

## Commentary

## What gas exchange data can tell us about photosynthesis

This title is a modification of the title of Long and Bernacchi (2003), who described the then current state of analysis of  $A/C_i$  curves, plots of photosynthetic  $\text{CO}_2$  assimilation versus  $\text{CO}_2$  inside the leaf (to remove any influence of stomata). These curves were made possible by the concept of a calculated  $\text{CO}_2$  partial pressure inside a leaf (Moss & Rawlins 1963) and have been studied since the 1970s when Graham Farquhar was a post-doc and I was a graduate student in Klaus Raschke's lab. In 1980, Farquhar, von Caemmerer and Berry (1980) published a seminal paper describing photosynthesis as either 'Rubisco limited' or 'RuBP limited', and soon, a third limitation was added, 'TPU limited' (Sharkey 1985).  $A/C_i$  curves are ideally suited to assess these three mechanisms that can set the upper limit to the rate of photosynthesis. Analyses of  $A/C_i$  curves and the tripartite model of photosynthesis limitations have been very useful for testing mechanistic models of photosynthetic metabolism and for predicting photosynthetic responses to global change (Wullschleger 1993). This issue of *Plant, Cell & Environment*, includes several papers related to the analysis of gas exchange data (Bellasio *et al.* 2015b; Bellasio *et al.* 2015a; Walker & Ort 2015). Here, I highlight the importance of these contributions and also announce an update to the PCE Calculator (version 2.0) that can be used for a simple analysis of gas exchange parameters from  $A/C_i$  curves. A separate sheet is provided to fit light response curves following the recommendations of Buckley and Diaz-Espejo (2015).

The success of the Farquhar, von Caemmerer and Berry (1980) model derives at least in part because it allows gas exchange measurements to be interpreted in terms of biochemical and biophysical processes. Soon after publication of the model, programs and algorithms for estimating the underlying parameters became available. One early program was provided by Dundee Scientific (Dundee, Scotland) and called 'Photosyn Assistant' (<http://www.ddsci.com/>). Several other methods were shared informally among researchers, including one by Carl Bernacchi that used linear versions of some of the equations so that critical parameters could be estimated from linear regressions.

In 2007, *PCE* made available an Excel spreadsheet to help estimate key parameters from  $A/C_i$  curves (Sharkey *et al.* 2007). The intent was to strike a balance between detailed information and ease of use. Theoretically, five parameters could be estimated:  $V_{\text{cmax}}$ ,  $J$ ,  $TPU$ ,  $R_d$  and  $g_m$  (the maximum carboxylation rate of Rubisco, maximum rate of electron transport for the given light intensity, maximum rate of triose phosphate use, day respiration, and mesophyll conductance to  $\text{CO}_2$  transfer, respectively). With five parameters that can be adjusted, some very good fits are possible, even if they are not always believable. Instead, each parameter should

be considered carefully, and when possible, parameters should be fixed using other data to improve estimates of the remaining parameters. Since the publication of this tool in 2007, a number of issues have become more clear about each of the parameters, as outlined in the following.

$V_{\text{cmax}}$  A particular problem with estimating  $V_{\text{cmax}}$  is that  $V_{\text{cmax}}$  and  $g_m$  are almost complimentary, so that very good fits can be had by varying these two parameters inversely over a wide range of values. It is therefore important to have data that constrains  $g_m$ , either independent measures of  $g_m$  from another method or many data points in the RuBP-regeneration-limited region of the  $A/C_i$  curve.

