

MASARYKOVA UNIVERZITA

Design sekvence PCR primerů

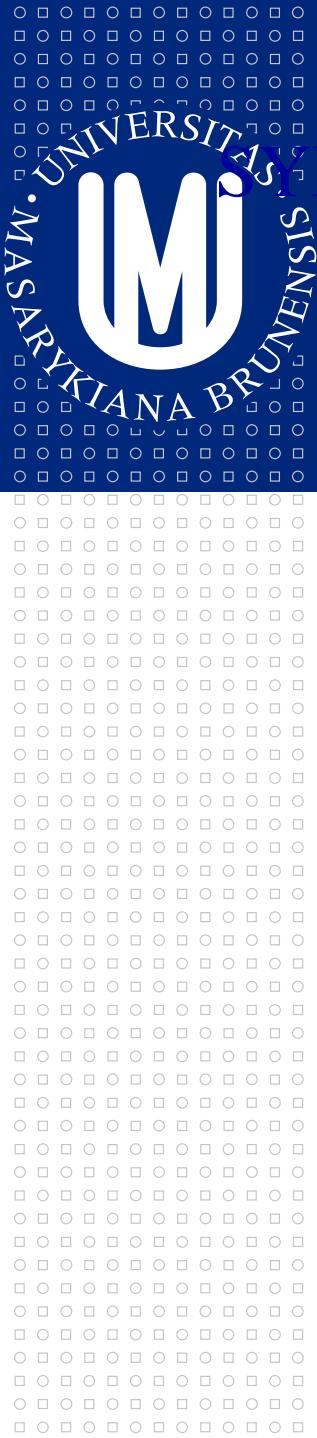
Hana Konečná

CEITEC - MU
Centrální laboratoř - Proteomika

Tento projekt je spolufinancován Evropským sociálním fondem a státním rozpočtem České republiky.



INVESTICE DO ROZVOJE VZDĚLÁVÁNÍ



SYNTETICKÉ OLIGONUKLEOTIDY

MASARYKOVA UNIVERZITA

- definice
- aplikace
- modifikace
- syntéza
- purifikace
- kontrola kvality

OLIGONUKLEOTIDY

- design sekvence
- zásady navrhování
- software OLIGO 7
- praktická ukázka

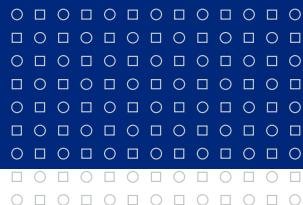
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MINISTERSTVO ŠKOLSTVÍ,
MLÁDEŽE A TĚLOVÝCHOVY

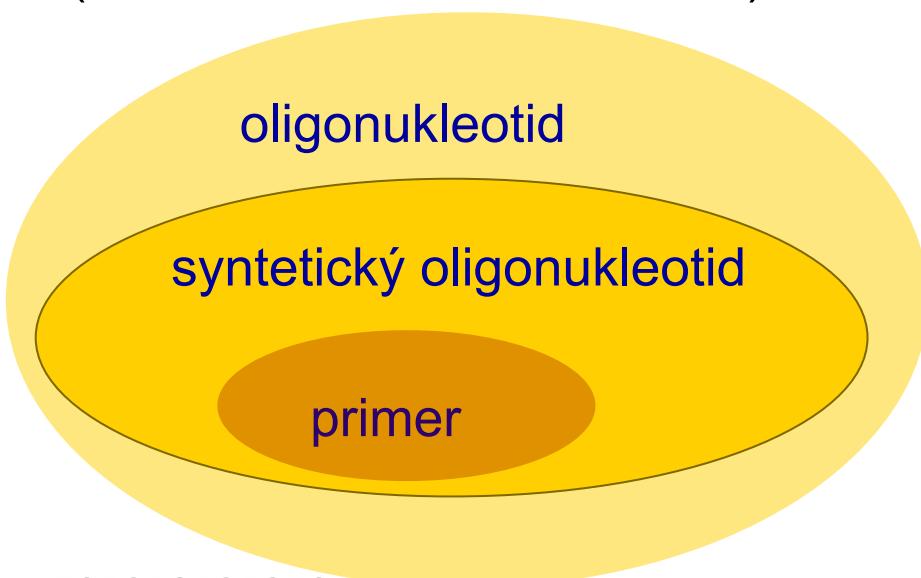


INVESTICE DO ROZVOJE VZDĚLÁVÁNÍ

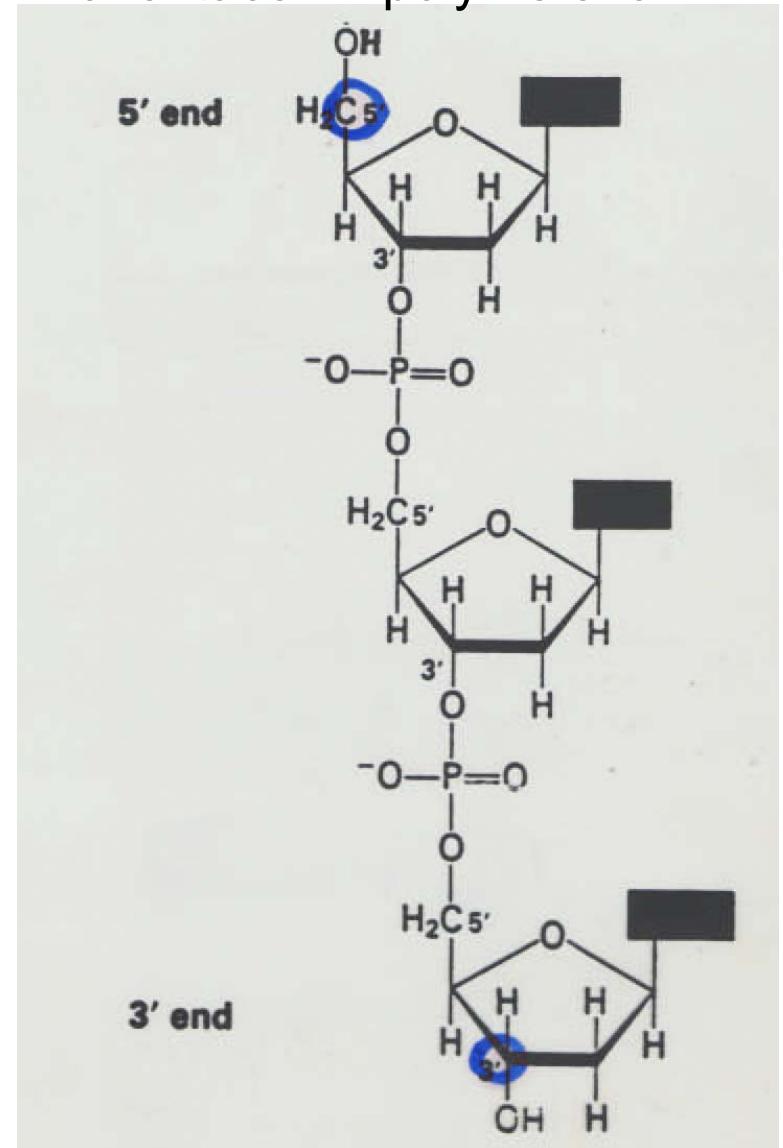


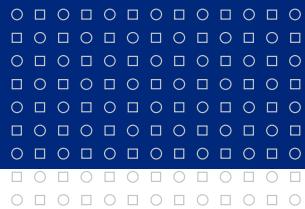
oligonukleotid

- krátká jednořetězcová struktura
- DNA nebo RNA (event. PNA)
- **hydroxyl** na obou koncích
(normálně na 5'- konci fosfát)



orientace! polymeráza!

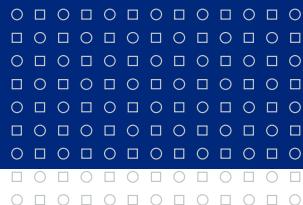




Aplikace syntetických oligonukleotidů

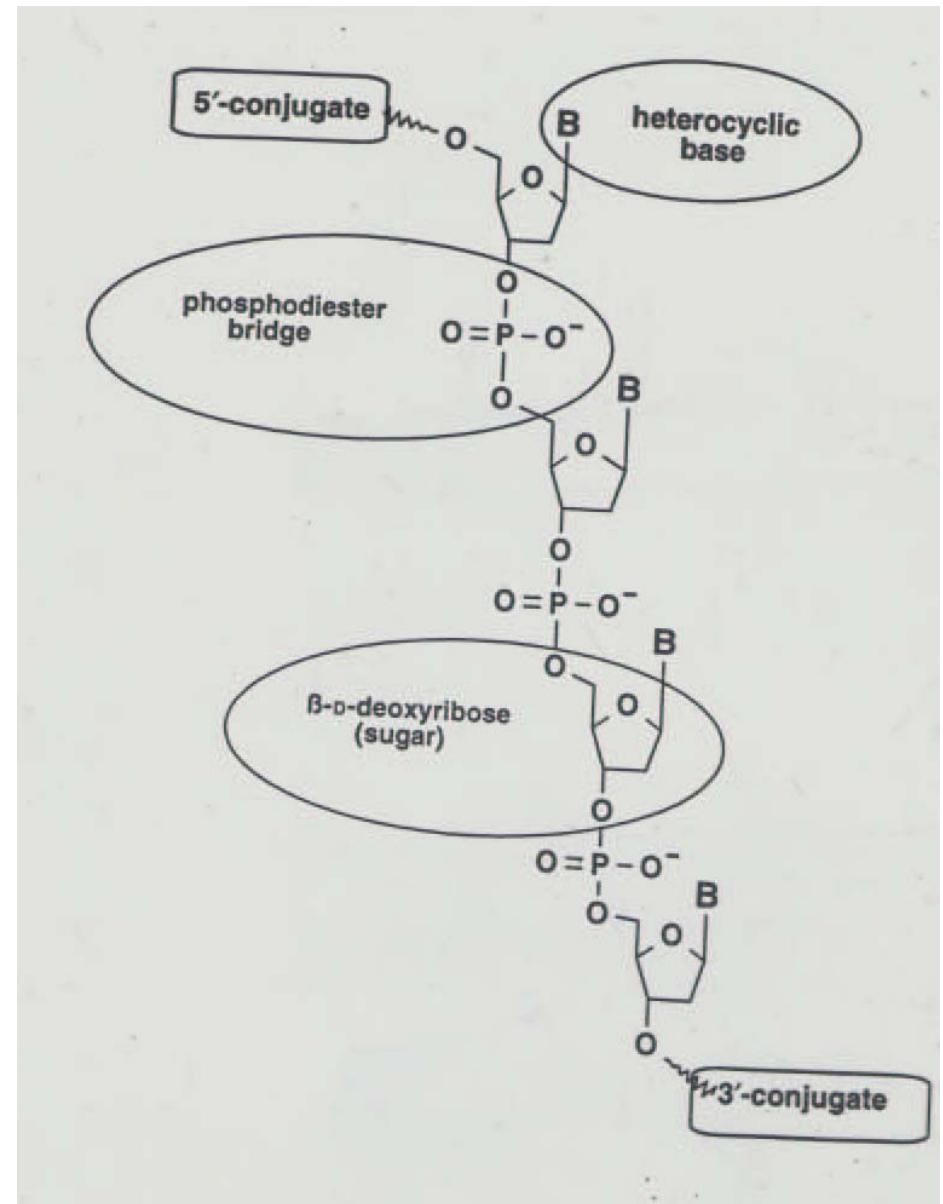
- primery pro syntézu komplementární DNA
PCR, Real-Time PCR
- syntéza genů a rekombinantní proteiny
- hybridizační sondy pro klonování
- místně cílená mutageneza
- sekvenování a genetické profilování
- diagnostika – testy a biosensory
- gene arrays
- blokace genové exprese *antisense oligo*
- potenciální léčiva a DNA vakcíny
- NMR studia interakcí DNA-protein
- strukturální rentgenová analýza NA

