

# Moderní analytická instrumentace pro genetický výzkum, lékařskou diagnostiku a molekulární identifikaci organismů

Karel Klepárník

*Oddělení bioanalytické instrumentace  
Ústav analytické chemie  
Akademie věd České republiky  
Brno*

**iac**  
**brno**



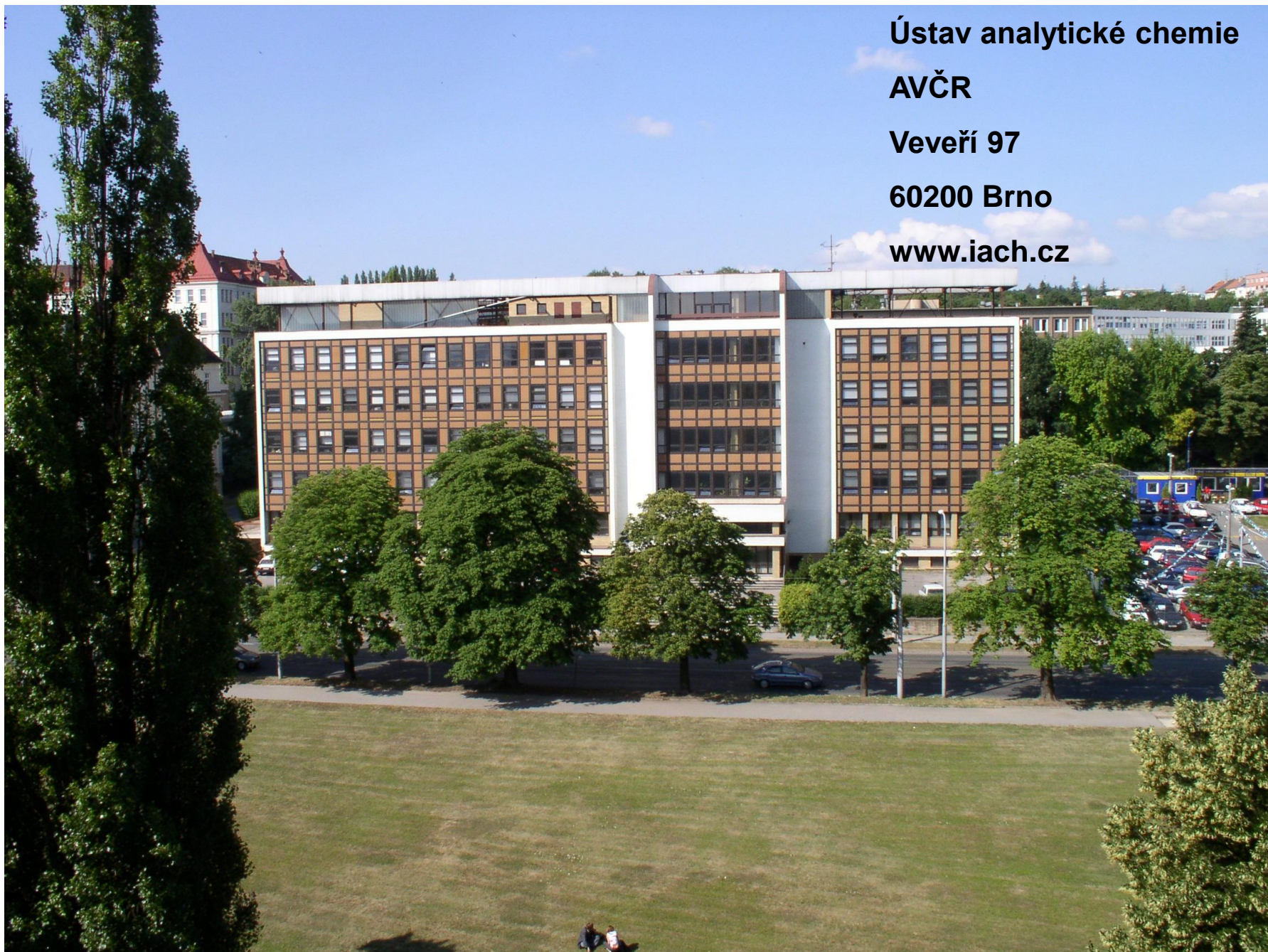
**Ústav analytické chemie**

**AVČR**

**Veveří 97**

**60200 Brno**

**[www.iach.cz](http://www.iach.cz)**



**Polymerase chain reaction**

**PCR amplification**



## **Kary B. Mullis**

born 1944

La Jolla, CA, USA

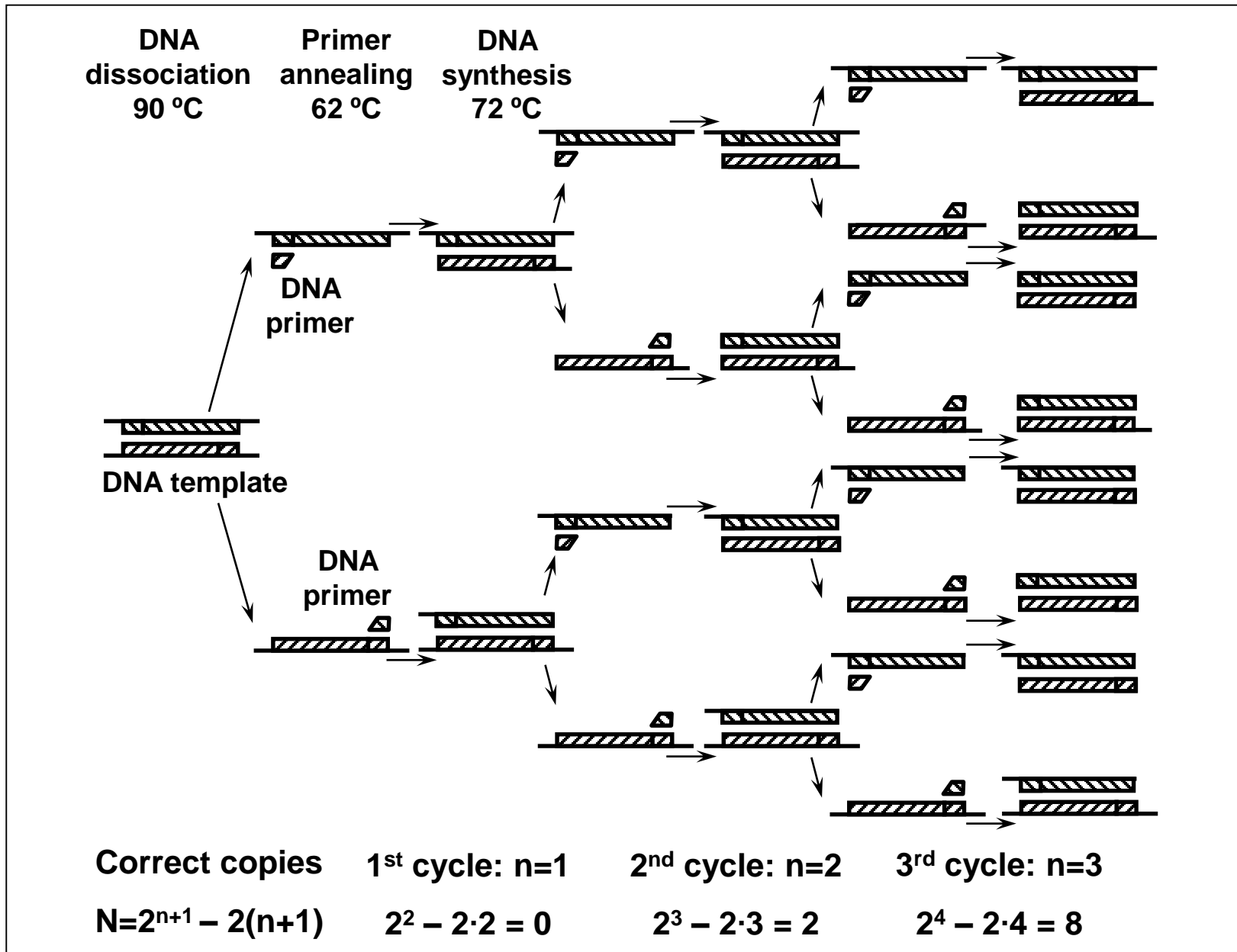
University of British Columbia



### **The Nobel Prize in Chemistry 1993**

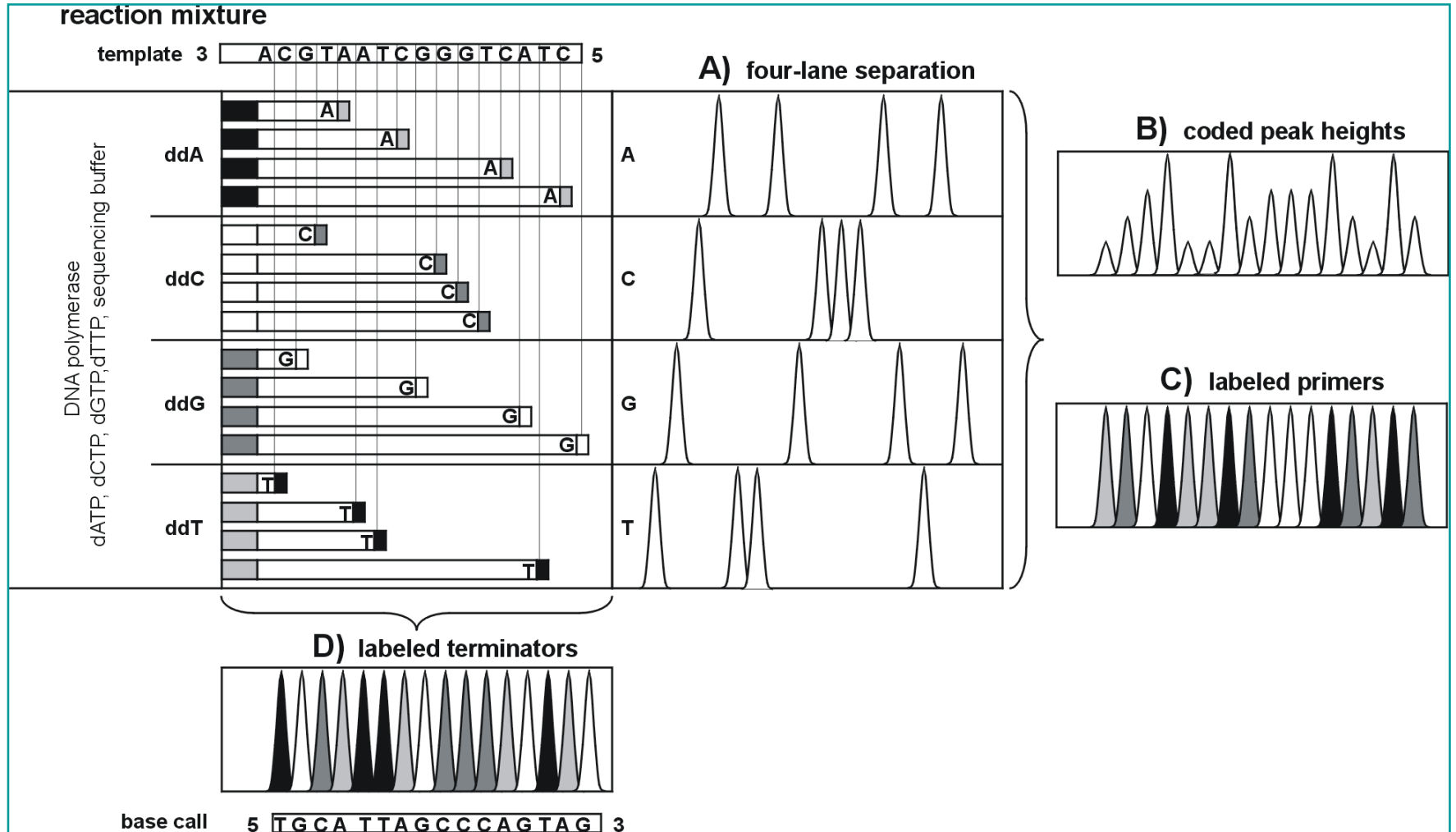
For his invention of the polymerase chain reaction (PCR) method

