UK



VALIDATION: results of our validation method on controversial HIV data



We have applied our approach to validate the map presented in the controversial work of MAO (1) and we have compared the results with the map reported by Subramaniam (2)

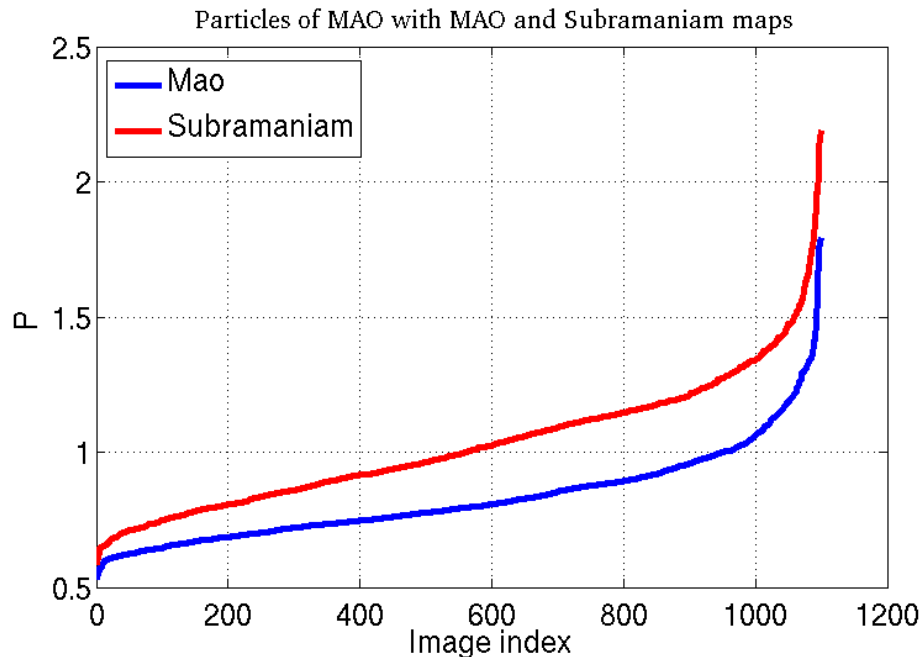
- (1) Mao Y, et al. (2013) Molecular architecture of the uncleaved HIV-1 envelope glycoprotein trimer. *Proc Natl Acad Sci USA* 110(39: 12428-12433
- (2) Subramaniam S (2013) Structure of trimetric HIV-1 envelope glycoproteins. *Proc natl Acad Sci USA* 110(E4172-E4174)



We have used the data deposited at EMDB EMPIAR 10008 (MAO) & EMPIAR 10004 Subramaniam, for the validation we have used approximately 1000 particles and the EMDB maps



For the validation approach we have used first the extracted particles of MAO, which has been compared with the two maps (MAO & Subramaniam)



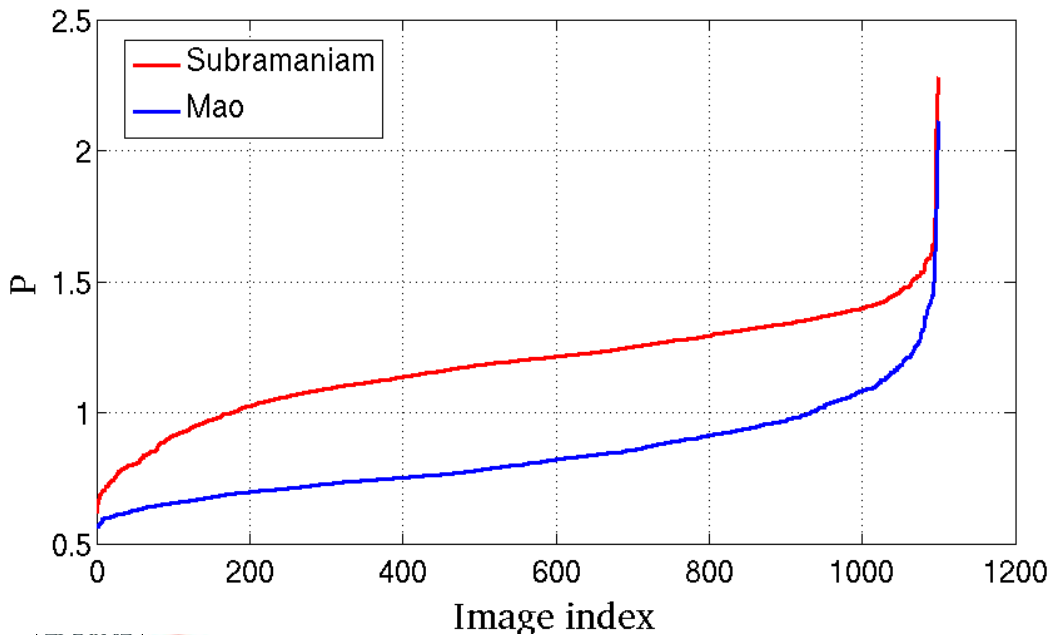
The quality parameters obtained are of

$Q_0 = 0.48$ (Subramaniam)

$Q_0 = 0.13$ (Mao)

For the validation approach we have used first the extracted particles of MAO, which has been compared with the two maps (MAO & Subramaniam)