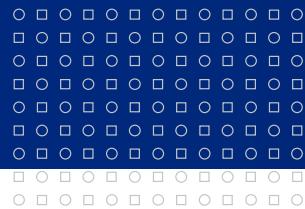




Modifikace

- degenerace
- konce řetězce
- báze
- fosfát
- cukr
- PNA





Degenerované oligonukleotidy

Příklady:

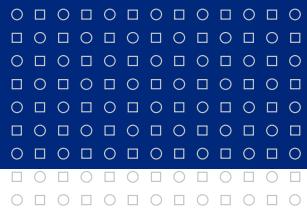
ACG TAC GTA CGT ACG TAC
nedegenerovaný

ACG TA**M** GTA CGT ACG TAC M = A/C

ACG TAC GTA **CDT** ACG TAC D = A/G/T

ACG TAC GTA CGT ACG **NAC** N = A/C/G/T

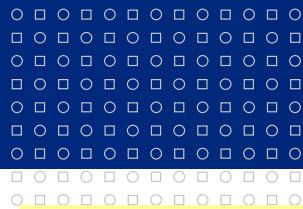




Degenerované oligonukleotidy

2-deoxyinosin

M	A or C
R	A or G
W	A or T
S	C or G
Y	C or T
K	G or T
V	A or C or G
H	A or C or T
D	A or G or T
B	C or G or T
N	G or A or T or C
X	G or A or T or C



Modifikace na 5'- konci

postsyntetické modifikace



5'

fosforylace

aminoskupina

thioskupina

digoxigenin

biotin

enzymy

psoralen

akridin

cholesterol

fluoresc. barviva

zhášedla

2,4-dinitrofenyl

TBR-chelát

spacer

větvení

blokáda

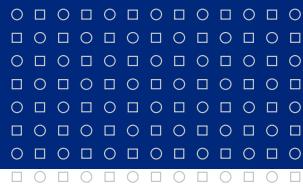
sekvenování



fragmentační analýza

gene arrays

Real-Time PCR



Modifikace na 3'- konci

derivatizovaná matrice



3'

fosfát

thioskupina

aminoskupina

spacer

akridin



biotin



fluoresc.barviva

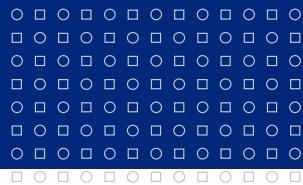


zhášedla

cholesterol

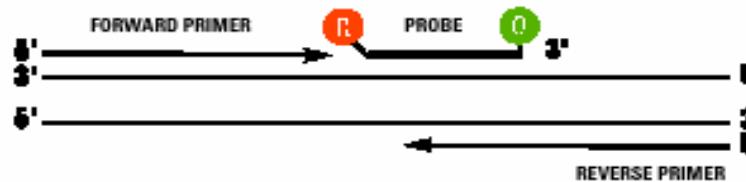
2,4-dinitrofenyl



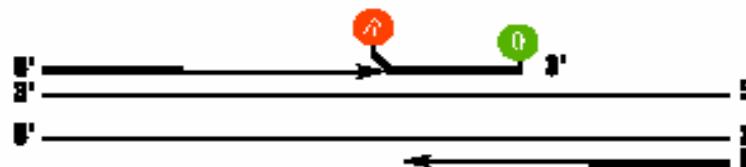


Real-Time PCR

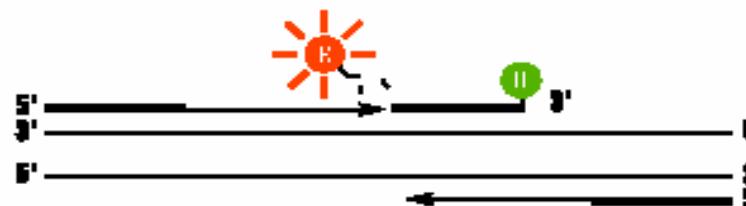
- 2x značená sonda
- REPORTER
- QUENCHER



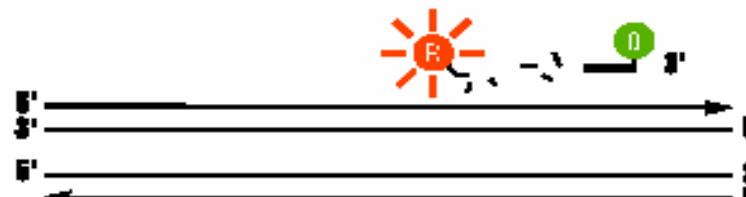
2. Strand displacement: When the probe is intact, the reporter dye emission is quenched.

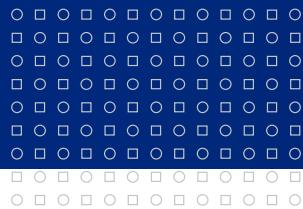


3. Cleavage: During each extension cycle, the DNA polymerase cleaves the reporter dye from the probe.



4. Polymerization completed: Once separated from the quencher, the reporter dye emits its characteristic fluorescence.





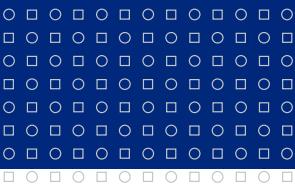
Další modifikace

fosforothioáty
fosforodithioáty
H-fosfonáty
methylfosfonáty

← páteř

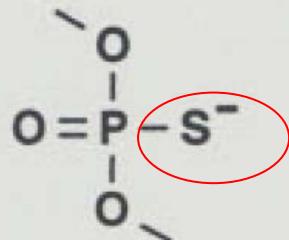
cukr →

modifikace v 2' pozici
modifikace ribózové jednotky

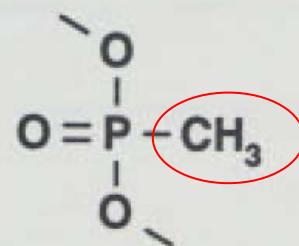


Terapeutika

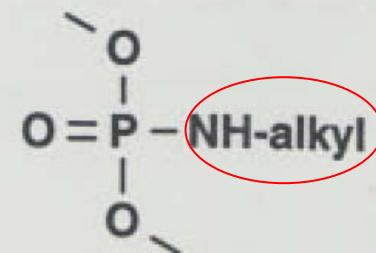
→ nedegradována nukleázami!
modifikace fosfodiesterové vazby



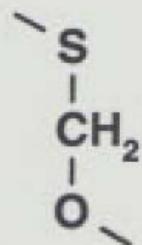
phosphorothioate



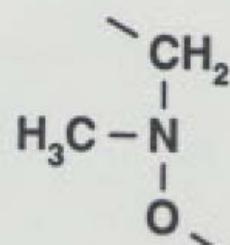
methylphosphonate



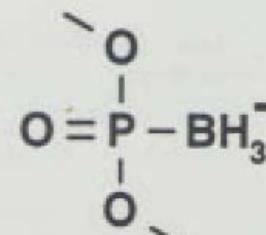
phosphoramidate



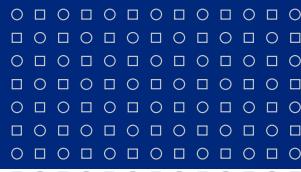
3'-thioformacetal



methylene(methylimino)

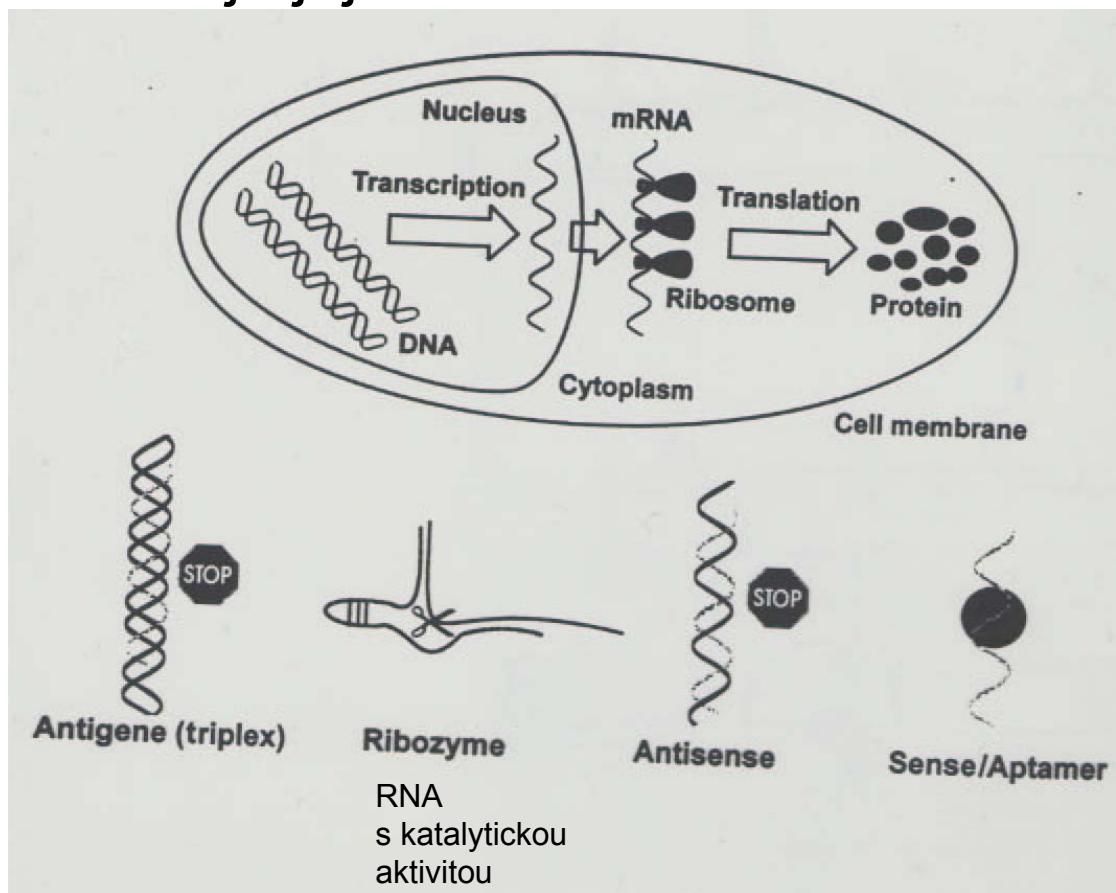


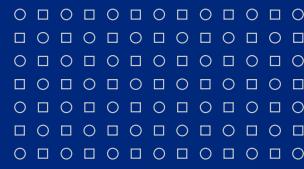
boranophosphate



ANTISENSE oligonukleotid

- oligonukleotid nebo analog
- komplementární k segmentu RNA nebo DNA
- vazbou inhibuje jejich normální funkci

