

# Study of Interactions and Protein Structure Determination by NMR

For Application to Protein Characterization

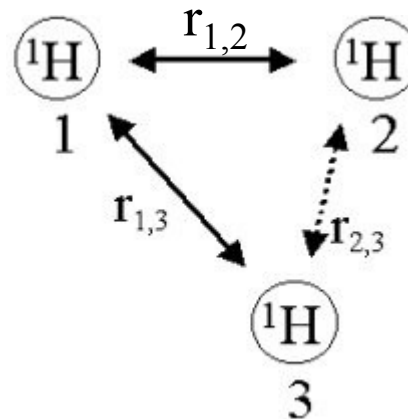
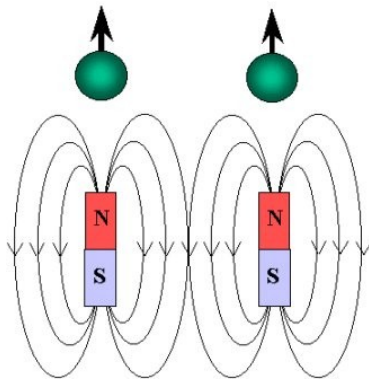
by

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CEITEC, Masaryk University

# NMR as a tool for study structure, dynamics and interactions of biomolecules

- 0) AA/NA sequence, resonance assignment, standard chemical shifts
- 1) Structure determination of proteins/NAs
- 2) NMR can provide detailed information about the structure at the atomic level resolution relying on the spatial proximity of two interacting protons – nuclear Overhauser enhancement (NOE)
- 3) Additional structural information can be obtained (residual dipolar couplings – RDCs,  $J$ -couplings, backbone chemical shifts - CSI)

NOE:



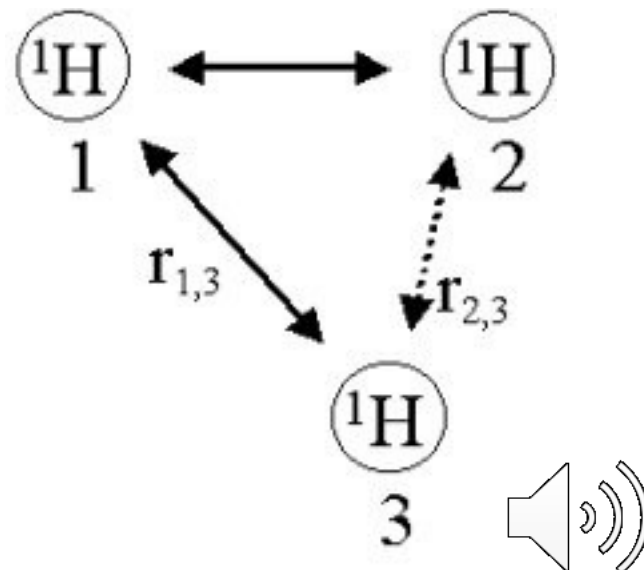
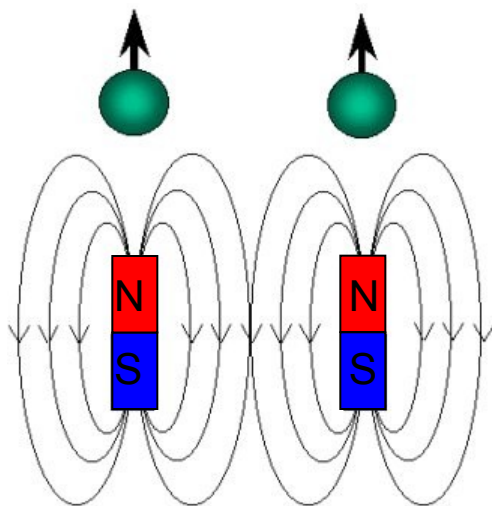
$$r_{1,2}; r_{1,3}; r_{2,3} \leq 6 \text{ \AA}$$

$$1 \text{ \AA} = 1.10^{-10} \text{ m}$$

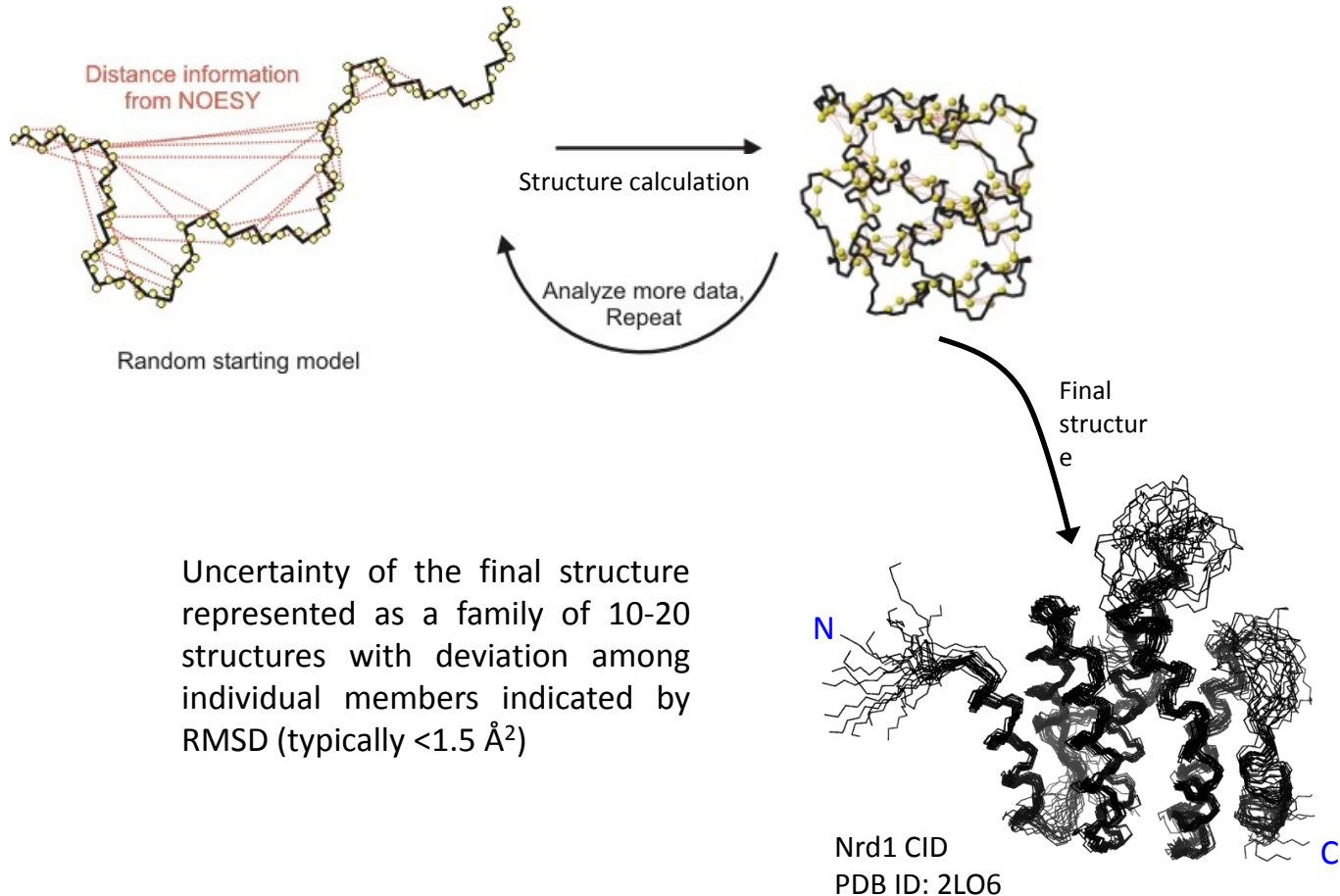


## Nuclear Overhauser Effect (Spectroscopy) = NOE(SY)

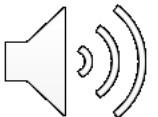
- i) caused by dipolar coupling between nuclei.
- ii) the local field at one nucleus is affected by the presence of another nucleus.
- iii) the result is a mutual modulation of resonance frequencies.
- iv) the NOE operates through space.
- v) the intensity of the interaction is a function of the distance between the nuclei according to the following equation:  $I = A(1/r^6)$ ,  $I$  is the intensity,  $A$  is a scaling constant, and  $r$  is the distance between the nuclei
- vi) the NOE provides a link between an experimentally measurable quantity,  $I$ , and internuclear distance
- vii) NOE is only observed up to  $\sim 6\text{\AA}$



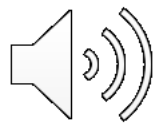
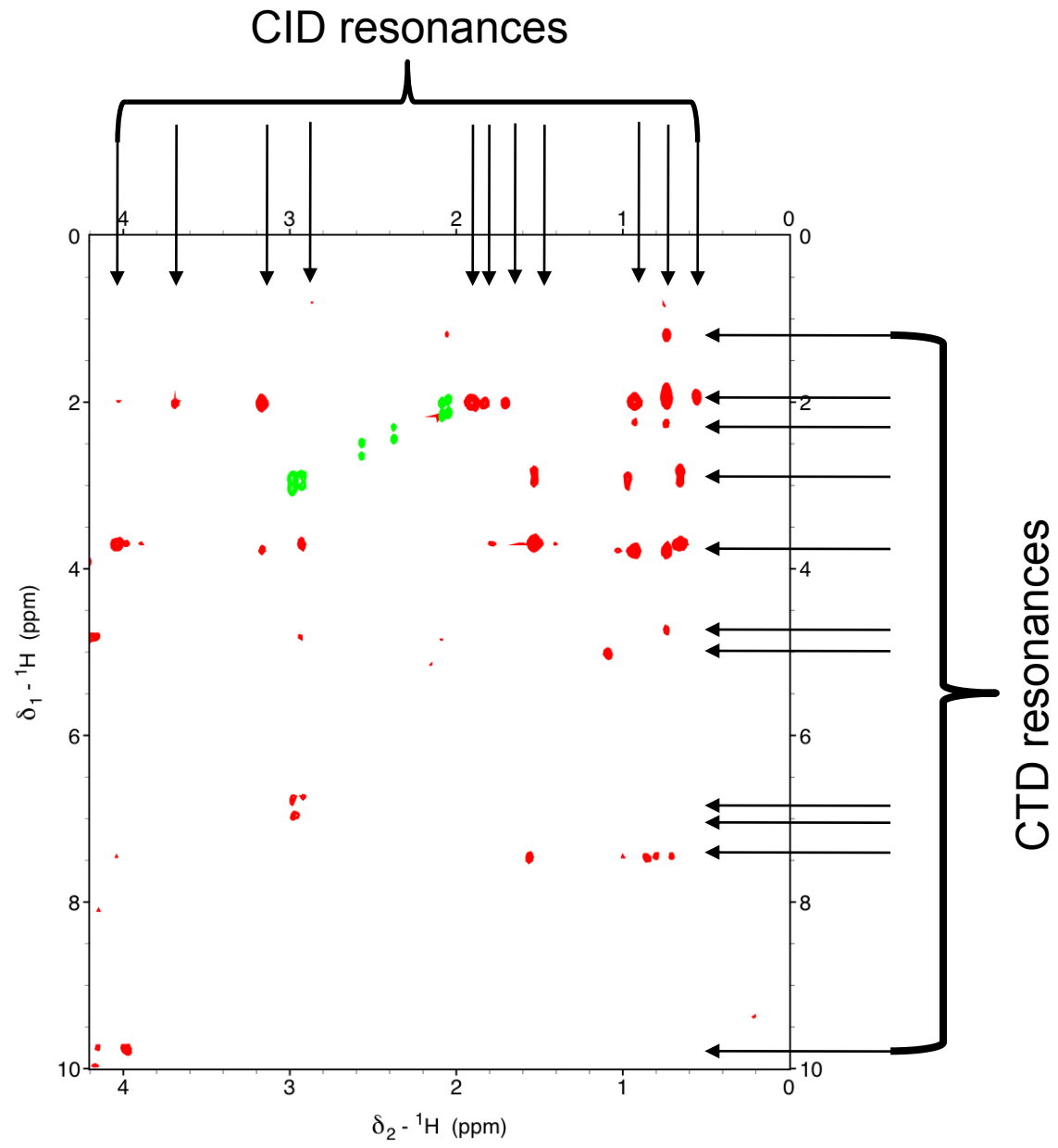
## Iterative procedure of structure determination by NMR



