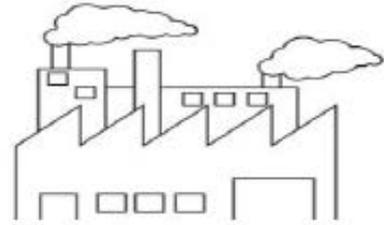


# Current issues in ecotoxicology research

**Luděk Bláha** ([blaha@sci.muni.cz](mailto:blaha@sci.muni.cz))

RECETOX PŘF MU

# PHARMACEUTICALS



R&D and Manufacturing

Storage ↓ Transport



Distribution

Storage ↓ Transport



Consumption

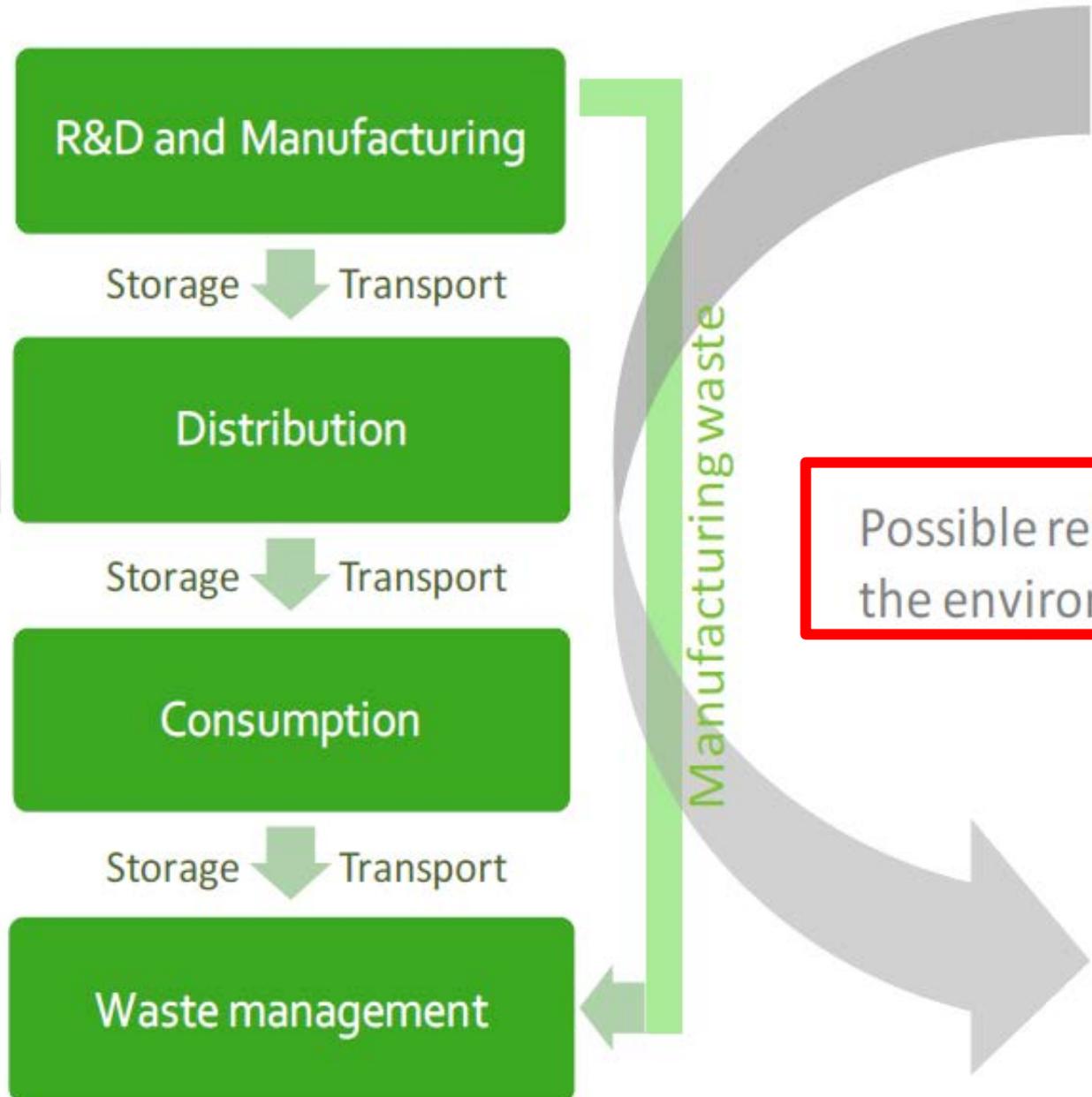
Storage ↓ Transport



Waste management

Manufacturing waste

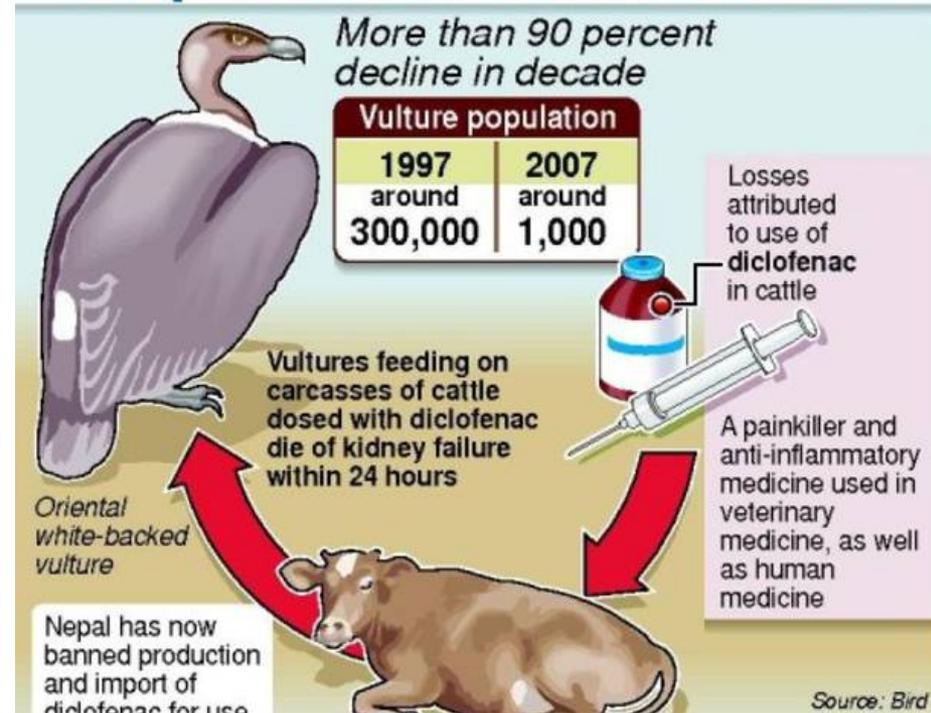
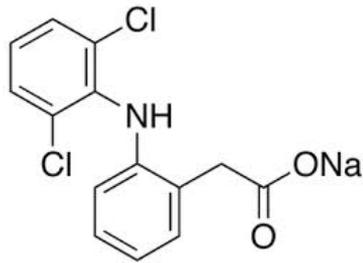
Possible releases to the environment



# Example 1 - DICLOFENAC

## Unexpected effects at NON-TARGET species

- **nephrotoxicity** at vultures
- Relevant also in EU (ESP, EL, CY)