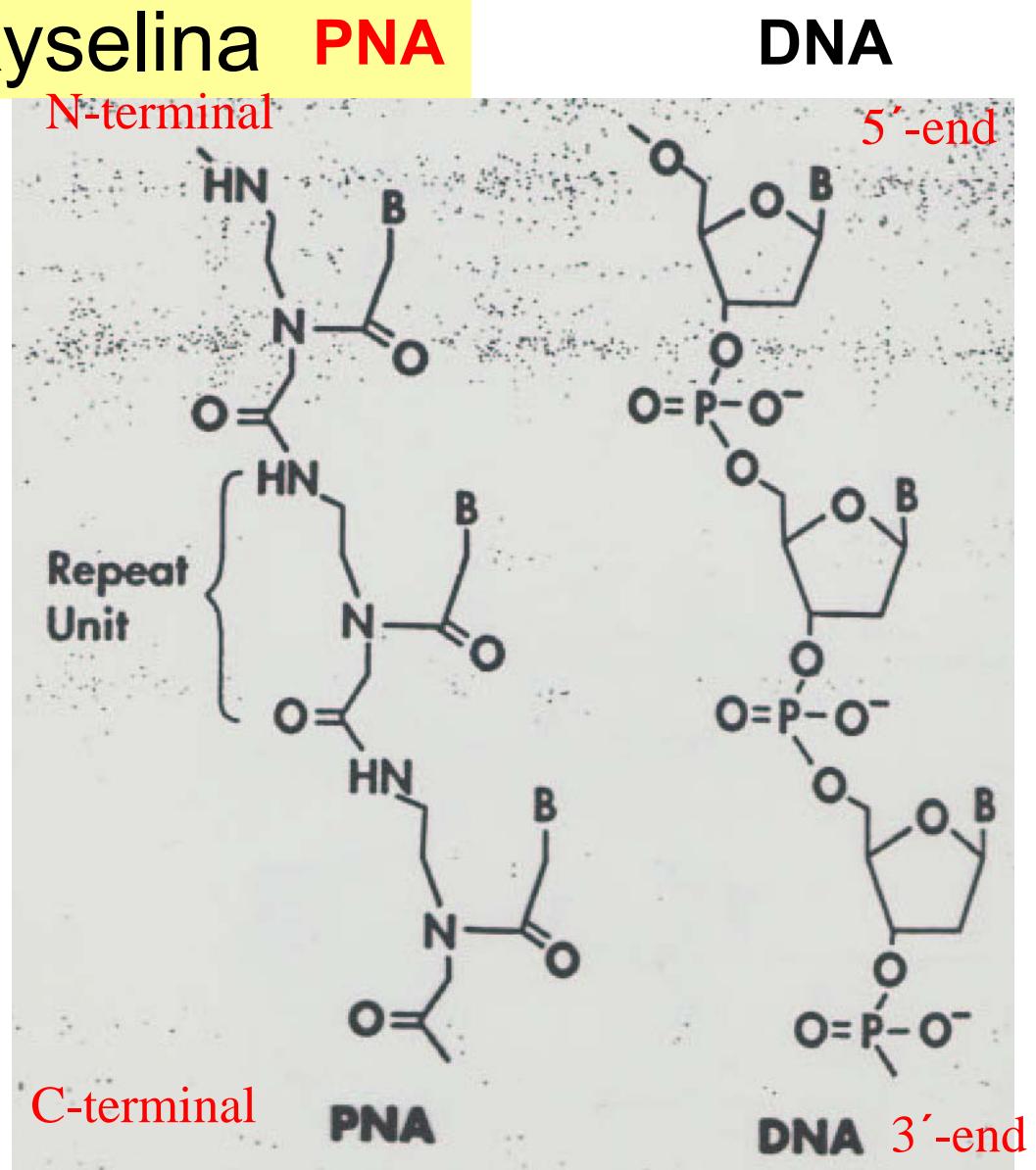


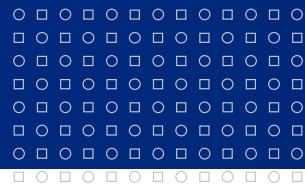


Peptidonukleová kyselina **PNA**

- nenabitá molekula
- vazba k DNA/RNA

N-(2-aminoethyl)-glycin →

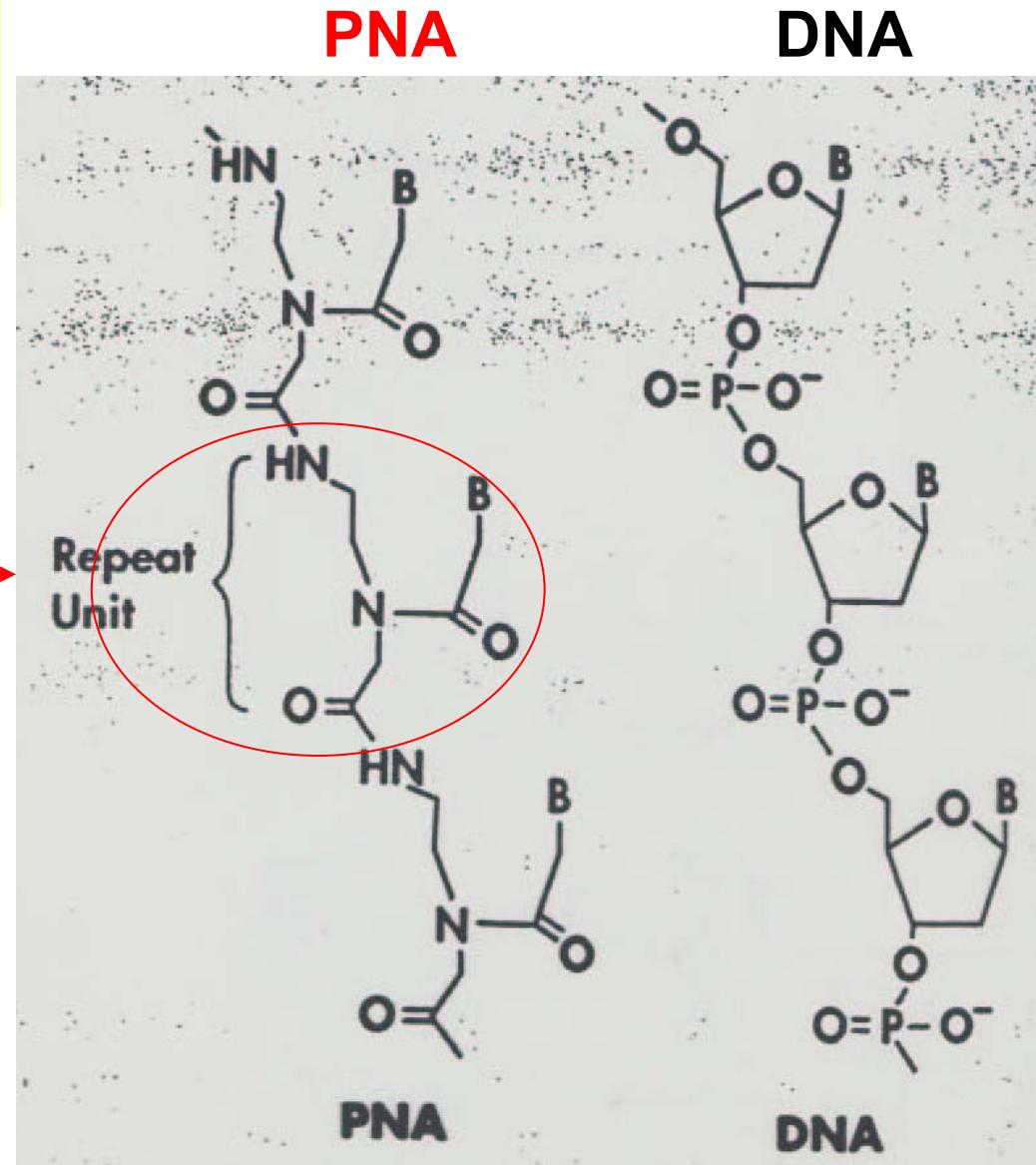


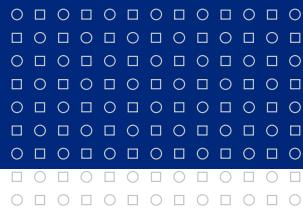


Peptidonukleová kyselina

- nenabitá molekula
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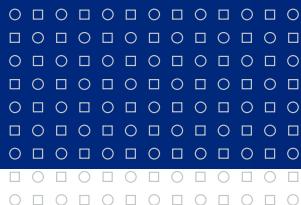




Vlastnosti PNA

- vysoká termostabilita
- T_m nezávisí na obsahu solí
- vyšší specificita
- vyšší afinita
- rezistentní k enzymům...

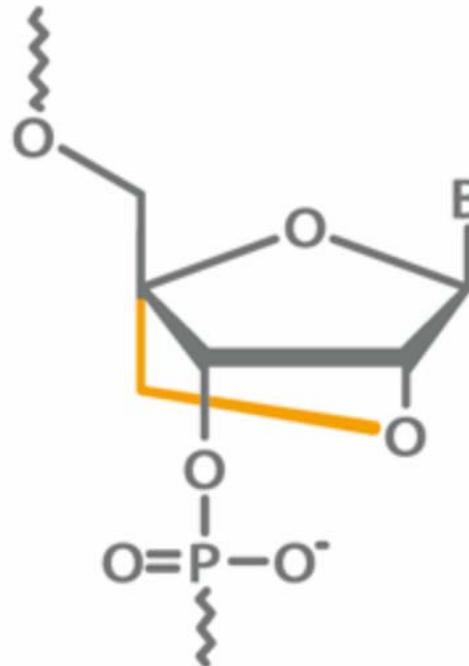


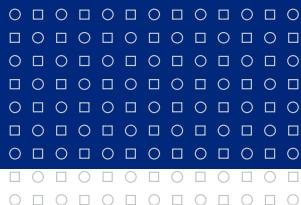


LNA

Locked Nucleic Acid

2'-O, 4'-C methylenový můstek
potlačená flexibilita ribofuranózového kruhu
struktura je **zamčena** do rigidní C3-endo konformace
zlepšená hybridizace
výjimečná biostabilita