$J$  There has been discussion of the meaning of  $J$ .  $J_{\text{max}}$ , as used in global models of photosynthesis, is a theoretical number requiring extrapolation to infinite light intensity. In global models,  $J$  is calculated using  $J_{\text{max}}$  and two other parameters.  $J_{\text{max}}$  should not be used for the maximum rate of electron transport at high light intensity, a parameter that has sometimes been called  $J_{\text{high}}$  (Buckley & Diaz-Espejo 2015). In the PCE Calculator version 1.0, the maximum  $J$  obtained over all  $C_i$  was just called  $J$ , not  $J_{\text{max}}$ , the maximum rate of electron transport over the  $A/C_i$  curve at that light intensity, despite the fact that the irradiance may or may not have been saturating, and of course was not infinite. In version 2.0 of the calculator, the maximum  $J$  is still reported as  $J$ , and a new sheet is provided to allow calculation of  $J_{\text{max}}$  from a light response curve. A true  $J_{\text{max}}$  would require using only RuBP-regeneration-limited points, but in most cases, measurements of  $A$  at high light will be limited by Rubisco or  $TPU$ .

$TPU$  The maximum rate of end product formation that can be observed sometimes reflects the maximum capacity and sometimes not. Wullschleger (1993) found this behaviour in 23 of 109 species he analysed. When the highest data points of the  $A/C_i$  curve do not increase with  $\text{CO}_2$ ,  $TPU$  limitation is assumed. However, sometimes  $A$  actually declines at high  $\text{CO}_2$ . For some time, this was attributed to changes in photorespiration and direct use of glycine and serine by export from leaves and use in protein synthesis (Harley & Sharkey 1991); the best equation for describing this effect was published by von Caemmerer (2000). However, in recent attempts to incorporate this equation into the PCE Calculator, it became clear that even if all of the glycine or serine produced in photorespiration were used directly, it would not give the degree of reverse sensitivity sometimes observed. An earlier explanation for reverse sensitivity now seems a more likely explanation. Phosphoglyceric acid can inhibit stromal phosphoglucoisomerase and, in so doing, limit the capacity for starch synthesis (Sharkey & Vasey 1989). Unfortunately, this effect is not easily described by an equation. It is proposed that the von Caemmerer (2000) equation still be used to model this phenomenon, but that the fitting parameter,

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$\alpha$ , be considered an arbitrary parameter useful for comparison of the degree of reverse sensitivity, but without a mechanistic basis (see calculator version 2.0 (R)).

$R_d$  This parameter has been perhaps the most problematic in many fittings of  $A/C_i$  curve data. The calculator often estimates unrealistically high values of  $R_d$ , a phenomenon whose origins are not clear. This parameter is the best candidate for finding some other method of determining a reliable estimate for  $R_d$  and then fixing the value in the calculator so that better estimates of the other parameters are found. The best available method is the method used to determine  $\Gamma^*$ . This is relatively involved and requires very accurate measurements of  $A$  at low  $\text{CO}_2$ . A simpler way to constrain  $R_d$  is to measure respiration in the dark and use that as an upper limit or set  $R_d$  to be some proportion of dark respiration, perhaps 50%.

$g_m$  There has been a tremendous interest in  $g_m$  in recent years (Sharkey 2012), and it is possible to obtain an estimate of  $g_m$  from the calculator. However, using the calculator is not as good as other methods specifically designed to measure this parameter. The calculator assumes that  $g_m$  is constant with  $\text{CO}_2$  even though many studies find it varies with  $\text{CO}_2$ . Even so, it gives an assimilation-weighted value that works reasonably well in predicting how photosynthesis will respond to changes in  $\text{CO}_2$ . However, if other estimates of  $g_m$  are available, it is best to use those instead of letting the calculator estimate it. Regardless, it should be noted that it has been argued that any reasonable estimate of  $g_m$  is preferable to assuming infinite mesophyll conductance (Sun *et al.* 2014a; Sun *et al.* 2014b).