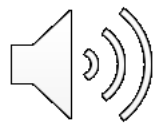
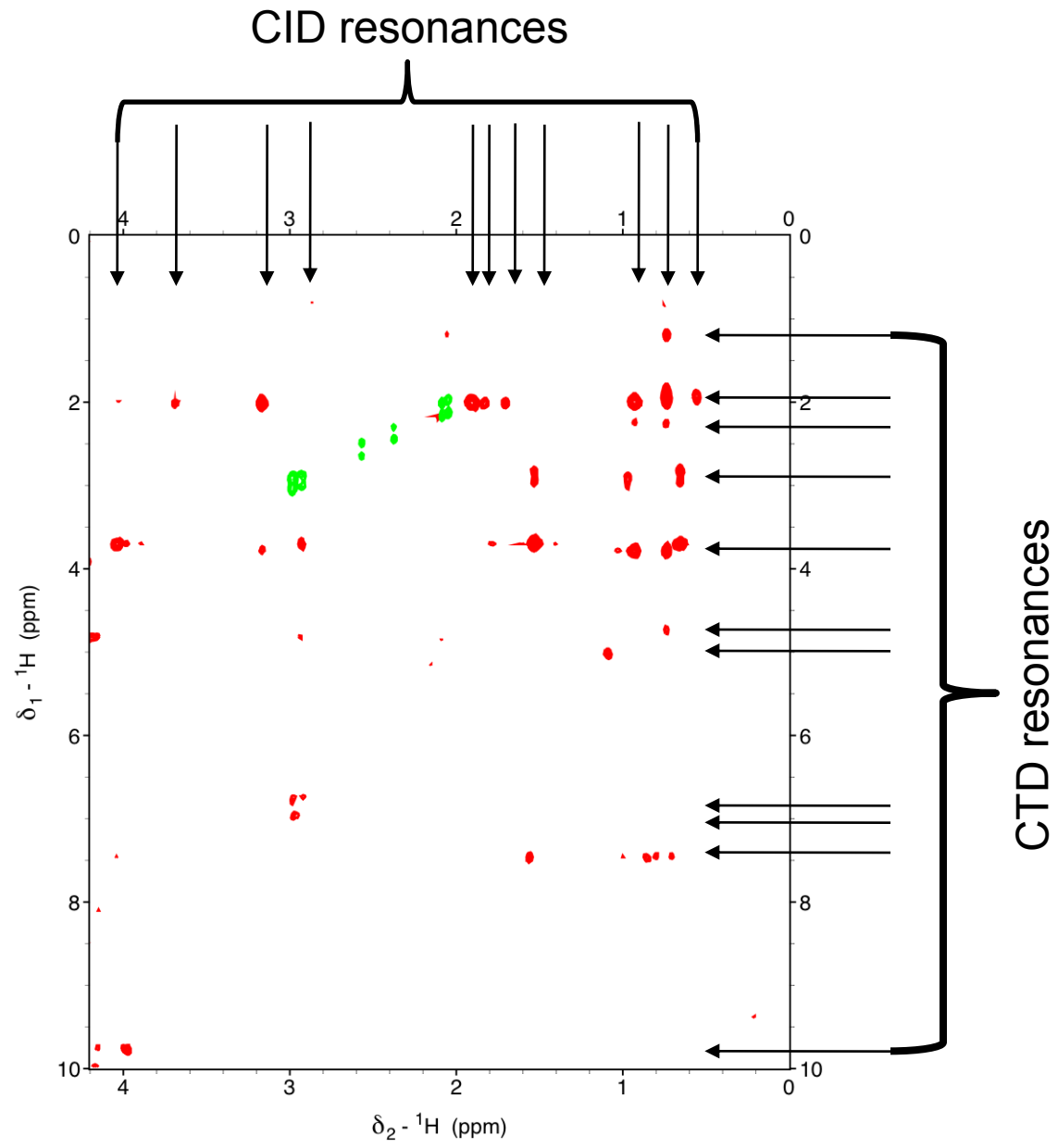
[http://www.fbregents.com/basics\\_nmr/9proteins.htm](http://www.fbregents.com/basics_nmr/9proteins.htm)



# Interligand NOEs between CID and CTD – 900MHz, 150ms, 293K



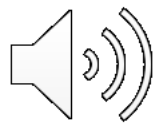
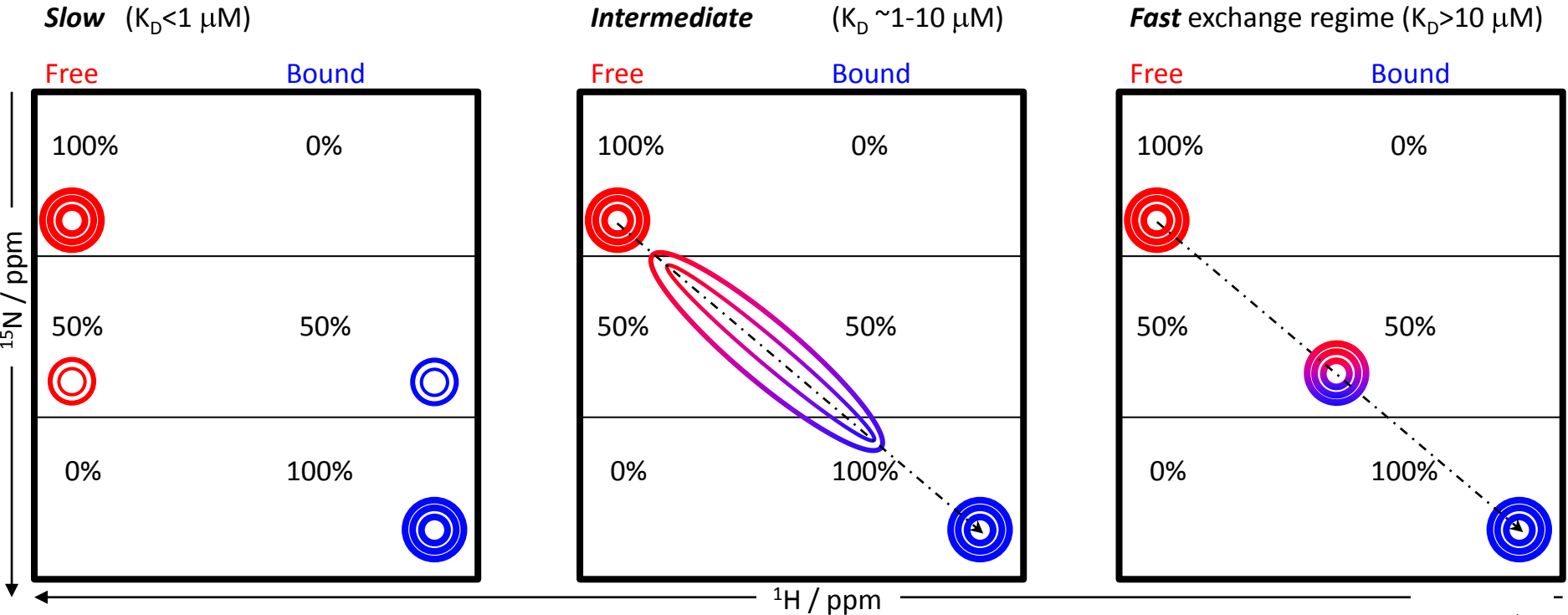
# Interligand NOEs between CID and CTD – 900MHz, 150ms, 293K



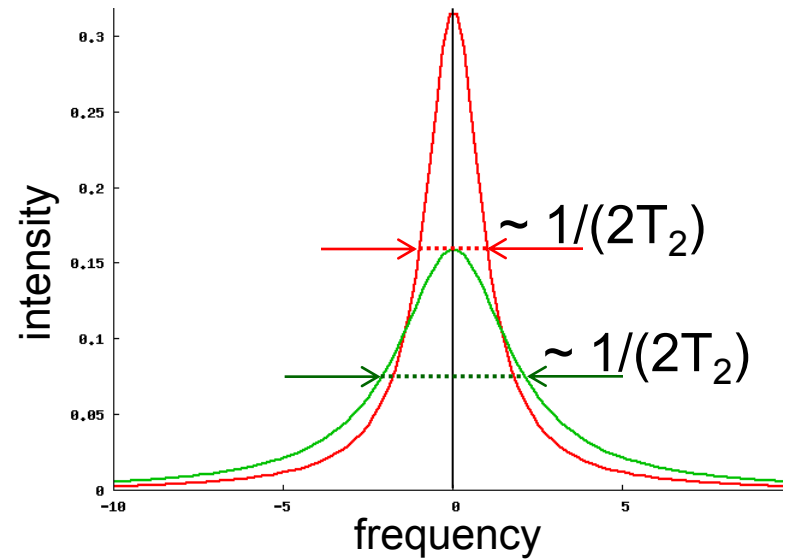
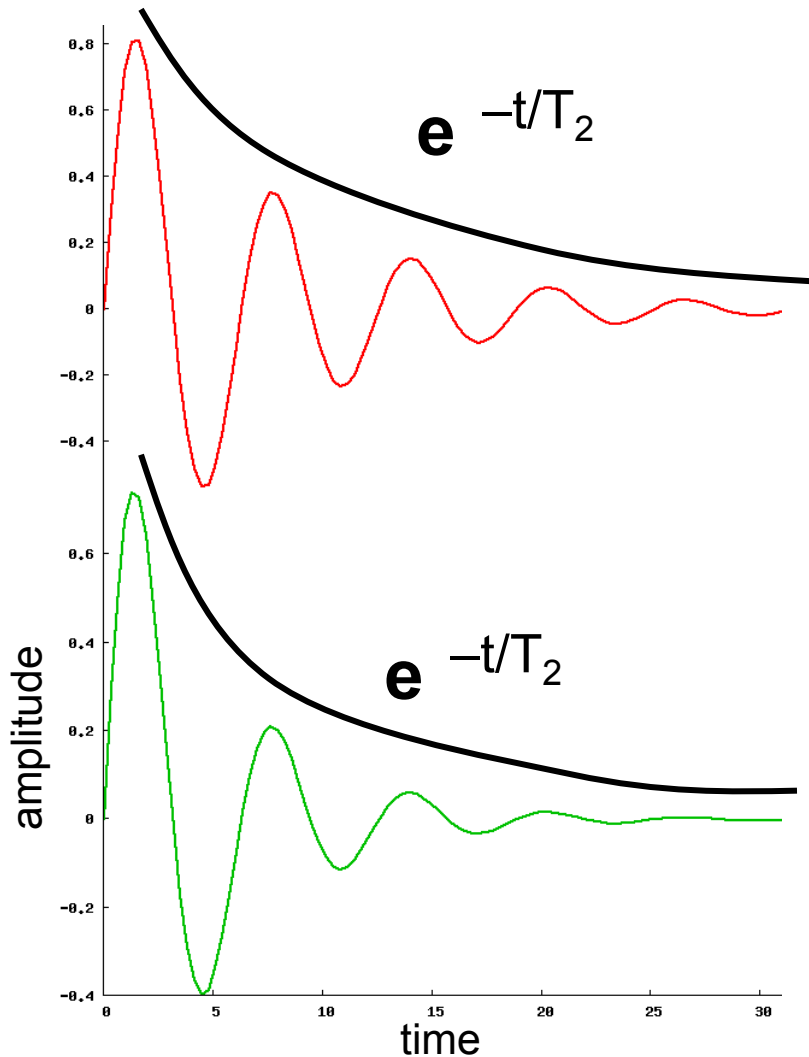
# Studying interactions by NMR titration