OLIGONUKLEOTIDY

design

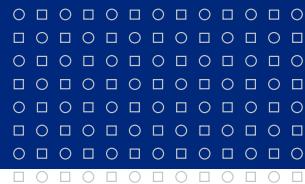
syntéza

purifikace



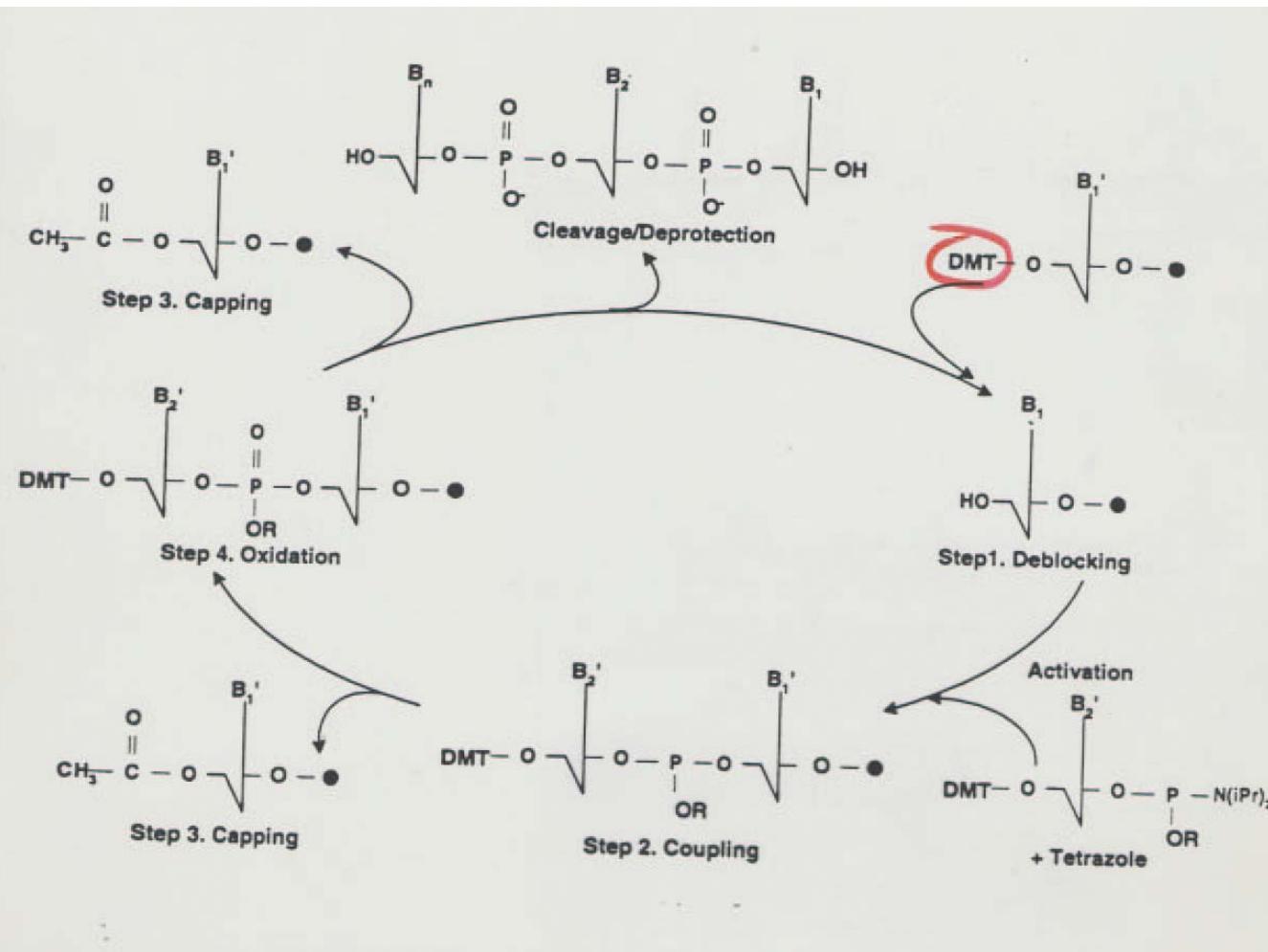
EXPEDITE 8909

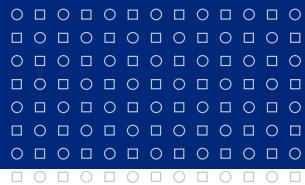




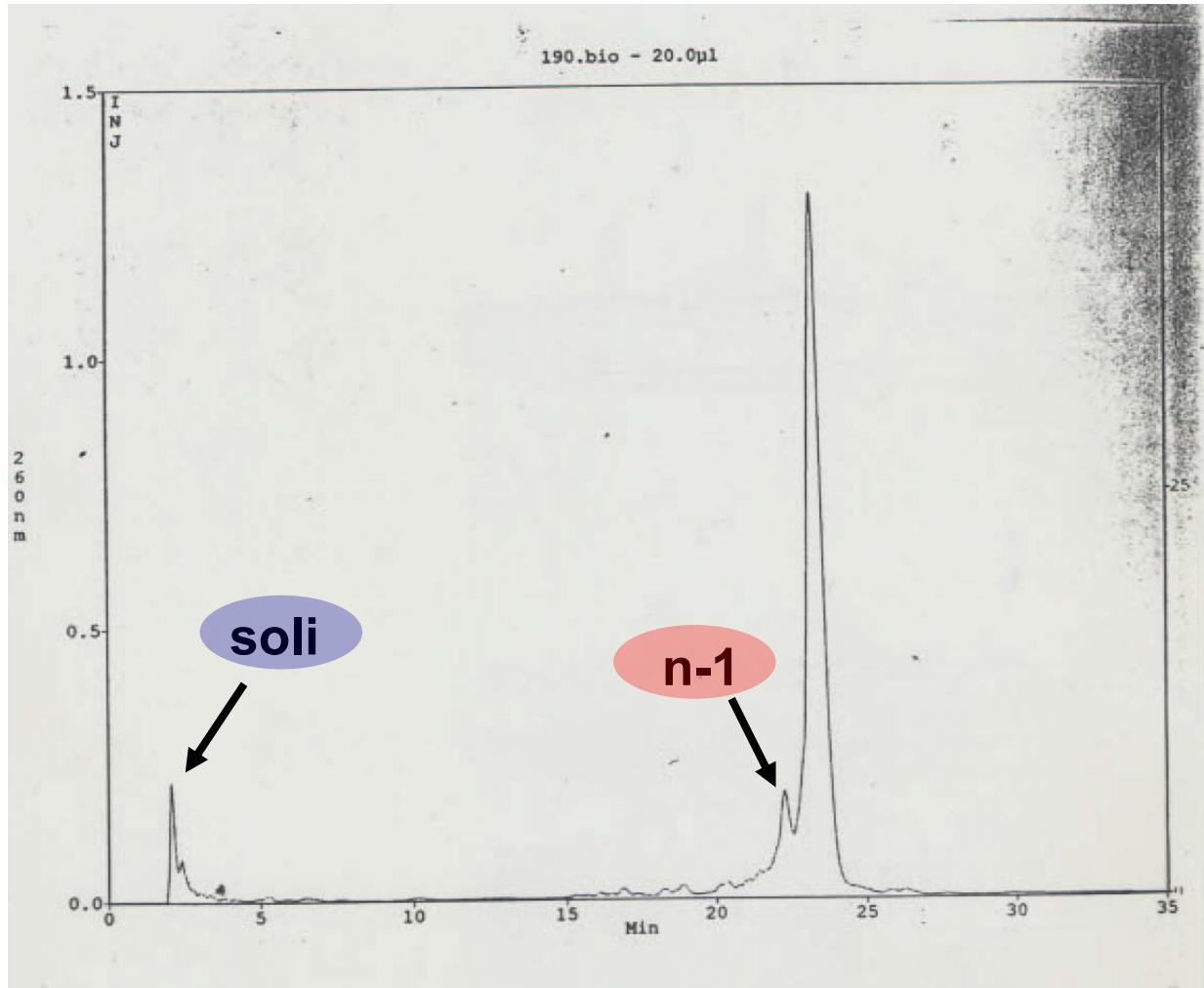
Syntéza oligonukleotidu

- syntéza na pevné fázi
- od 3'-konce k 5'-konci
- bezvodé prostředí



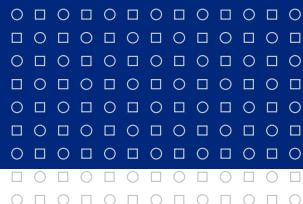


Kontrola kvality



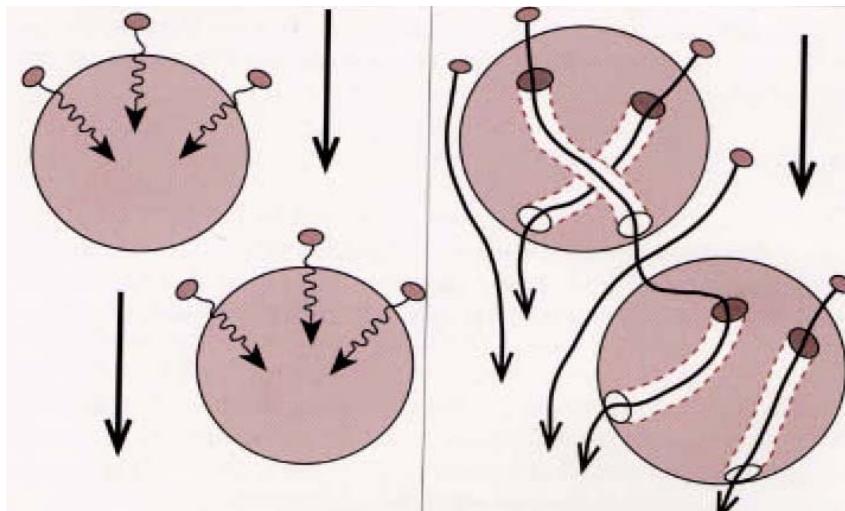
- HPLC
- Perfúzní chromatografie

- anex
- RP

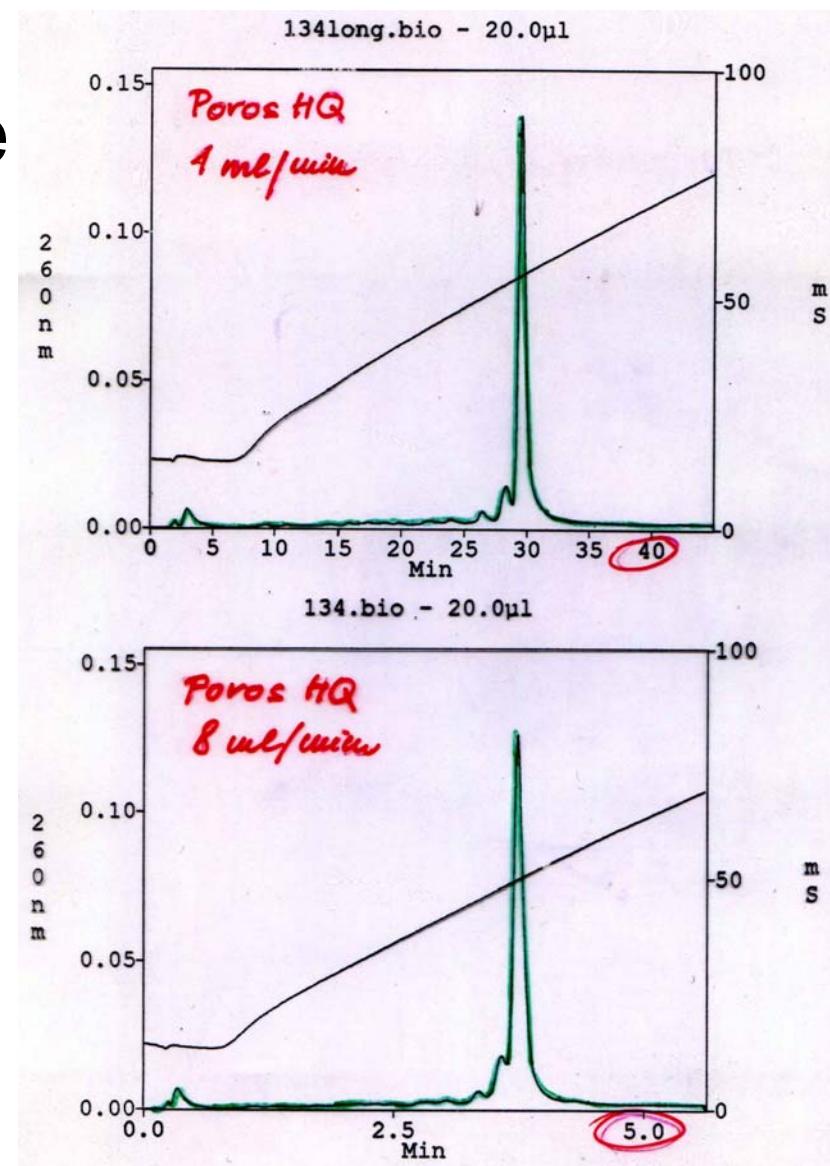


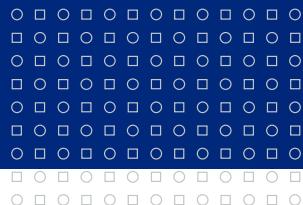
Perfúzní chromatografie

klasický sorbent



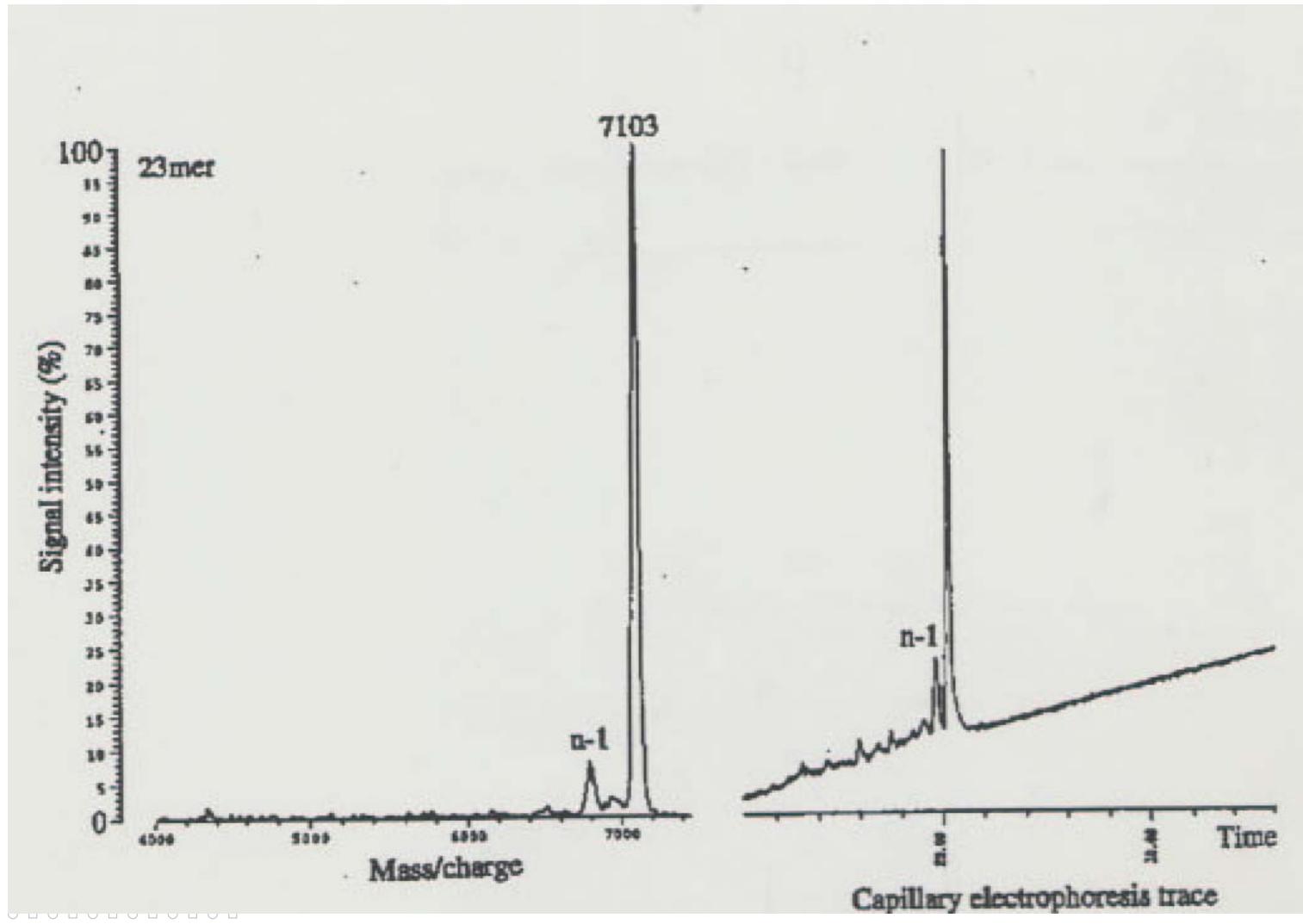
POROS

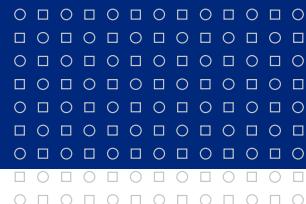




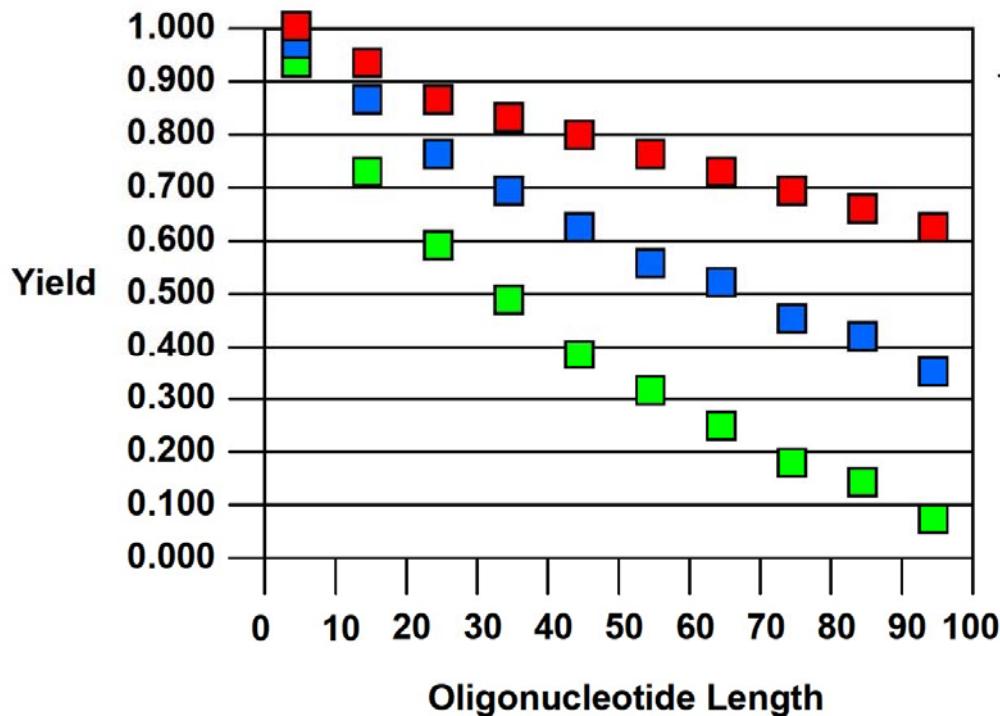