Particles Subramaniam with MAO and Subramaniam maps

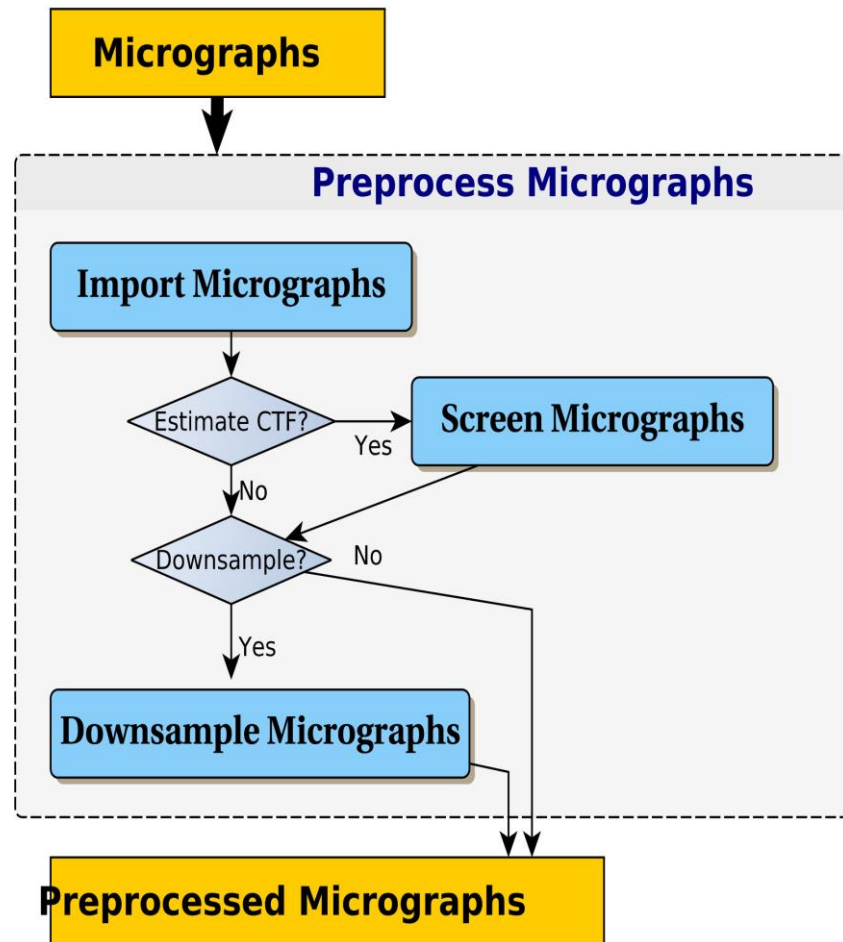


The quality parameters obtained are of

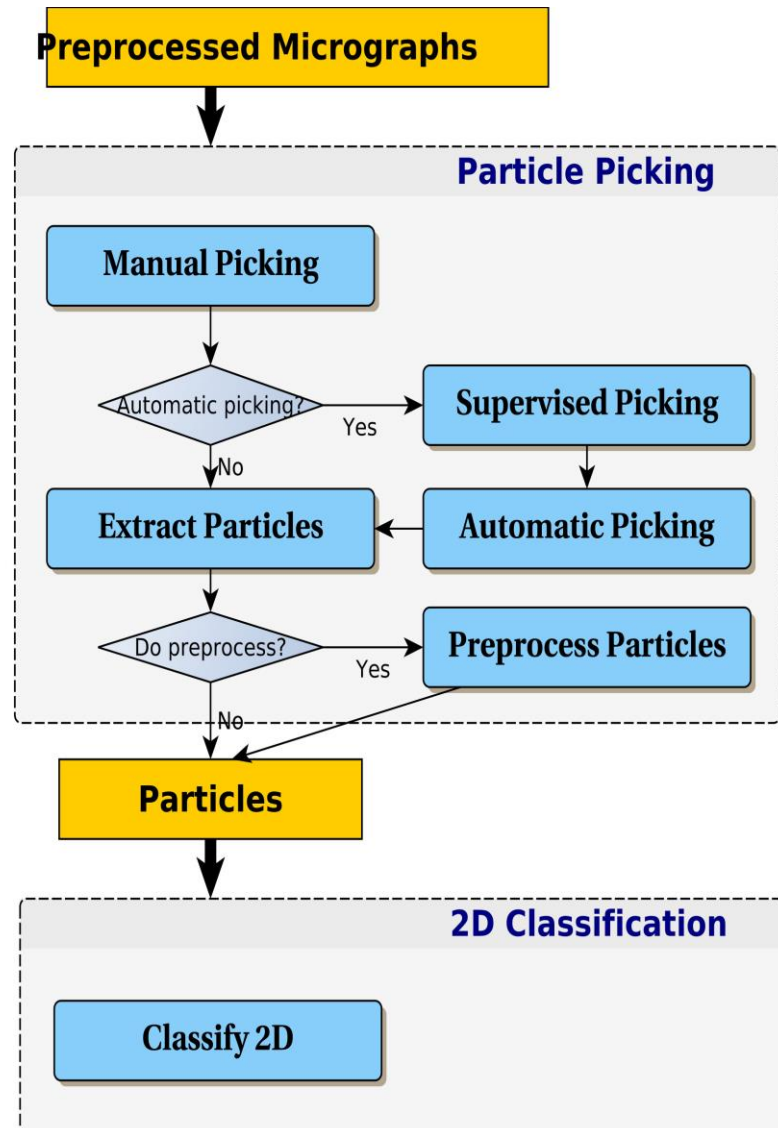
$Q_0 = 0.84$ (Subramaniam)

$Q_0 = 0.15$ (Mao)

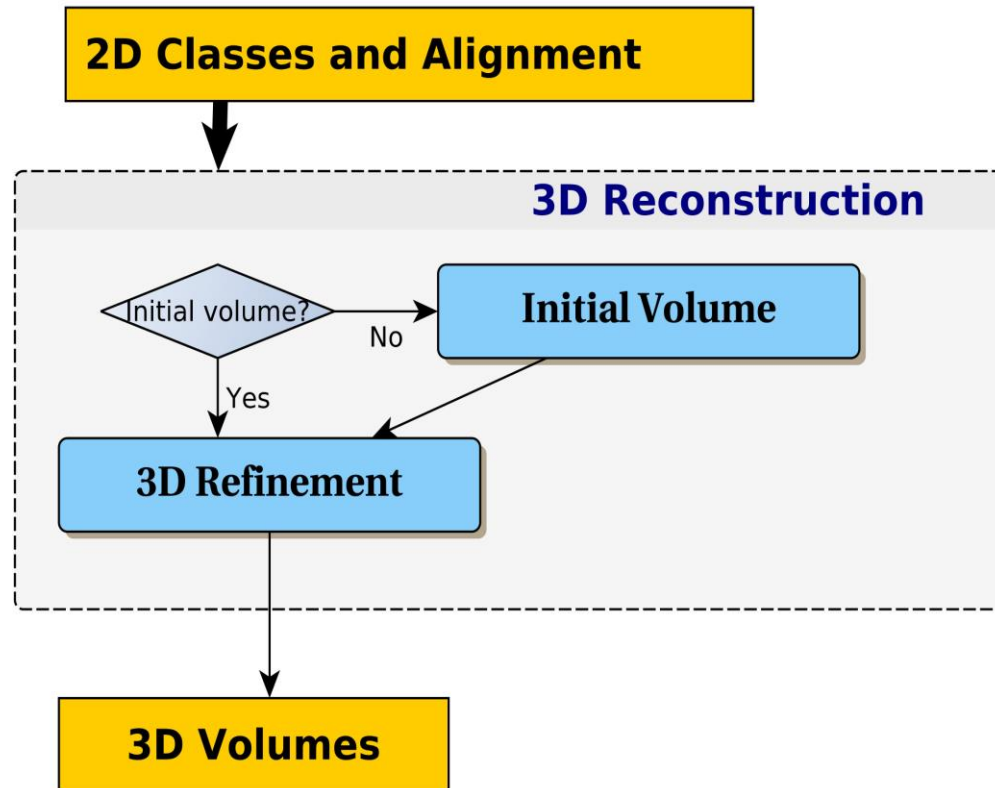
Workflows: How do we do it in practice?