# PCR amplification scheme



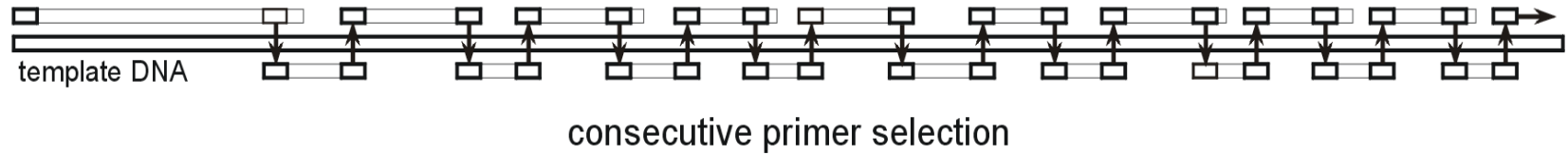
# **DNA sequencing**

# Analysis of Sanger sequencing fragments

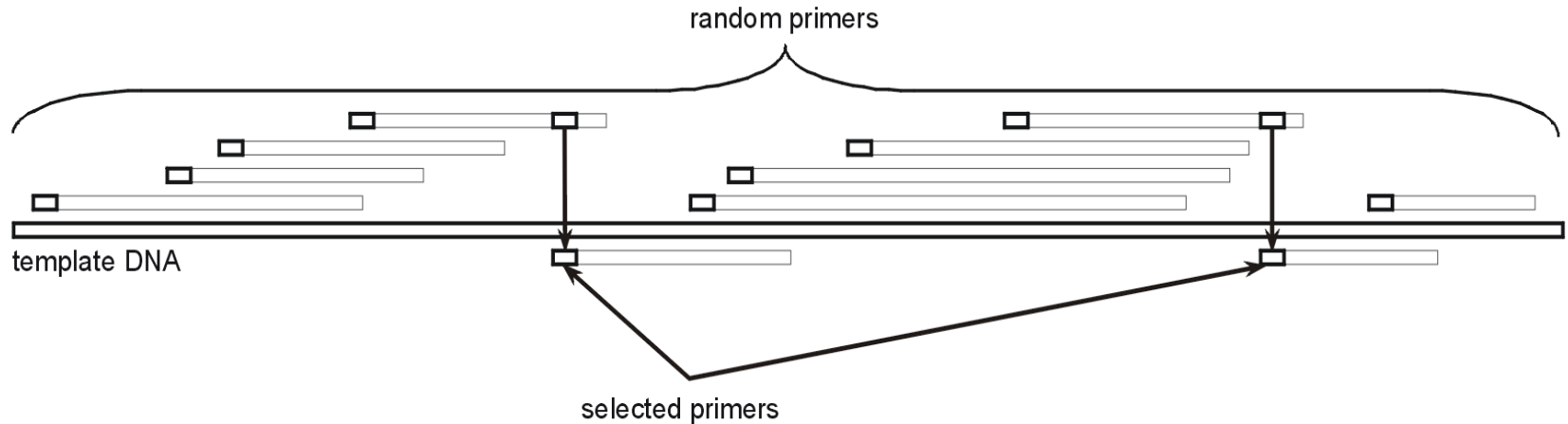


# DNA sequencing strategy

## Primer walking



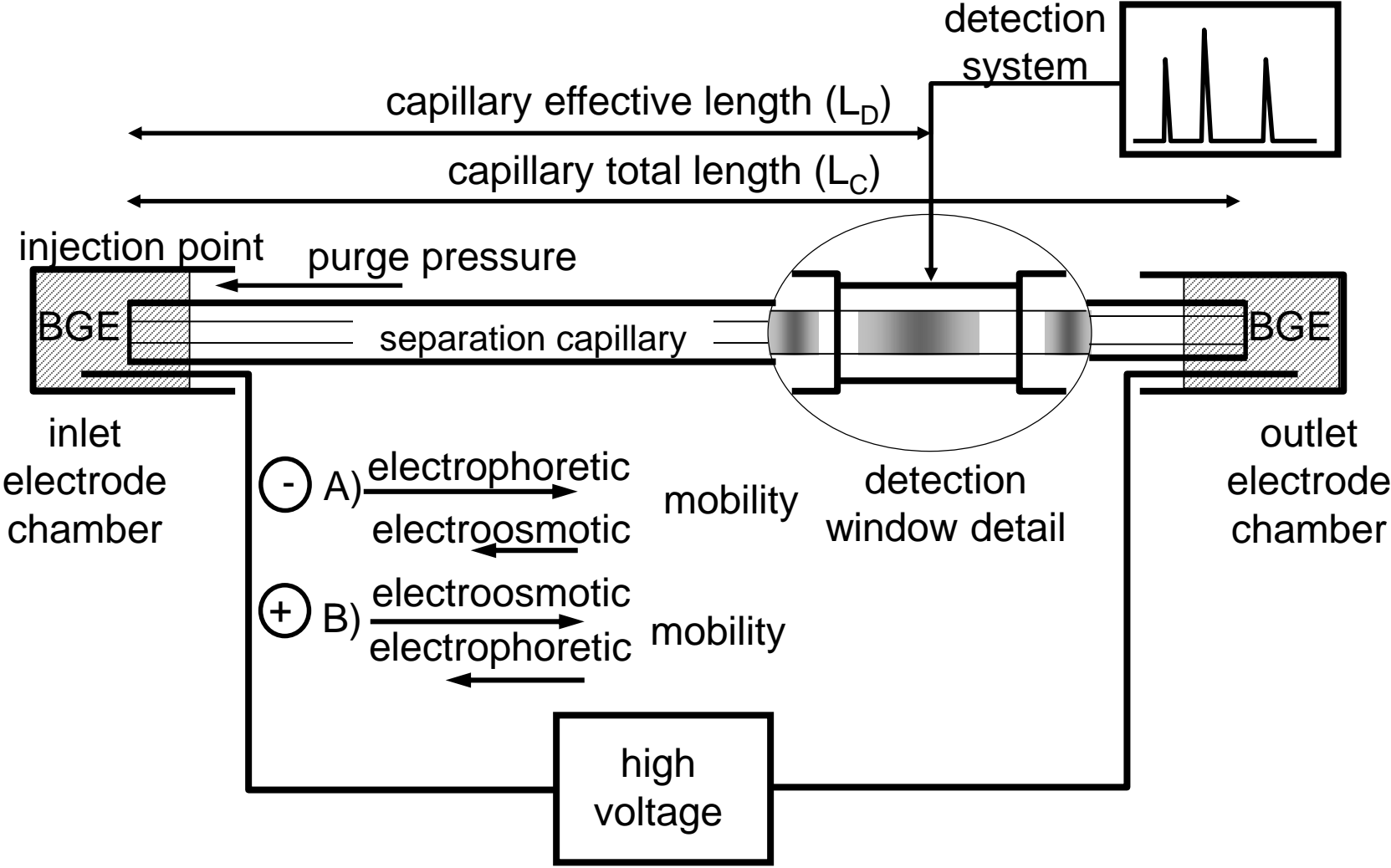
## Shotgun



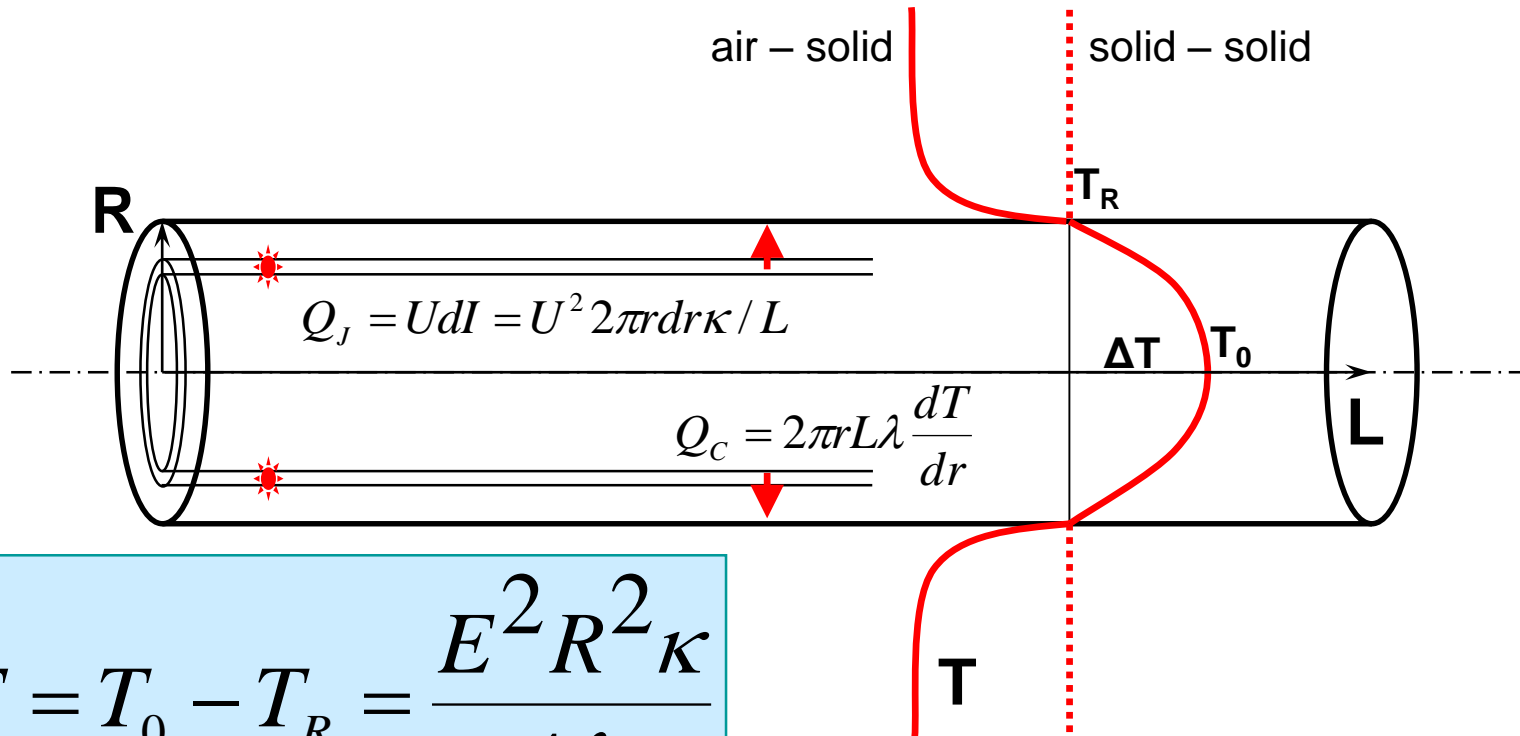


**Separation methods**  
**Capillary electrophoresis**  
**CE**

# Capillary electrophoresis scheme



# Why capillary electrophoresis?



$$\Delta T = T_0 - T_R = \frac{E^2 R^2 \kappa}{4\lambda}$$

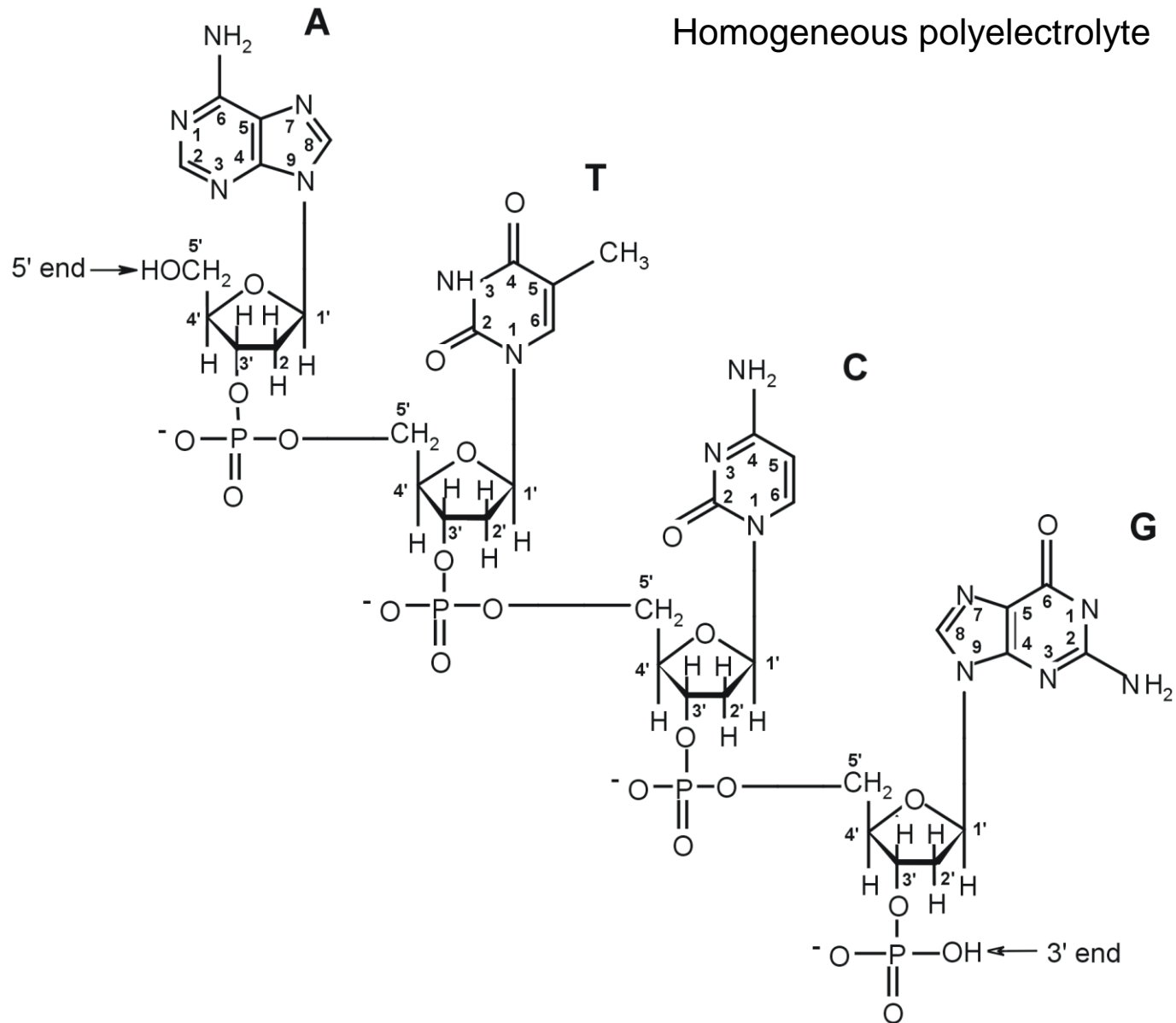
**Miniature capillary:** low  $R \Rightarrow$  fast separation

- 1) high resistivity  $\Downarrow$  low current at high voltage  $\Downarrow$  low heat production
- 2) efficient heat transport  $\Downarrow$  low temperature difference inside the capillary

# **DNA electromigration**

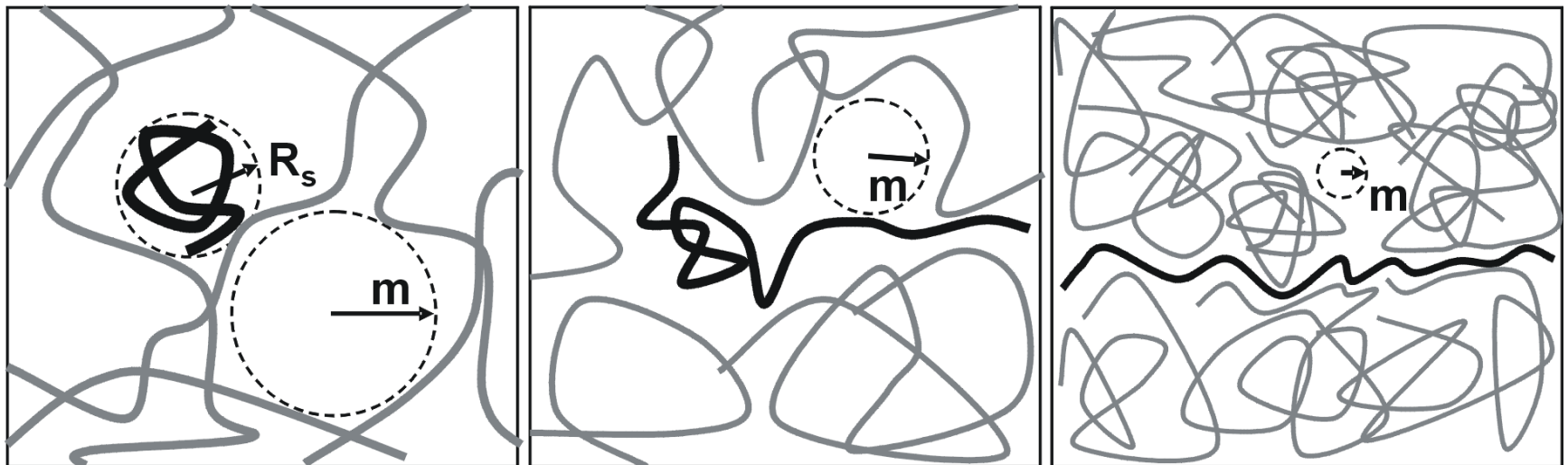
K. Klepárník, P. Boček, DNA diagnostics by Capillary Electrophoresis  
Chemical Reviews 107, 5279 – 5317, 2007.

# DNA primary structure



# DNA electromigration regimes in sieving media

Size separations of homogeneous polyelectrolytes are impossible in free solutions



**a**  $R_s < m$

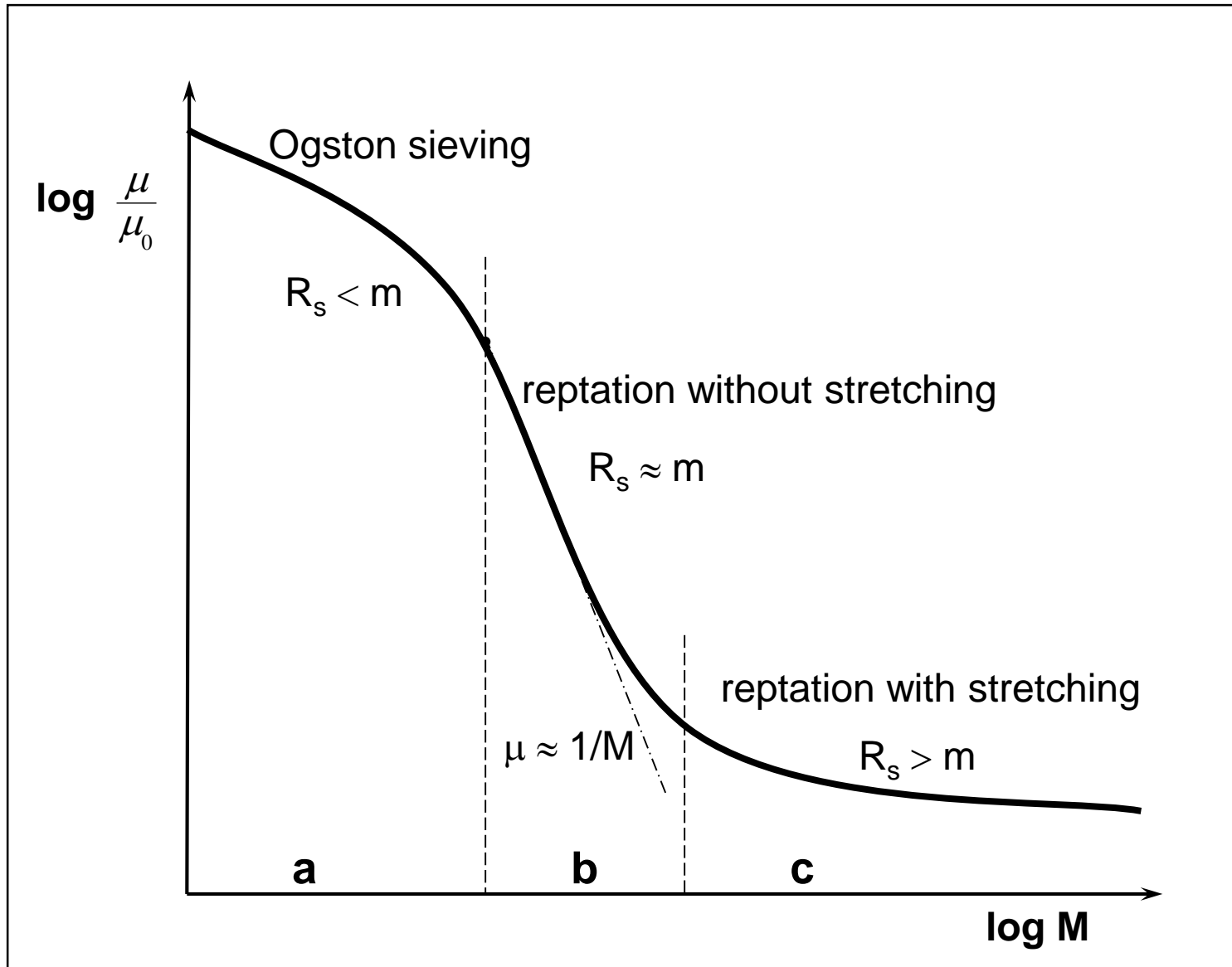
**b**  $R_s \sim m$

**c**  $R_s > m$

Short DNA fragments  
Low concentration of media

Long DNA fragments  
High concentration of media

# Dependence of DNA electrophoretic mobility on molecular mass



# **Human Genome Project**



**J. CRAIG VENTER, Ph.D., PRESIDENT, CELERA GENOMICS**  
**REMARKS AT THE HUMAN GENOME ANNOUNCEMENT**  
**THE WHITE HOUSE**  
**MONDAY, JUNE 26, 2000**

Mr. President, Honorable members of the Cabinet, Honorable members of Congress, distinguished guests. Today, June 26, 2000 marks an historic point in the 100,000-year record of humanity. We are announcing today that for the first time our species can read the chemical letters of its genetic code. At 12:30 p.m. today, in a joint press conference with the public genome effort, Celera Genomics will describe **the first assembly of the human genetic code** from the whole genome shotgun sequencing method. **Starting only nine months ago** on September 8, 1999, eighteen miles from the White House, a small team of scientists headed by myself, Hamilton O. Smith, Mark Adams, Gene Myers and Granger Sutton began sequencing the DNA of the human genome using a novel method pioneered by essentially the same team five years earlier at The Institute for Genomic Research in Rockville, Maryland. The method used by Celera has determined the **genetic code of five individuals...**

...There would be no announcement today if it were not for the more than **\$1 billion that PE Biosystems invested in Celera and in the development of the automated DNA sequencer** that both Celera and the public effort used to sequence the genome...



