

Tato prezentace je spolufinancována Evropským sociálním fondem a státním rozpočtem České republiky



Genomika 05	
 Zdrojová literatura 	
 Surpin, M. and Raikhel, N. (2004) Traffic jams a signal transduction. Nature Reviews/Molecular Ce 	affect plant development and Il Biology 5,100-109
 Zouhar, J., Hicks, G.R. and Raikhel, N.V. (2004 Chemical compounds to study vacuolar sorting in the National Academy of Sciences of the U.S.A., 1 	4) Sorting inhibitors (Sortins): n Arabidopsis. Proceedings of 101, 9497–9501
 Nevo-Dinur, K., Nussbaum-Shochat, A., Ben-Yeh O. (2011). Translation-independent localization of 1081-1084. 	nuda, S., and Amster-Choder, mRNA in E. coli. Science 331,
 Lecuyer, E., Yoshida, H., Parthasarathy, N., Alm, C., Babak, T., Cerovina, T., Hughes, T.R., Tomancak, P., and Krause, H.M. (2007). Global analysis of mRNA localization reveals a prominent role in organizing cellular architecture and function. Cell 131, 174-187. 	
 Schonberger, J., Hammes, U.Z., and Dresselhaus, T. (2012). In vivo visualization of RNA in plants cells using the lambdaN(22) system and a GATEWAY-compatible vector series for candidate RNAs. The Plant journal : for cell and molecular biology 71, 173-181. 	

































































Polar localization of mRNA can be achieved by three different mechanisms: (i) local stabilization and regulated degradation of mRNA; (ii) local trapping of an RNA that is diffusing through the cytoplasm; and (iii) active and directed transport of mRNA. Although there are examples for all three mechanisms, the latter is the most common mechanism employed. The transported mRNP complexes form larger structures, which are referred to as RNA transport granules. These granules are transported by motor proteins along microtubules and/or the actin cytoskeleton to their final destination. In addition to the association with the cytoskeleton there is also evidence that mRNA transport and ER trafficking may be coupled (Schmid et al., 2006; Aronov et al., 2007). Before mRNPs reach their final subcellular destination, translation is usually delayed by the recruitment of translational repressors (Schoenberger et al., Plant J., 2012).

Figure 2. Anterior/Posterior Patterns and Functional Enrichments (A–E) Sagittal views of entire embryos (A and B) or of the posterior region (C–E) between stages 2–5, following FISH with probes to bcd (A), asp (B), osk (C), orb (D), or grp (E) transcripts (mRNA green/nuclei red). (A and B) Varieties of anterior patterns, with bcd mRNA (A) showing tight anterior localization and asp transcripts (B), a more diffuse anterior enrichment. (C–E) Early and late posterior localization patterns. While both osk and orb transcripts localize to the posterior pole plasm in stage 1–2 embryos ([C and D] arrowheads), orb mRNA forms distinctive rings around pole cell nuclei at stage 3 ([D] arrow).



Figure 4. Membrane-AssociatedPatterns (A–F) Surface plane (upper panels) and sagittal (lower panels) views of stage 3 (A and B), 4 (C), 5 (E and F), and 6 (D) embryos hybridized with probes for the transcripts indicated in lower panels (mRNA green/nuclei red). cno transcripts localize within cortical polygonal networks ([A] arrowhead), while anillin mRNA is first perinuclear ([B] arrowhead) and then evolves into a cell junction type pattern (C). (D–F) Patj, dlg1, and mira transcripts localize at different positions along the lateral membrane (arrowheads).



Figure 5. Cell Division and Nuclei-Associated Transcripts (A–P) Surface plane (A–L and O) or sagittal (M, N, and P) views of stage 1–5 embryos hybridized with the indicated probes (mRNA green/nuclei red). (A–H) Examples of mRNAs that localize to different sections of the cell division apparatus, including spindle poles (A), microtubule networks and centrosomes (B, C, E, and F), the spindle midzone (G and H), or in proximity to metaphase chromosomes (D). (I and J) Doc-element transcripts localize to centromeric chromatin regions on polar body chromosomes (I) and during mitosis in diploid nuclei (J). (K and L) Ste12DOR transcripts localize in chromatin-associated foci during metaphase (K), which then become telomeric during anaphase (L).



(A-F) Stage 4 (B-E), 5 (A) and 9 (F) embryos hybridized with probes for the indicated apical (A), cell division-associated (B and C), membrane-associated (D), and nuclear retained (E and F) mRNAs (red signal, left panels) and colabeled with antibodies against the indicated

protein products (green signal, middle panels). Overlaid mRNA and protein signals are shown in the right panels. Nuclei are shown in blue in the left and middle panels in (A)–(E). Arrowheads in (E) and (F) indicate nuclei showing an accumulation of kuk and cas mRNAs, respectively.



Subcellular localization of mRNA transcripts correlates with subsequent localization of their protein products. (A) Fluorescence microscopy images of cells expressing MS2-GFP (green) and transcripts containing or lacking binding sites for MS2-GFP. "6xbs" after a gene's name denotes that the MS2 binding sites were introduced immediately next to this gene. Cells shown in (I) were supplemented with the fluorescent membrane stain FM4-64. Transcripts were plasmid-encoded, unless otherwise stated. Scale bar, 1 mm.

(C) (a and b) Fluorescence microscopy images of cells expressing MS2-GFP (green) and a BglF-mCherry fusion protein red), whose transcript contains the 6xbs. (c) An overlay of the signals. Scale bar, 1 mm. The possibility of false detection due to fluorescence leakage between the filters was ruled out. The white arrows point at a cell in which the 6xbs-tagged bglF-mCherry transcripts are detected (a) and the BglF mCherry protein is not (b) (see also the results in fig. S18).



Figure 1. Visualization of Reporter mRNAs in Live Cells (A) Schematic describing the constructs used in this approach. The system is comprised of two components, a reporter mRNA and a GFP-MS2 fusion protein. The GFP-MS2 was expressed under the control of the constitutive GPD promoter, while the reporter mRNA was under the control of the GAL promoter. The reporter mRNA contains six binding sites for the coat protein of the bacterial phage MS2. To avoid possible interference with translation and the function of the 3'UTR, the MS2-binding sites were introduced immediately after a translation termination codon. The 3'UTRs were either from the ASH1 gene, to induce mRNA localization at the bud tip, or from the ADHII gene, as control. In addition, a nuclear localization signal (NLS) followed by an HA tag was introduced at the N terminus of the fusion protein, so that that only the GFP protein that is bound to its target mRNA would be present in the cytoplasm. (B) Live cells expressing the GFP-MS2 fusion protein and the lacZMS2-ASH1 reporter mRNA. Arrows indicate some of the particles, usually in the bud. Bar, 5 mm.















Tato prezentace je spolufinancována Evropským sociálním fondem a státním rozpočtem České republiky