Workflows: How do we do it in practice?

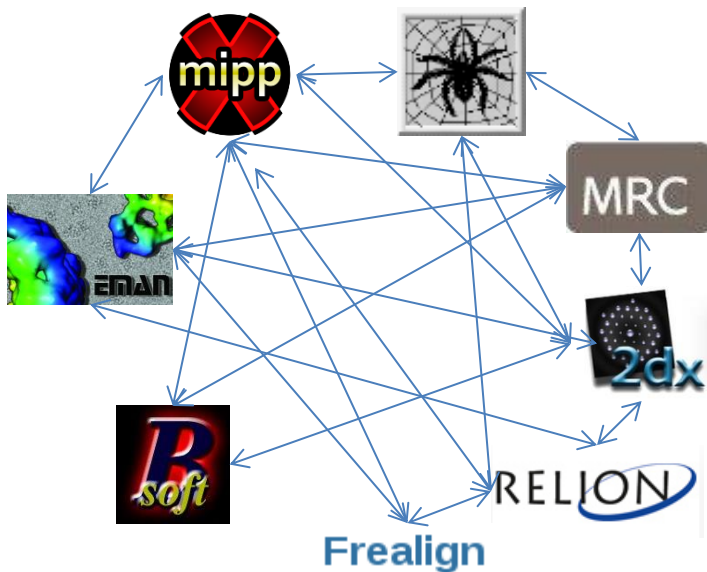


Workflows: How do we do it in practice?



Workflows: How do we do it in practice?

Using different EM software packages is now like the tower of Babel



Project TutorialIntro

Project Help

SCIPION

Protocols | Data | Hosts

View: Protocols SPA

- import PDB
- import movies
- import averages
- import masks
- export to EMX
- Micrographs**
 - Preprocess
 - xmipp3 - movie extract partic
 - xmipp3 - movie alignment
 - xmipp3 - preprocess microgr
 - CTF estimation
 - grigoriefflab - ctffind3
 - xmipp3 - ctf estimation
 - scipion - ctf assign
 - xmipp3 - ctf discrepancy
- Particles**
 - Set operations
 - Picking**
 - xmipp3 - consensus picking
 - xmipp3 - automatic picking
 - eman2 - boxer
 - bsoft - particle picking
 - appion - dogpicker
 - xmipp3 - supervised picking
 - xmipp3 - tilt pairs particle pic
 - Extract
 - Process

Edit Copy Delete Steps Db Collapse
 View: Tree Refresh

```

    graph TD
      PROJECT[PROJECT] --> 1[1. import mics saved]
      PROJECT --> 6[6. import vol saved]
      1 --> 2[2. downsample x5 saved]
      6 --> 2
      2 --> 3[3. ctffind3 saved]
      2 --> 4a[4a. xmipp picking saved]
      2 --> 4b[4b. eman boxing saved]
      3 --> 5[5. extract particles saved]
      4a --> 5
      4b --> 5
      5 --> 7[7. relion refine 3D saved]
  
```

Analyze Results

Summary Methods Output Log

Input
Output

SUMMARY
Output SetOfMicrographs not ready yet.



Importing micrographs

Protocol Run: ProtImportMicrographs

Protocol: scipion - import micrographs [Cite](#) [Help](#)

Run

Run name: 1. import mics [✎](#) Comment: [✎](#)

Execution Mode: Resume [?](#) Host: localhost [?](#)

Use queue? Yes No [?](#)

Expert Level: Normal [?](#)

Import

Import from: files [?](#)

Files directory: ests/xmipp_tutorial/micrographs/*.mrc [?](#)

Pattern: [?](#)

Acquisition info

Microscope voltage (kV): 300.0 [?](#)

Spherical aberration (mm): 2.0 [?](#)

Amplitude Contrast: 0.1 [?](#)

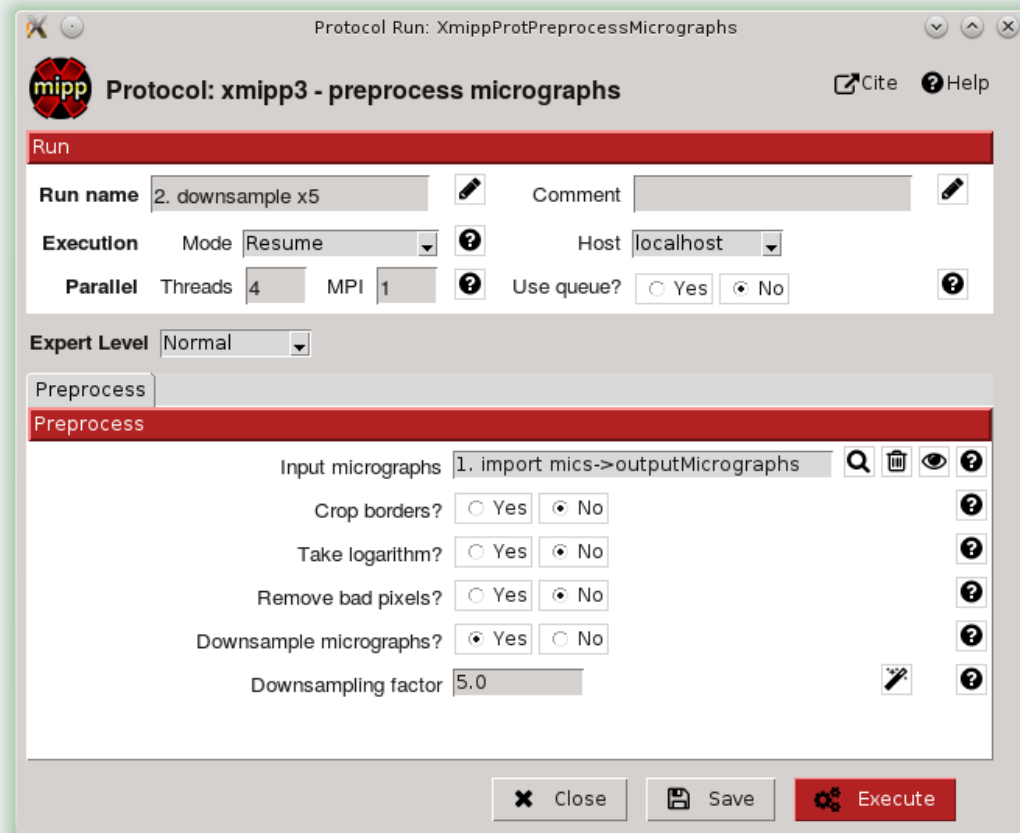
Magnification rate: 50000 [?](#)

Sampling rate mode: From image [?](#)