## **J. Craig Venter**

The Institute for Genomic Research  
**(TIGR)**

The first president of **Celera Genomics**

The completed sequence of the human genome was published in February 2001 in *Science*.

Venter, C. J. et al. *Science* **2001**, 291, 1304-1351.

# **Fluorescence chemistry**

# Lloyd M. Smith

Born 1954

A.B. 1976, University of California - Berkeley

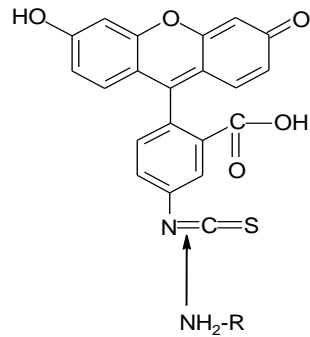
Ph.D. 1981, Stanford University

University of Wisconsin - Madison

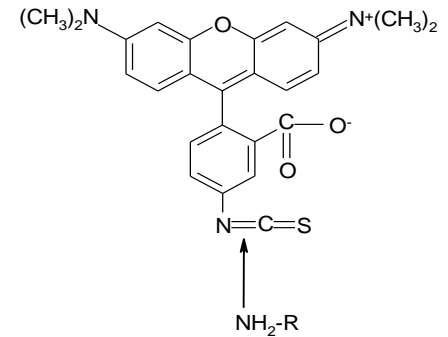
Smith, L. M., Sanders, J. Z., Kaiser, R. J., Hughes, P., Dodd, C., Connell, C. R., Heiner, C., Kent, S. B. H. and Hood, L. E.  
Fluorescence detection in automated DNA sequence analysis  
*Nature*, 321, 674-679, 1986.



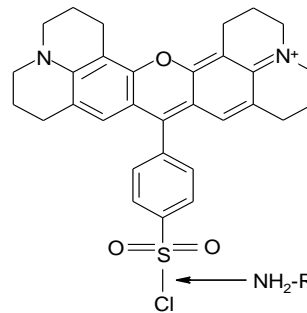
# Fluorescent labels



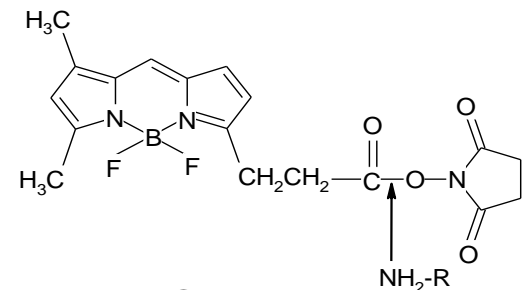
Fluorescein



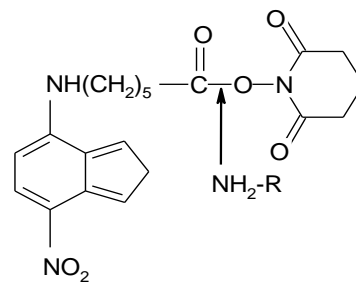
Rhodamine



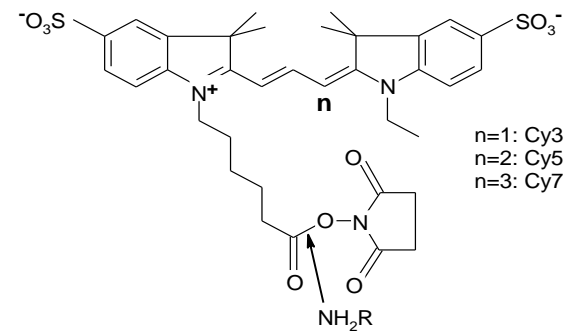
Texas Red



BODIPY



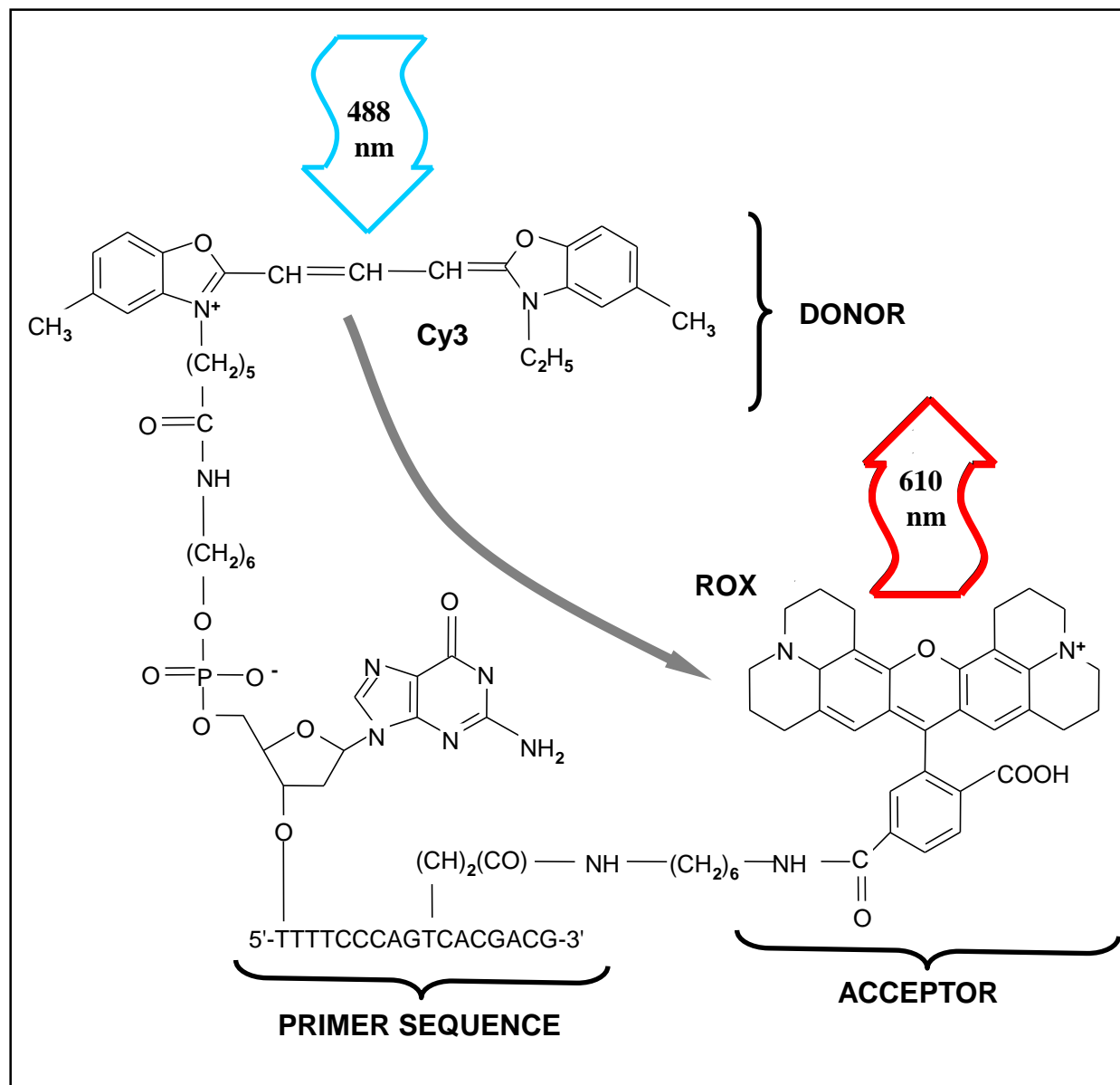
NBD



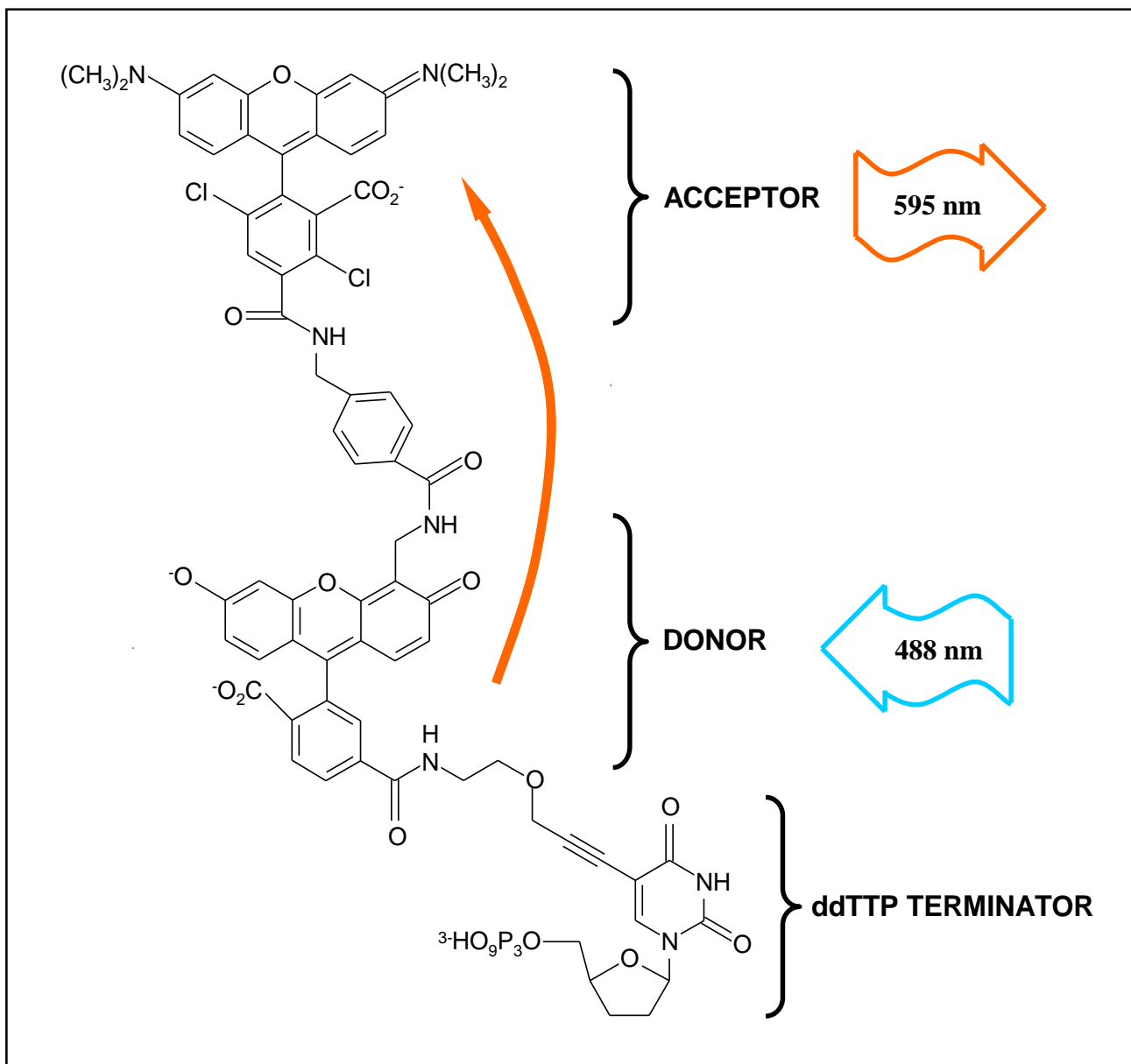
n=1: Cy3  
n=2: Cy5  
n=3: Cy7

Cy3,5,7

# Sequencing primer attached to Fluorescence Resonance Energy Transfer



# Dideoxy terminator attached to Fluorescence Resonance Energy Transfer

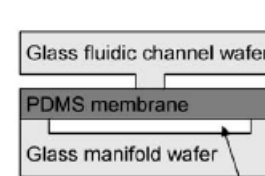


# Prof. Richard A. Mathies

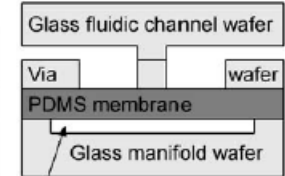
University of California at Berkeley  
 Department of Chemistry  
 Berkeley, CA



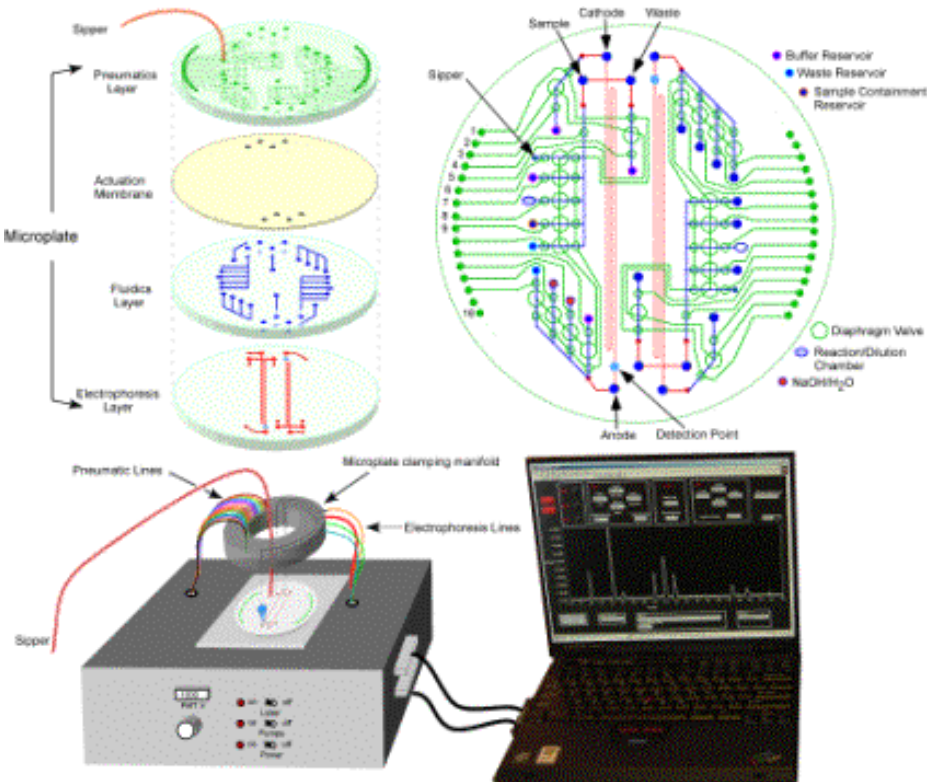
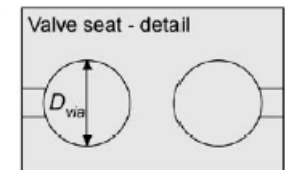
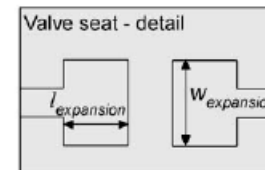
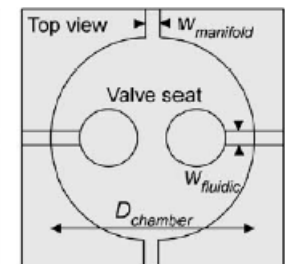
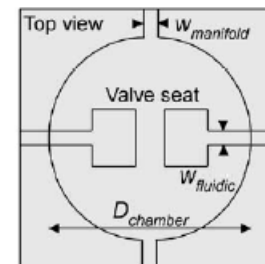
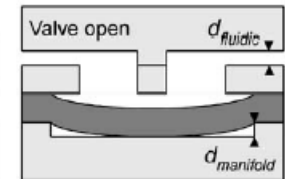
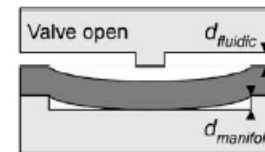
(A) 3-layer valve