- 1) **Slow** exch. regime (on the NMR timescale)
- 2) **Intermediate** exchange regime
- 3) **Fast** exchange regime

- individual peaks for each of the studied states (e.g. free / complexed forms of a protein), peak intensity representing population of a given state
- single peak whose chemical shift position is given by the molar ratio of the states present in solution

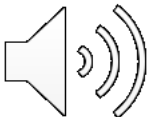


# Not all molecules relax (decay) with the same rate



**Bigger molecules** (higher molecular weight) relax faster  $\Rightarrow$  **broad peaks**

**Small molecules** relax slower  $\Rightarrow$  **narrow peaks**





**Size**

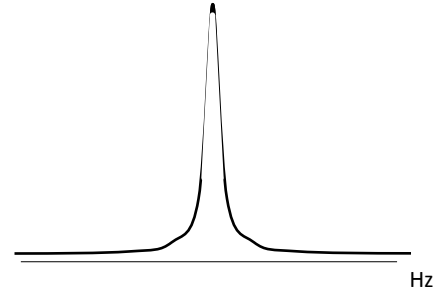
**Relaxation**

**FID**

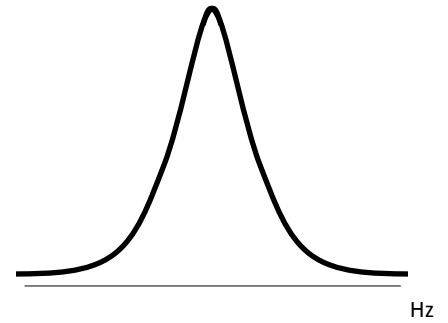
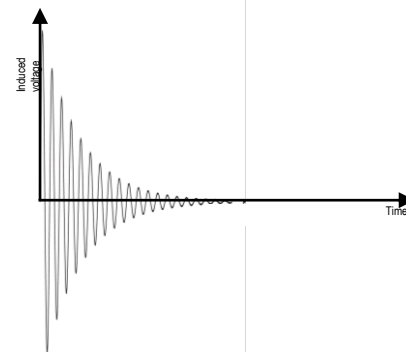
**NMR line(width) after FT**



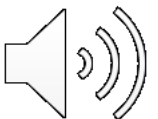
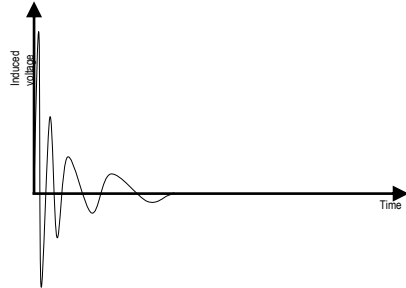
slow (i.e. long  $t_2$  time)



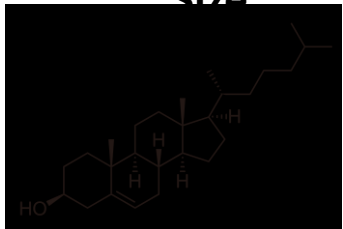
medium



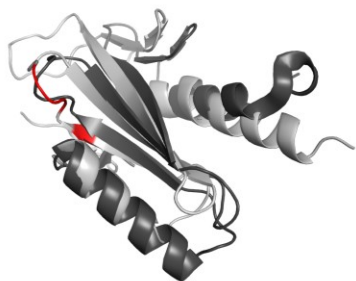
fast



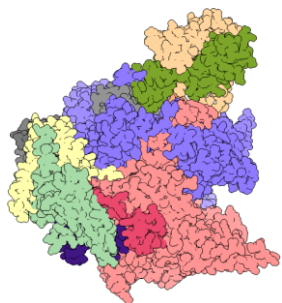
### Size



e.g. Cholesterol



Biomolecules 5-30 kDa



Large molecules 50+ kDa

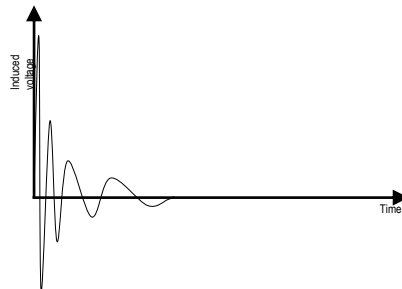
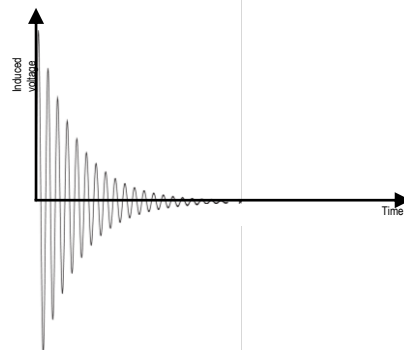
### Relaxation

slow (i.e. long  $t_2$  time)

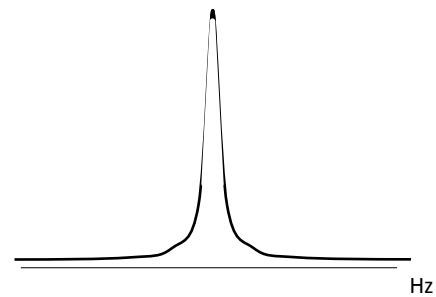
medium

fast

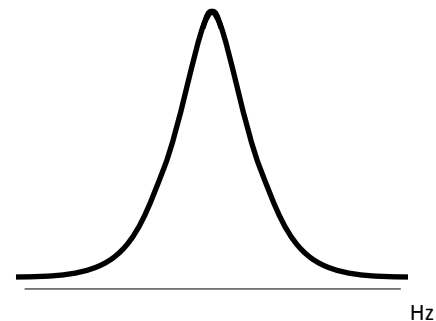
### FID



### NMR line(width) after FT



Hz



Hz



# **Protein – metal ion interaction**

slow exchange case



# DR1885 from *deinococcus radiodurans*

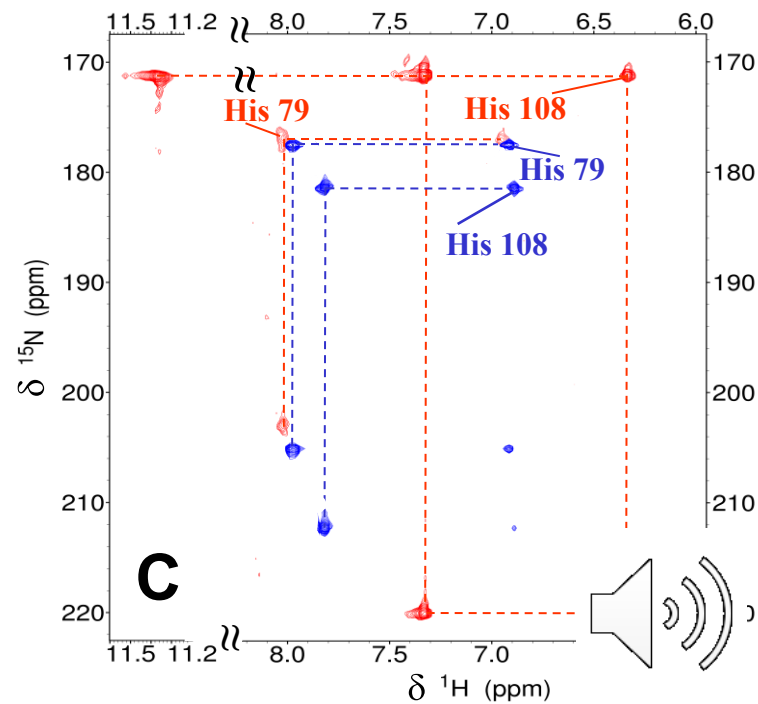
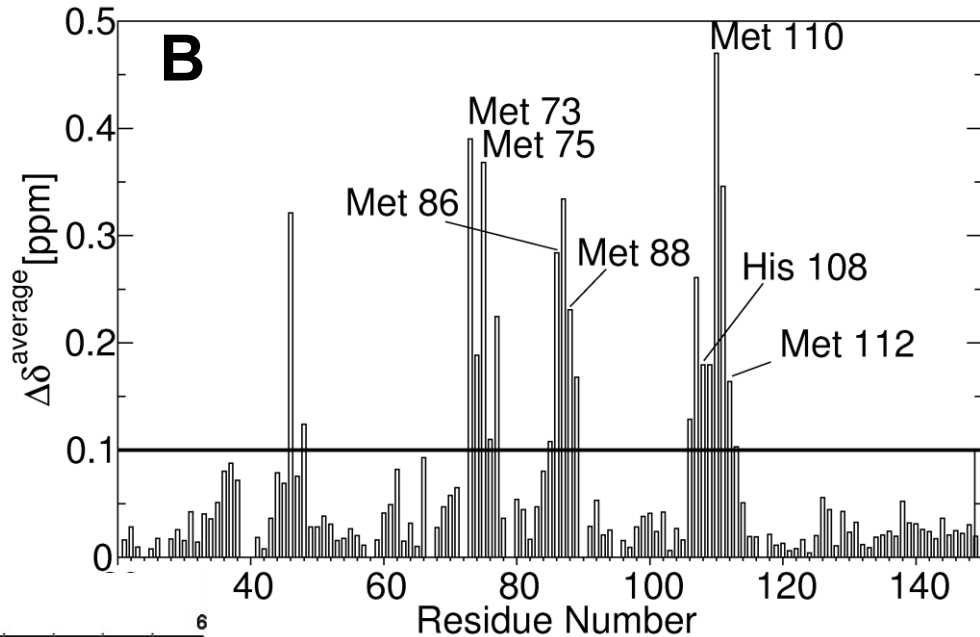
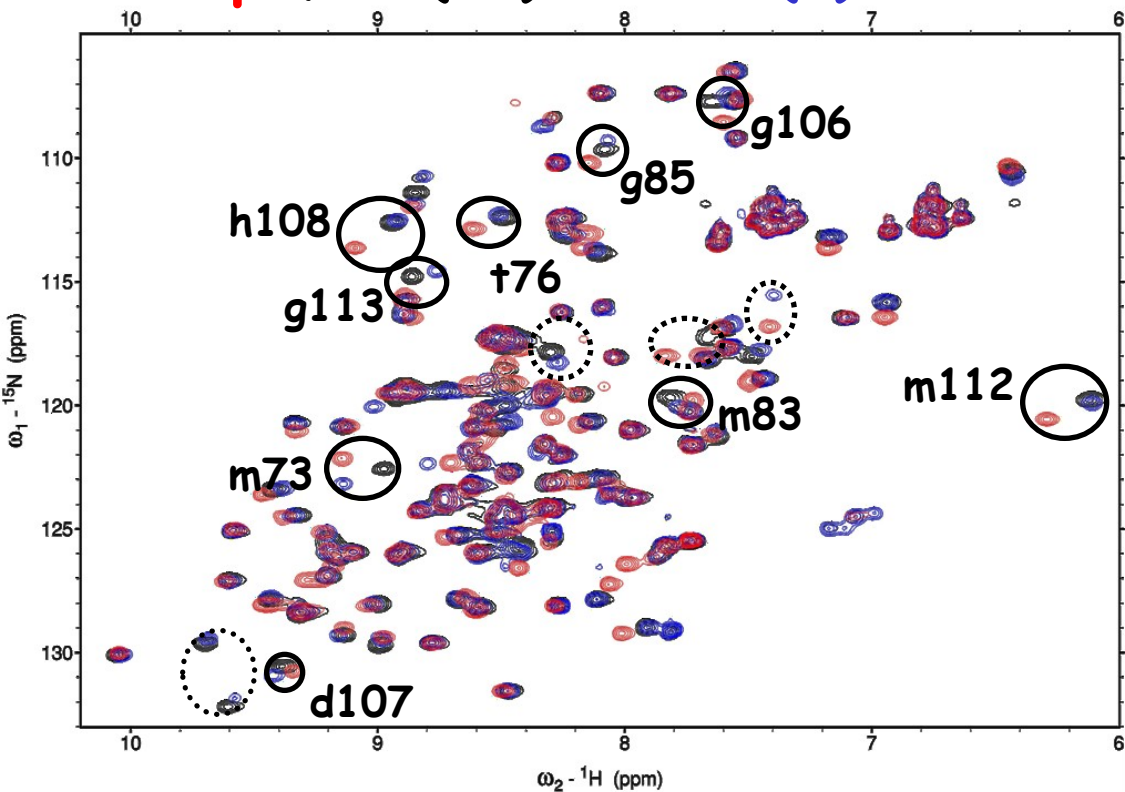
The image displays a 50x50 grid of amino acid sequences for DR1885 from *deinococcus radiodurans*. The grid is annotated with several elements:

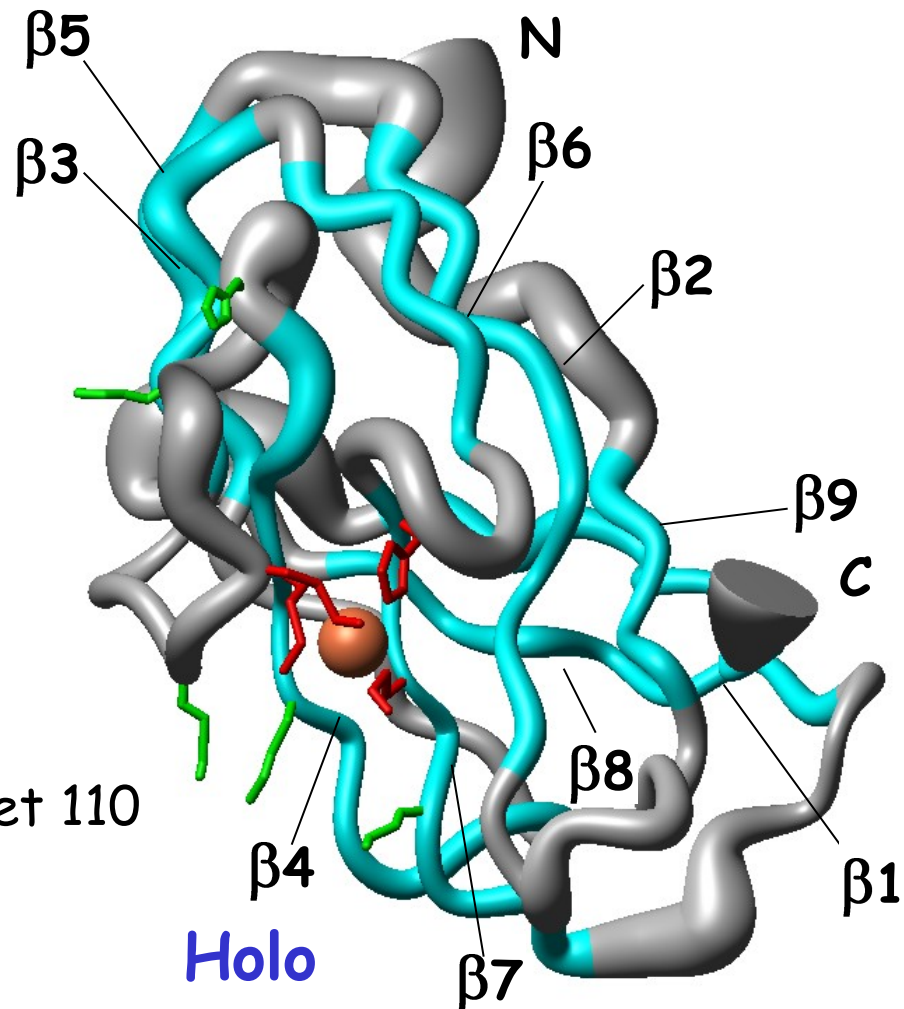
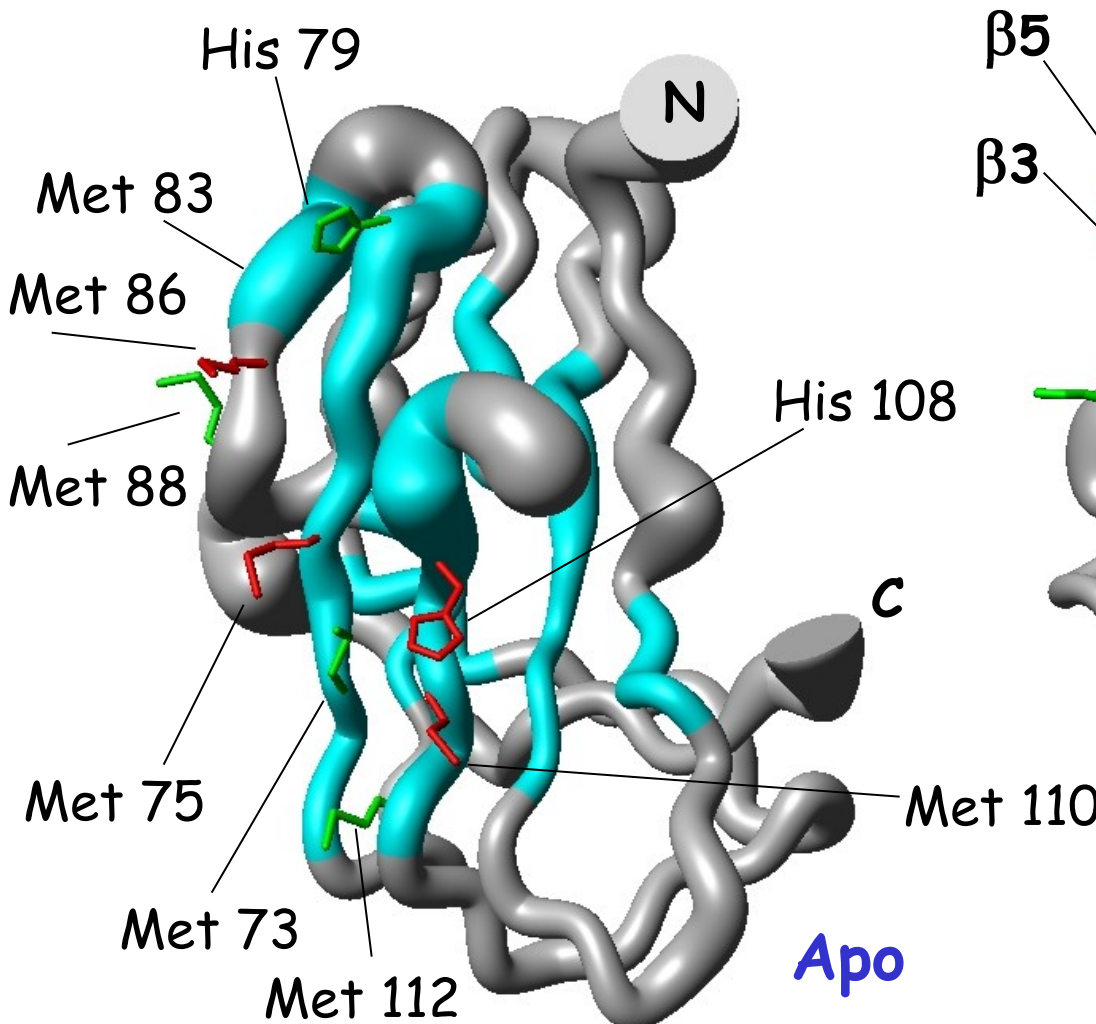
- A red 'M' in a black circle is located at the top left, with a blue oval highlighting the 'MT' sequence in row 1, column 39.
- Four black boxes with white letters 'H', 'M', 'H', 'M' and numbers '75', '86', '108', '110' are positioned at the bottom of the grid.
- Vertical red and yellow boxes highlight specific columns in the grid.
- A speaker icon is located in the bottom right corner.