Pixel size ("sampling rate") (A/px): 1.237 [?](#)


[Close](#) [Save](#) [Execute](#)

Preprocessing micrographs







Calculating CTF



Protocol Run: ProtCTFFind


 **Protocol: grigoriefflab - ctffind3** [Cite](#) [Help](#)

Run

Run name 3. ctffind3  **Comment** 




Execution Mode **Resume**  **Host** localhost 



Parallel Threads **4** MPI **1**  **Use queue?** Yes No 



Expert Level Normal 

CTF Estimation

CTF Estimation

Input Micrographs 2. downsample x5->outputMicrographs   

CTF Downsampling factor 1.0  

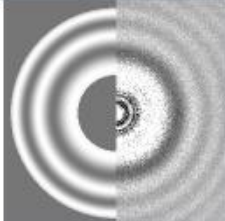
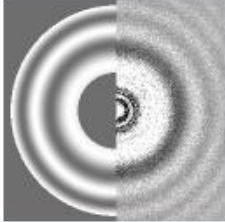
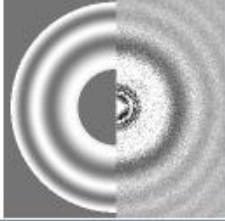
Resolution *Lowest* 0.14 *Highest* 0.46  

Typical CTF displays

File Display Help

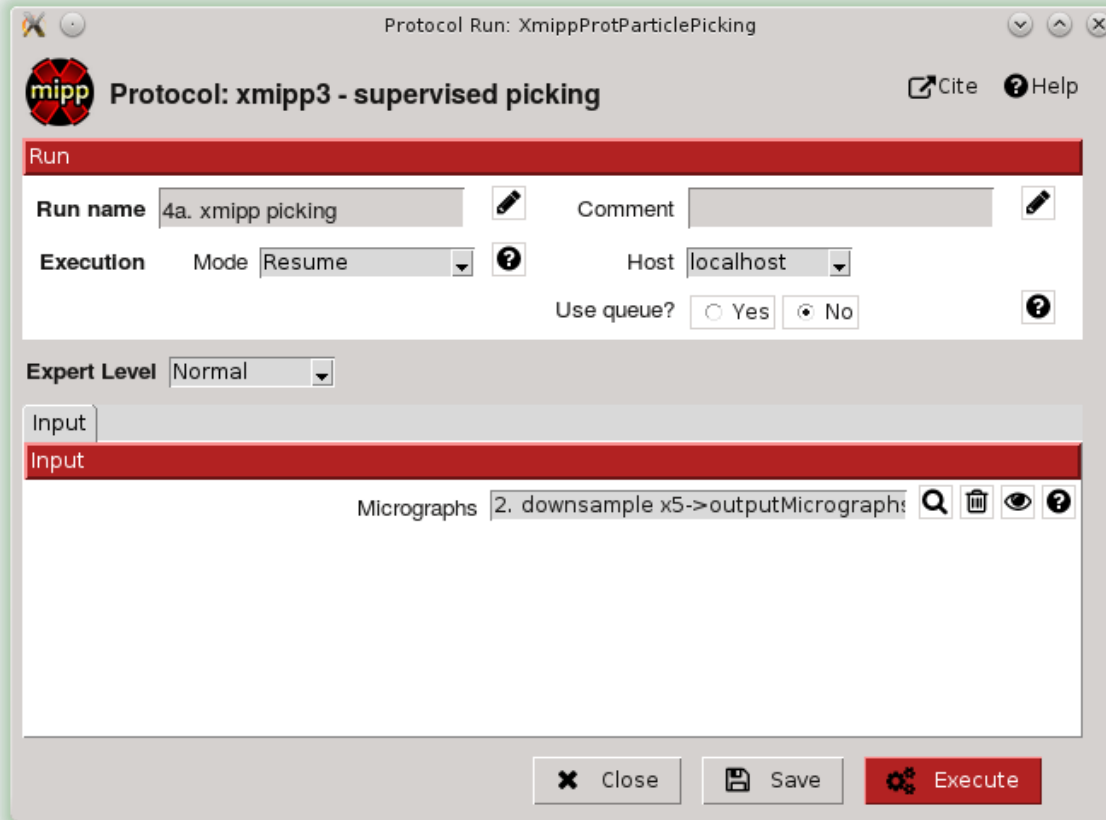
50 1 Cols 8 Rows 3

Block CTFModels

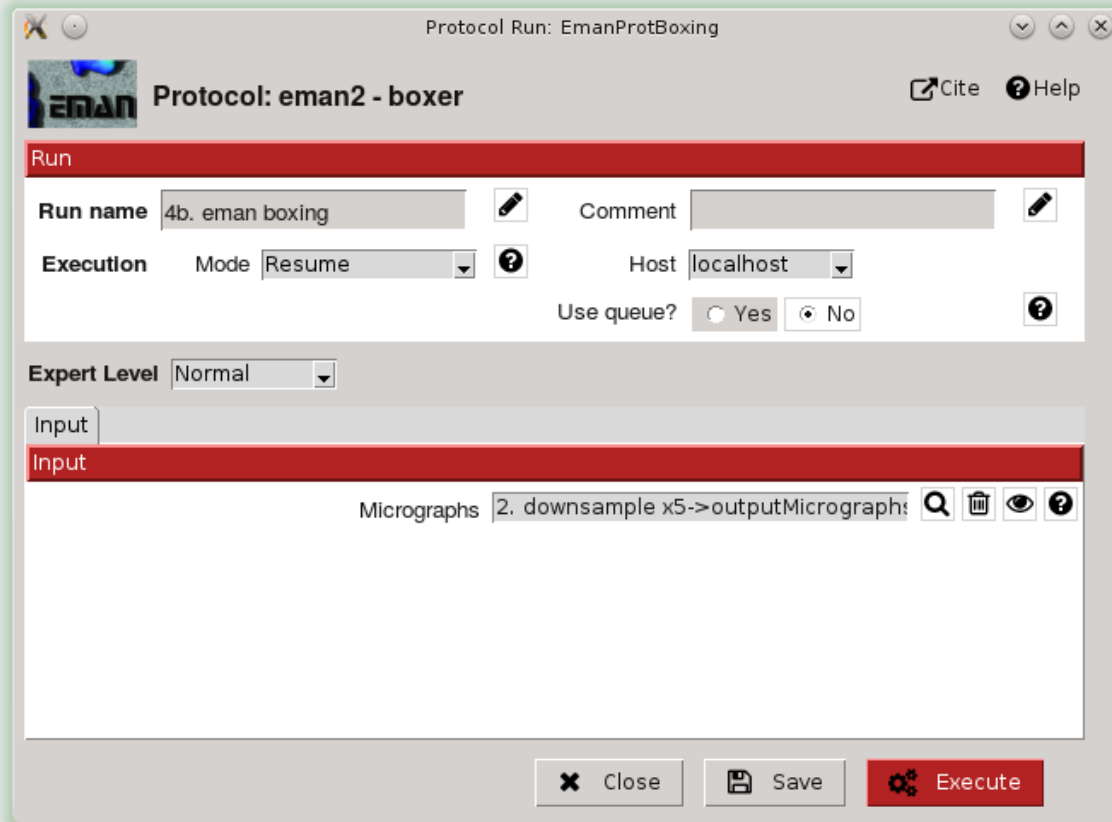
	id	enabled	comment	_psdFile	_defocusU	_defocusV	_defocusAngle	
1	1	<input checked="" type="checkbox"/>			23751,9395	23901,1504	-36,8300	Runs/000070_XmippPro
2	2	<input checked="" type="checkbox"/>			21755,3906	21923,6309	-31,6300	Runs/000070_XmippPro
3	3	<input checked="" type="checkbox"/>			22199,0195	22382,5801	-33,5800	Runs/000070_XmippPro