(B) 4-layer valve



Displacement chamber

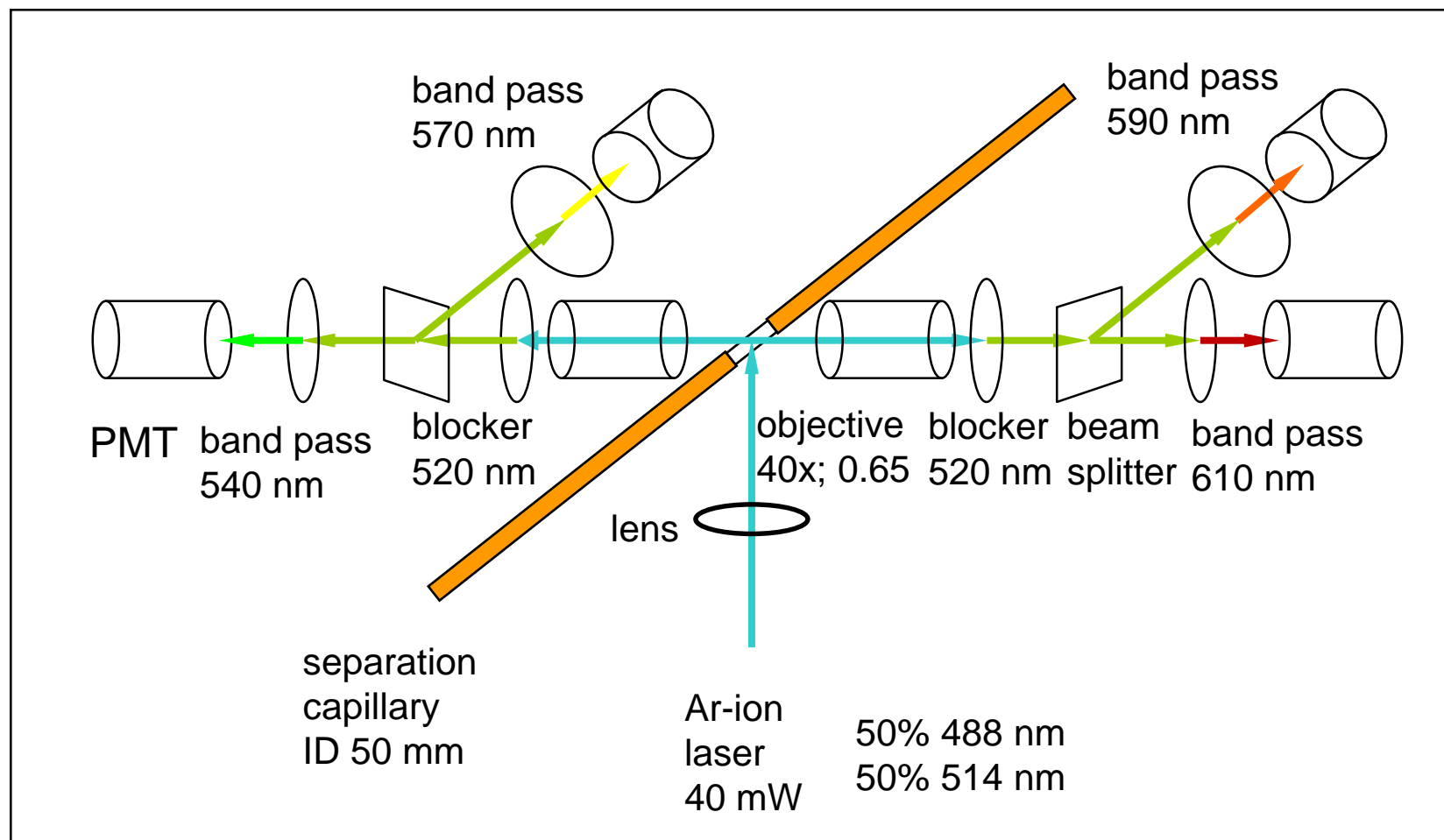




**LIF detection**

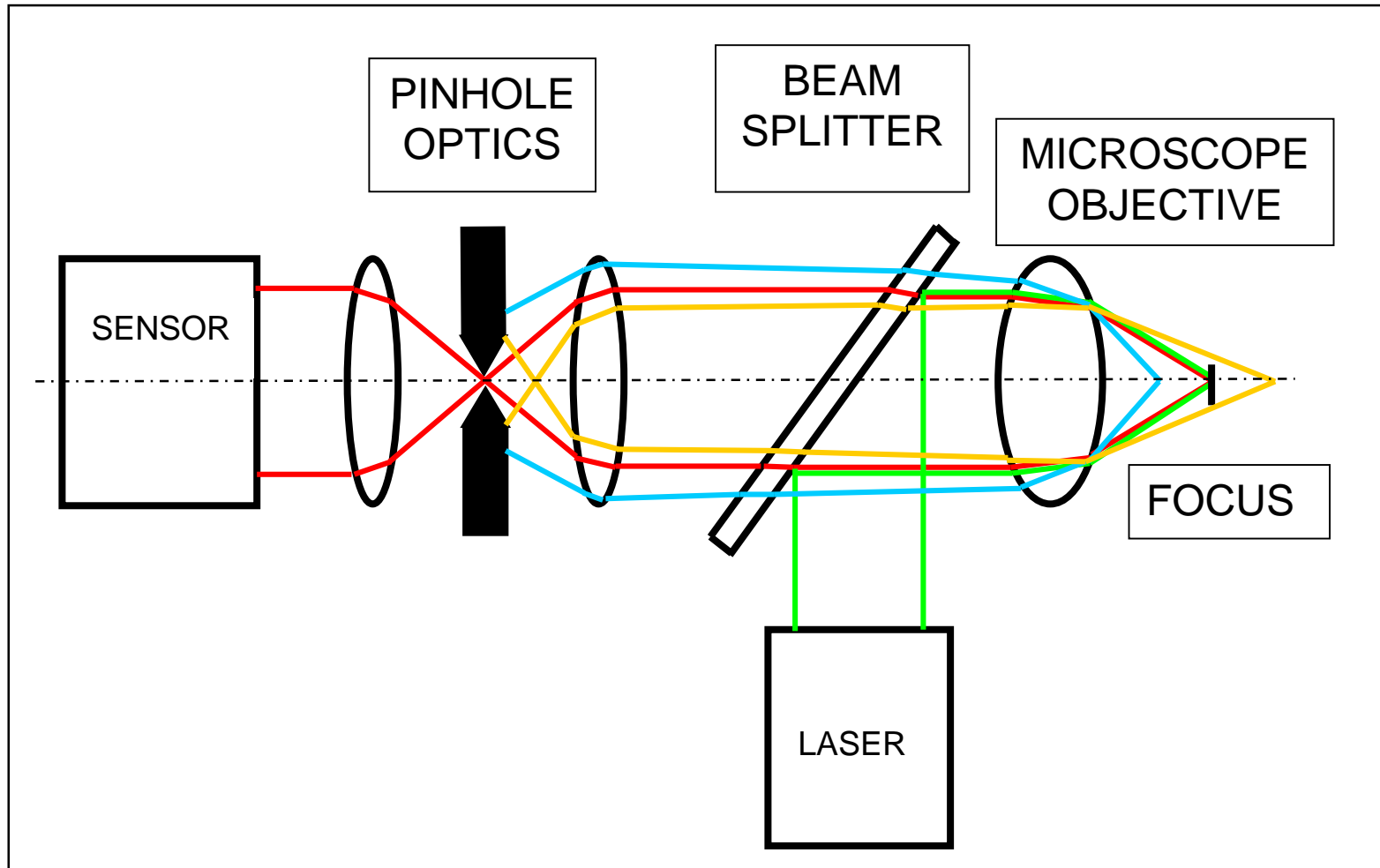
# Spectral filtering

## Four channel LIF detection arrangement



# Space filtering

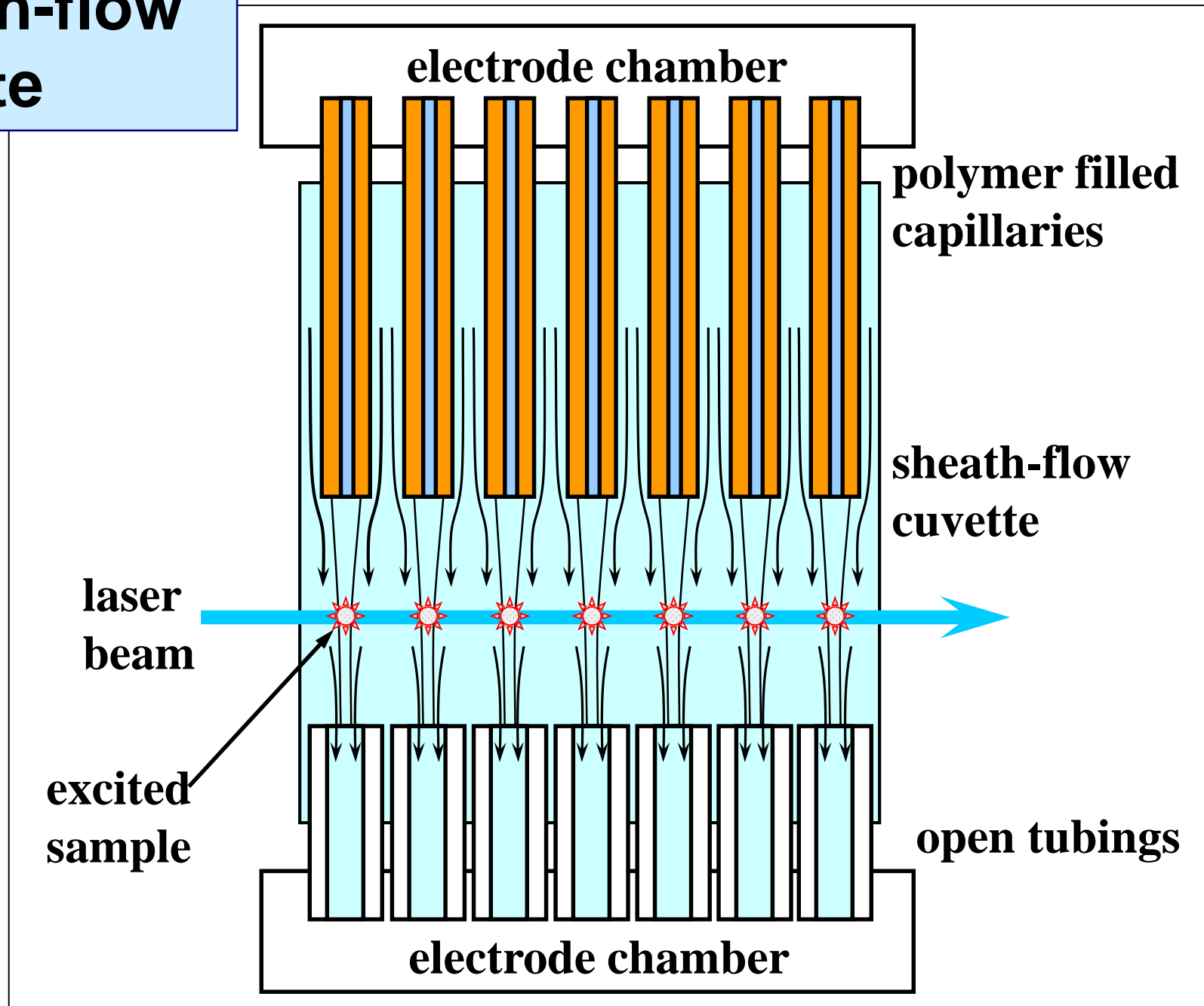
## SCHEME OF CONFOCAL DETECTOR





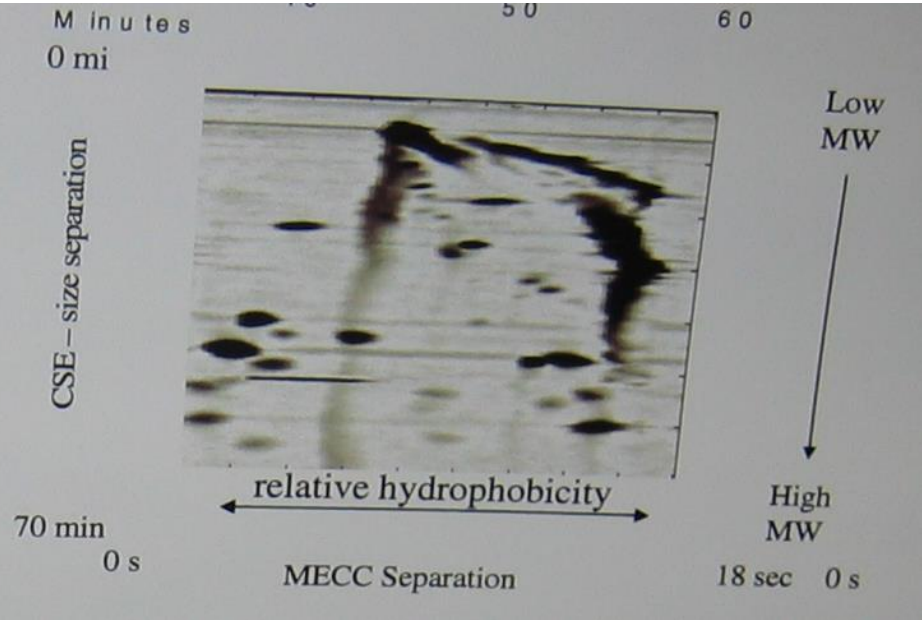
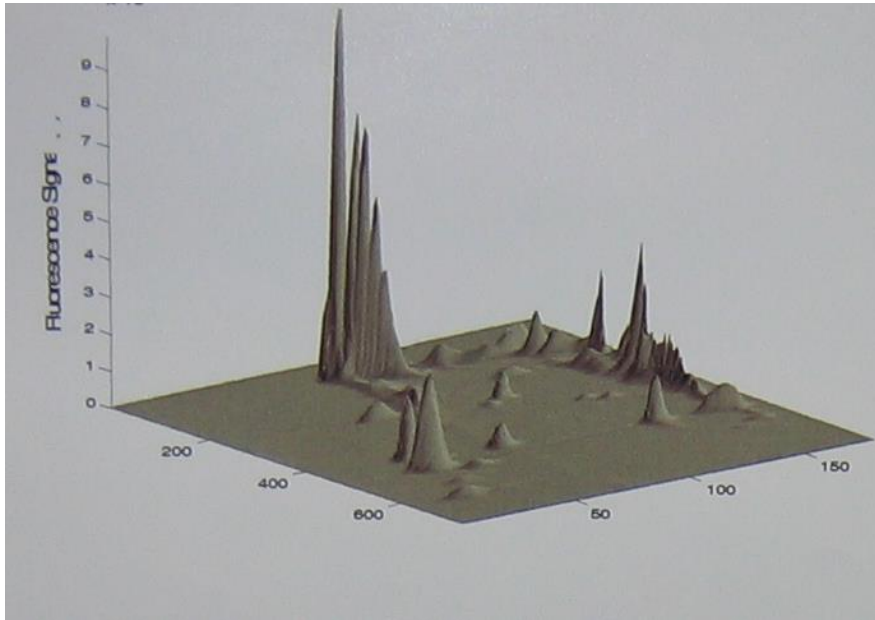
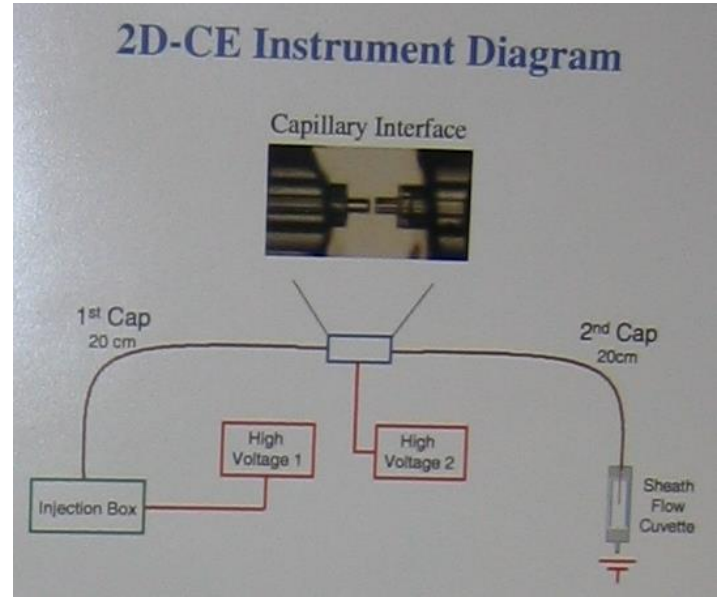
**Prof. Edward S. Yeung**  
Ames Laboratory  
U.S. Department of Energy  
Iowa State University.