Maldi-TOF MS

CE





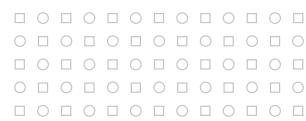
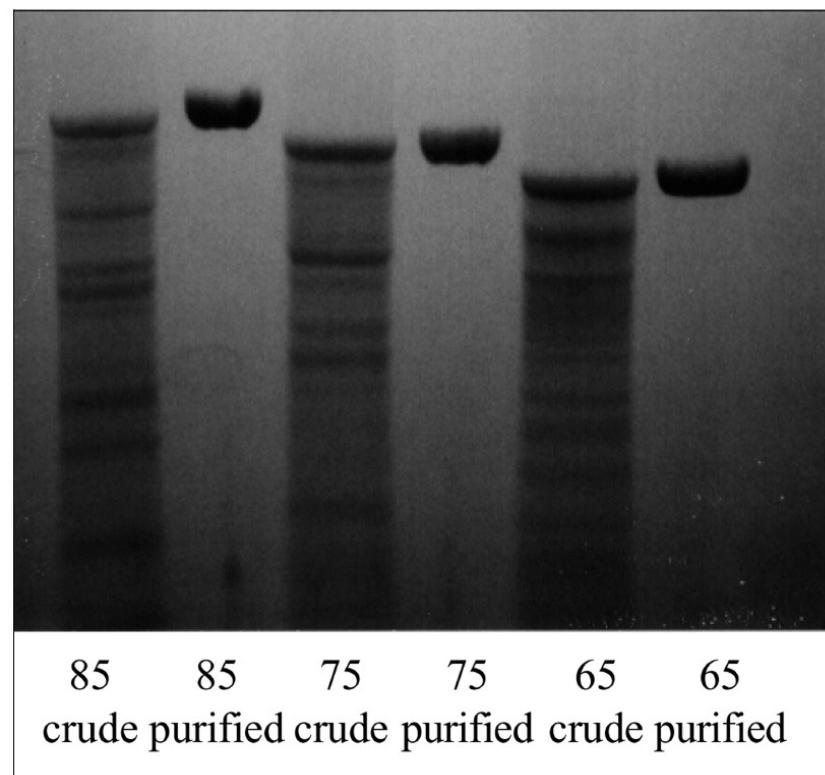
VÝTĚŽEK

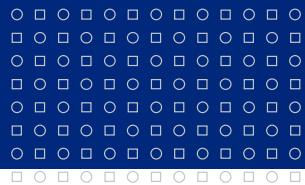


Efficiency

- 0.995
- 0.990
- 0.980

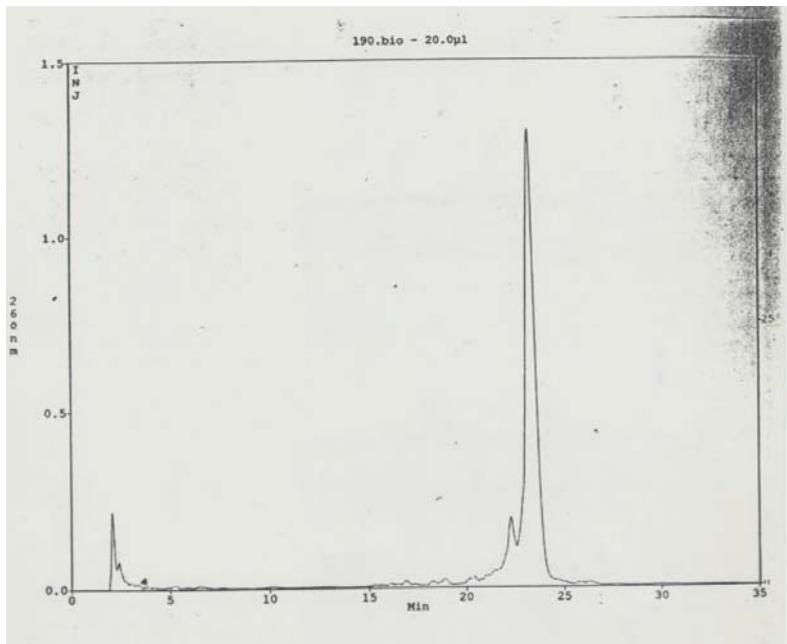
PAGE

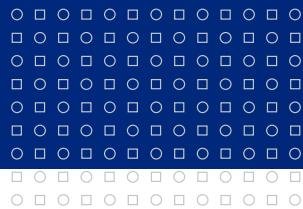




PURIFIKACE

- Sephadex
- RP cartridge
- HPLC





DESIGN OLIGONUKLEOTIDU

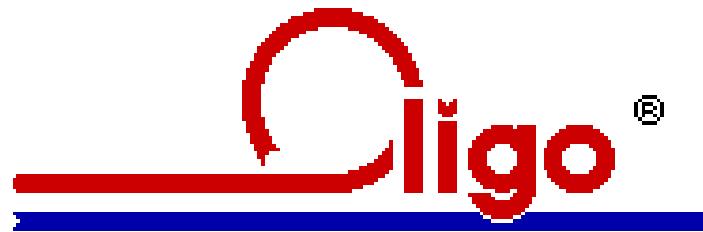
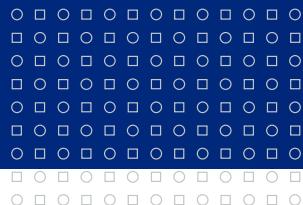
- manuální
- počítačový

www.protocol-online.org/prot/Research_Tools/Online_Tools/Oligo_Design/index.html

Hlavní kritéria pro sekvenci PCR primeru

- vysoce specifické
- netvoří dimery a vlásenky
- stabilní duplexy s aktivní sekvencí
- nepříliš stabilní 3'-konec