Close Micrographs Recalculate CTFs

Particle picking



Particle picking



Typical particle picking display

The screenshot displays the Xmipp Particle Picker software interface. The main window shows a micrograph of a biological sample with a large circular region of interest outlined. The interface includes several control panels:

- File Filters Window Help**: Contains search and filter options.
- Shape**: Includes checkboxes for Circle, Rectangle, and Center.
- Autopick**: Includes a Autopick checkbox and a **Threshold** slider set to 0.
- Micrographs**: A table listing micrographs with columns for Name, Particles, and State.
- Manual: 0 Automatic: 0**: A status indicator.
- Buttons**: Includes **Reset Micrograph**, **Save**, and **Create Coordinates**.

A **B&C** (Brightness and Contrast) dialog box is open, showing a histogram and sliders for **Minimum** (81.05), **Maximum** (142.90), **Brightness**, and **Contrast**. It also features **Auto**, **Reset**, **Set**, and **Apply** buttons.

	Name	Particles	State
1	BPV_1386	0	Available
2	BPV_1387	0	Available
3	BPV_1388	0	Available

Extracting particles

Protocol Run: XmippProtExtractParticles

mipp Protocol: xmipp3 - extract particles [Cite](#) [Help](#)

Run

Run name: 5. extract particles [✎](#) Comment: [✎](#)

Execution Mode: Resume [?](#) Host: localhost [?](#)

Parallel Threads: 4 MPI: 1 [?](#) Use queue? Yes No [?](#)

Expert Level: Normal [?](#)

Input Preprocess

Input

Coordinates: 4a. xmipp picking->outputCoordinates [?](#) [?](#) [?](#)

Downsampling type: same as picking [?](#)

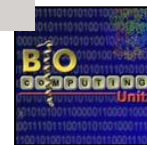
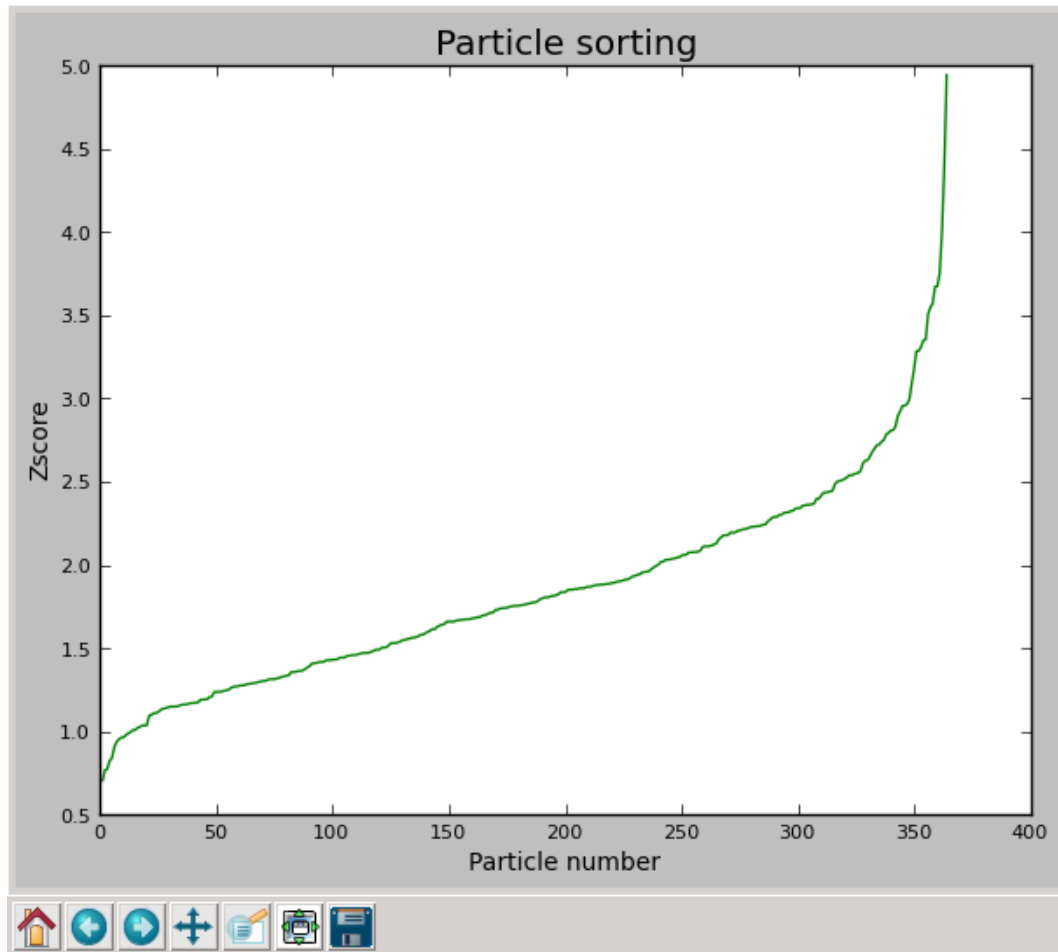
CTF estimation: 3. ctffind3->outputCTF [?](#) [?](#)

Particle box size: 110 [?](#)

Perform sort by statistics: Yes No [?](#)