# Sheath-flow cuvette





**Prof. Norman Dovichi**  
**University of Washington**  
**Seattle, WA, USA**



**Prof. Hideki Kambara**

senior chief scientist

Hitachi Central Research Laboratory

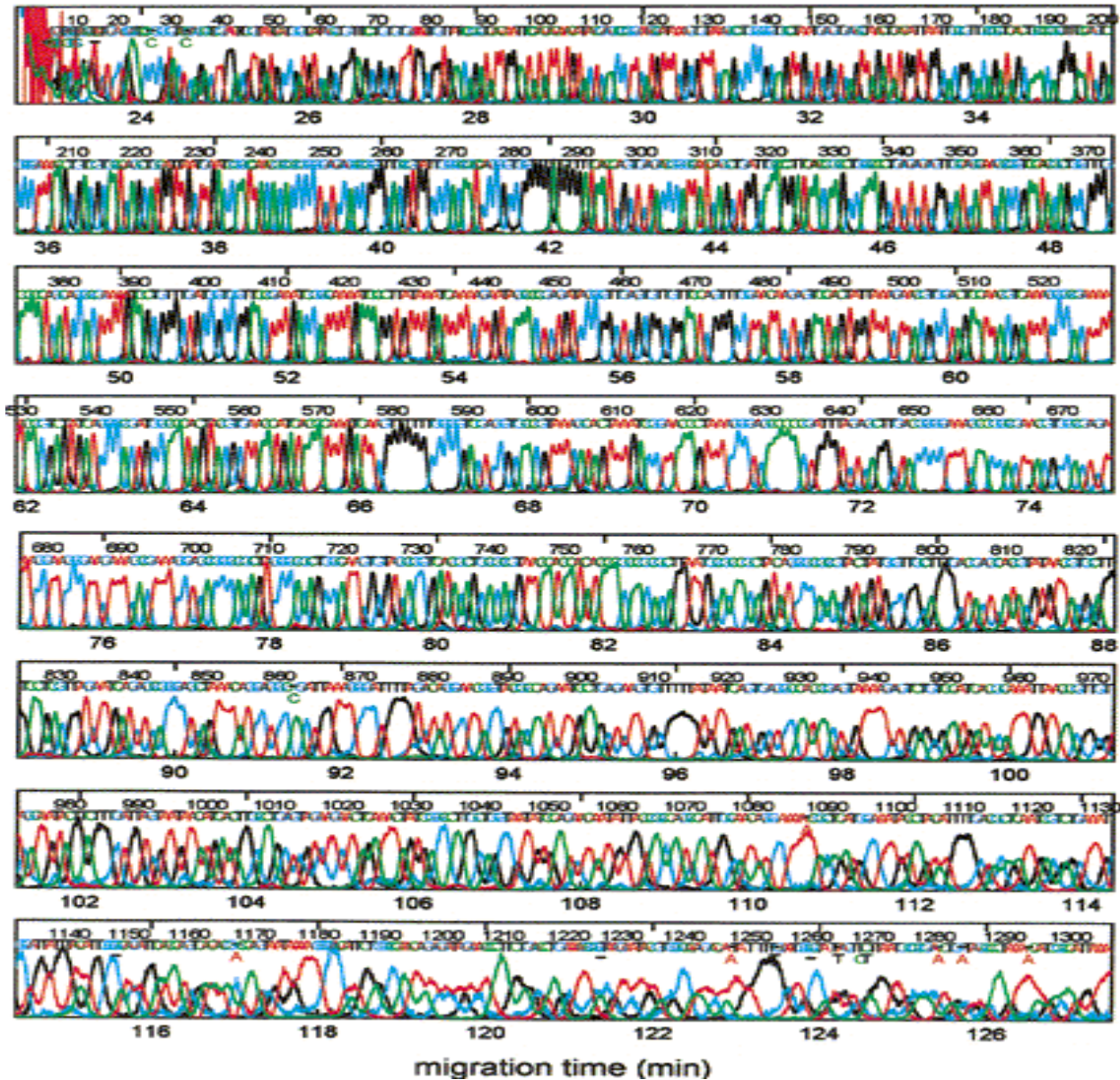
Tokyo, Japan



# DNA sequencing up to 1300 bases in 2 hours

Separation matrix: LPA 2.0% (w/w) 17 MDa, 0.5% (w/w) 270 kDa

E: 125 V/cm, T: 70 °C





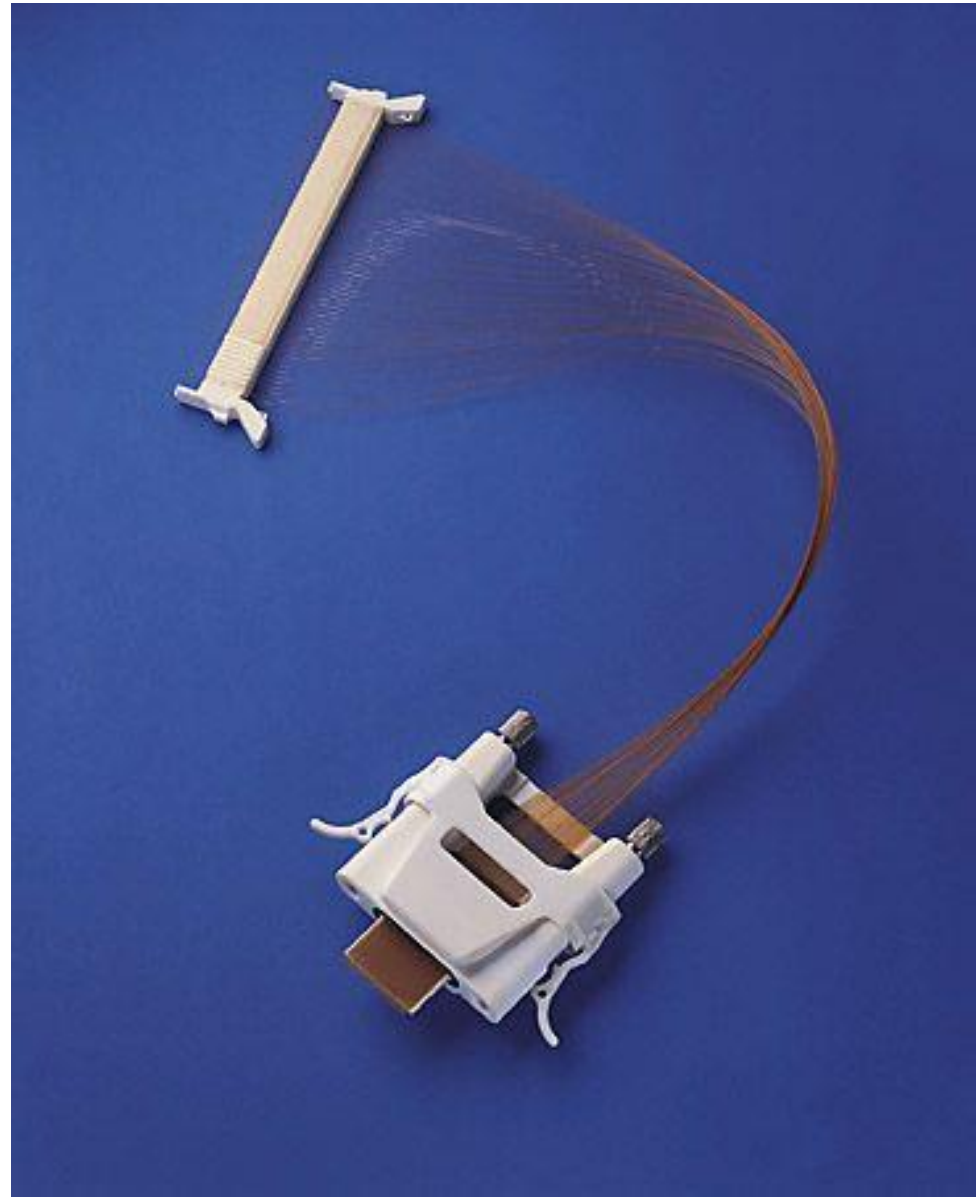


**Barry L. Karger  
James L. Waters  
Professor of Analytical  
Chemistry**

The Barnett Institute  
Northeastern University  
Boston MA

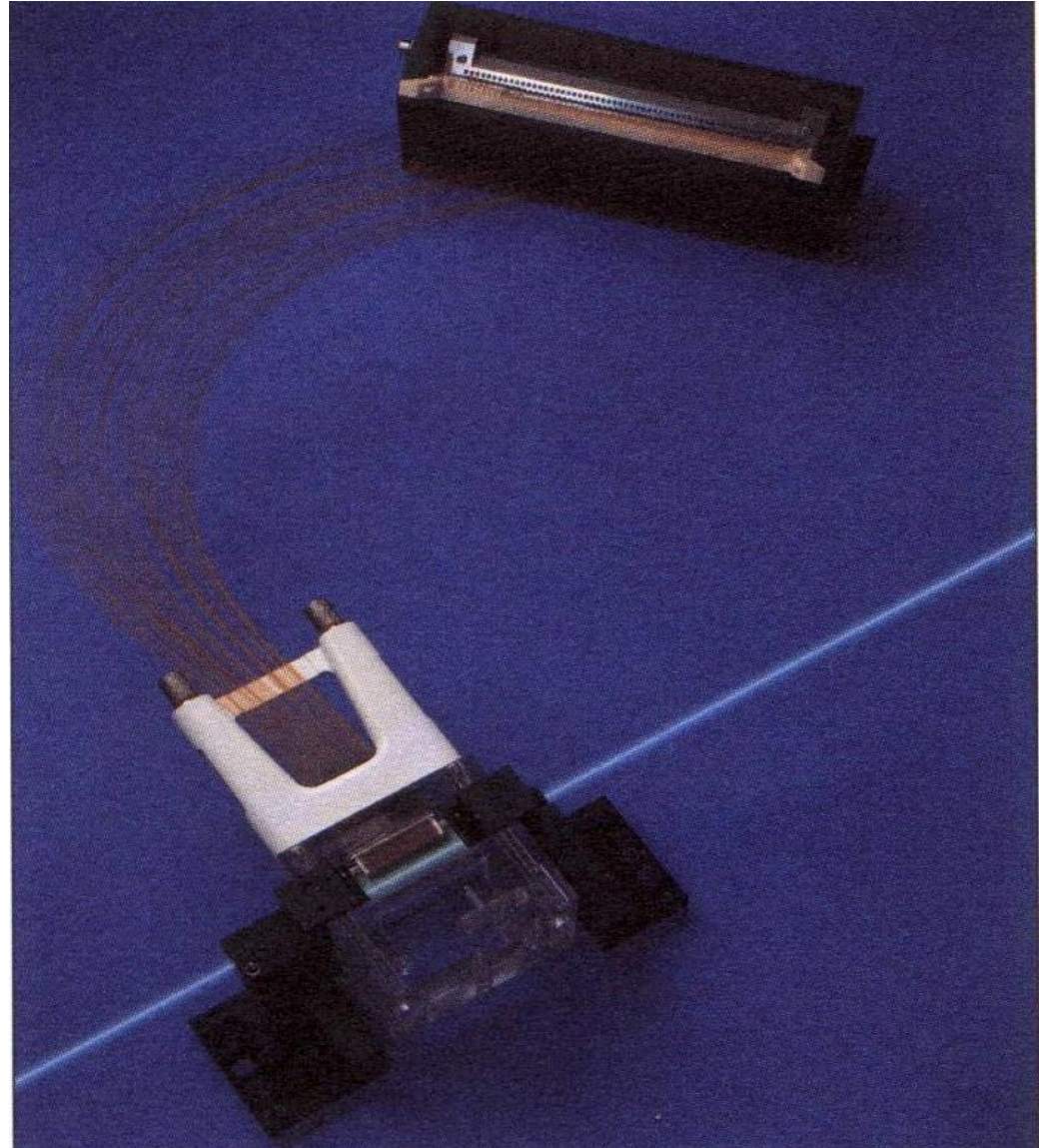
# ABI PRISM® 3700 DNA Analyzer

**96 active  
eight reserve capillaries**

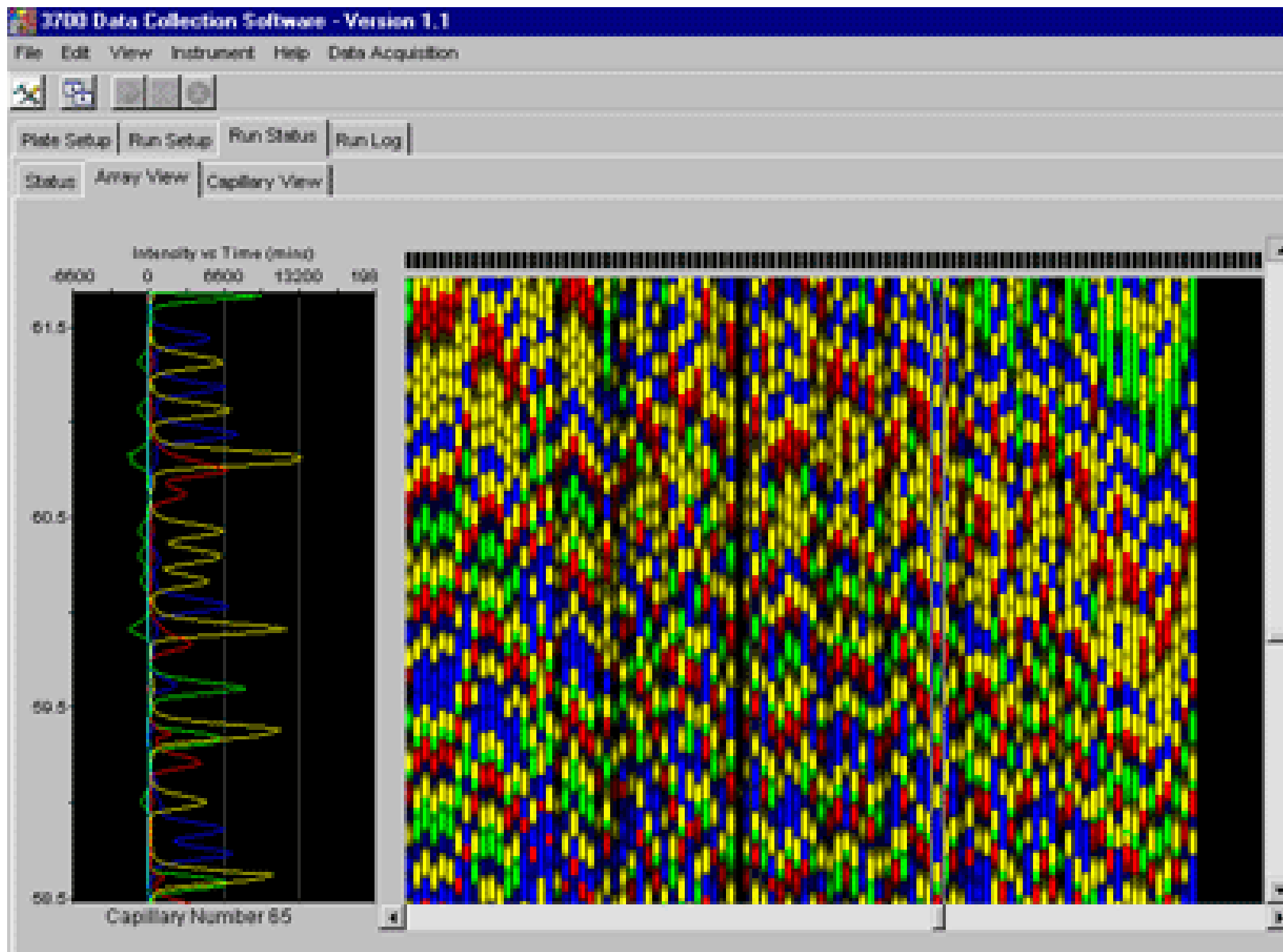


# ABI PRISM® 3700 DNA Analyzer

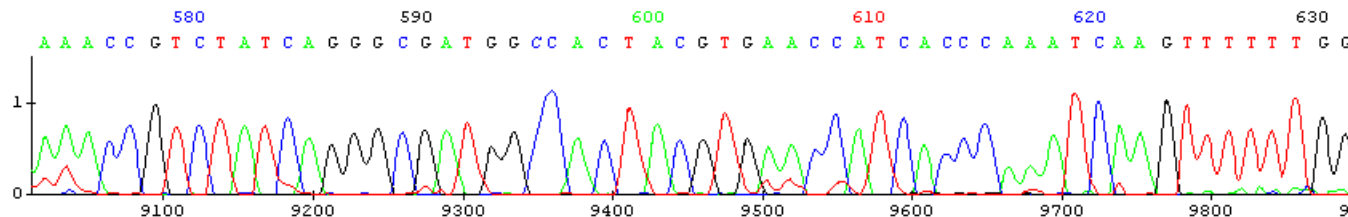
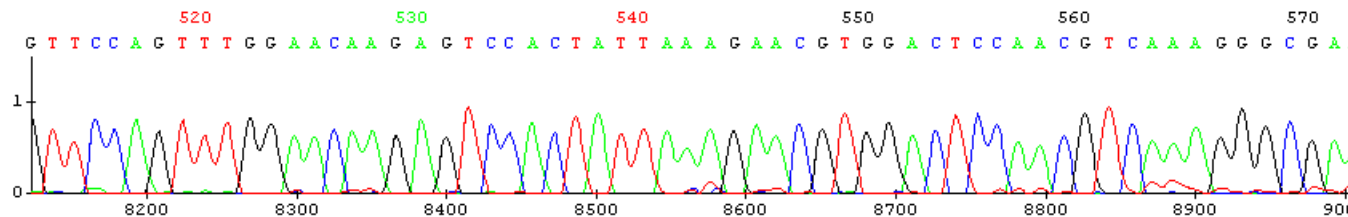
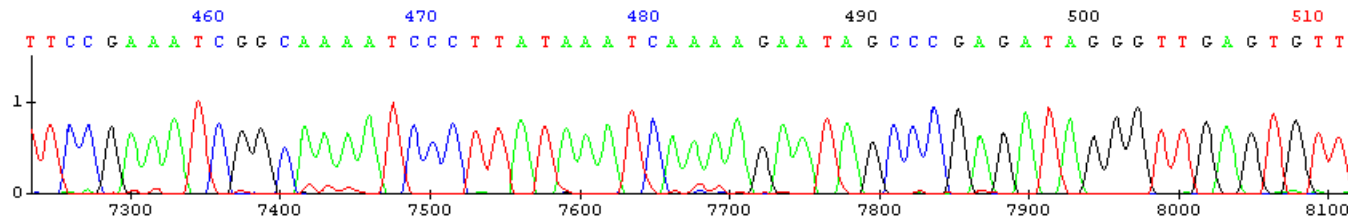
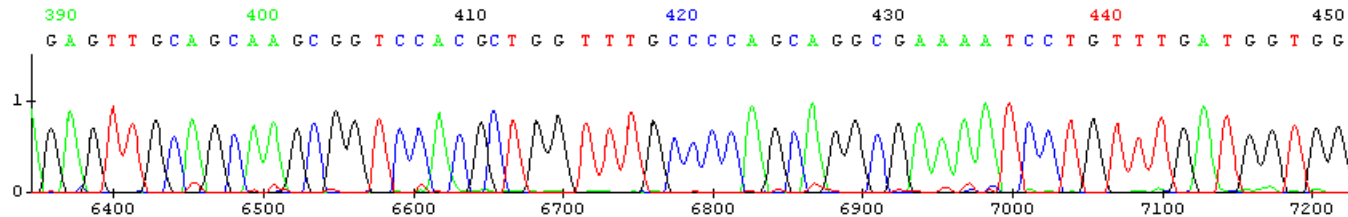
**Sheath flow cuvette**



# ABI PRISM® 3700 DNA Analyzer



# DNA sequencing record



# PE Applied Biosystems

ABI PRISM 3700

accuracy > 98.5% to 550 base  
96 samples per run in 3 hours  
laser Ar-ion 488 and 514.5 nm  
detection in sheath flow  
concave spectrograph and cooled CCD