# Interaction of DR1885 with copper

-titration (A,B)  
- $^2J$  HSQC (C)

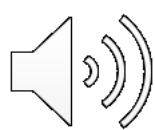
**Apo**, **Cu(II)** and **Cu(I)**





— Conserved **Met&His**  
— Other **Met&His** residues

—  $\beta$ -sheets  
— random

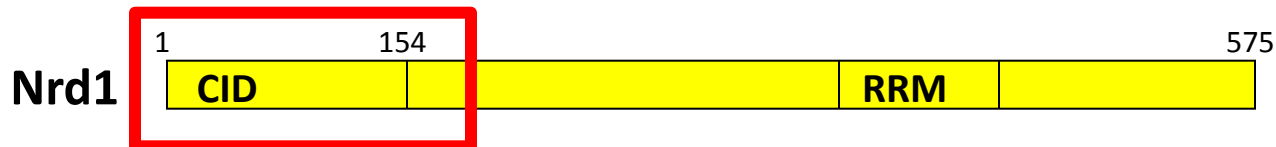
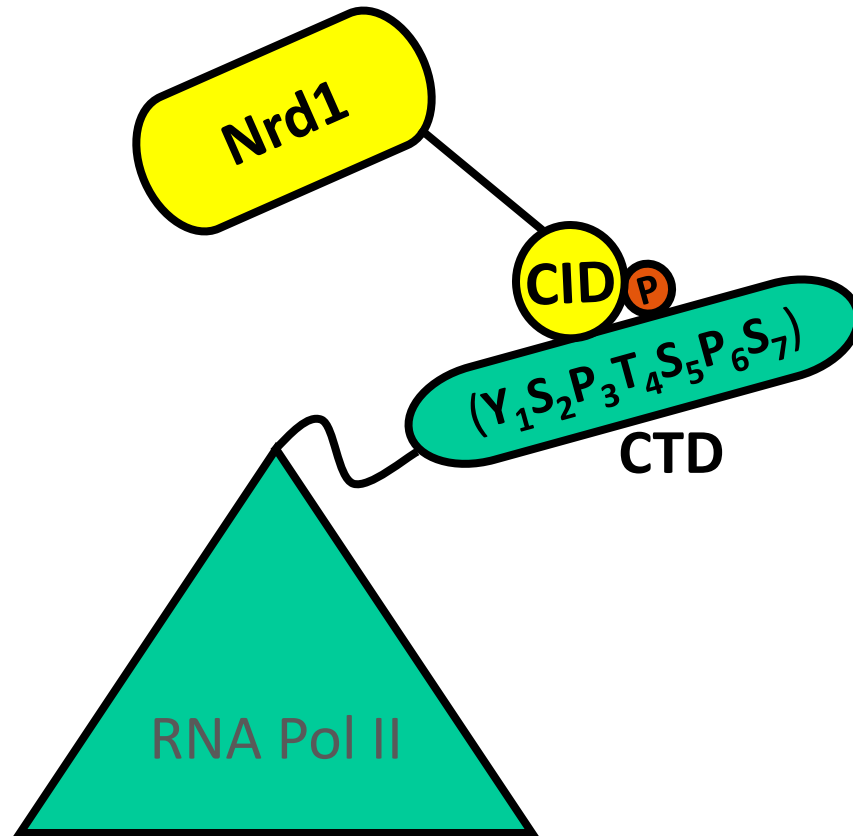


# **Protein – peptide interaction**

fast exchange case



# Interaction of Nrd1-CID with CTD

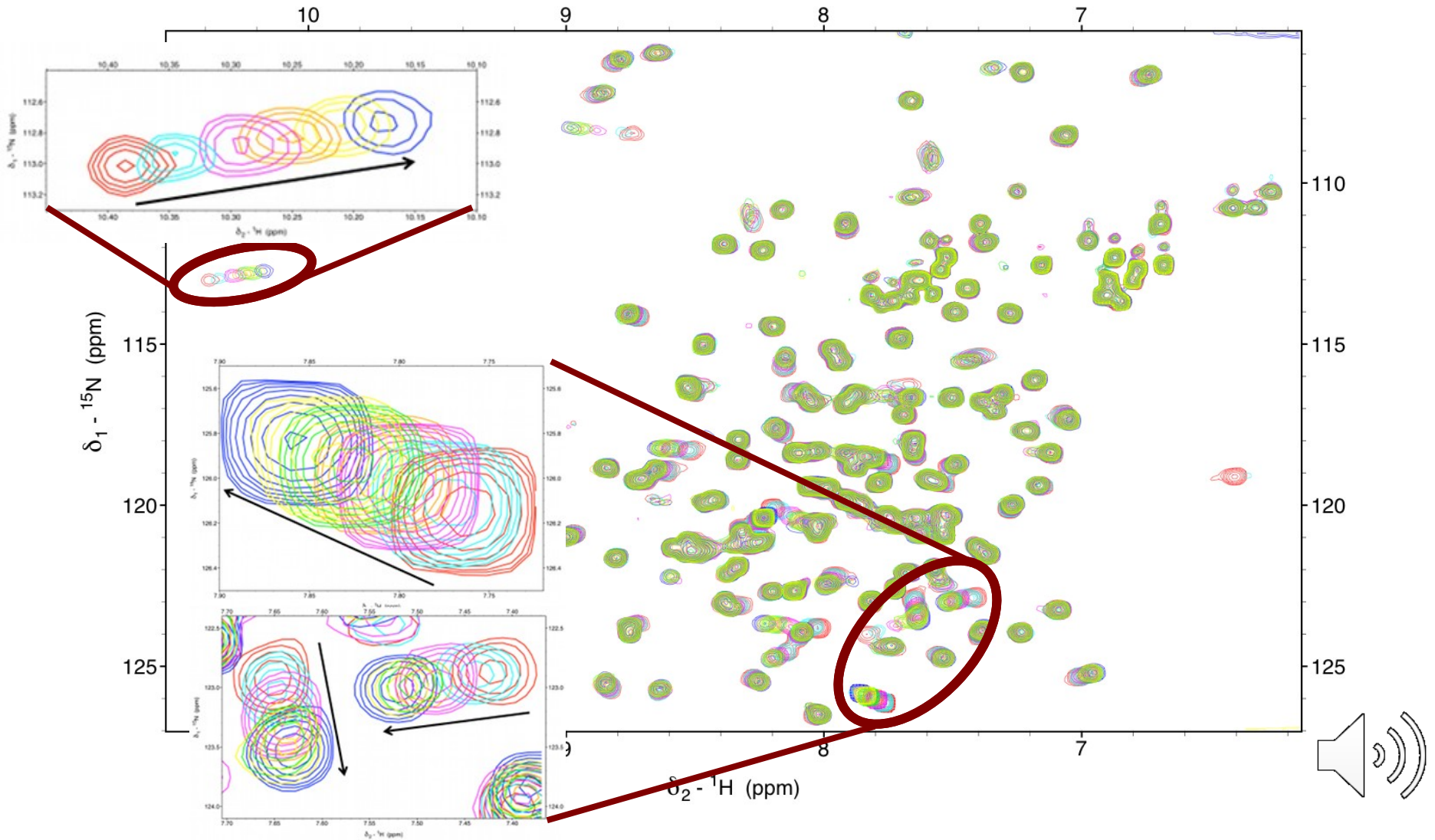


RRM: RNA recognition motif; CID: CTD interaction domain; CTD: C-terminal domain



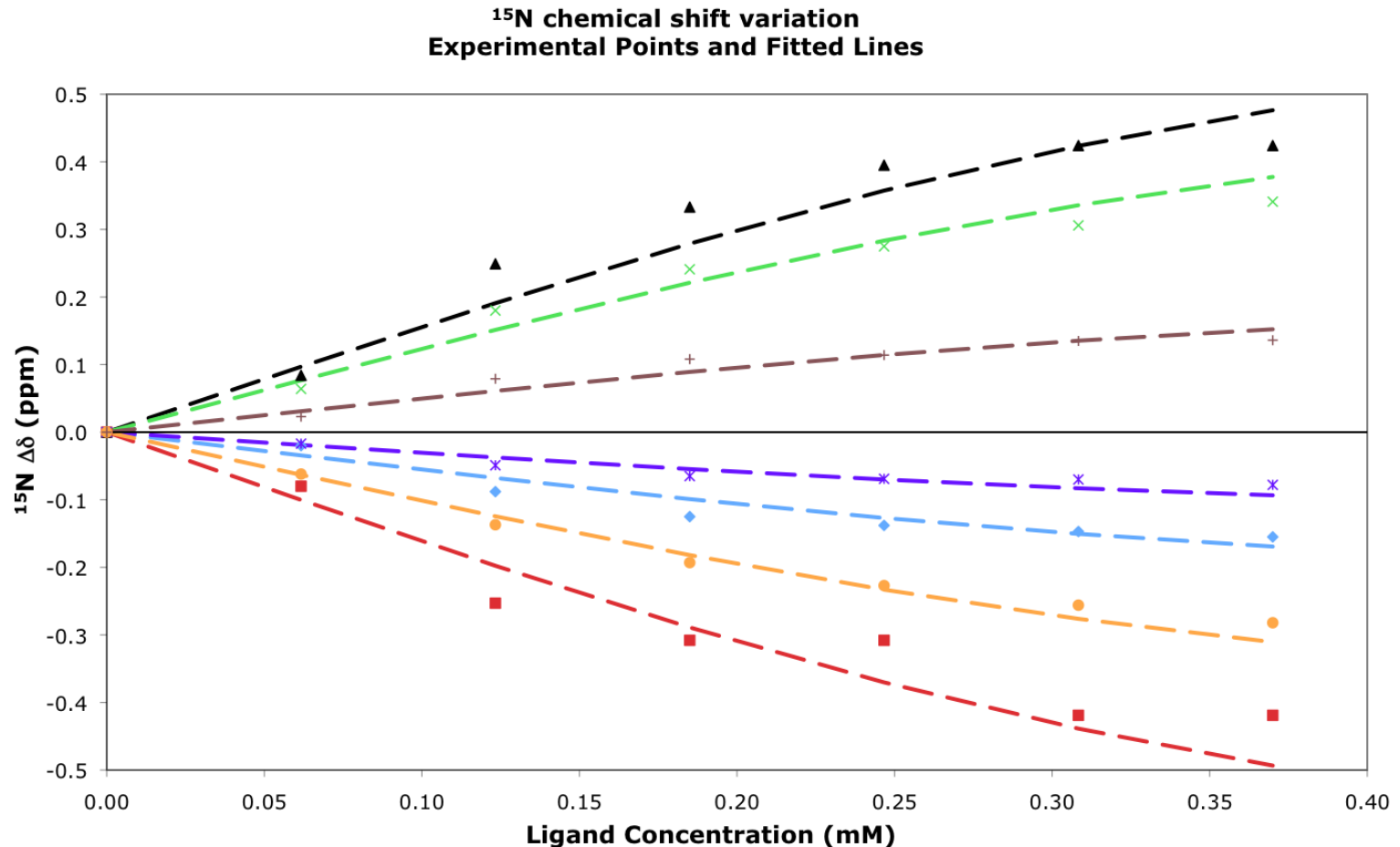


Interaction of  $^{15}\text{N}$  enriched CID with unlabeled CTD-Ser5P in  $n$ -steps,  $n=6$  in our case - peaks corresponding to the interacting residues of CID change their chemical shift (position in the spectrum) => interaction surface, binding constant, stoichiometry