[✕](#) Close [💾](#) Save [⚙️](#) Execute

Sorting particles



Typical sorting display

File Display Help

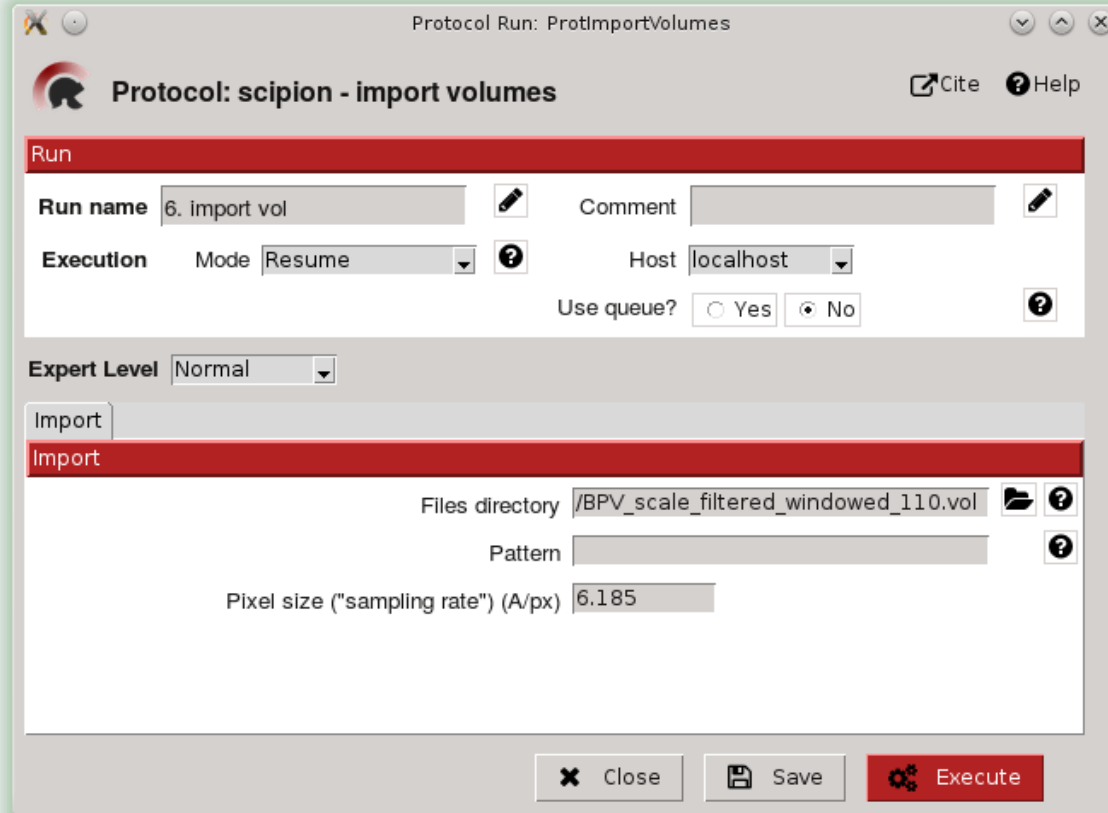
100 179 Cols 7 Rows 52

Block **Particles**

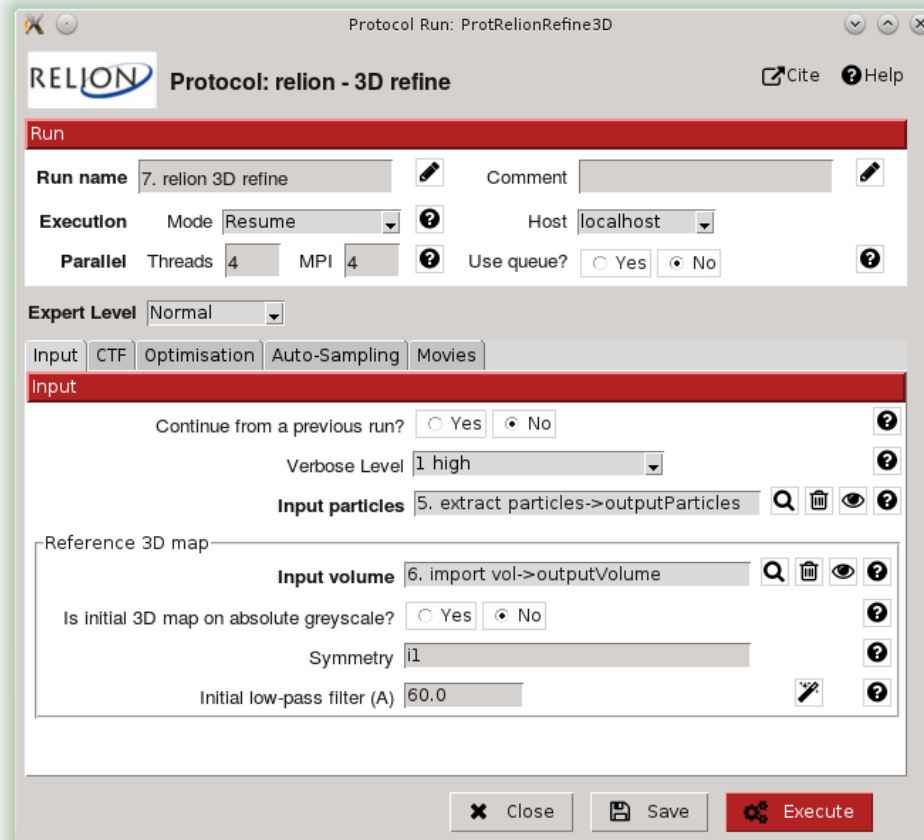
	1	2	3	4	5	6	7
48							
49							
50							
51							
52							

Close Particles

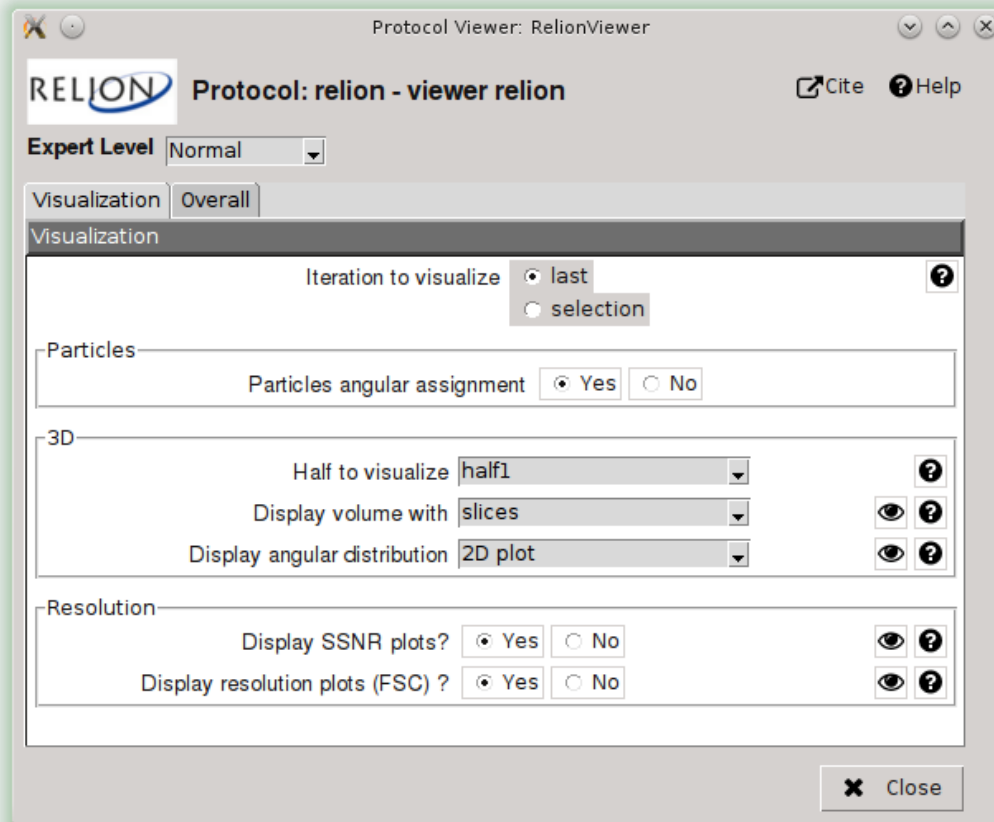
Importing Initial Volume (and the “Problem”?)