# Molecular Dynamics

MEGABACE 1000

accuracy > 98.5% to 550 base  
96 samples per run in 2 hours  
laser Ar-ion 488 nm  
energy transfer dyes  
confocal scanning with 4 filters and 2  
PMTs



# **DNA mutation analysis**

# Restriction (amplification) fragment length polymorphism

## RFLP (AFLP)

Size based separation of ds or ss DNA fragments

Resolution:

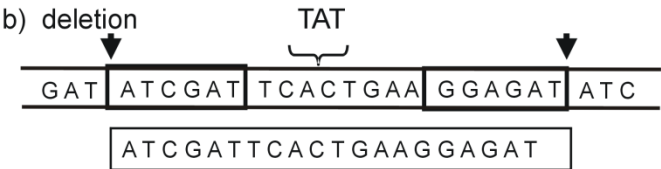
ss > 1000

ds > 400

a) wild type sequence



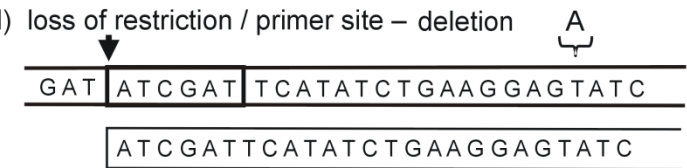
b) deletion



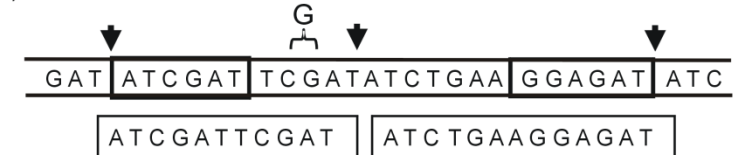
c) insertion



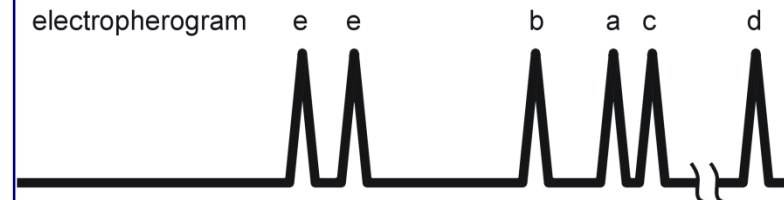
d) loss of restriction / primer site – deletion



e) creation of restriction site – insertion



electropherogram

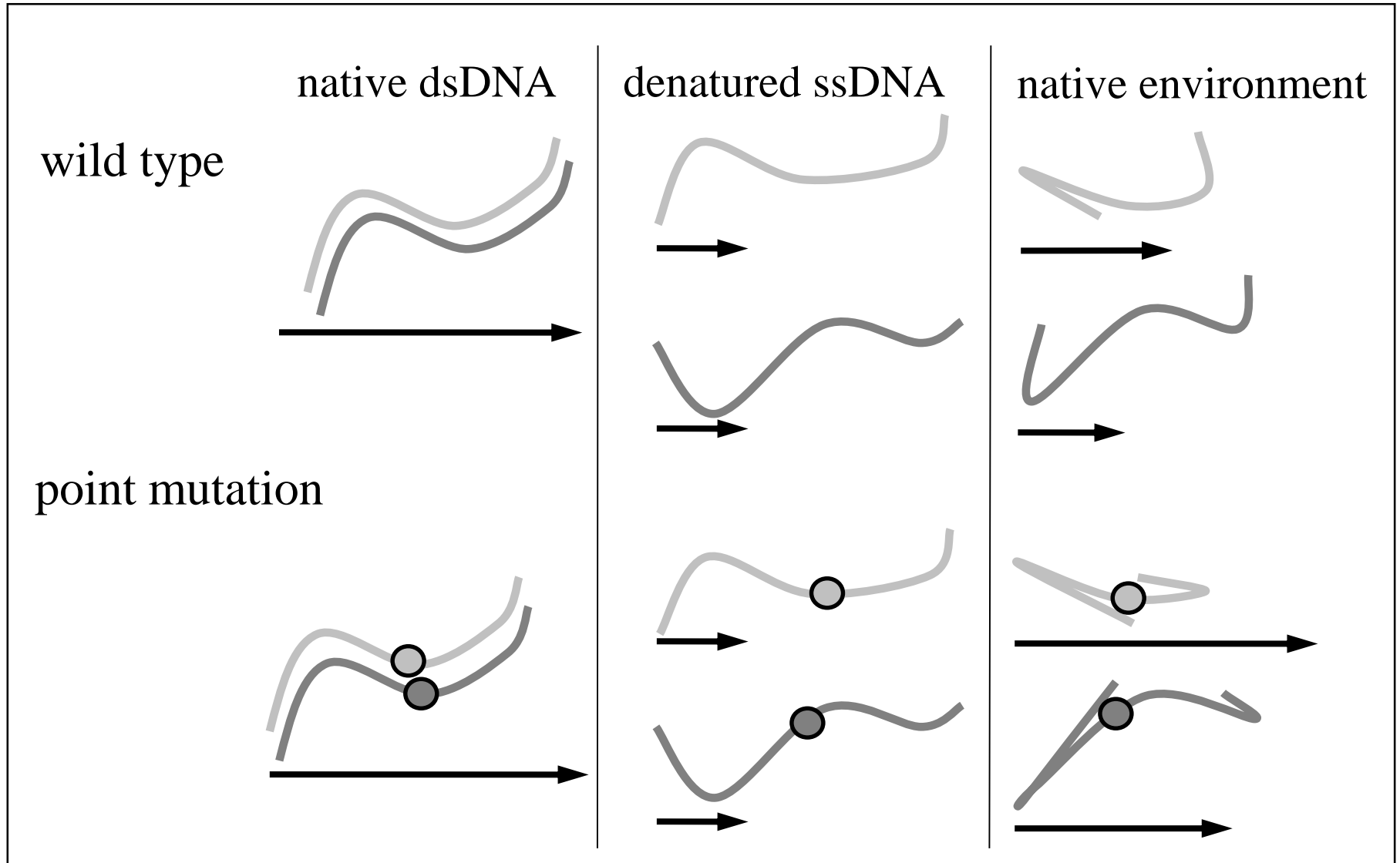




# **Single Strand Conformation Polymorphism**

**SSCP**

# Principle of SSCP technique



# Phenylketonuria

## SSCP analysis

Detection of point mutation C > T in phenylalanine hydroxylase gene on chromosome 12

## Separation conditions:

2% solution of agarose SeaPrep in 1xTBE with 10% formamide

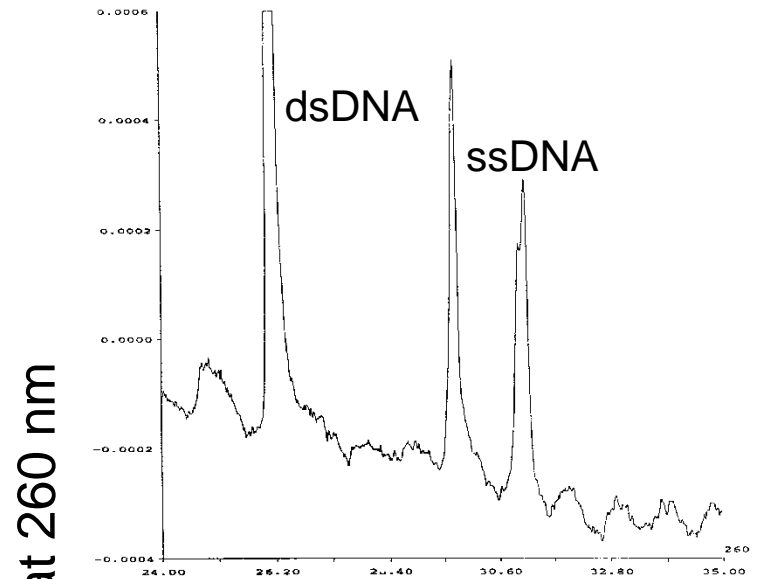
$T$  - 30 °C

$LC$  - 55 cm

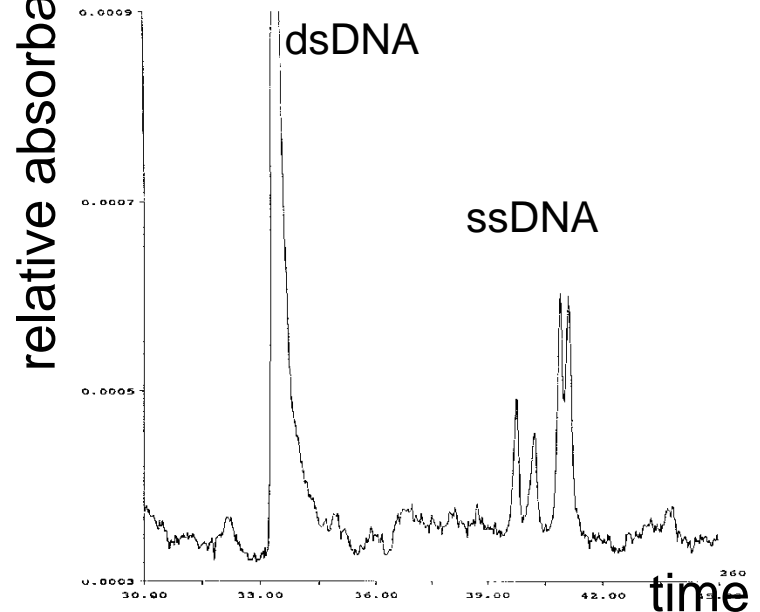
$LD$  - 50 cm

$E$  – a) 183 V/cm, b) 135 V/cm.