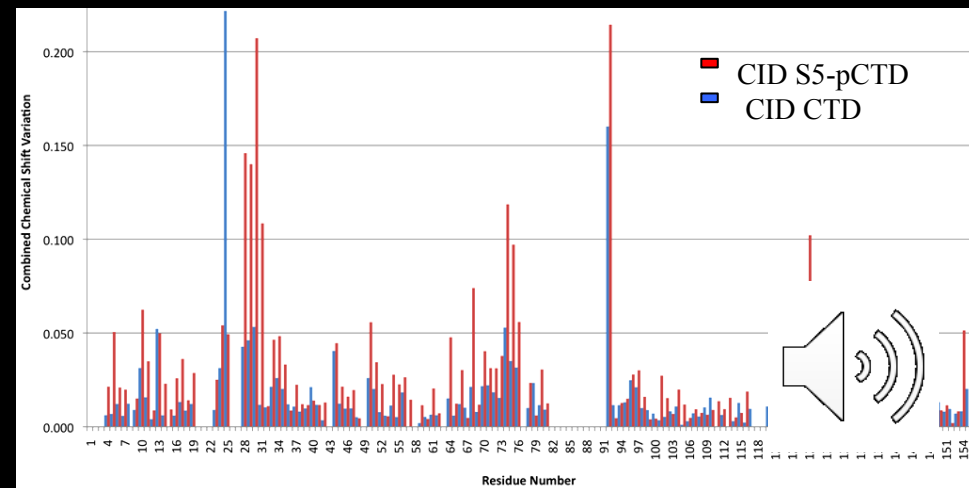
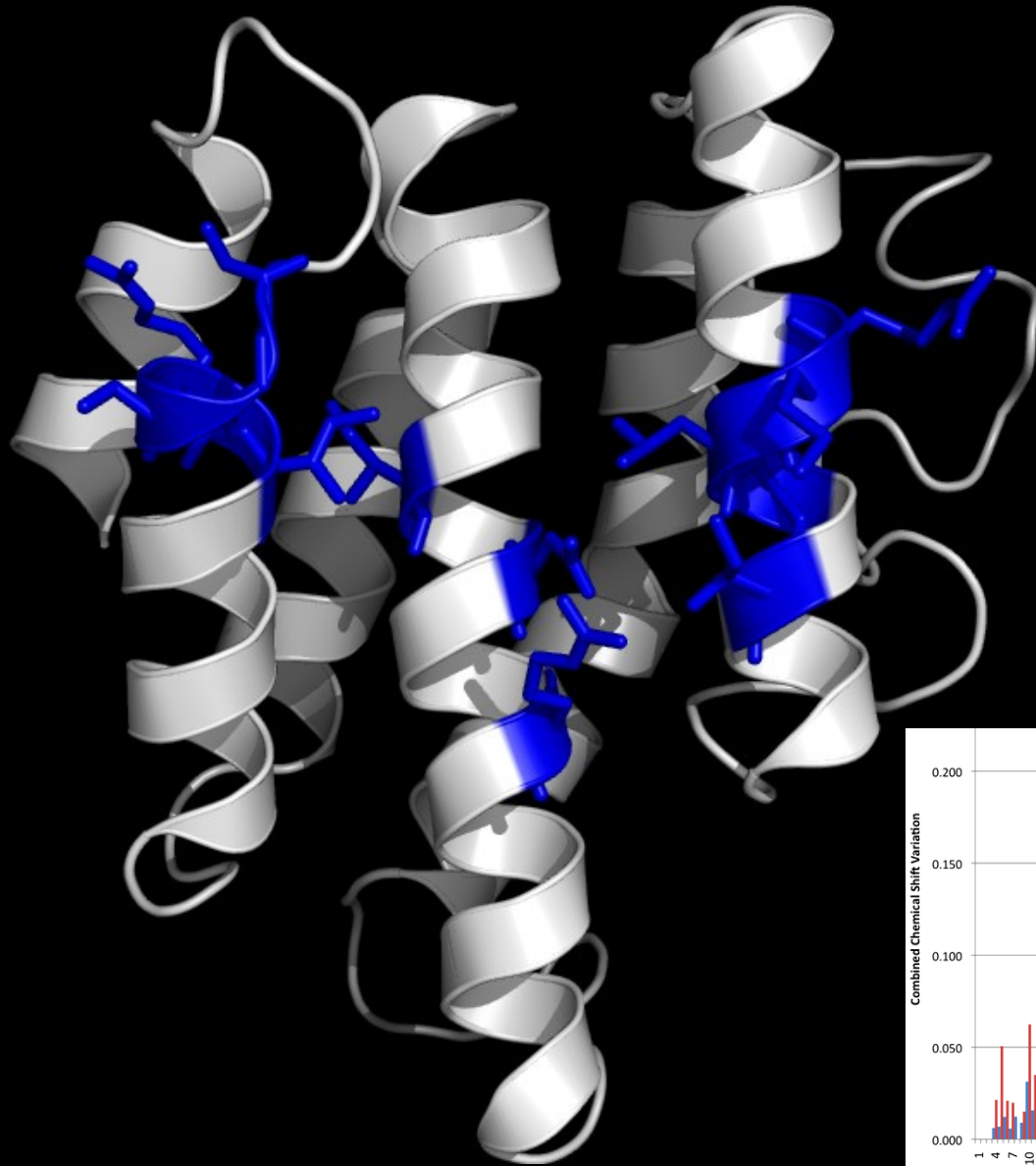


# Interaction of Nrd1-CID with CTD

- NMR Titration:
- ~ 0.6 mM  $^{15}\text{N}$  enriched CID + ~ 0.8mM (YSPT<sub>p</sub>SPS)<sub>2</sub>
  - ~ 0.6 mM  $^{15}\text{N}$  enriched CID + ~ 0.8mM (YSPT<sub>S</sub>SPS)<sub>2</sub>
  - $\mu\text{M}$ -mM range of interaction ->
  - > fast exchange regime on NMR time-scale
  - NMR-derived  $K_d$ =0.080mM and 35mM



# Nrd1 CID interaction surface — CID residues experiencing the largest chemical shift variations upon the interaction with 5-phospho-Ser CTD shown in blue with side-chains in stick representation

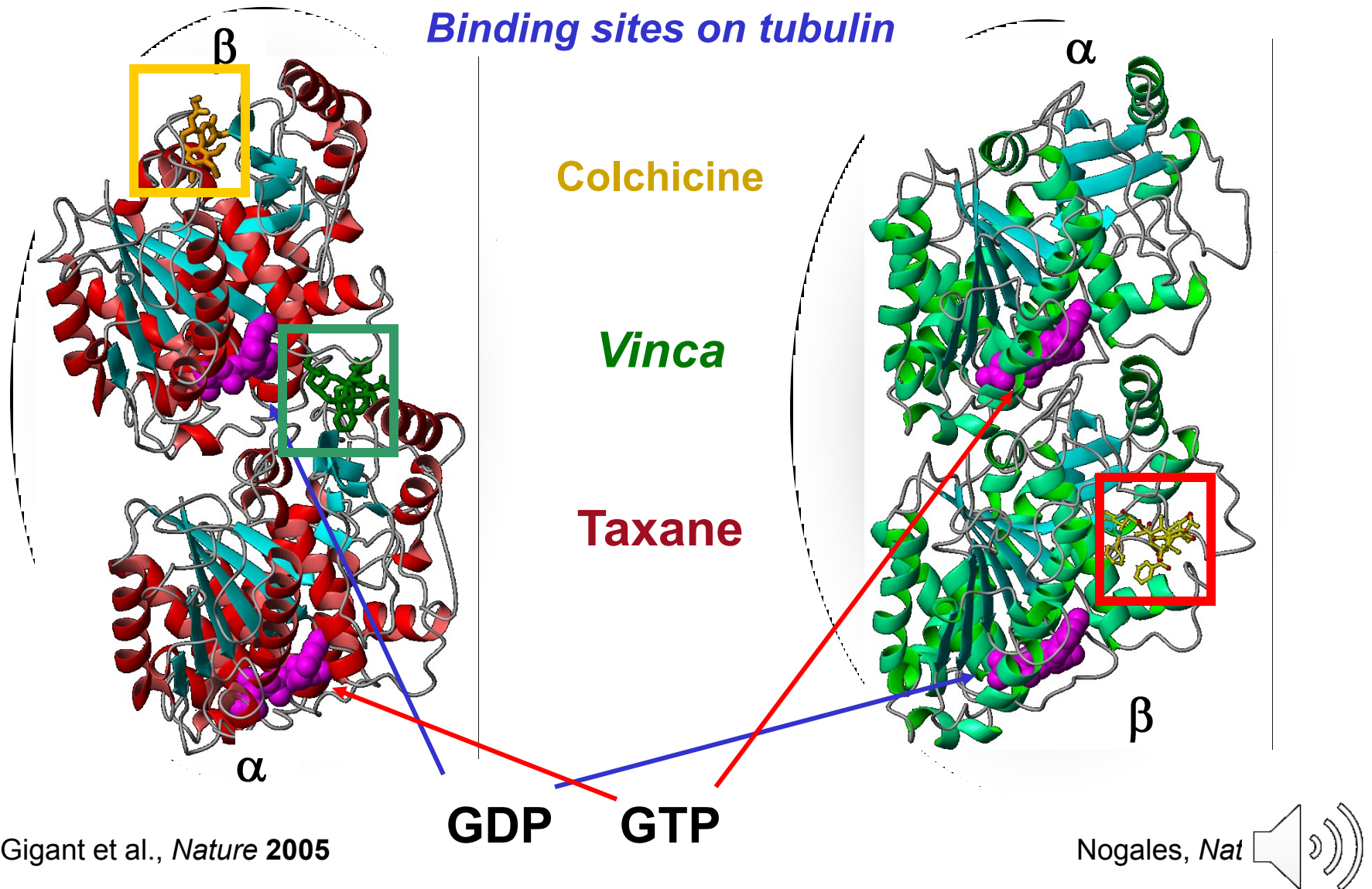


# **Protein – peptide interaction**

drug-receptor case



# *Tubulin* - successful target for anticancer therapy

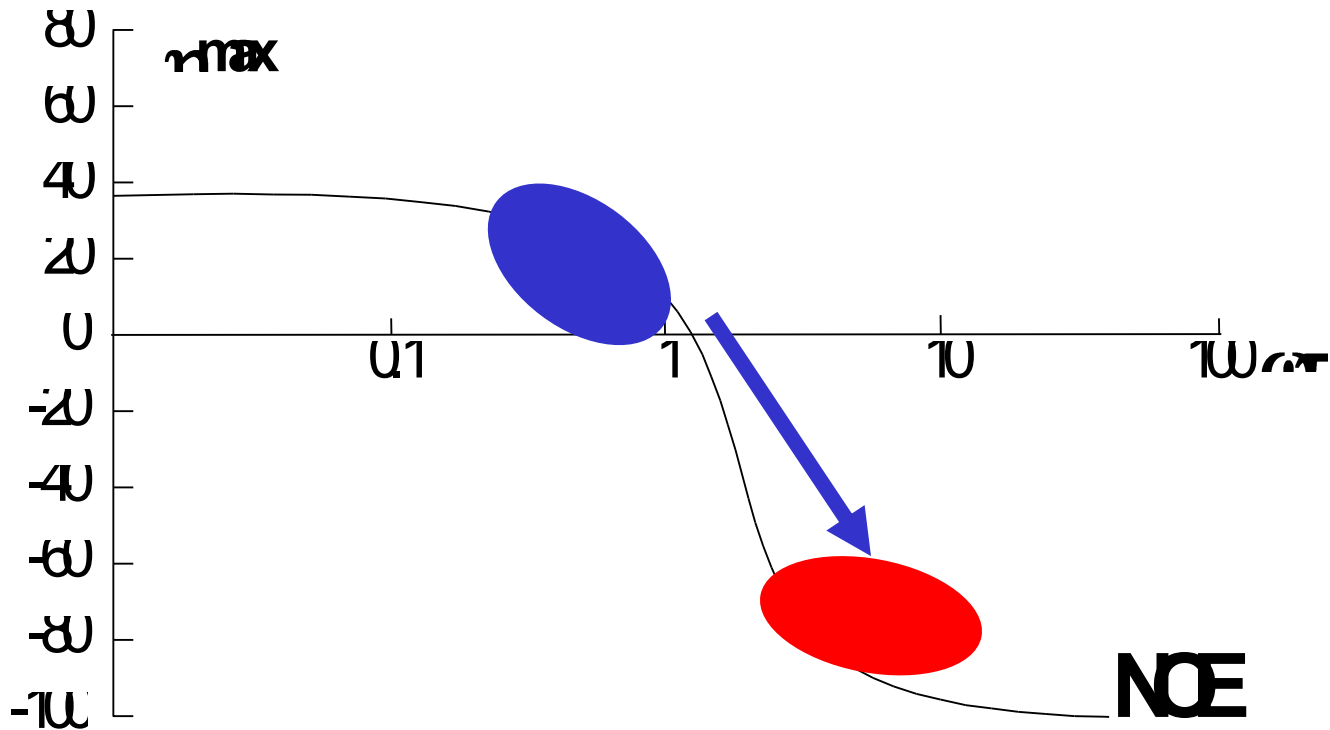


# Transferred-NOE

$$\text{NOE} = p_{\text{bound}} \cdot \text{NOE}_{\text{bound}} + p_{\text{free}} \cdot \text{NOE}_{\text{free}}$$