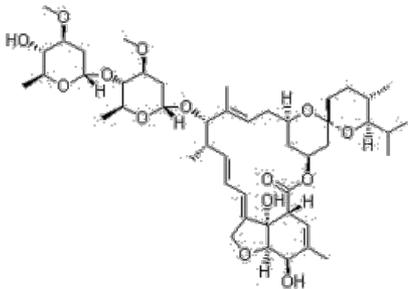
# Example 2 – AVERMECTIN-like antiparasitics

**Moxidectin** – used e.g. in home „spot on” products



**Ivermectin** – antiparasitics in large herds

- Used **2-times per season** per sheep/cow
- **Kills 100% parasites** in sheep
- Released in dung - **kills 80-90% larvae of dung flies**
- High concentrations in dung (released 2 days post application)
- **Persistent in the soil** (half-life 30 days)
- Can be washed into adjacent streams (highly toxic to water insects)



# MIXTURE EFFECTS ... VERY LOW CONCENTRATIONS

## International ring test (2012-13)

Testing comparability of existing and innovative bioassays for water quality assessment

### Main questions:

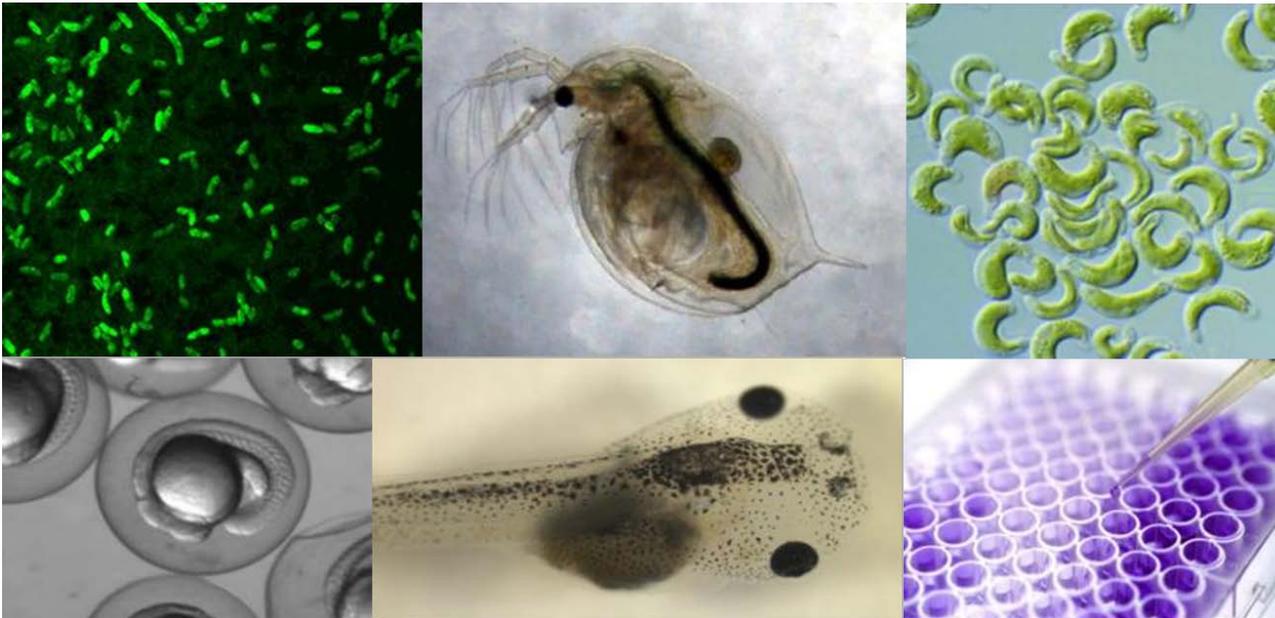
Are current limits (for individual compounds) safe?

Relevance of “**Something from Nothing**” phenomenon ?

### 3 samples

→ 12 European laboratories – different bioassays

→ ČR – RECETOX: 11 bioassays



Carvalho, R. et al. (2014) Mixtures of chemical pollutants at European legislation safety concentrations: how safe are they? *Toxicol Sci* **141(1): 218-233**

# MIXTURE EFFECTS ... VERY LOW CONCENTRATIONS

EU WFD  
priority  
substances

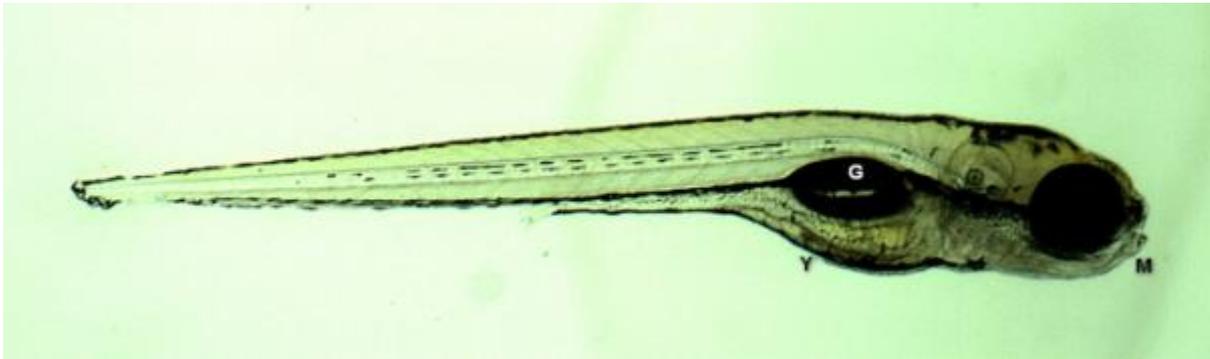
Different  
concentrations

EQS  
= limit  
(Environmental  
Quality  
Standard)

	RM 1 <sup>a</sup>	RM 2 <sup>a</sup>	RM 3 <sup>a</sup>
<i>Priority substances</i> mg/L	around <u>or</u> >EQS	< EQS	< EQS
<b>Atrazine</b>	6	0.6	0.6
<b>BaP</b>	0.0017	0.00017	0.00017
<b>Cadmium<sup>b</sup></b>	0.8	0.08	0.08
<b>Chlorfenvinphos</b>	1	0.1	0.1
<b>Chlorpyrifos</b>	0.3	0.03	0.03
<b>DEHP (Bis(2-ethylhexyl) phthalate)</b>	13	1.3	1.3
<b>Diclofenac</b>	1	0.1	0.1
<b>diuron</b>	2	0.2	0.2
<b>17beta-estradiol</b>	0.004	0.0004	0.0004
<b>fluoranthene</b>	0.063	0.0063	0.0063
<b>Isoproturon</b>	3	0.3	0.3
<b>Ni<sup>b</sup></b>	40	4	4
<b>4-Nonylphenol</b>	3	0.3	0.3
<b>Simazine</b>	10	1	1
<b>Carbamazepine</b>	-	-	0.5
<b>Sulfamethoxazole</b>	-	-	0.6
<b>Triclosan (Irgasan)</b>	-	-	0.02
<b>DEET</b>	-	-	41
<b>Bisphenol A</b>	-	-	1.5