## a) health homozygote



## b) heterozygote



# Single nucleotide primer extension

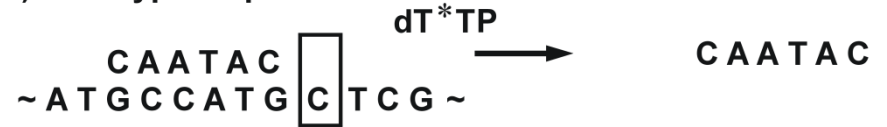
Minisequencing

## SNuPE

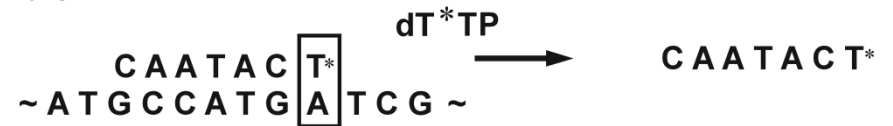
### SNuPE reaction

### SNuPE products

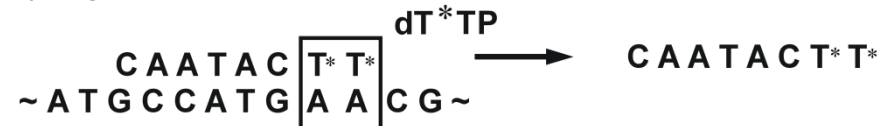
a) wild type sequence



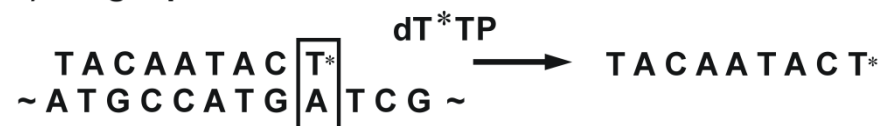
b) point mutation



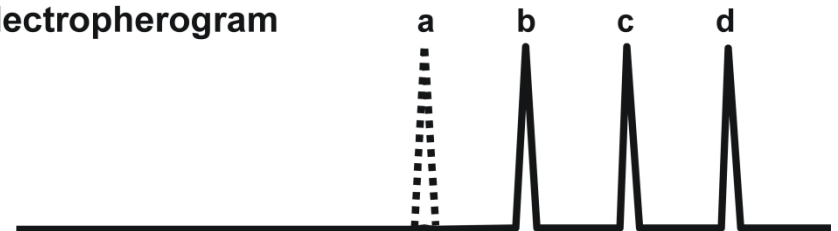
c) repetitive nucleotides



d) longer primer



electropherogram



**Next generation  
sequencing**

**Single molecule  
detection**

# **Single molecule reaction monitoring**

# Parallel single molecule sequencing by synthesis

## Helicos

### The HeliScope™ Sequencer

**$2 \cdot 10^9$  b/day**

**$10^9$  reads/run**

**25 – 55 bp read lengths**



**454** LIFE  
SCIENCES

### Genome Sequencer FLX System

**$3 \cdot 10^8$  b/day**

**100 Mb/7.5 hour run**

**400 000 reads/7.5 hour**

**200 – 300 bp read lengths**



## Solexa

### Illumina Genome Analyzer

**$6 \cdot 10^8$  b / day**

**$3 \cdot 10^9$  b / 5 days run**

**$50 \cdot 10^6$  oligo clusters**

**36 – 50 bp read lengths**



# Photocleavable dideoxy nucleotides

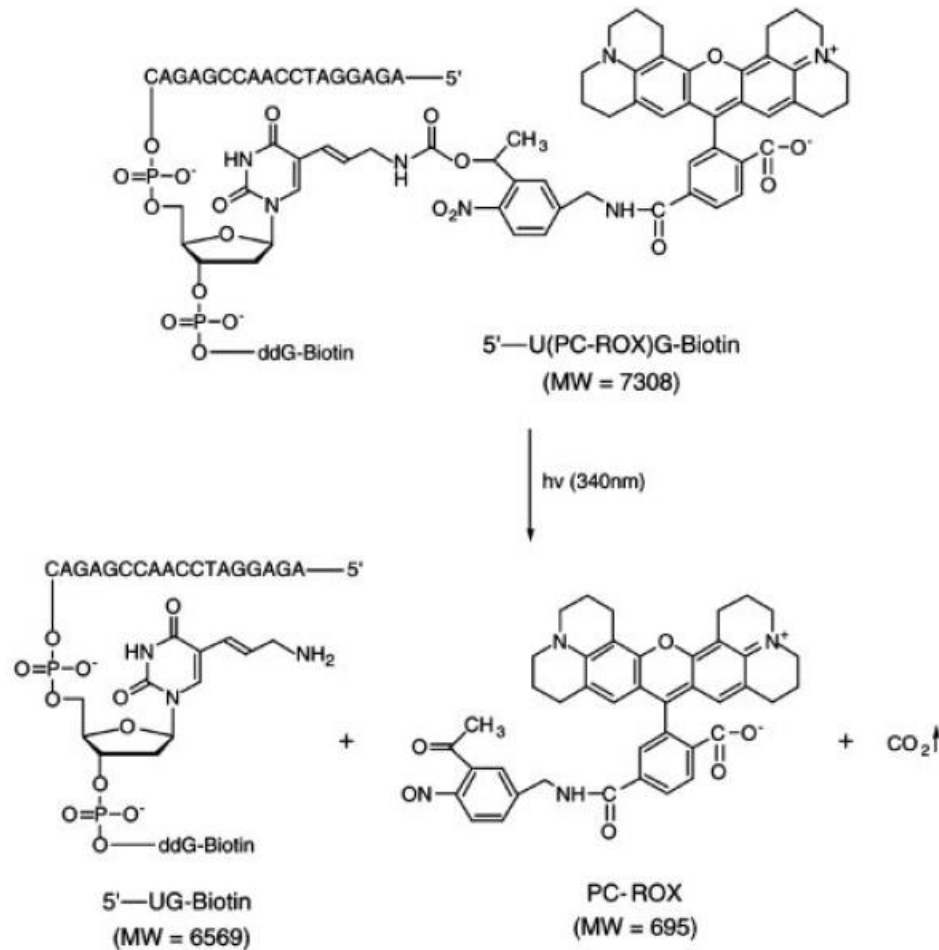


Fig. 2. DNA product 5'-U(PC-ROX)G-biotin, formed by incorporating a dUTP-PC-ROX into a primer in a polymerase reaction and its photocleavage, producing DNA fragment 5'-UG-biotin and PC-ROX. MW, molecular weight.



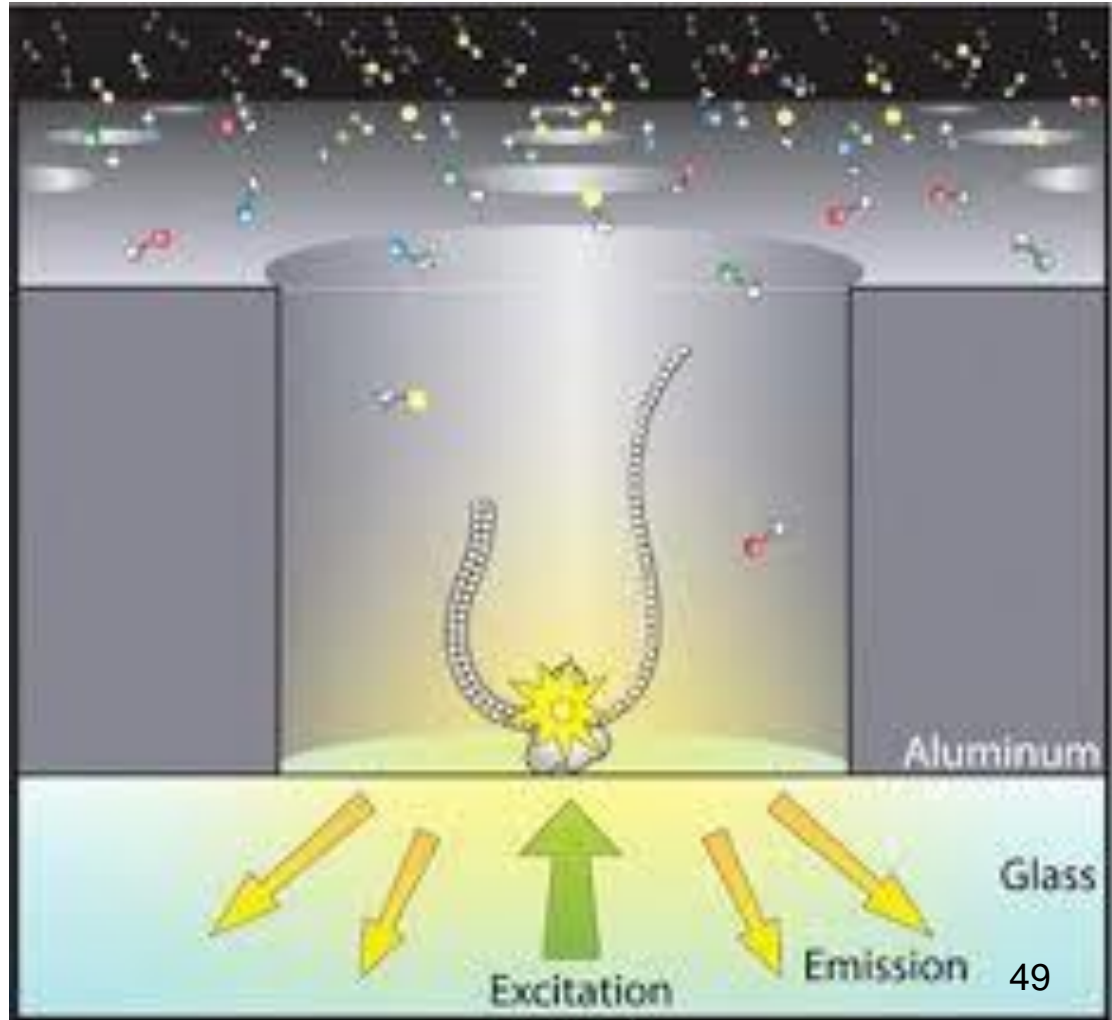
# Next generation DNA sequencing

## Single molecule real time sequencing (SMRT™)

### Pacific Biosciences

DNA sequencing – DNA polymerase  
RNA sequencing – reverse transcriptase  
Codone-resolved translation elongation by single ribosomes

Tens of nucleotide peaks in 1 sec  
Read length 1 – 15 kb  
80 000 detection points  
15 min/genome:  $50 \text{ n/s} * 80\,000 \text{ points}$   
 $* 15 \text{ min} * 60 \text{ s} = 3.6 \text{ Gb}$   
DNA polymerase 529 processivity 20  
kB – 400 b/s  
Some enzymes are not processive  
\$ 100/genome



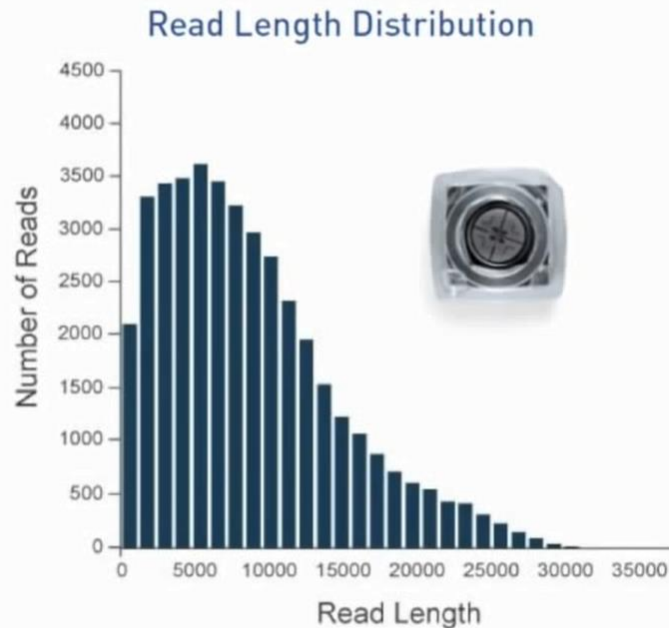
# *PacBio RS instrument*





# Pacific Biosciences Read Length

## New P5-C3 Sequencing Chemistry



## Typical Results

### Read Length:

Average: ~ 8.5 kb  
Maximum: > 30 kb  
Top 5% of reads: > 18 kb  
Half of data in reads: > 10 kb

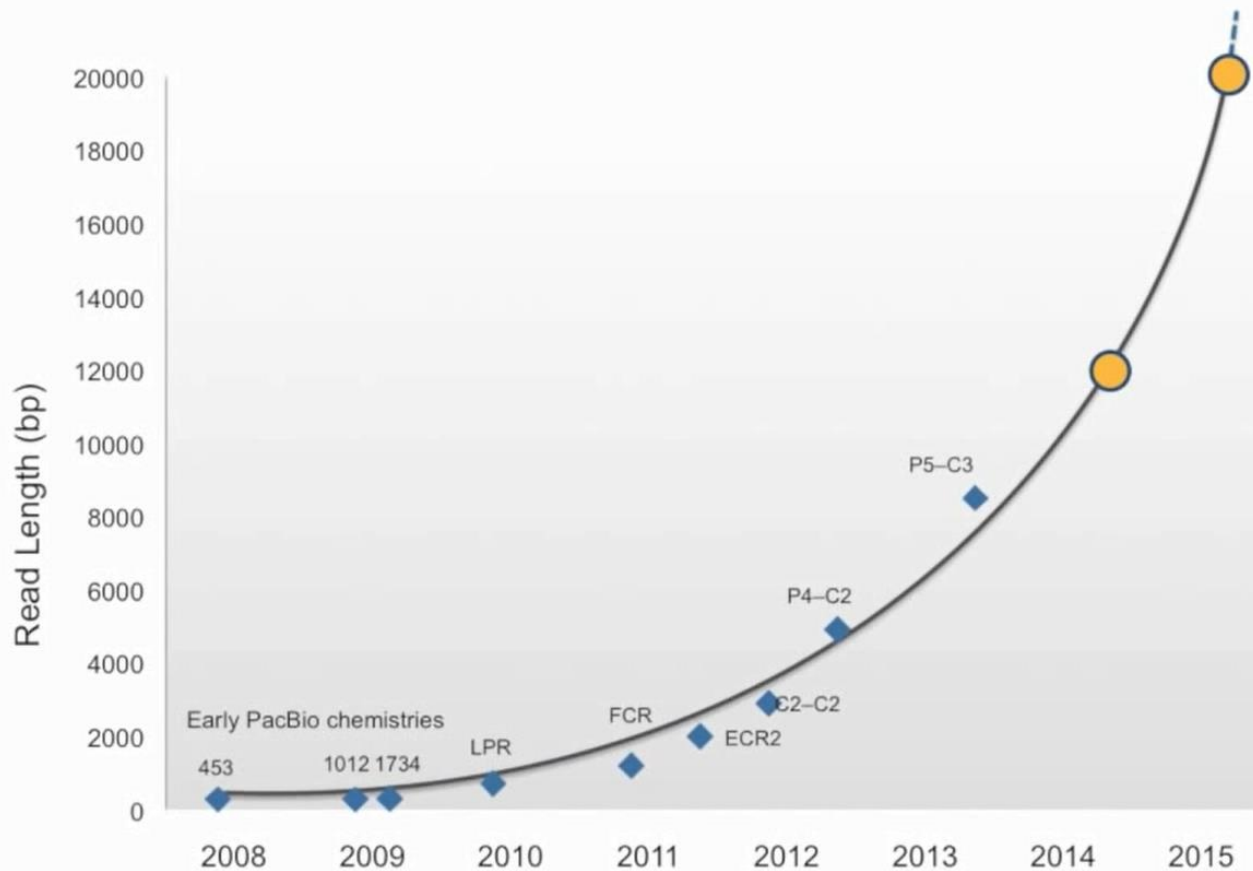
Data per SMRT® Cell: ~ 375 Mb

Based on data from a 20 kb size-selected *E. coli* library using a 180-minute movie.  
Each SMRT Cell yields ~ 50,000 reads.

# Pacific Biosciences Read Length

PacBio Technology Roadmap for 2014

## PacBio® Advances in Read Length



12

# Single molecule real time sequencing SMRT™

[www.pacificbiosciences.com](http://www.pacificbiosciences.com)

DNA sequencing – DNA polymerase

RNA sequencing – reverse transcriptase

Codone-resolved translation elongation by single ribosomes

Tens of nucleotide peaks in 1 sec

Read length 1 – 15 kb

80 000 detection points

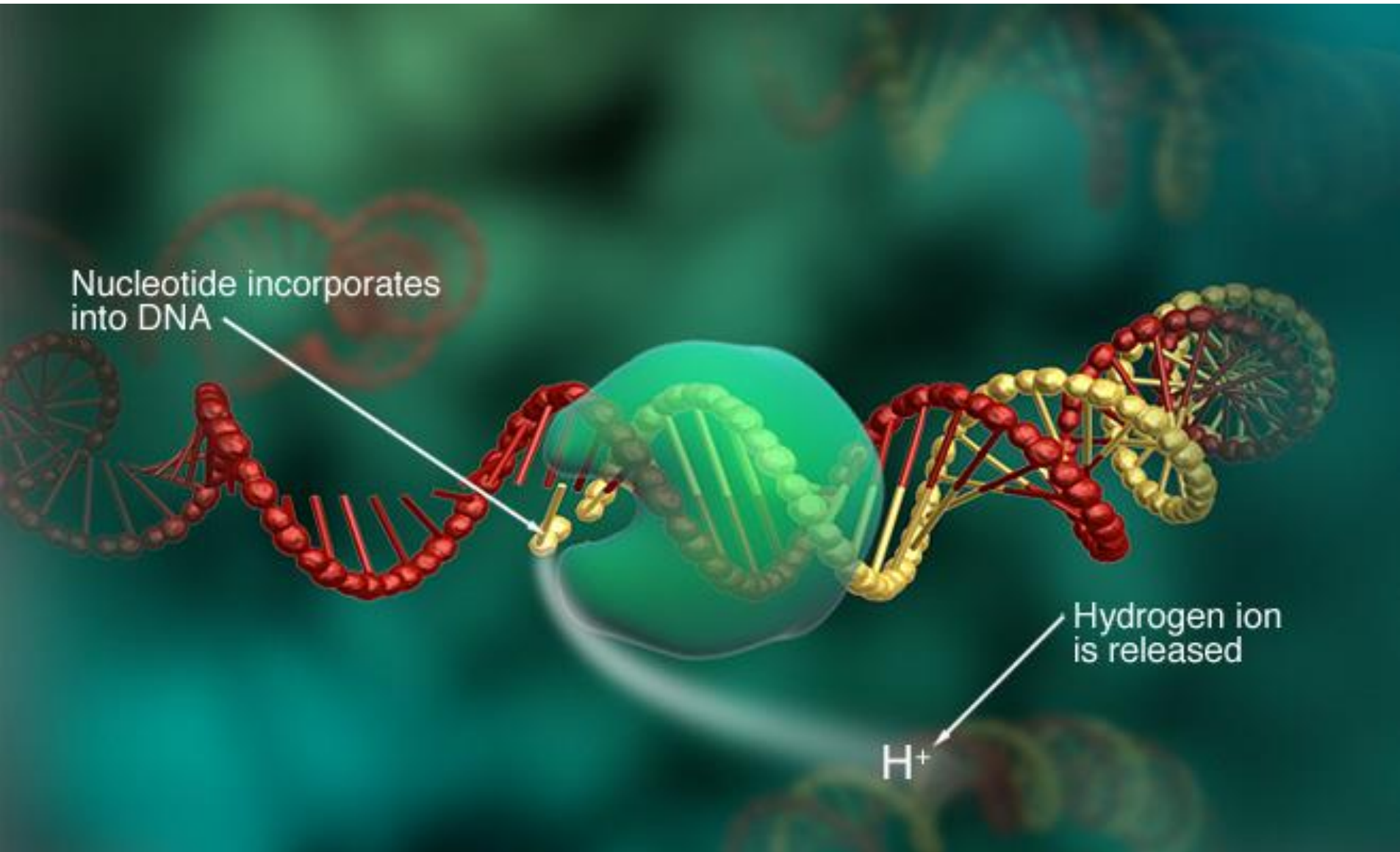
15 min/genome:  $50 \text{ n/s} * 80\,000 \text{ points} * 15 \text{ min} * 60 \text{ s} = 3.6 \text{ Gb}$