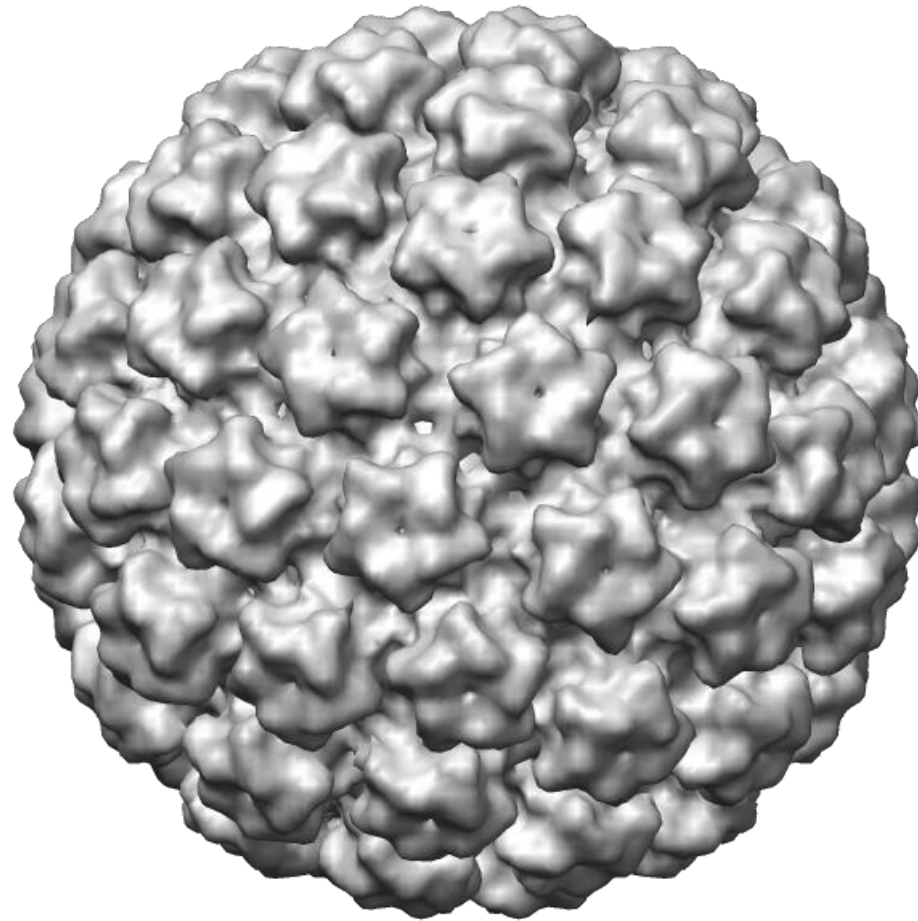
Finding Projection Geometry



Typical Relion 3D display



Typical Volume display



The Initial Volume Problem (in the Web)

Obtain your initial volume

This webportal allows you to obtain an initial 3D map from your average images. Several methods are available from different packages (Xmipp3 and Eman2). At the end of the processing you will be able to compare the output volumes from the different methods, downloading the selected ones.

It is recommended to [read the tutorial](#) before using this portal.

Choose your project

Create an empty project

Create

Create a project with Test data

Test data

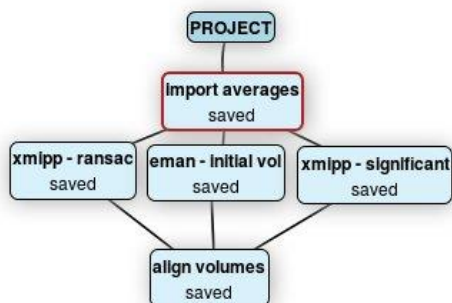
Explore project with Test data (read-only)

Explore

The Initial Volume Problem (in the Web)

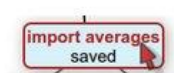
Edit Copy Delete

List Refresh Help




Let's start!

Step 1: Upload and import your files to the web portal clicking on the graph on the left hand side of this panel, on the **Import Averages** box.



Finally press "Execute"!

Step 2: Select a method to obtain your initial map . You can choose among **Xmipp - RANSAC**, **EMAN initial volume** and **Xmipp - Significant**.



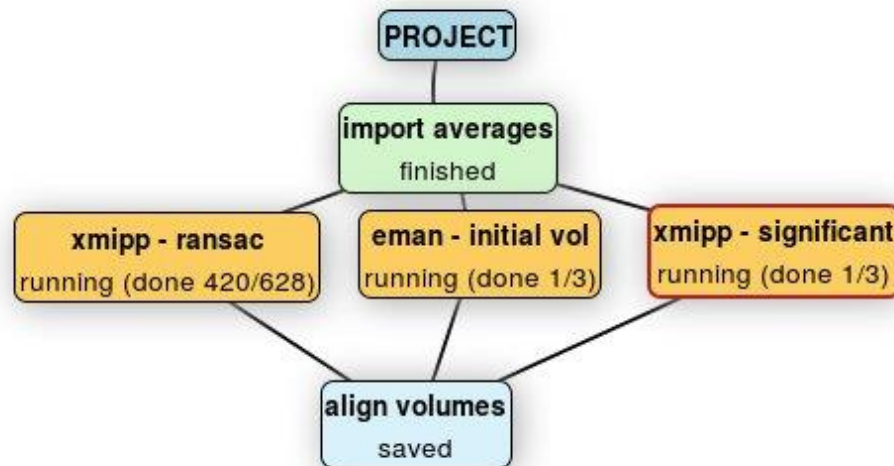
Summary Methods Output Logs Analyze Results Download Results