# MIXTURE EFFECTS ... VERY LOW CONCENTRATIONS

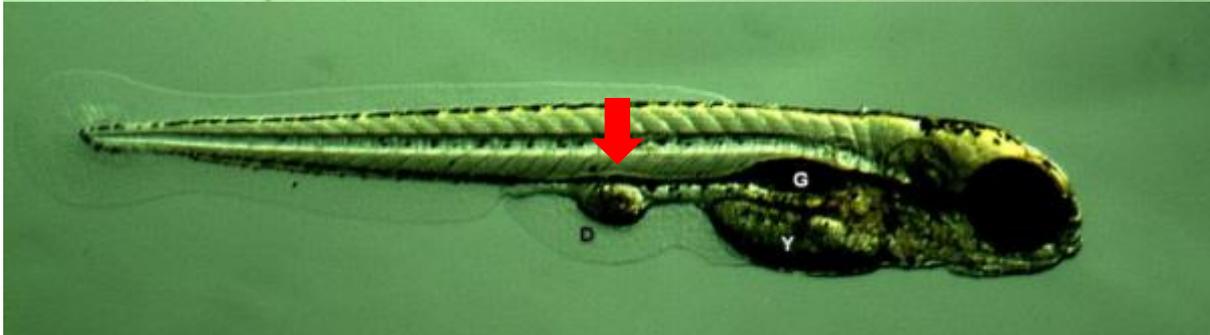
Example: Effects of mixtures on *D. rerio* fish embryos



Control



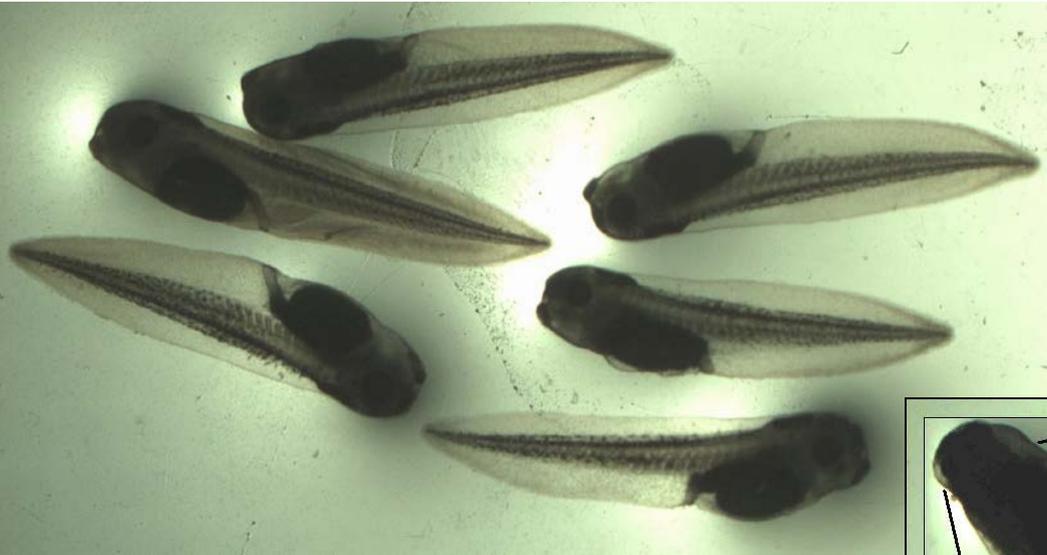
Effects of RM 3 (i.e. safe) mixtures



Carvalho, R. et al. (2014) Mixtures of chemical pollutants at European legislation safety concentrations: how safe are they? *Toxicol Sci* **141(1): 218-233**

# MIXTURE EFFECTS ... VERY LOW CONCENTRATIONS

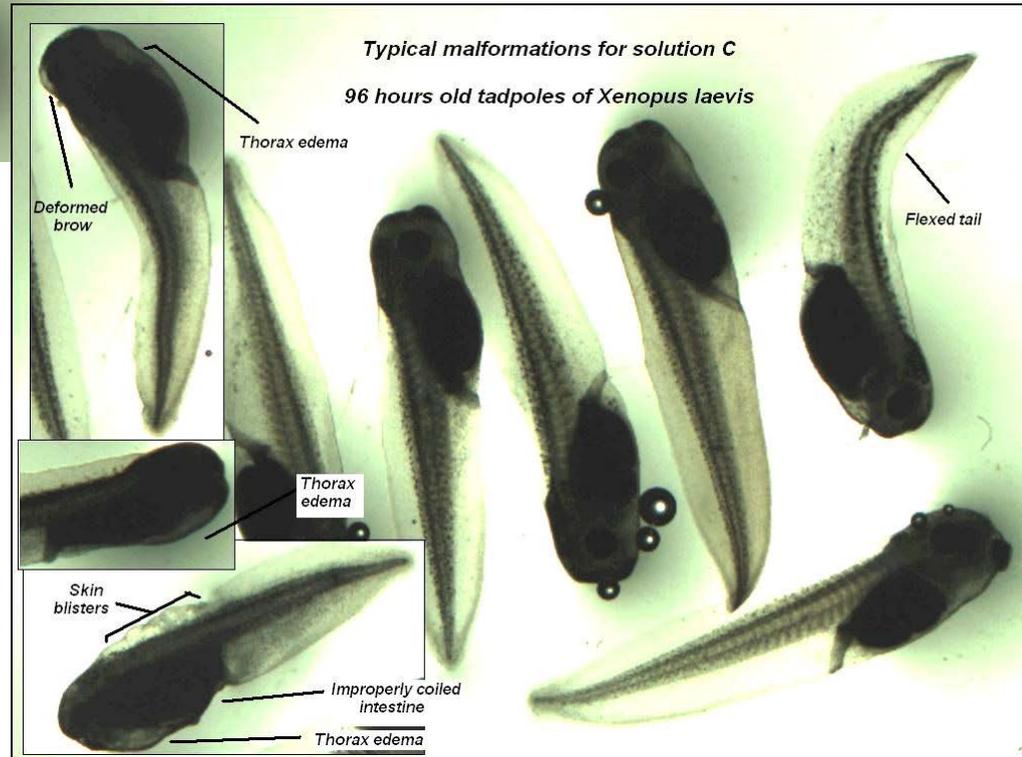
Example: Effects of mixtures on *X. laevis* frog embryos



Controls

Carvalho, R. et al. (2014) Mixtures of chemical pollutants at European legislation safety concentrations: how safe are they? *Toxicol Sci* **141(1): 218-233**