The simplicity of the calculator is both its strength and weakness. A number of parameters need to be specified, for example the affinity ( $K_m$ ) of Rubisco for  $\text{O}_2$  and  $\text{CO}_2$ . The parameter values in the calculator are mostly from tobacco (Bernacchi *et al.* 2002), but in this issue of *PC&E*, several papers show how more precise values can be obtained. One critical parameter is  $\Gamma^*$ , the Rubisco  $\text{CO}_2$  compensation point. This varies among species, and it is among the easier parameters to measure. One paper in this issue directly addresses this. The commonly used technique for assessing  $\Gamma^*$  was first suggested by Agu Laik and made popular by Brooks and Farquhar (1985). This method requires measurement of  $A/C_i$  curves over a range of limiting light intensities. Theoretically, these should intersect at  $C_i^*$ , which is related to  $\Gamma^*$  by the equation

$$\Gamma^* = C_i^* + \frac{A}{g_m}$$

This technique has been used by many people, but what is not clear in the publications is that the lines often do not intersect that well, creating uncertainty in the intersection point. In this issue of *PC&E*, Walker and Ort (2015) provide a method for analysing  $\Gamma^*$  data so that the effects of small errors and uncertainties are minimized, giving a more robust estimate of  $\Gamma^*$ .

A new estimate of  $\Gamma^*$  over a range of temperature has been determined for *Arabidopsis thaliana* (Weise *et al.* 2015), and a new spreadsheet in the PCE Calculator with an *Arabidopsis thaliana*  $\Gamma^*$  value is now available (version 2.0 (A)). When

reverse sensitivity is observed at high  $\text{CO}_2$  in an  $A/C_i$  curve, a second new sheet (2.0 (R)) is provided that estimates  $\alpha$ , an arbitrary parameter useful for describing the degree of decline of photosynthesis with increasing  $\text{CO}_2$  at high  $\text{CO}_2$ .

As before, the PCE Calculator requires the user to assign which points are controlled by Rubisco, which by RuBP regeneration and which by TPU. The most informative data points are the RuBP-regeneration-limited data points, and so, investigators should be sure to include many points in this region. The fitting program of Gu *et al.* (2010) estimates these transitions within the program and allows users to share data at the website leafweb.ornl.gov.

Now an even more detailed approach is available in the supplemental material of Bellasio *et al.* (2015b), who provide a series of worksheets that allows estimation of many more parameters. This is a comprehensive analysis of photosynthesis making use of  $A/C_i$  curves at normal and low oxygen and light response curves. Carrying out such a comprehensive analysis will likely be challenging, but also very rewarding because of the rich dataset that will be obtained. Finally, Bellasio *et al.* (2015a) extend this type of  $A/C_i$  curve analysis to  $C_4$  plants. Because the  $C_4$  pump obscures many of the  $C_3$  processes, the  $C_4$  analysis is less mechanistic, but very interesting data can still come from this analysis.

It was not obvious in 1980 that the mechanistic model of photosynthesis would have such far-reaching implications. The original paper has been cited over 4900 times according to Google Scholar, and the peak in the number of citations came in 2013, 33 years after its publication. One reason for its popularity is that underlying biochemical mechanisms can be estimated from leaf gas exchange characteristics. The papers in this issue (Bellasio *et al.* 2015b; Bellasio *et al.* 2015a; Walker & Ort 2015) and the updated version of the PCE Calculator should continue the usefulness of analysis of  $A/C_i$  curves and allow ever greater information to be obtained by gas analysis of photosynthesis.

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## REFERENCES

- Bellasio C., Beerling D.J. & Griffiths H. (2015a) Deriving  $C_4$  photosynthetic parameters from combined gas exchange and chlorophyll fluorescence using an Excel tool: theory and practice. *Plant, Cell & Environment* **in press**.
- Bellasio C., Beerling D.J. & Griffiths H. (2015b) An Excel tool for deriving key photosynthetic parameters from combined gas exchange and chlorophyll fluorescence: theory and practice. *Plant, Cell & Environment* **in press**.
- Bernacchi C.J., Portis A.R., Nakano H., von Caemmerer S. & Long S.P. (2002) Temperature response of mesophyll conductance. Implications for the