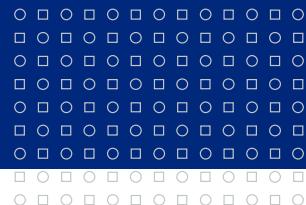
OLIGO 6

- PCR primery,
- hybridizační sondy
- sekvenační primery

OLIGO 7 (od roku 2008)

- TaqMan sondy
- primery pro *nested PCR*
- *molecular beacons*
- siRNA





Terminologie PCR primerů

forward primer... část sekvence + vlákna
reverse primer... část sekvence - vlákna



UPPER - FORWARD - LEFT

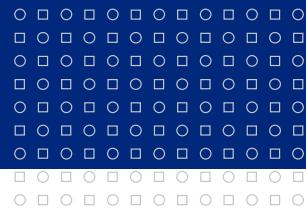
5' →

+ 5' → 3'

- 3' ← 5'

LOWER - REVERSE - RIGHT





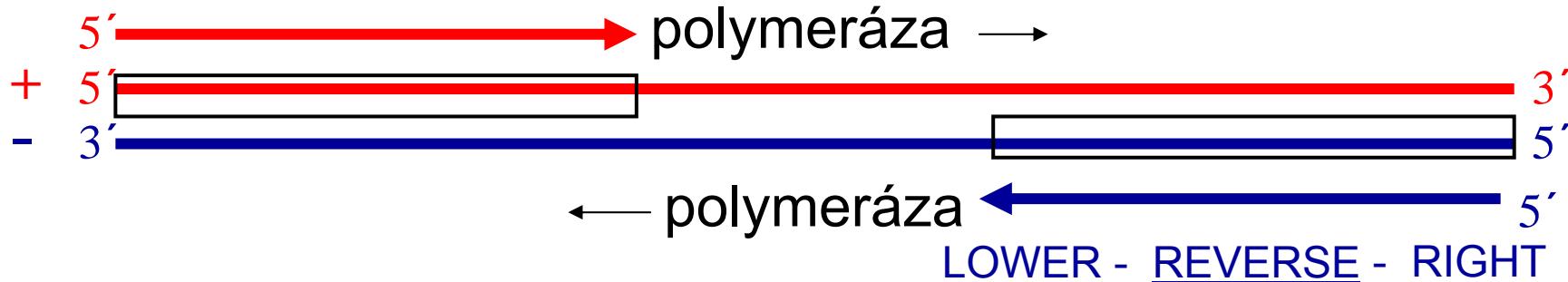
Terminologie

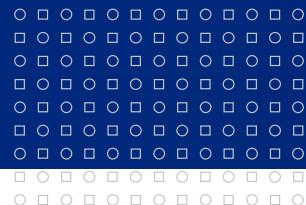
forward primer... část sekvence + vlákna

reverse primer... část sekvence - vlákna



UPPER - FORWARD - LEFT

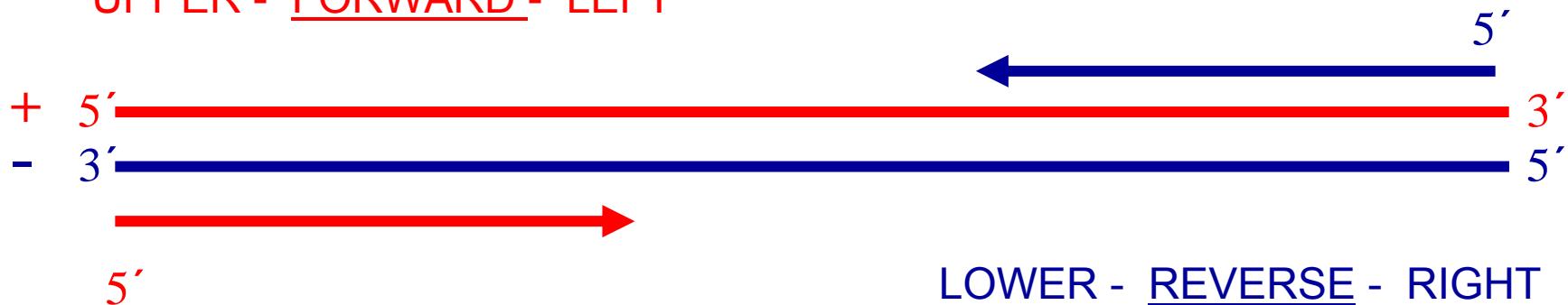


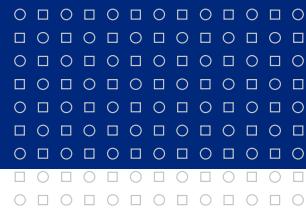


Nasedání PCR primerů



UPPER - FORWARD - LEFT





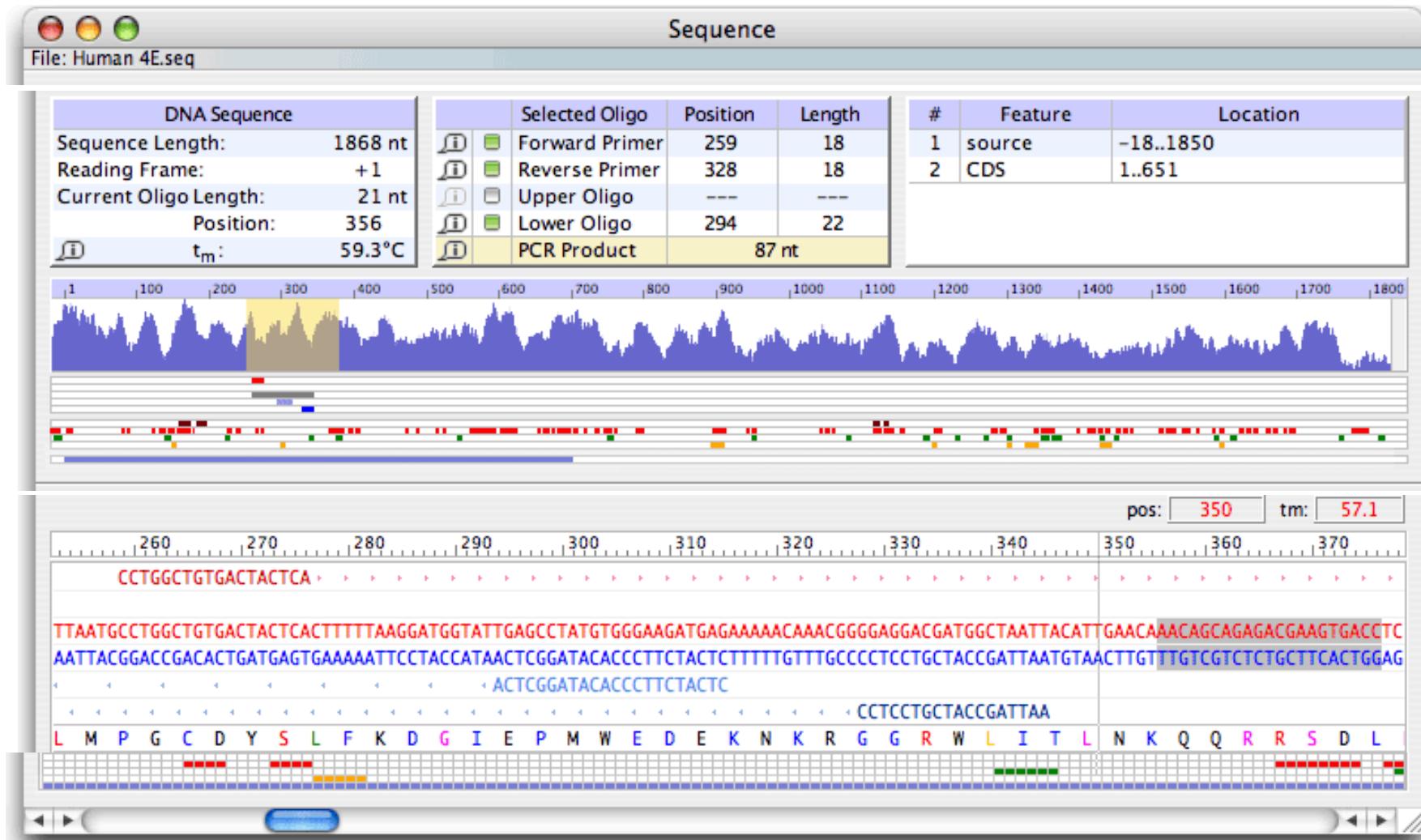
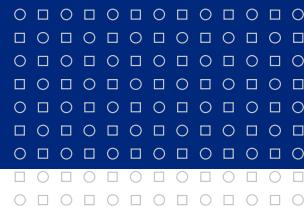
5' CTT CTG CTC AAT CTT TCT AC 3' FORWARD

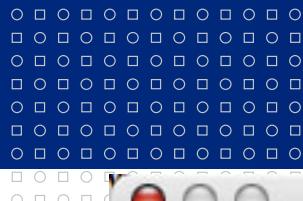
+ 5'

1 ATGGCTTCTG CTCAACTTT CTACAAACCAA AGCTCTGTCT TGAAAATCAA
51 TGTCACTGGTT GTGGACGATG ATCATGTTT CCTTGATATC ATGTCACGCA
101 TGCTTCAACA CTCCAAATAAC AGAGGTAATT AAATATTATT ATCATATTAT
151 ATATAATATG TTATTGATT TTTGTTTGTG ATTCATTAA GATTTTTATT
201 TCTATGATT CTTAGCATGA AATACAATT TTGGAGAAC AACTAGCAGT
251 TTTAAAAACAA AAACTTGAAT TTTGAGAAAT TCAAAAGATGT TATATATATA
301 TGTCAAAATT TAACAATTAT TCTTCTAAAT CATCCGGATT CCGTTTACAT
351 GTACACATCT ACAATTTCA ATTGAGGTAT TCTTGTGTTG ATGCCTTGA
401 GACGAATAGT TTGATTGATA AAAAAAATTCA TAACCAATAT GATATATAAA
451 GTTTATTTTC TTTTGTCAA ACCATACTTT ATACTATGTA ACTTTTTAA
501 GAGATTATTG AAAATAGTTT ATTTATAAAA TAGTAACCTA TTGTTGAATT
551 AAAAAAAAATTTGT AAATCGTGT TGCAAACGAC ATGTGATTAA
601 TCTTAGTTA AAACTAGCTG ATATTCTTCA AATCGACTGT TCTTATAAGT
651 AATCAACCAA TTAGCATCAA TCACAATAAA TTGTAAACAC TTCAATGAAA
701 ATGGTGATT TAAAGAATAT GTTTACTTA TGTTATGAAC TATCTCAAAT
751 TTGTGAAATA TTTCATAACT AATGTGGAAA ACTATATAAC CCCTCCATAC
801 AAAACGTAAG TAAAATTAT GAAATCCTAT CATTAAA GGTAAACCA
851 ATCAAAAAGT AATAATTCTT GGTACTTGCA ATATTTTGT CATTATATT
901 TAGTTTATTA ATTTTATTTT GATTAATGG TTTTAGATCC ATCAGTTATG
951 GAGATCGCAG TTATAGCTGT AGACGATCCG AAGAAAGCAT TATCTACTCT
1001 AAAAATTCAA CGAGACAATA TAGATCTCAT AATCACAGAT TATTATATGC
1051 CTGGTATGAA CGGTTTACAA CTAAAAAC AAATCACTCA GGAATTGGAA
1101 AATTACCAGG TCTTAGGTAA CATTGTTGT TCTTACAAC TTAAATTAAA

3'