Input

Output



The Initial Volume Problem (in the Web)


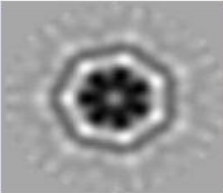
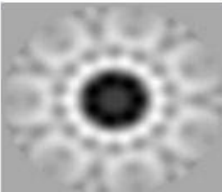


The Initial Volume Problem (in the Web)

 **SCIPION** Xmipp Showj Web

200 1 Select Block Volumes

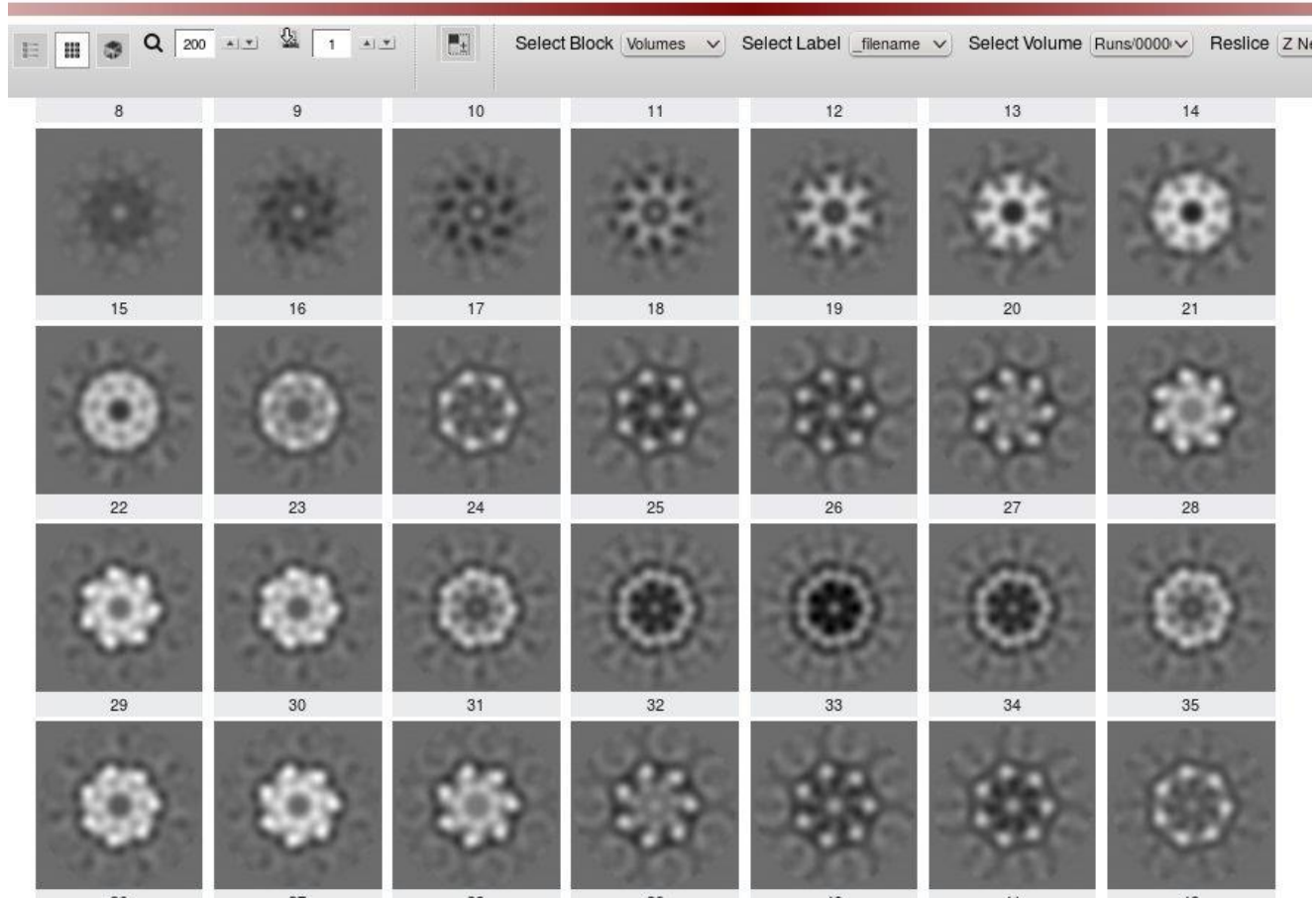
Search:

id	enabled	comment	_filename
1	<input checked="" type="checkbox"/>	ransac volume 01	
2	<input checked="" type="checkbox"/>	ransac volume 02	
3	<input checked="" type="checkbox"/>	ransac volume 03	

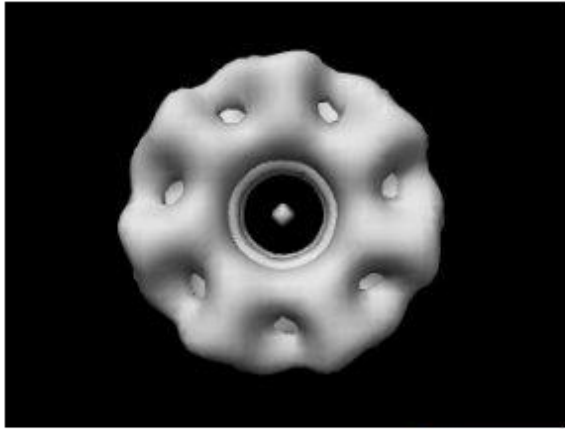


Typical 3D displays

 **SCIPION** Xmipp Showj Web



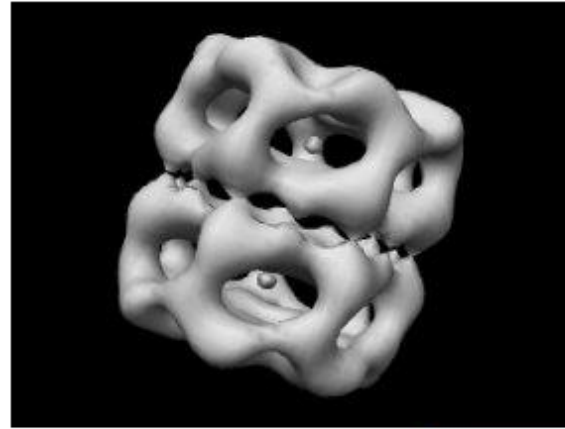
Typical 3D displays



Threshold

Refresh

Min [-0.213], Max [0.284]



Threshold

Refresh

Min [-0.213], Max [0.284]



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13th Feb 2014

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