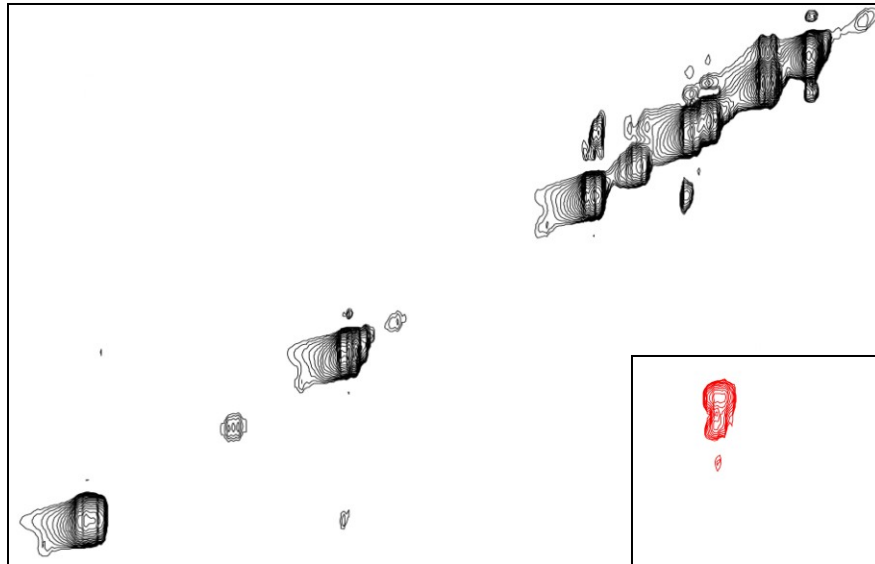
$$\tau_{c,\text{bound}} \gg \tau_{c,\text{free}} \quad (\text{and} \quad p_{L,\text{free}} \gg p_{L,\text{bound}})$$

$$\text{NOE}_{\text{bound}} > \text{NOE}_{\text{free}}$$

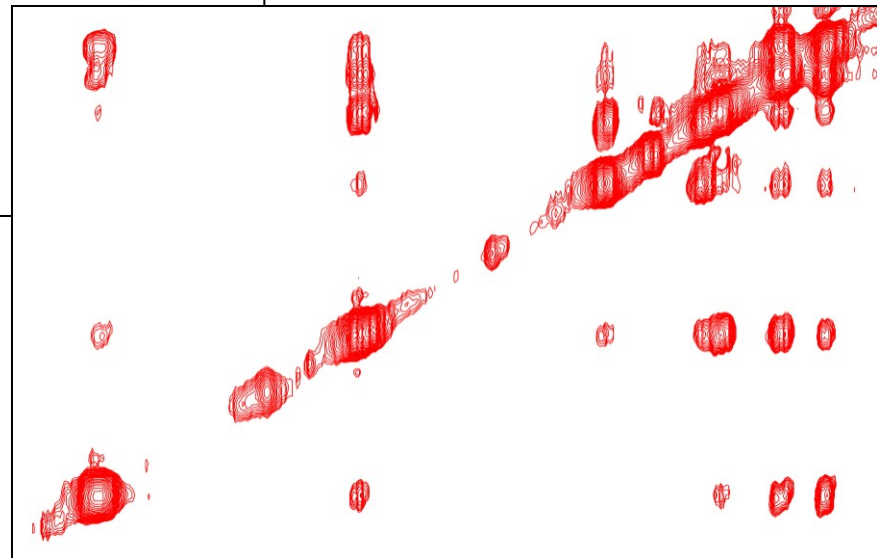


# Transferred NOE Experiments

tr-NOESY ~500 $\mu$ M tubulysin (TBS) **without** and **with** ~10 $\mu$ M tubulin



900MHz,  
mixing time=100ms

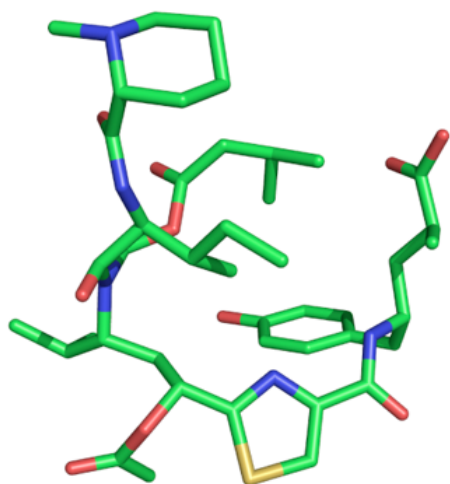


↑  
tbs w/o tub

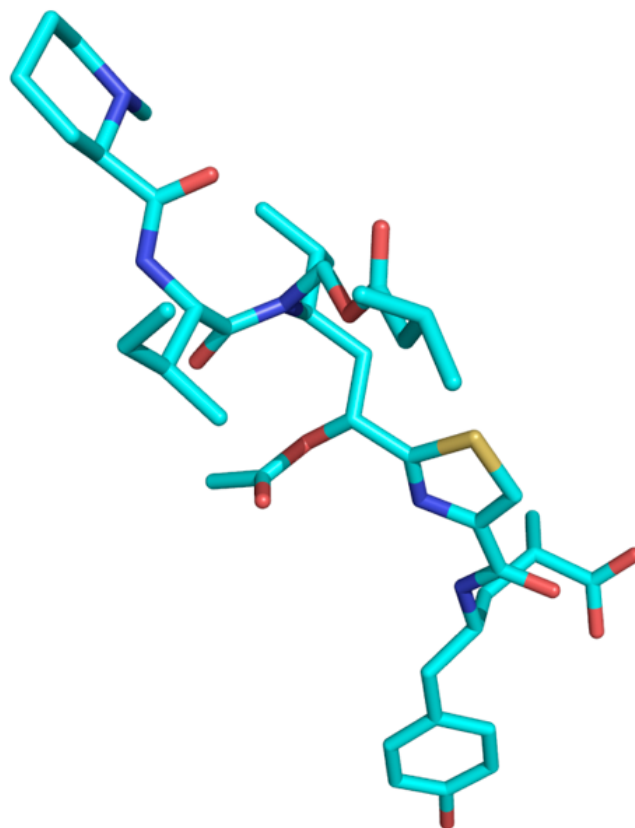
tbs:tub 50:1



A



B



Conformation of the **tubulin-bound** - NMR (**A**) and **free** – X-Ray TDDFT (**B**)



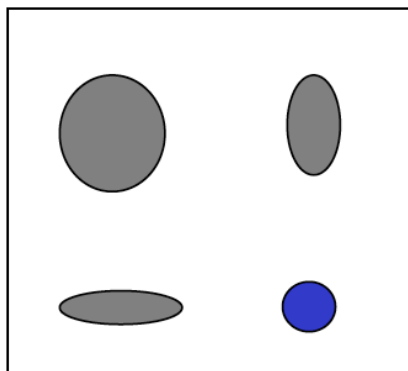


# **Large biomolecules and their interactions**

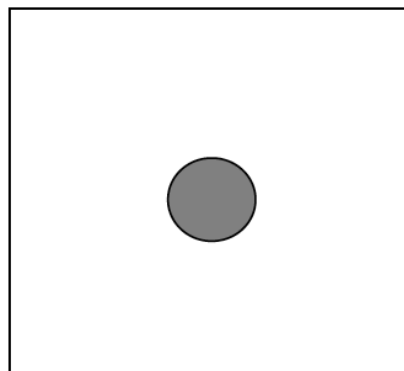


# TROSY-based NMR experiments to study complexes up to 900kDa

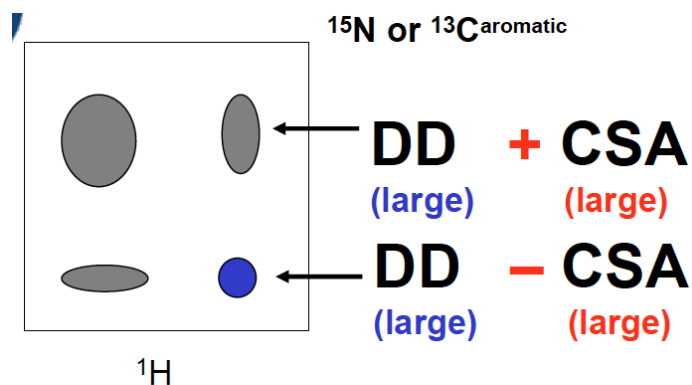
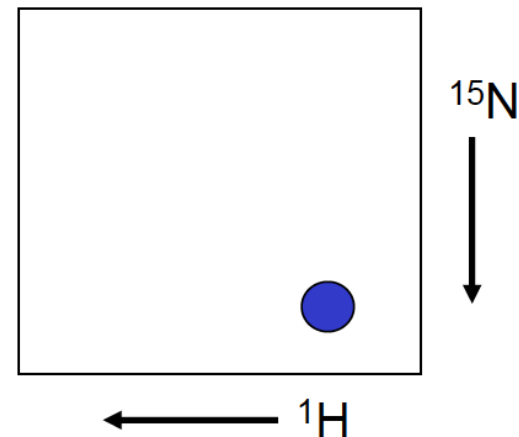
Coupled HSQC