- determination of Rubisco enzyme kinetics and for limitations to photosynthesis in vivo. *Plant Physiology* **130**, 1992–1998.
- Brooks A. & Farquhar G.D. (1985) Effects of temperature on the O<sub>2</sub>/CO<sub>2</sub> specificity of ribulose-1,5-bisphosphate carboxylase/oxygenase and the rate of respiration in the light. Estimates from gas exchange measurements on spinach. *Planta* **165**, 397–406.
- Buckley T.N. & Diaz-Espejo A. (2015) Reporting estimates of maximum potential electron transport rate. *New Phytologist* **205**, 14–17.
- von Caemmerer S. (2000) *Biochemical Models of Leaf Photosynthesis*, pp. 1–165. CSIRO Publishing, Collingwood, Victoria, Australia.
- Farquhar G.D., von Caemmerer S. & Berry J.A. (1980) A biochemical model of photosynthetic CO<sub>2</sub> assimilation in leaves of C<sub>3</sub> species. *Planta* **149**, 78–90.
- Gu L., Pallardy S.G., Tu K., Law B.E. & Wullschlegel S.D. (2010) Reliable estimation of biochemical parameters from C<sub>3</sub> leaf photosynthesis–intercellular carbon dioxide response curves. *Plant, Cell & Environment* **33**, 1852–1874.
- Harley P.C. & Sharkey T.D. (1991) An improved model of C<sub>3</sub> photosynthesis at high CO<sub>2</sub>: reversed O<sub>2</sub> sensitivity explained by lack of glycerate reentry into the chloroplast. *Photosynthesis Research* **27**, 169–178.
- Long S.P. & Bernacchi C.J. (2003) Gas exchange measurements, what can they tell us about the underlying limitations to photosynthesis? Procedures and sources of error. *Journal of Experimental Botany* **54**, 2393–2401.
- Moss D.N. & Rawlins S.L. (1963) Concentration of carbon dioxide inside leaves. *Nature* **197**, 1320–1321.
- Sharkey T.D. (1985) Photosynthesis in intact leaves of C<sub>3</sub> plants: physics, physiology and rate limitations. *The Botanical Review* **51**, 53–105.
- Sharkey T.D. (2012) Mesophyll conductance: constraint on carbon acquisition by C<sub>3</sub> plants. *Plant, Cell and Environment* **35**, 1881–1883.
- Sharkey T.D., Bernacchi C.J., Farquhar G.D. & Singsaas E.L. (2007) Fitting photosynthetic carbon dioxide response curves for C<sub>3</sub> leaves. *Plant, Cell & Environment* **30**, 1035–1040.
- Sharkey T.D. & Vassey T.L. (1989) Low oxygen inhibition of photosynthesis is caused by inhibition of starch synthesis. *Plant Physiology* **90**, 385–387.
- Sun Y., Gu L., Dickinson R.E., Norby R.J., Pallardy S.G. & Hoffman F.M. (2014a) Impact of mesophyll diffusion on estimated global land CO<sub>2</sub> fertilization. *Proceedings of the National Academy of Sciences* **111**, 15774–15779.
- Sun Y., Gu L., Dickinson R.E., Pallardy S.G., Baker J. Cao Y., ... Winter K. (2014b) Asymmetrical effects of mesophyll conductance on fundamental photosynthetic parameters and their relationships estimated from leaf gas exchange measurements. *Plant, Cell & Environment* **37**, 978–994.
- Walker B.J. & Ort D.R. (2015) Improved method for measuring the apparent CO<sub>2</sub> photocompensation point resolves the impact of multiple internal conductances to CO<sub>2</sub> to net gas exchange. *Plant, Cell & Environment* in press.
- Weise S.E., Carr D.J., Bourke A.M., Hanson D.T. & Sharkey T.D. (2015) The *arc* mutants of *Arabidopsis* with fewer large chloroplasts have a lower mesophyll conductance. *Photosynthesis Research* **124**, 117–126.
- Wullschlegel S.D. (1993) Biochemical limitations to carbon assimilation in C<sub>3</sub> plants—a retrospective analysis of the A/C<sub>i</sub> curves from 109 species. *Journal of Experimental Botany* **44**, 907–920.

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