Effects of RM 3 (i.e. safe) mixtures

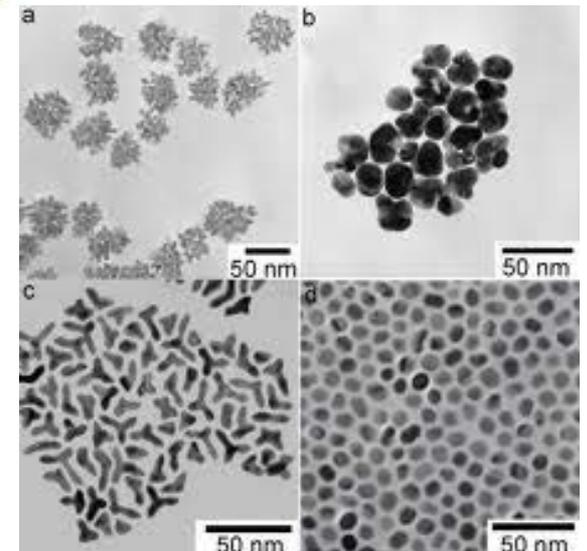
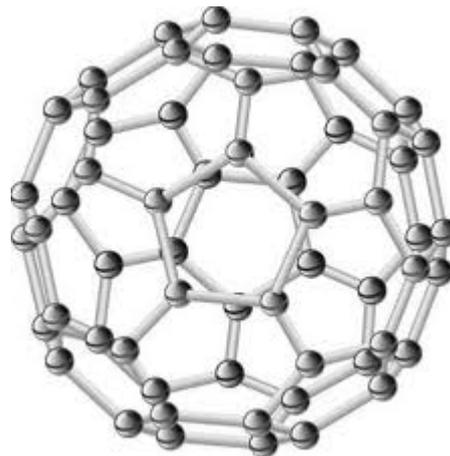
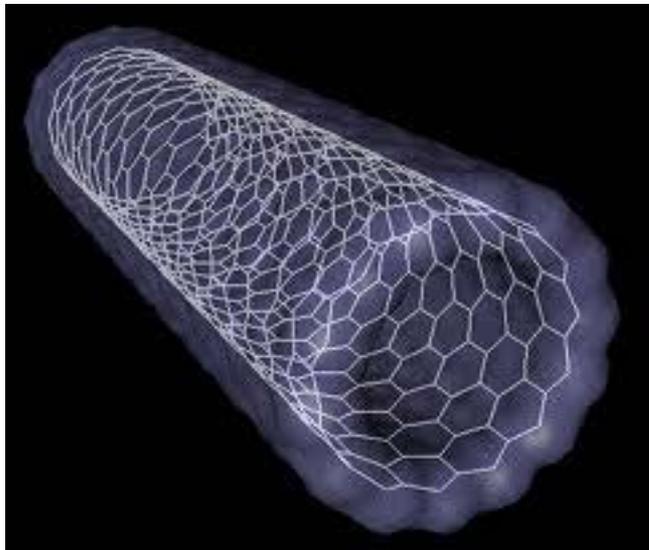
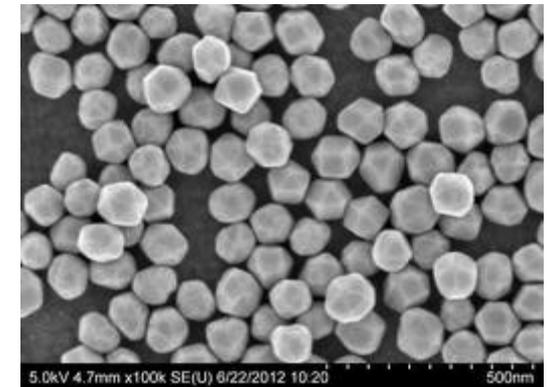
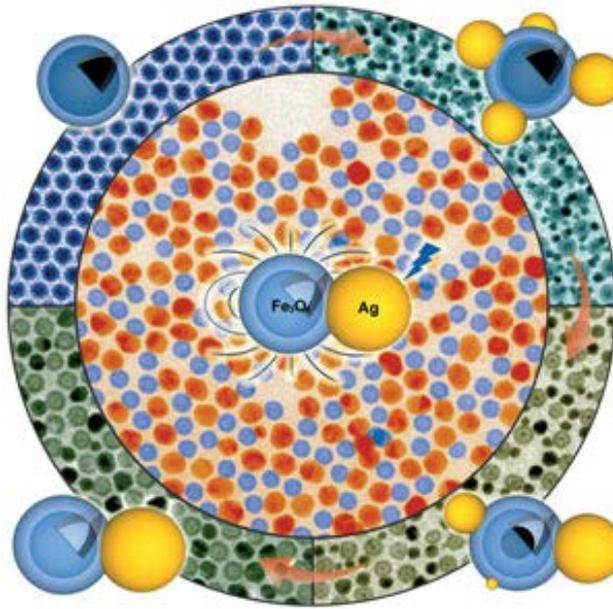
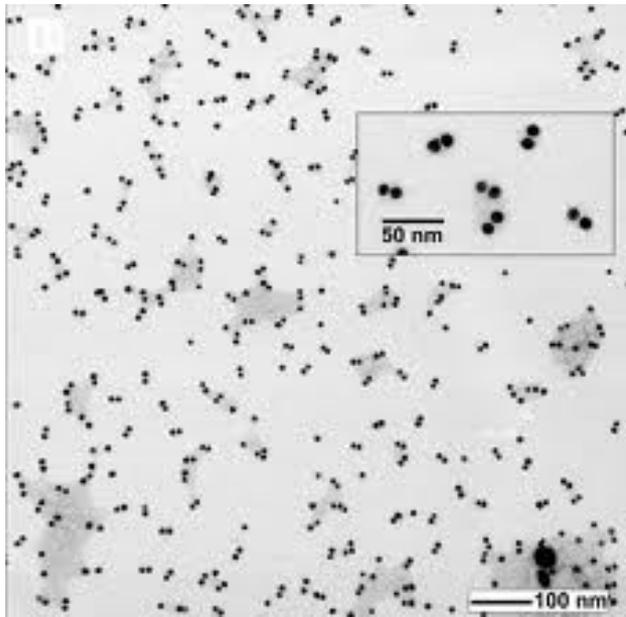


<b>Biotest</b>	<b>A</b>	<b>B</b>	<b>C</b>
<b>Microtox</b>	26 and 36% stimulation of luminescence in 15 and 30 mins of exposure, respectively	18 and 35% stimulation of luminescence in 15 and 30 mins of exposure, respectively	22 and 39% stimulation of luminescence in 15 and 30 mins of exposure, respectively
<b>Algae growth inhibition test 96-h exposure</b> 	31% inhibition of growth compared to solvent control	20% inhibition of growth compared to solvent control	16% inhibition of growth compared to solvent control
<b>Acute immobilization test with <i>D. magna</i></b>	90% immobilization after 48 hours of exposure; 25% immobilization occurred in 50% concentration - not statistically significant	no effect observed	no effect observed
<b>Reproduction test with <i>D. magna</i> (21-d exposure)</b>	100% mortality after 3 days of the test, no reproduction could be evaluated	31 +/- 37 % inhibition of reproduction, not statistically significant	23 +/- 24 % inhibition of reproduction, not statistically significant
<b>FETAX (96-h exposure)</b> 	62 +/- 10 % of malformed embryos; no effect on embryo length observed	43 +/- 12 % of malformed embryos; no effect on embryo length observed	34 +/- 14 % of malformed embryos; no effect on embryo length observed
<b>FET (120-h exposure)</b>	effects observed in number of defected embryos - absence of gas bladder, (head) deformities and underdeveloped embryos were observed the most often. 	no significant effects observed	effects observed in number of defected embryos, number of underdeveloped embryos and length 
<b>In vitro - cytotoxicity</b>	no effect observed compared to solvent control	no effect observed compared to solvent control	no effect observed compared to solvent control
<b>In vitro - estrogenicity</b>	effect under LOQ	effect under LOQ	effect under LOQ
<b>In vitro - dioxin-like toxicity</b>	effect under LOQ	effect under LOQ	effect under LOQ
<b>In vitro - androgenicity</b>	effect under LOQ	effect under LOQ	effect under LOQ
<b>In vitro - antiandrogenicity</b>	effect under LOQ	effect under LOQ	effect under LOQ