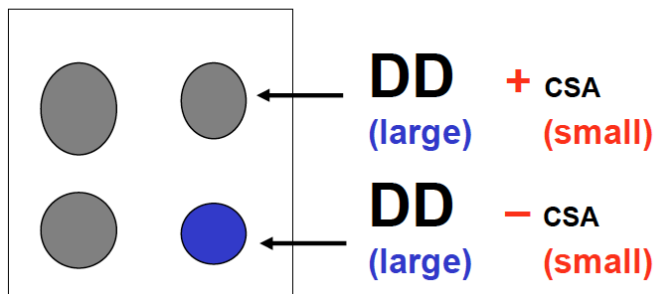
Decoupled HSQC



TROSY-HSQC



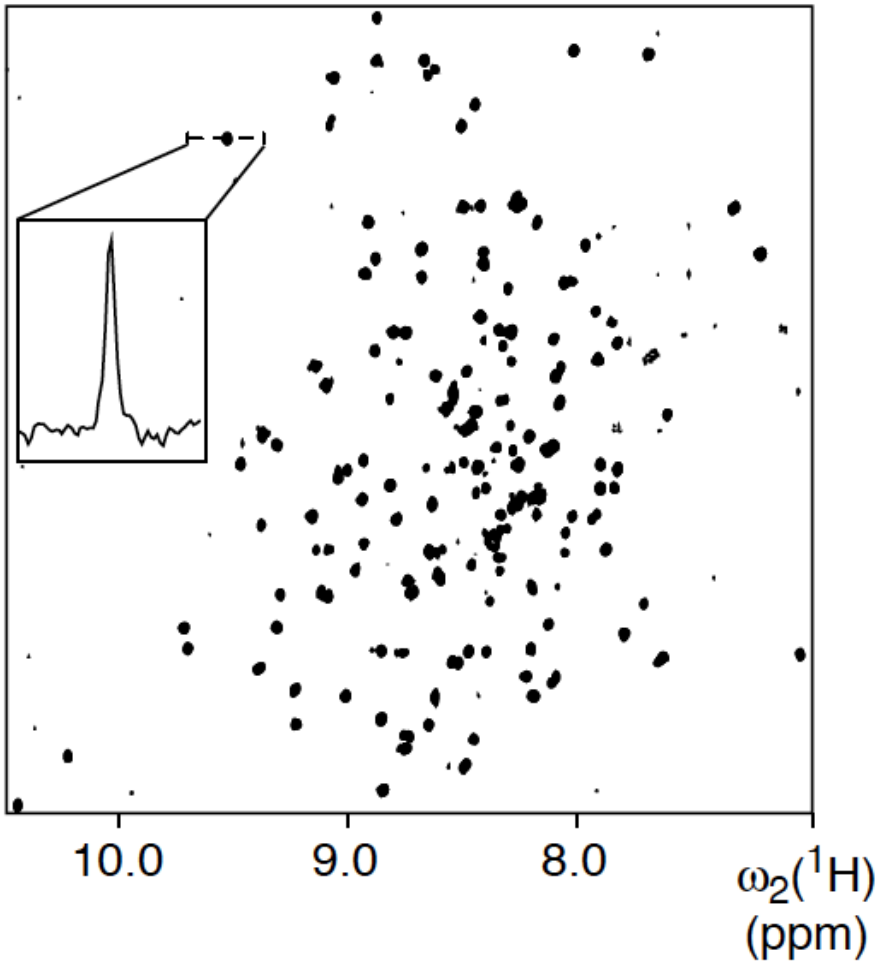
high mg. field >700 MHz



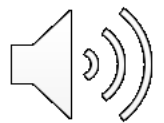
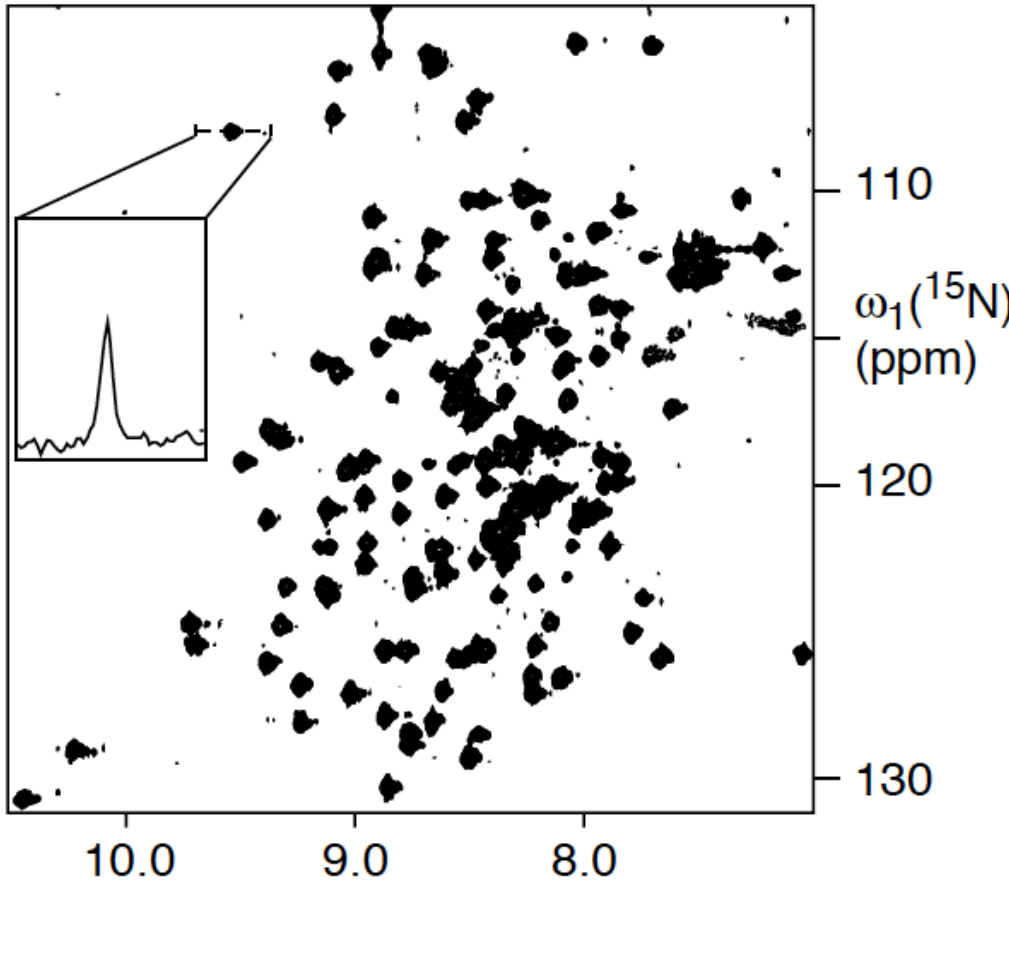
low mg. field <700 MHz



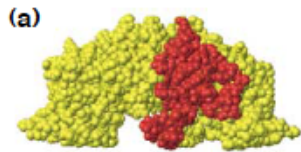
**(a)** 2D [ $^{15}\text{N}$ ,  $^1\text{H}$ ]-TROSY



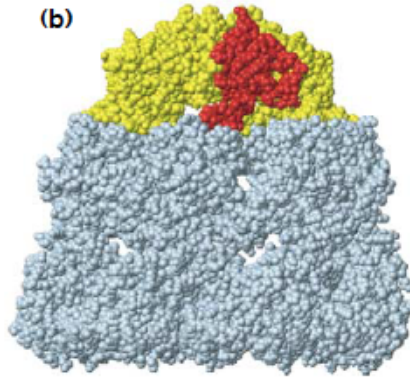
**(b)** 2D [ $^{15}\text{N}$ ,  $^1\text{H}$ ]-COSY



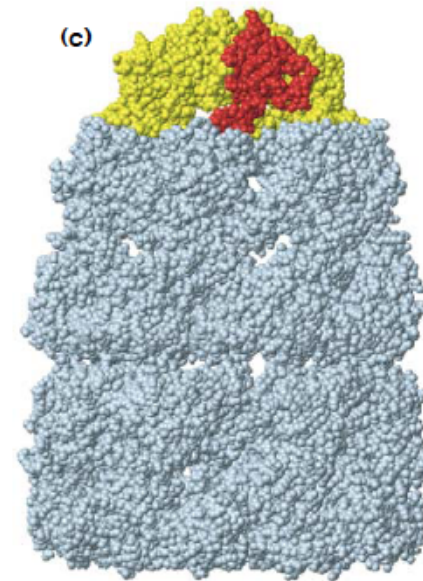
70%  $^2\text{H}$ ,  $^{13}\text{C}$ ,  $^{15}\text{N}$  sample in  $\text{H}_2\text{O}$ , 700MHz  
CRINEPT-TROSY



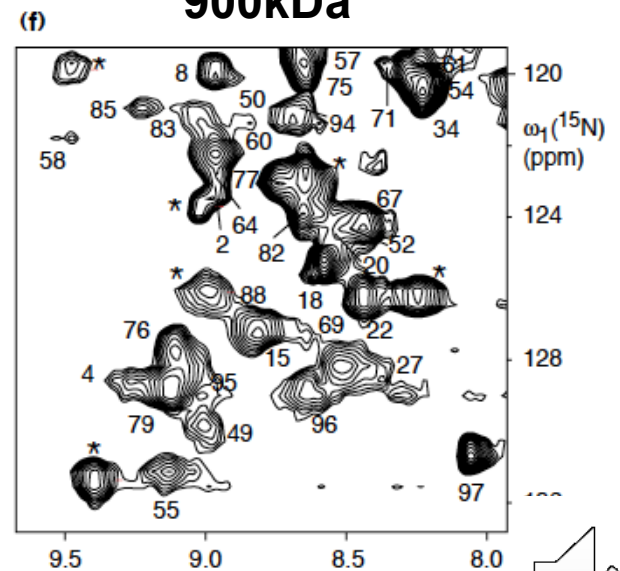
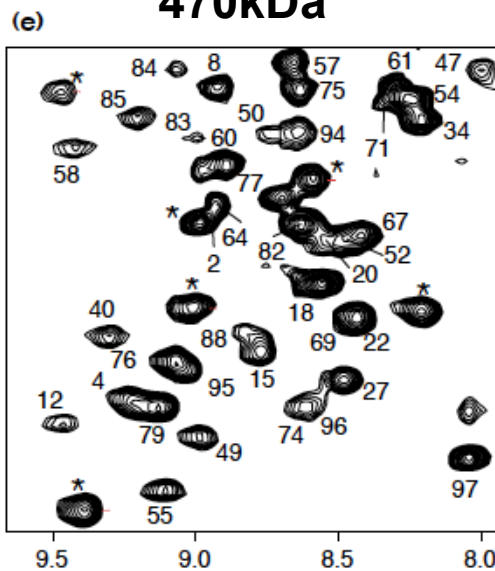
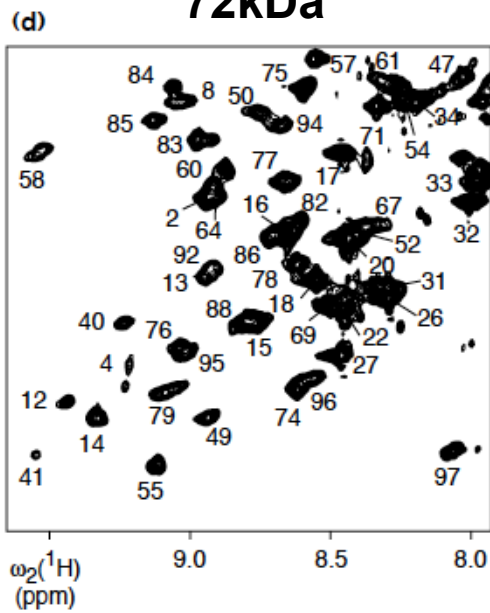
72kDa



470kDa



900kDa



# Summary

NMR is a robust tool for studying structural properties and interaction properties of biomolecules of variable molecular size at various levels of resolution.



# Thank you for your attention

For Application to Protein Characterization

by

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