DNA polymerase 529 processivity 20 kB – 400 b/s

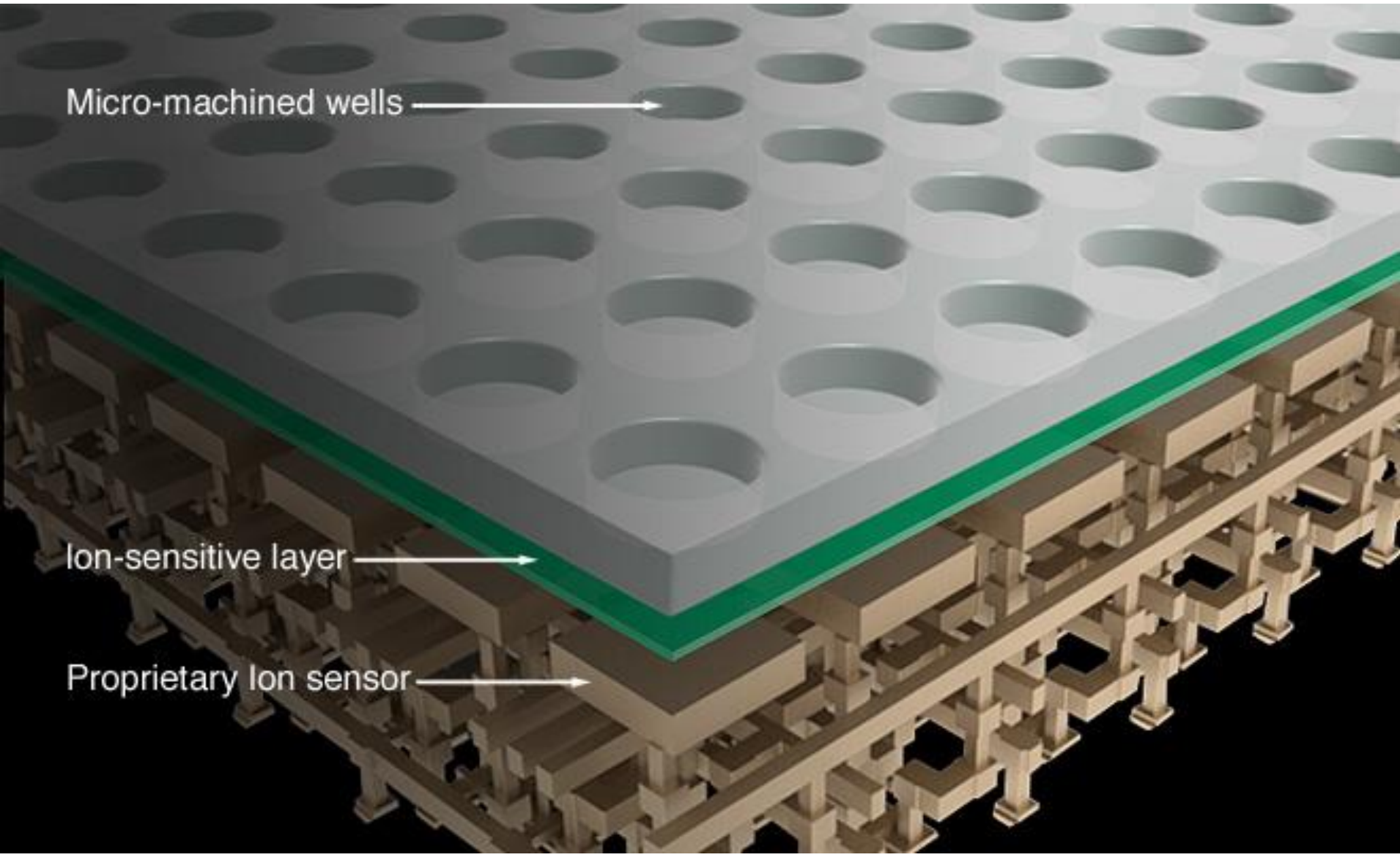
Some enzymes are not processive

\$ 100/genome

Hydrogen ion is released as a byproduct when a nucleotide is incorporated into a strand of DNA by a polymerase

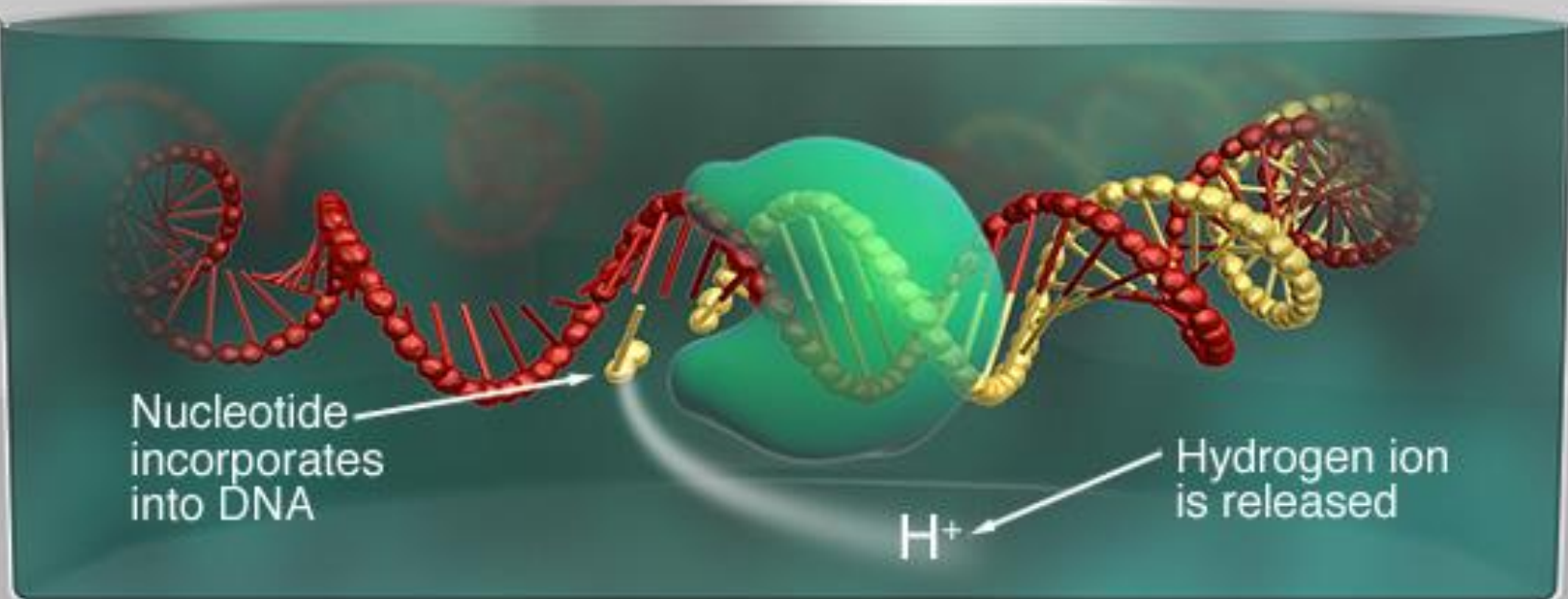


High-density array of micro-machined wells. Each well holds a different DNA template. Beneath the wells is an ion-sensitive layer and a proprietary ion sensor.

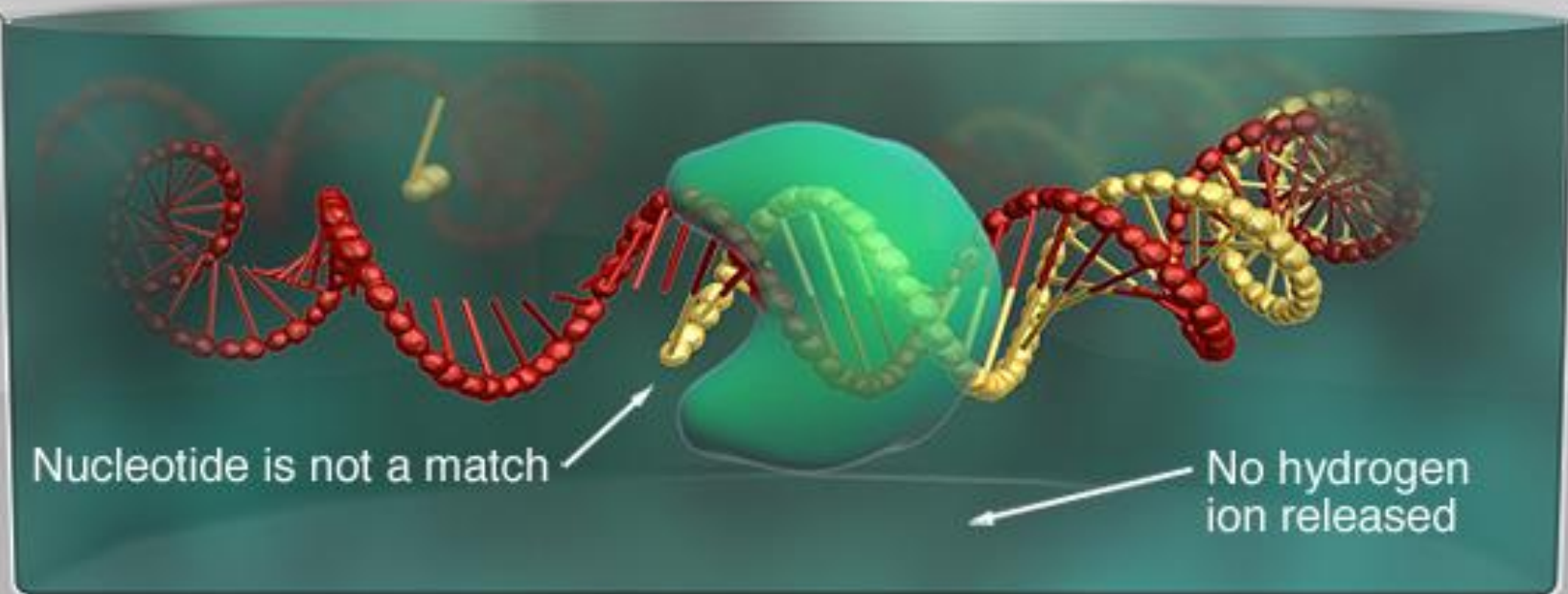




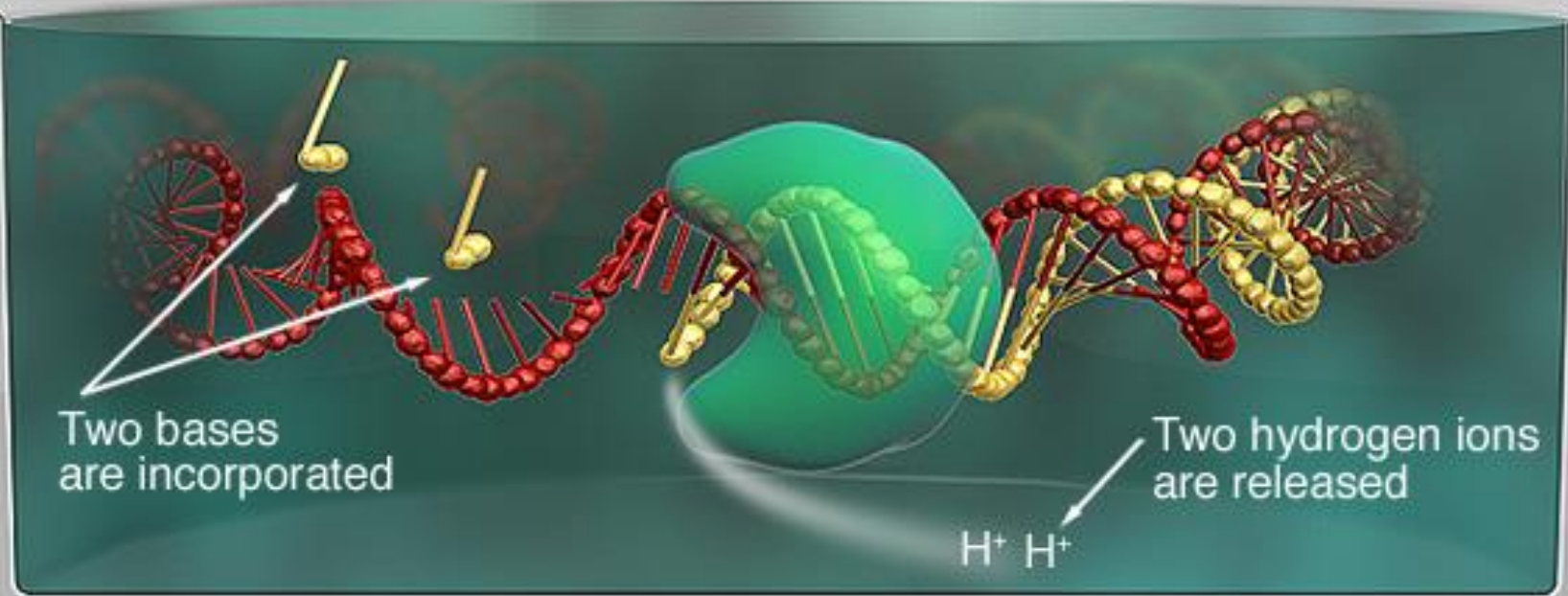
If a nucleotide is added to a DNA template and is then incorporated into a strand of DNA, a hydrogen ion is released. The charge from that ion will change the pH of the solution. The world's smallest solid-state pH meter—will call the base.



The sequencer sequentially floods the chip with one nucleotide after another. If the next nucleotide that floods the chip is not a match, no voltage change will be recorded.



If there are two identical bases on the DNA strand, the voltage is double, and the chip records two identical bases.



# Ion Torrent

The Ion Personal Genome Machine (PGM™) sequencer

<http://www.iontorrent.com/>

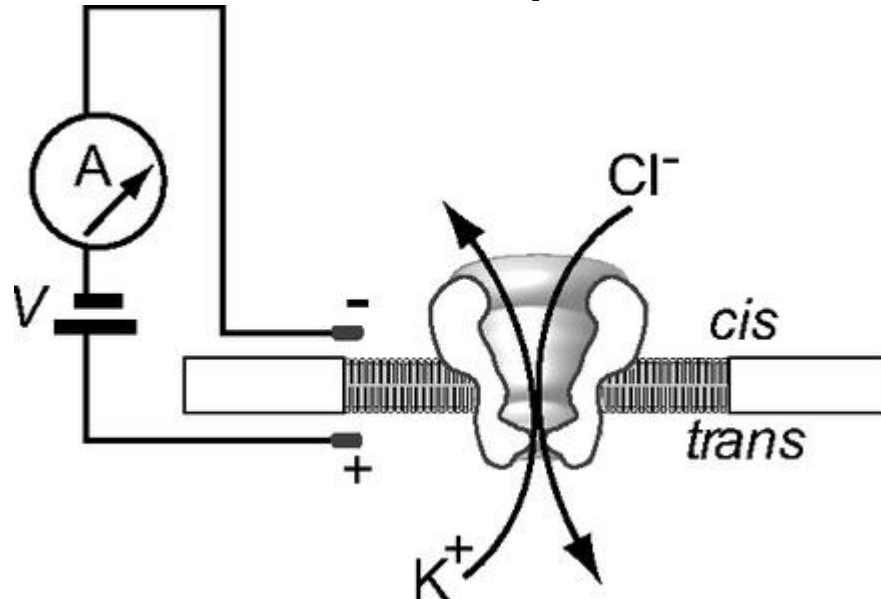
- ❖ Different templates in microwells
- ❖ Washing steps by individual nucleotides G, C, T, A
- ❖ The world's smallest solid-state pH meter
- ❖ Digital output

**Single molecule passage  
through a pore**

# Oxford Nanopore Technologies



Schematic of the nanopore device.



# DNA sequencing development

2001: Genome draft of 5 individuals in 9 months  
– more than billion \$

2015: Complete human genome in an hour – ~100 \$