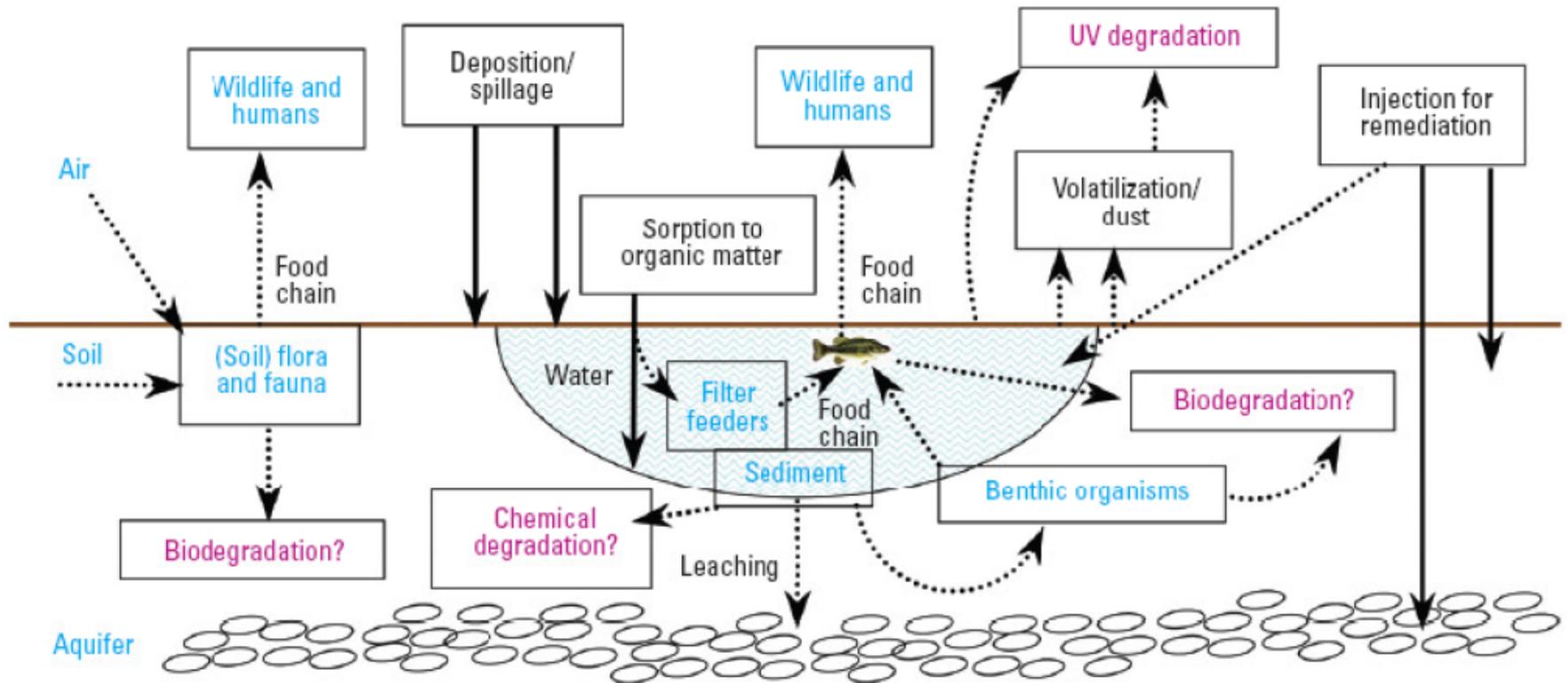
**Kde „tradiční“ ekotoxikologie  
nestačí**

**Nano-eco-toxicology**

# Nanoparticles - examples

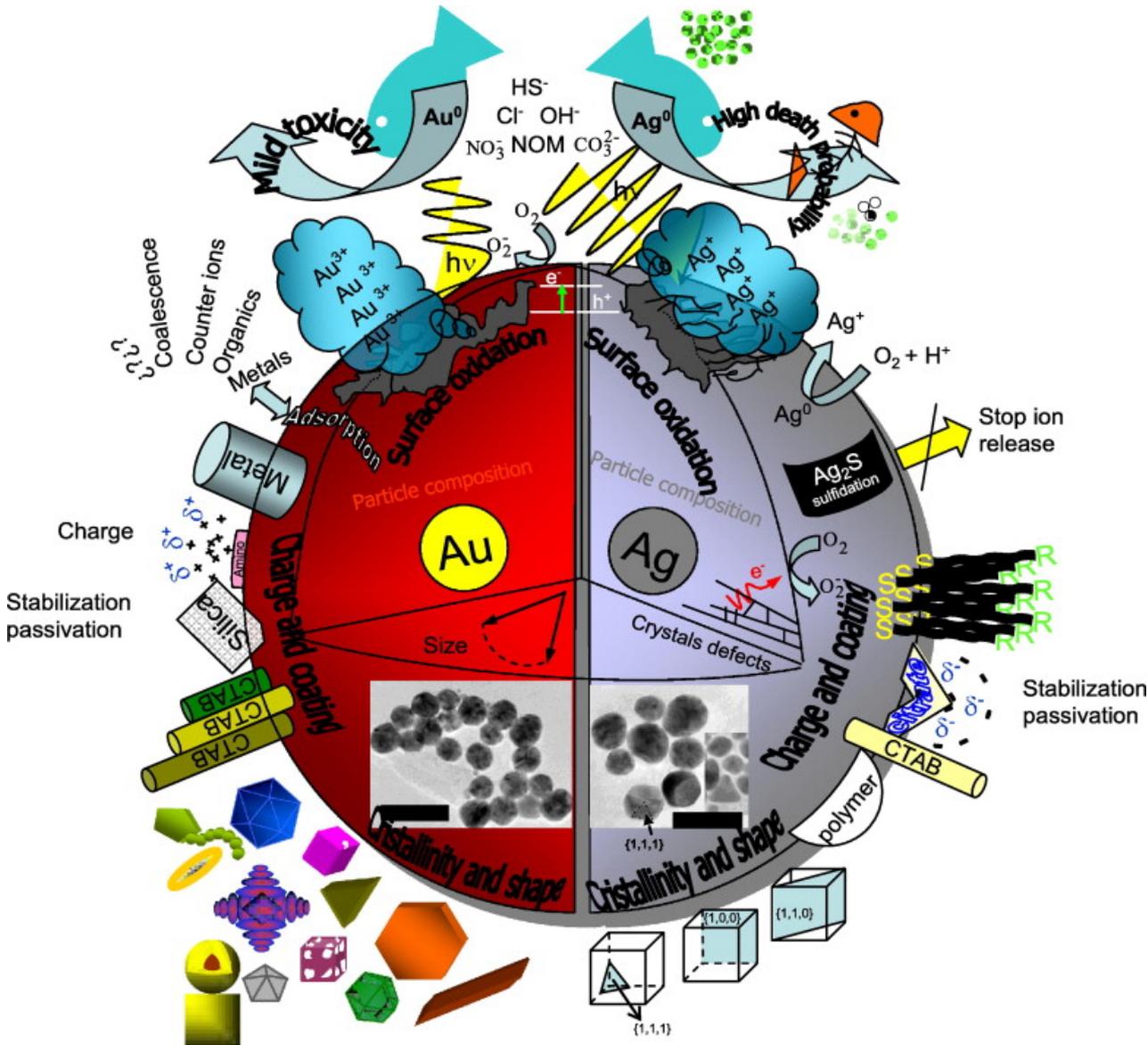


# Nanoparticle movement through the environment



**Figure 5.** Routes of exposure, uptake, distribution, and degradation of NSPs in the environment. Solid lines indicate routes that have been demonstrated in the laboratory or field or that are currently in use (remediation). Magenta lettering indicates possible degradation routes, and blue lettering indicates possible sinks and sources of NSPs.