5' TGA AGA ATA TCA GCT AGT TT 3' REVERSE





Search for Primers & Probes

Search Options Subsearches

Search in: + Strand - Strand

Search Mode: Select Verify

Complex Substrate

PCR Primers

Compatible with the Forward Primer Reverse Primer

TaqMan Probes & PCR Pairs

Compatible with the Upper Probe Lower Probe

Molecular Beacons & PCR Pairs

Nested Primers

Sequencing Primers

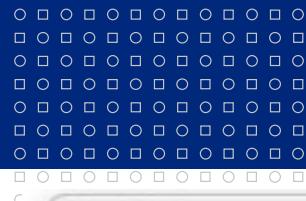
Hybridization Probes

siRNA Probes

After successfull search show: All Results

Search Cancel Apply

Parameters Ranges Defaults



Search for Primers & Probes

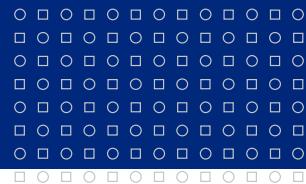
Search Options Subsearches

Search method: Compatible Pairs

Eliminate Ambiguous Bases
 Duplex-free Oligonucleotides
 Highly Specific Oligonucleotides (3'-end Stability)
 5'-end Stability
 siRNA Internal Stability
 Oligonucleotides with GC Clamp
 Oligonucleotides within Selected Tm Limits
 Hairpin-free Oligonucleotides
 Eliminate Mono- and Di-Nucleotide Repeats
 Detect Sequence Repeats
 Eliminate Frequent Oligonucleotides
 Omit High Secondary Structure Regions in the Template
 Check Primers/Probe Sequence Constraints
 Restrict the Number of G Bases
 Eliminate False Priming Oligonucleotides
and Continue Above Search in Other File(s)
 Consensus Primers

Search Cancel Apply

Parameters Ranges Defaults



PCR

File: Human 4E.seq

Optimal Annealing Temperature: 50.8 °C (Max: 66.3 °C)

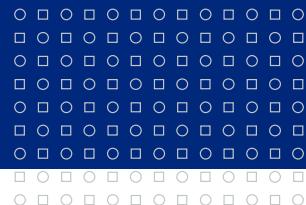
	Position and Length	T _m [°C]	GC [%]	P.E.#	Score
Product	862	78.9	29.6	n/a	697
Forward Primer	918 22	56.9	45.5	471 / 471	840
Reverse Primer	1753 27	55.3	29.6	489 / 489	834
Upper Oligo	979 24	56.5	33.3	479 / 479	917
Lower Oligo	1694 23	55.4	39.1	457 / 457	841

Product T_m – Reverse Primer T_m: 23.6 °C
Primers T_m difference: 1.6 °C

Comments:

	Concentration
Forward Primer	200.0 nM
Reverse Primer	200.0 nM
Upper Oligo	200.0 nM
Lower Oligo	200.0 nM
Monovalent Cation	50.0 mM
Free Mg[2+]	0.7 mM

Total Na[+] Equivalent: 155.8 mM



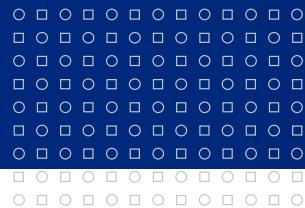
Selected Primers

File: BRCA2 gene.seq

AY436640:15438F22		AY436640:15917R20		
5' CAATATATACCGTAGTCCCCCTA 3'				
Length:	22-mer	Length:	20-mer	
Score:	802 points	Score:	914 points	
5' Position:	15438	3' Position:	15917	
T _m /t _m :	53.4	52.6 °C	53.1	53.8 °C
ΔG/Δg (25 °C):	-30.5	-29.2 kcal/mol	-28.6	-28.5 kcal/mol
ΔS/Δs:	-472.1	-449.5 cal/°K * mol	-430.5	-419.6 cal/°K * mol
ΔH/Δh:	-171.3	-163.2 kcal/mol	-157.0	-153.6 kcal/mol
3'ΔG:		-6.5 kcal/mol		-6.9 kcal/mol
Degeneracy:	1		1	
P.E.#:	443/443		477/477	
1/E:	4.63 nmol/A ₂₆₀		5.05 nmol/A ₂₆₀	
	31.1 µg/A ₂₆₀		31.0 µg/A ₂₆₀	

Priming Efficiency PE Score





- HAIRPIN intramolekulární
- DIMER intermolekulární



File: BRCA2 gene.seq

Current Oligo 21-mer [5042]

[Current+ Oligo] – The most stable 3'-dimer: # of hydrogen bonds = 10; $\Delta G = -0.7$ kcal/mol



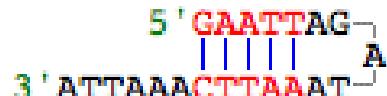
[Current- Oligo] – The most stable 3'-dimer: # of hydrogen bonds = 10; $\Delta G = -7.3$ kcal/mol; $T_m = 2.9^\circ\text{C}$

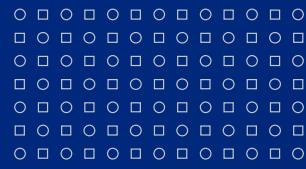


The most stable dimer overall: # of hydrogen bonds = 10; $\Delta G = -7.4$ kcal/mol; $T_m = 2.2^\circ\text{C}$



Hairpin: loop = 5 nt; $\Delta G = -3.0$ kcal/mol; $T_m = 54.6^\circ\text{C}$





Current Oligo Hairpin Stems

File: BRCA2 gene.seq

Current Oligo 21-mer [5042]

1. # of paired bases = 5; loop = 5 nt; $\Delta G = -3.0 \text{ kcal/mol}$; $T_m = 54.6 \text{ }^\circ\text{C}$

5042 GAATT 5046
| | | |
5056 CTTAA 5052



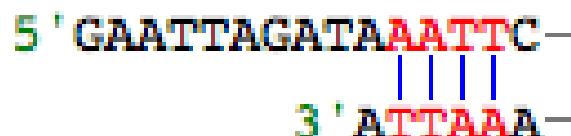
2. # of paired bases = 6; loop = 5 nt; $\Delta G = 0.2 \text{ kcal/mol}$; $T_m = 21.7 \text{ }^\circ\text{C}$

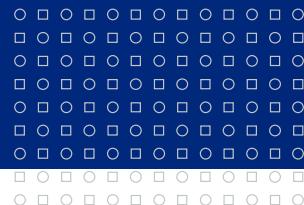
5043 AATTAGA 5049
| | | | |
5061 TTAAACT 5055



3. # of paired bases = 4; loop = 2 nt; $\Delta G = 0.9 \text{ kcal/mol}$; $T_m = 8.7 \text{ }^\circ\text{C}$

5052 AATT 5055
| | | |
5061 TCAA 5058





Reverse Primer False Priming Sites

File: M13MP18

Reverse Primer M13MP18:6310R19 (positive strand)

Priming efficiency of the perfect match is 482 (above the threshold)

Priming efficiency: 482 (above the threshold)

5' (6328) GGTTTTCCCAGTCACGACG (6310)3'

3' (6328) ccaaaaagggtcaagtgctgc (6310)5'

Priming efficiency: 244 (above the threshold)

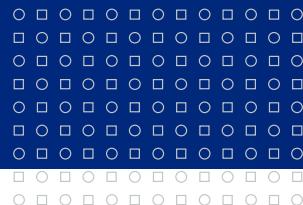
5' (6328) GGTTTTCCCAGTCACGACG (6310)3'

3' (626) agcaaatggtc--tgctgc (610)5'

Priming efficiency: 193 (above the threshold)

5' (6328) GGTTTTCCCAGTCACGACG (6310)3'

3' (5125) tctaagggtcaagtg-tgc (5108)5'

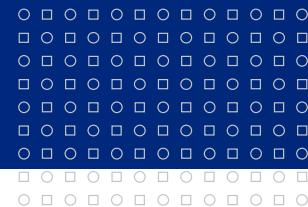


 Forward Primer Composition
File: BRCA2 gene.seq
Forward Primer AY436640:6275F19

T _d	64.2°	[nearest neighbor method]
T _m	56.5°	[nearest neighbor method]
T _m	70.8°	[%GC method]
T _m	56°	[2(A+T)° + 4(G+C)° method]
T _m (RNA)[1M Na]	81°	[%GC method]
T _m (DNA:RNA)[1M Na]	74.7°	[%GC method]
A ₂₆₀ /A ₂₈₀	1.59	[single strand]
Molecular Weight	5.8K	[one strand]
Molecular Weight	11.7K	[two strands]
µg/OD	47.4	[dsDNA]

Base	Number & %
A	2 [10.5%]
C	5 [26.3%]
G	4 [21.1%]
T	8 [42.1%]
A + T	10 [52.6%]
G + C	9 [47.4%]





Oligonucleotide Database
File: NewDatabase.odb # of Records: 29

#	Date	ID Number	Sequence	3'-Dim. ΔG	P.E. / p.e.	Tm / t _m	
1							
21	12/02/06	AY436640:5916R19	AATGCCCTGCCTTAGTCTG	-	SC 430	430 54.1	54.5
22	12/02/06	AY436640:5916R20	CAATGCCCTGCCTCTAGTCTG	0.3	SC 366	450 50.9	57.2
23	12/02/06	AY436640:5937R21	TCAATTCTTTAGCTTGGCAT	0.3	SC 449	449 54.7	53.1
<input checked="" type="checkbox"/> 24	12/02/06	AY436640:5937R22	TTCAATTCTTTAGCTTGGCAT	0.3	SC 458	458 55.9	53.8
25	12/02/06	AY436640:4695U22	TGCCTTAACAAAAGTAATCCAT	0.3	SC 432	432 54.5	53.0
26	12/02/06	AY436640:5325U22	AATTACGTCTTCCTTATGCCAA	0.3	SC 453	453 53.3	53.0
27	12/02/06	AY436640:5786L23	CTCTGCCCTAGAACATTATCACTC	-0.3	SC 451	451 54.8	55.0
28	12/02/06	AY436640:5860L19	AACAACCAAAGCCAACCTG	-0.9	SC 444	444 55.3	55.9