# Ecotoxicity of nanoparticles ...



(Mostly unknown)  
Parameters may  
Affect ecotoxicity

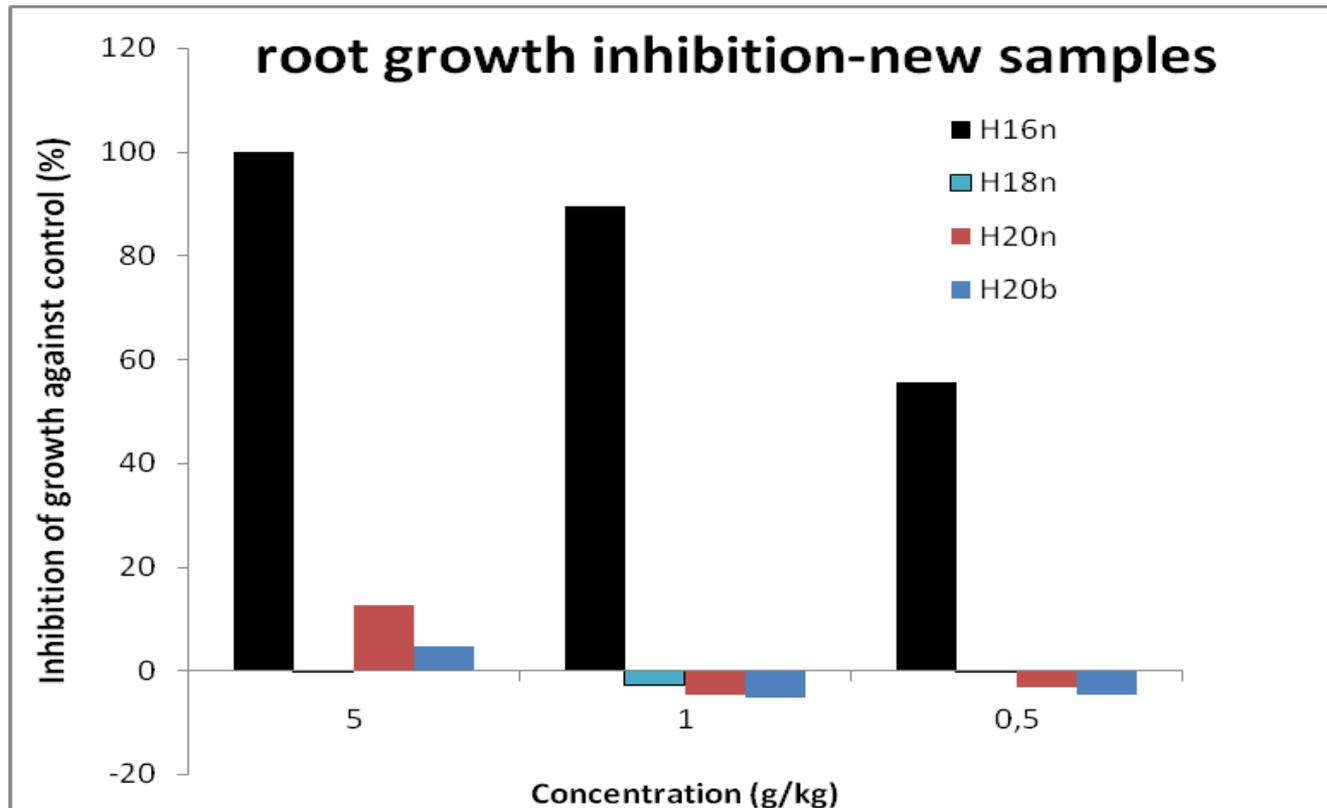
Composition (chemical)  
Surface (size, area)  
Charge  
Reactivity  
Interactions with ions,  
other chemicals...

→ Effects on  
environmental Fate  
and toxicity

# Ecotoxicity of nanoparticles – RECETOX example

Comparison of toxicity - 4 „appeared to be the same“ particles  
(one producer – 4 different lots)  
(zerovalent iron – ZVI – Fe<sup>0</sup>)

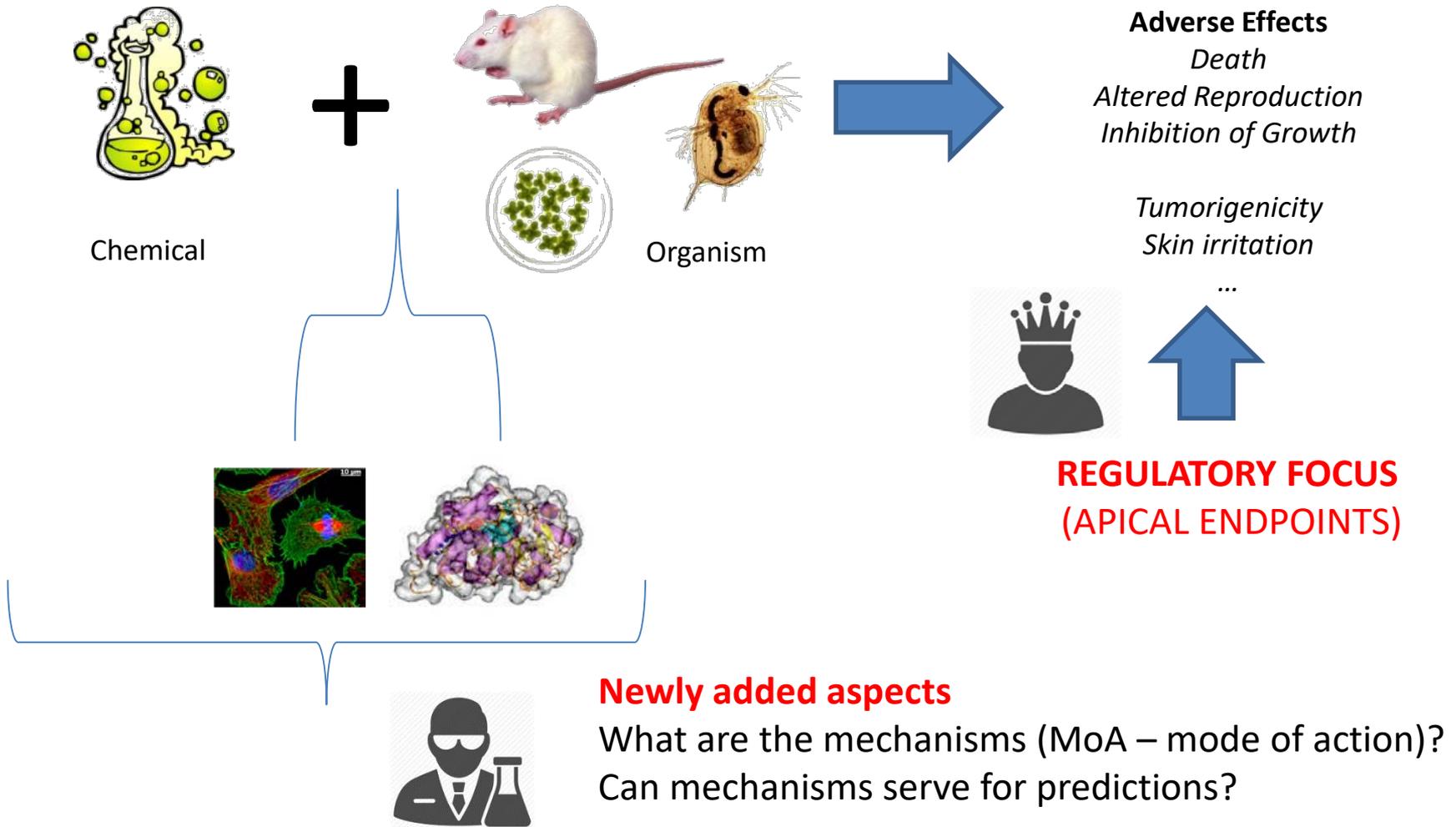
?? *Why is H16 so toxic ??*



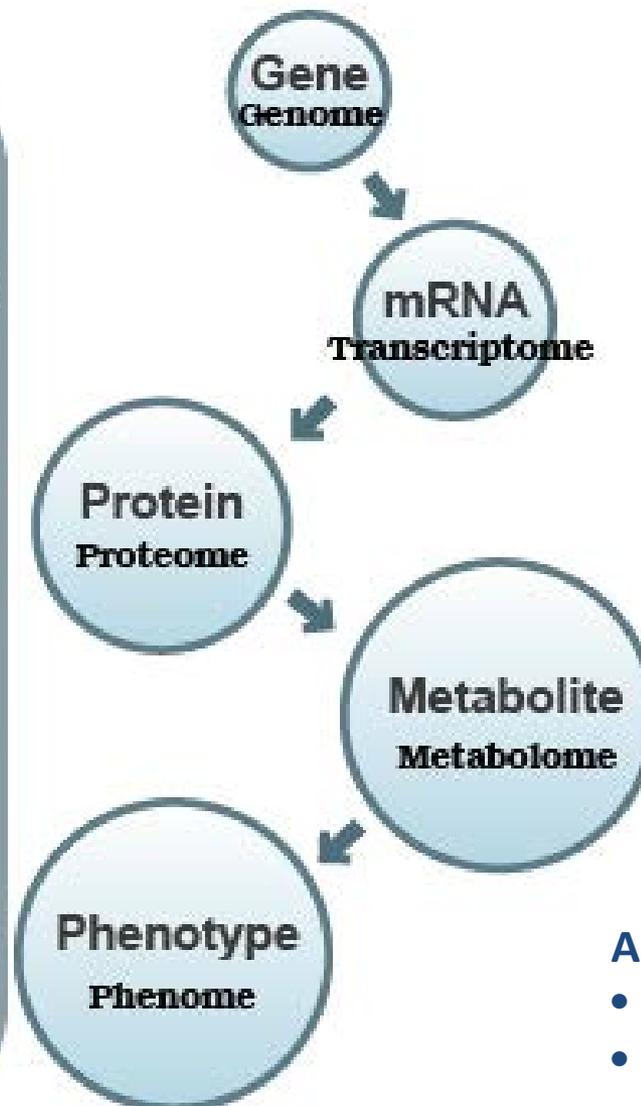
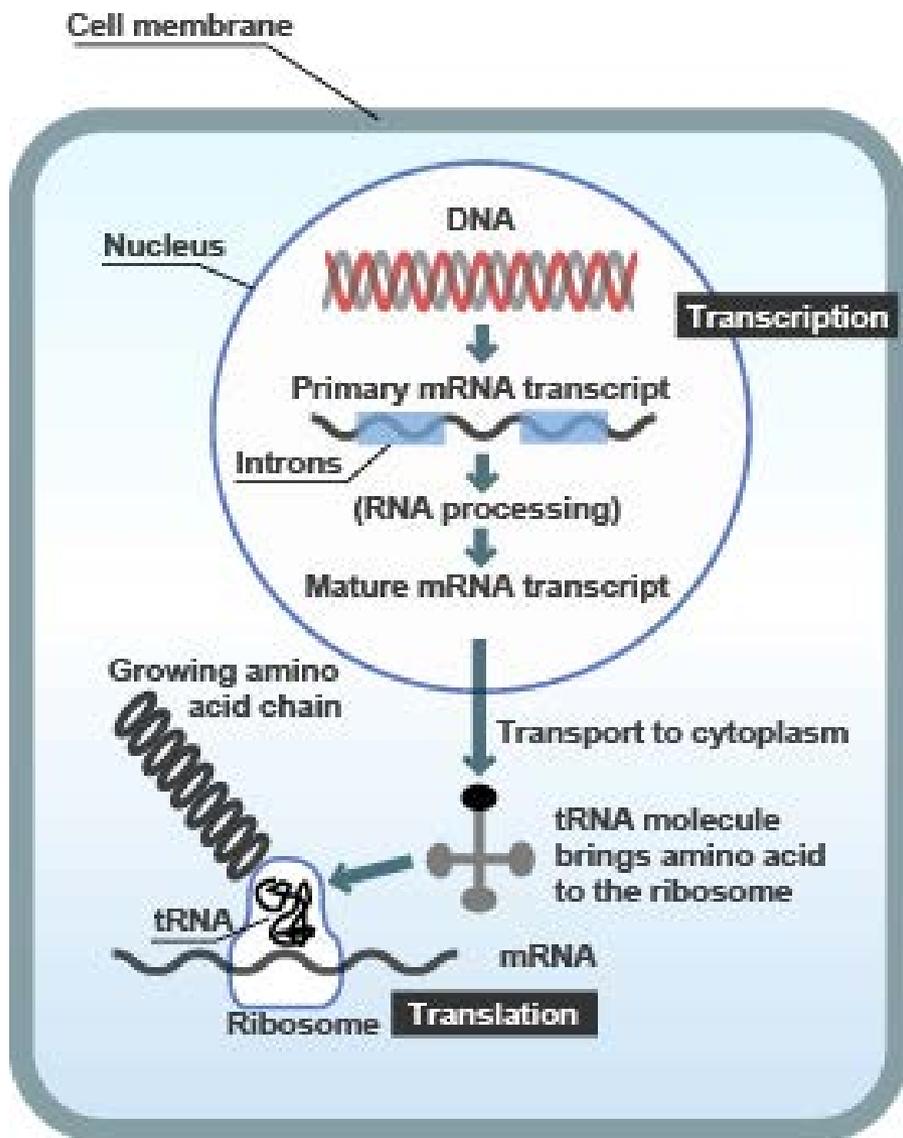
# **Mechanistic and Computational (ECO)TOXICOLOGY**

# Hazard assessment

**Traditionally** – Evaluation of adverse effects using the whole organism models



# Extrémní rozvoj analytických technologií → „OMICS“



A další „ómy“

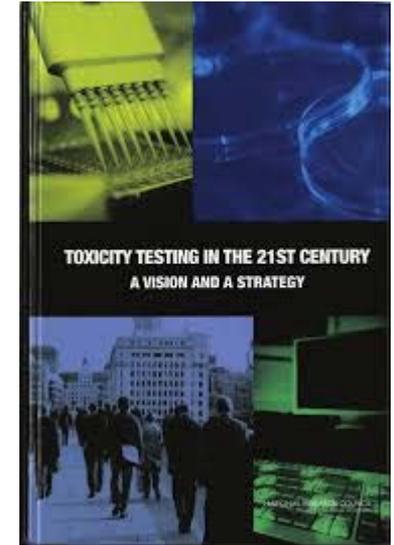
- Lipidóm
- Mikrobiom ...