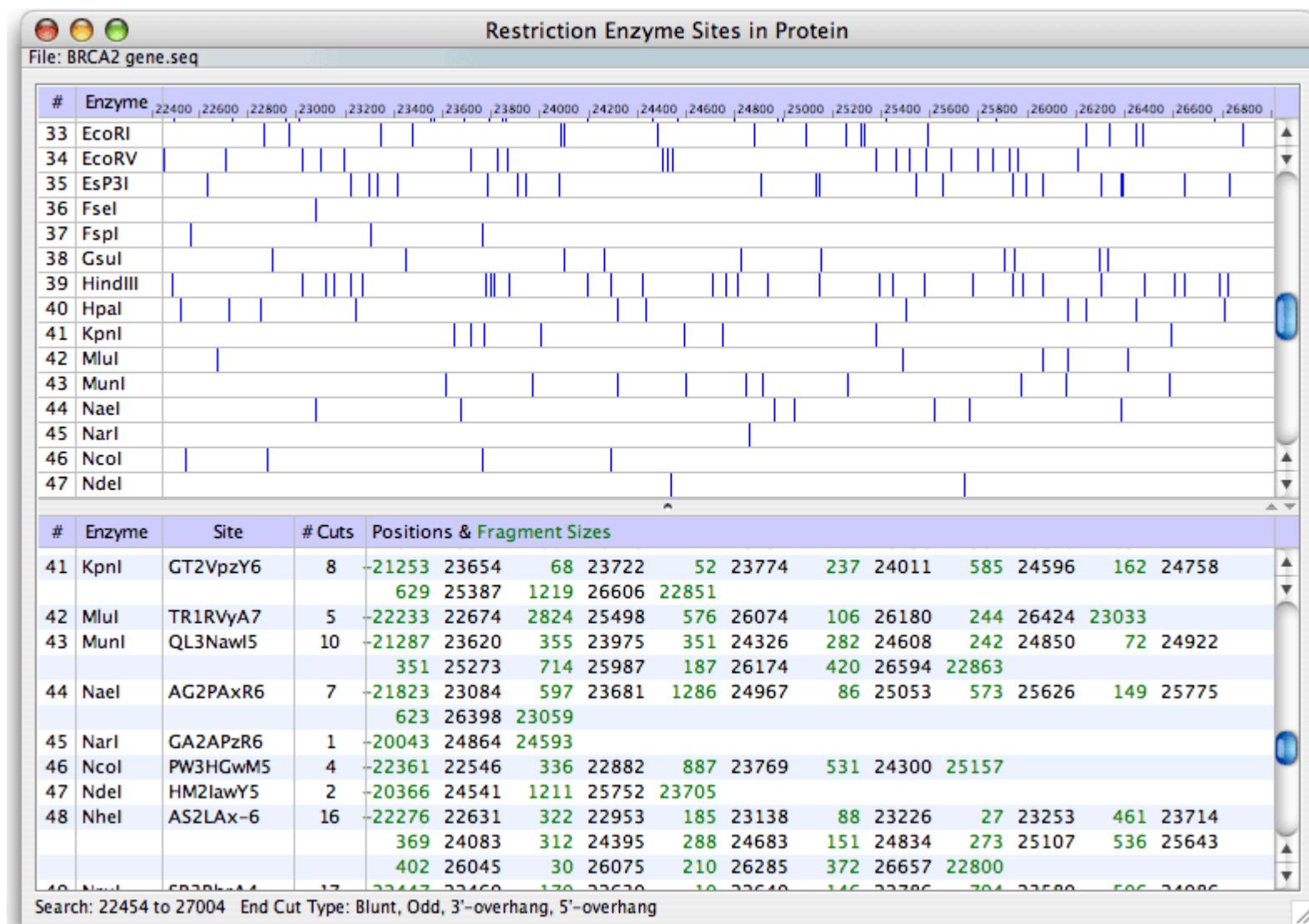
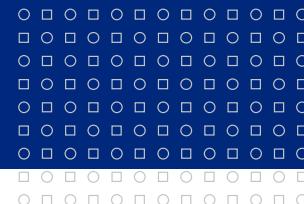
Selected oligo

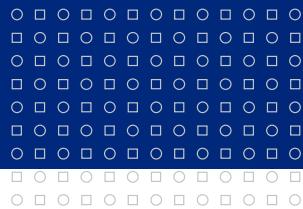
Oligonucleotide Sets (64)

#	Forward Primer	Reverse Primer	Upper Oligo	Lower Oligo
	1	2	3	4
36	8	23	25	28
42	8	24	25	28
<input checked="" type="checkbox"/> 47	9	14	25	27
39	9	15	25	27
33	9	16	25	27
61	9	17	25	27
48	9	18	25	27

Checked Set of nested primers

This database is linked to BRCA2 gene.seq





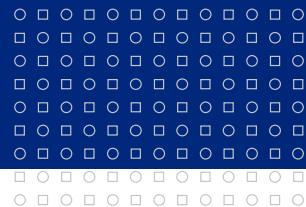
 Hybridization Time
File: M13MP18

DNA Length: nt.

Concentration: nM
 µg/mL

 $T_{\frac{1}{2}} = 45.4 \text{ sec}$
 $T = 3 \text{ min } 47 \text{ sec}$





Concentrations

File: BRCA2 gene.seq



Constant Concentration

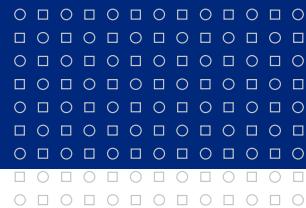


Constant Volume

- Current +Oligo: 5.08 nmol/OD, 32.5 µg/OD
- Current -Oligo: 4.67 nmol/OD, 30.9 µg/OD
- Entire Sequence (ds): 0.001 nmol/OD, 48.1 µg/OD
- Forward Primer: 5.98 nmol/OD, 35.0 µg/OD
- Reverse Primer: 5.31 nmol/OD, 34.0 µg/OD
- PCR Product (ds): 0.146 nmol/OD, 48.1 µg/OD
- Upper Oligo: 4.83 nmol/OD, 31.2 µg/OD
- Lower Oligo: 4.67 nmol/OD, 30.9 µg/OD

32.5 µg
or 1.0 OD(260)
or 5.084 nmol
in 508.4 µL
yields 10.0 µM

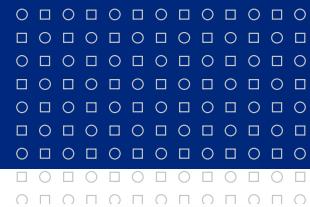


**AHP2 cDNA (TAIR database)**

Sequence: AT3G29350.1 Date last modified 2007-04-17 Name AT3G29350.1 Tair Accession Sequence:4010737427 Sequence Length (bp) 827

1 ACAATTAGCG AGAAAGACAA AACACAAGTT TCTTCTTCTT GGGATTGGCT
51 ATTTCCAGAA ATCCAAGTCA ATAATCAAAG TCCAAACAAA AAAATCCTCT
101 CCCAATCTCC GCTTCACTCT TCTCATGGAC GCTCTCATTG CTCAGCTTCA
151 GAGACAATT CGTGATTACA CCATTCTCT CTACCAACAG GGGTTTTGG
201 ATGATCAATT TACTGAGTTG AAAAAAGCTAC AAGATGATGG AAGTCCTGAT
251 TTTGTGTCTG AAGTGCTTTC ACTTTCTTT GAAGATTGTG TGAAGCTTAT
301 CAGTAACATG GCTAGAGCTT TGGACACGAC AGGAACGTGTA GATTTAGTC
351 AGGTAGGTGC TAGTGTGCAT CAATTGAAGG GTAGTAGCTC AAGTGTGGT
401 GCCAAGAGGG TCAAAACTTT GTGTGTTAGC TTCAAGGAAT GTTGTGAAGC
451 TAAGAACTAC GAAGGGTGTG TGAGATGTT GCAGCAAGTG GATATTGAGT
501 ACAAGGCGTT AAAGACAAAG CTTCAAGATA TGTTCAATCT TGAGAAACAG
551 ATCATTCAAG CTGGTGGTAT AGTTCTCAA GTGGATATTA ACTAAAGAGA
601 CTAGTCCATA AGAAGAAAAA AGATGATGAC TTTCTTCTT TAGTTCTCT
651 TCTAAATTAT TTTGGATTTG GTGTTGCTC AAAAACTCAA TAAAATATGT
701 GCAAAAAGAA ACAAAAACAA GTGATGGTTG TTTATAAAATC AGTAGTATGT
751 ATTGTTGAT CTCATCCGAG AAAATTGAAA CCATTGGACT AATGAATGTG
801 ATGATAATAT ATATTGGTT GCTTCTG





101 CCCAATCTCC GCTTCACTCT TCTC**ATGGAC** GCTCTCATTG CTCAGCTTCA
151 GAGACAATT CGTGATTACA CCATTTCTCT CTACCAACAG GGGTTTTGG
201 ATGATCAATT TACTGAGTTG AAAAAGCTAC AAGATGATGG AAGTCCTGAT
251 TTTGTGTCTG AAGTGCTTTC ACTTTCTTT GAAGATTGTG TGAAGCTTAT
301 CAGTAACATG GCTAGAGCTT TGGACACGAC AGGAACGTGTA GATTTAGTC
351 AGGTAGGTGC TAGTGTGCAT CAATTGAAGG GTAGTAGCTC AAGTGTGGT
401 GCCAAGAGGG TCAAAACTTT GTGTGTTAGC TTCAAGGAAT GTTGTGAAGC
451 TAAGAACTAC GAAGGGTGTG TGAGATGTTT GCAGCAAGTG GATATTGAGT
501 ACAAGGCGTT AAAGACAAAG CTTCAAGATA TGTCAATCT TGAGAAACAG
551 ATCATTCAAG CTGGTGGTAT AGTTCCCTCAA GTGGATATTA ACTAAAGAGA

EcoRI restriction site

5'.....G|AATTC.....3'
3'.....CTTAA|G.....5'

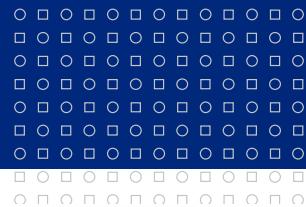
Design of primers

AHP2ex_up

5'- CCG **GAA TTC** ATG GAC GCT CTC ATT GCT CAG – 3'

AHP2ex_low

5'- CCG **GAA TTC** TTA GTT AAT ATC CAC TTG AGG – 3'



101 CCCAATCTCC GCTTCACTCT TCTC **ATGGAC GCTCTCATTG CTCAGCTTCA**
151 GAGACAATT CGTGATTACA CCATTTCTCT CTACCAACAG GGGTTTTGG
201 ATGATCAATT TACTGAGTTG AAAAAAGCTAC AAGATGATGG AAGTCCTGAT
251 TTTGTGTCTG AAGTGCTTTC ACTTTCTTT GAAGATTGTG TGAAGCTTAT
301 CAGTAACATG GCTAGAGCTT TGGACACGAC AGGAACGTGTA GATTTAGTC
351 AGGTAGGTGC TAGTGTGCAT CAATTGAAGG GTAGTAGCTC AAGTGTGGT
401 GCCAAGAGGG TCAAAACTTT GTGTGTTAGC TTCAAGGAAT GTTGTGAAGC
451 TAAGAACTAC GAAGGGTGTG TGAGATGTTT GCAGCAAGTG GATATTGAGT
501 ACAAGGCGTT AAAGACAAAG CTTCAAGATA TGTCAATCT TGAGAAACAG
551 ATCATTCAAG CTGGTGGTAT AGTTCCTCAA GTGGATATTA ACTAAAGAGA

EcoRI restriction site

5'.....G|AATTC.....3'
3'.....CTTAA|G.....5'

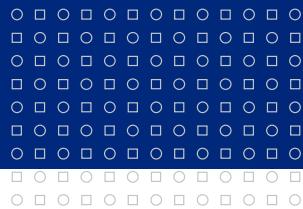
Design of primers

AHP2ex_up

5'- CCG **GAA TTC** ATG GAC GCT CTC ATT GCT CAG – 3'

AHP2ex_low

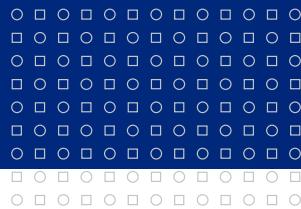
5'- CCG **GAA TTC** TTA GTT AAT ATC CAC TTG AGG – 3'



LITERATURA

- Artificial DNA: Methods and Applications; Khudyakov, Y.E., Fields, W.A., Ed. (2003)
- PCR Primer: A Laboratory Manual (2003)
- OLIGO Primer analysis software, Version 7





Discovery is not in seeking new landscapes,
but in having new eyes...

Marcel Proust

Tato prezentace vznikla s podporou projektu **OP VK „Rozvoj týmu pro výuku, výzkum a aplikace v oblasti funkční genomiky a proteomiky“**(CZ.1.07/2.3.00/09.0132)

Tento projekt je spolufinancován Evropským sociálním fondem a státním rozpočtem České republiky.



INVESTICE DO ROZVOJE VZDĚLÁVÁNÍ