# Sběr omics podporují strategické dokumenty & projekty

## Toxicity Testing in the 21st Century: A Vision and a Strategy

US National Academies of Sciences

<http://www.nap.edu/catalog/11970.html>



LEARN THE ISSUES | SCIENCE & TECHNOLOGY | LAWS & REGULATIONS | ABOUT EPA

### Computational Toxicology Research

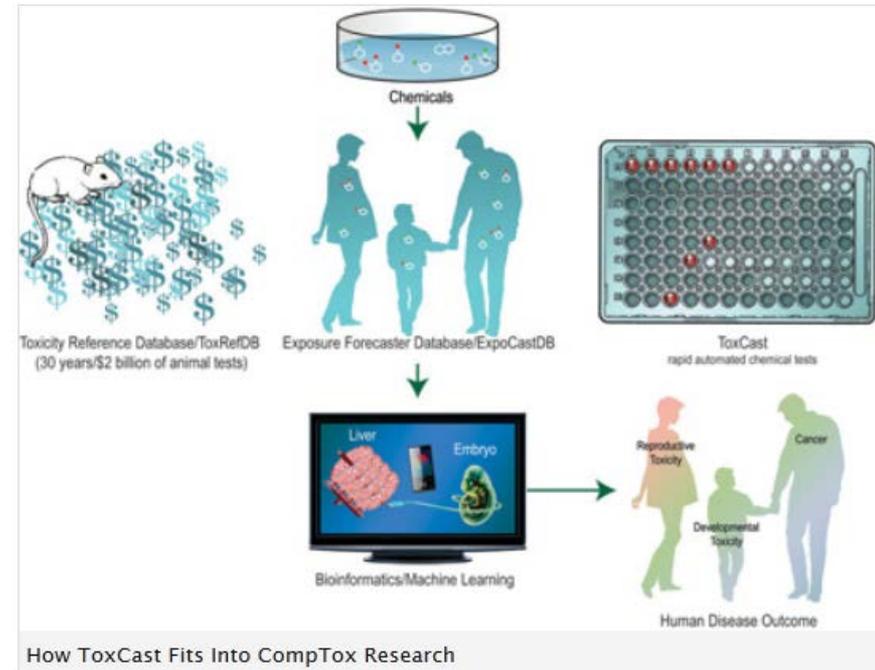
You are here: [EPA Home](#) » [Research & Development](#) » [CompTox](#) » [ToxCast™](#)

#### Key Links

<a href="#">CompTox Home</a>	<a href="#">Research Projects</a>	R
<a href="#">Basic Information</a>	<a href="#">Chemical Databases</a>	S
<a href="#">Organization</a>	<a href="#">CompTox Events</a>	C

## ToxCast™

Screening Chemicals to Predict Toxicity Faster and Better

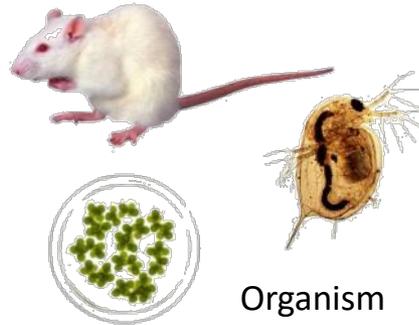


# Hazard assessment

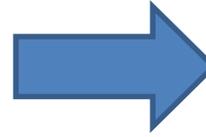
**Traditionally** – Evaluation of adverse effects using the whole organism models



Chemical

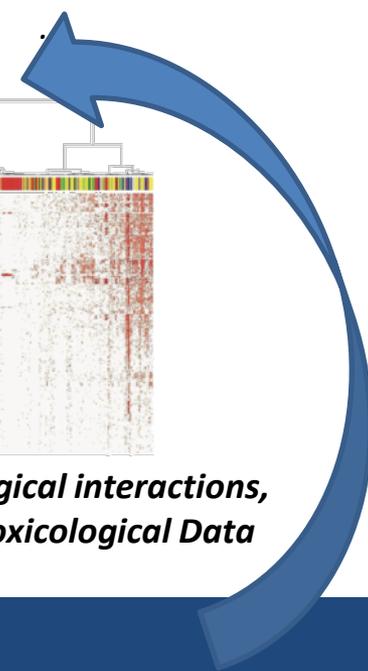


Organism

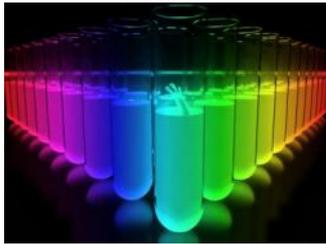


**Adverse Effects (EC50)**

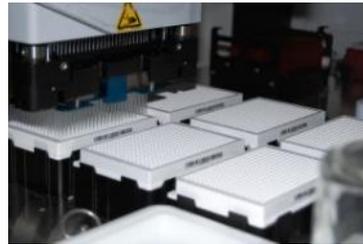
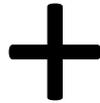
*Death*  
*Inhibition of Growth*  
*Altered Reproduction*  
*Tumor*  
*Skin irritation*



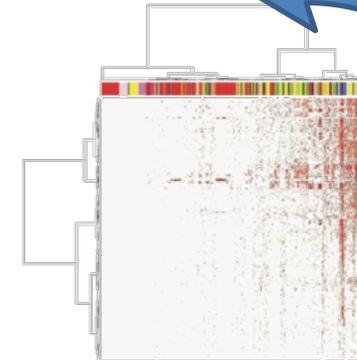
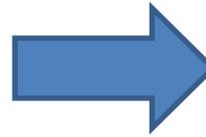
**New** – Ex vivo / in vitro / In chemico / In silico Methods



10<sup>4</sup> Chemicals



HTS



***Chemical-biological interactions,***  
***Mechanistic Toxicological Data***

**Key task/question:**

How to link MECHANISTIC INFORMATION with APICAL ENDPOINTS ?

# Kvantitativní mechanistické modelování



 OPEN ACCESS  PEER-REVIEWED

RESEARCH ARTICLE

## A Computational Model of the Rainbow Trout Hypothalamus-Pituitary-Ovary-Liver Axis

Kendall Gillies, Stephen M. Krone, James J. Nagler, Irvin R. Schultz 

Published: April 20, 2016 • <https://doi.org/10.1371/journal.pcbi.1004874>

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### Abstract

Reproduction in fishes and other vertebrates represents the timely coordination of many endocrine factors that culminate in the production of mature, viable gametes. In recent years

 Check for updates

**Subject Areas**

# Kvantitativní mechanistické modelování

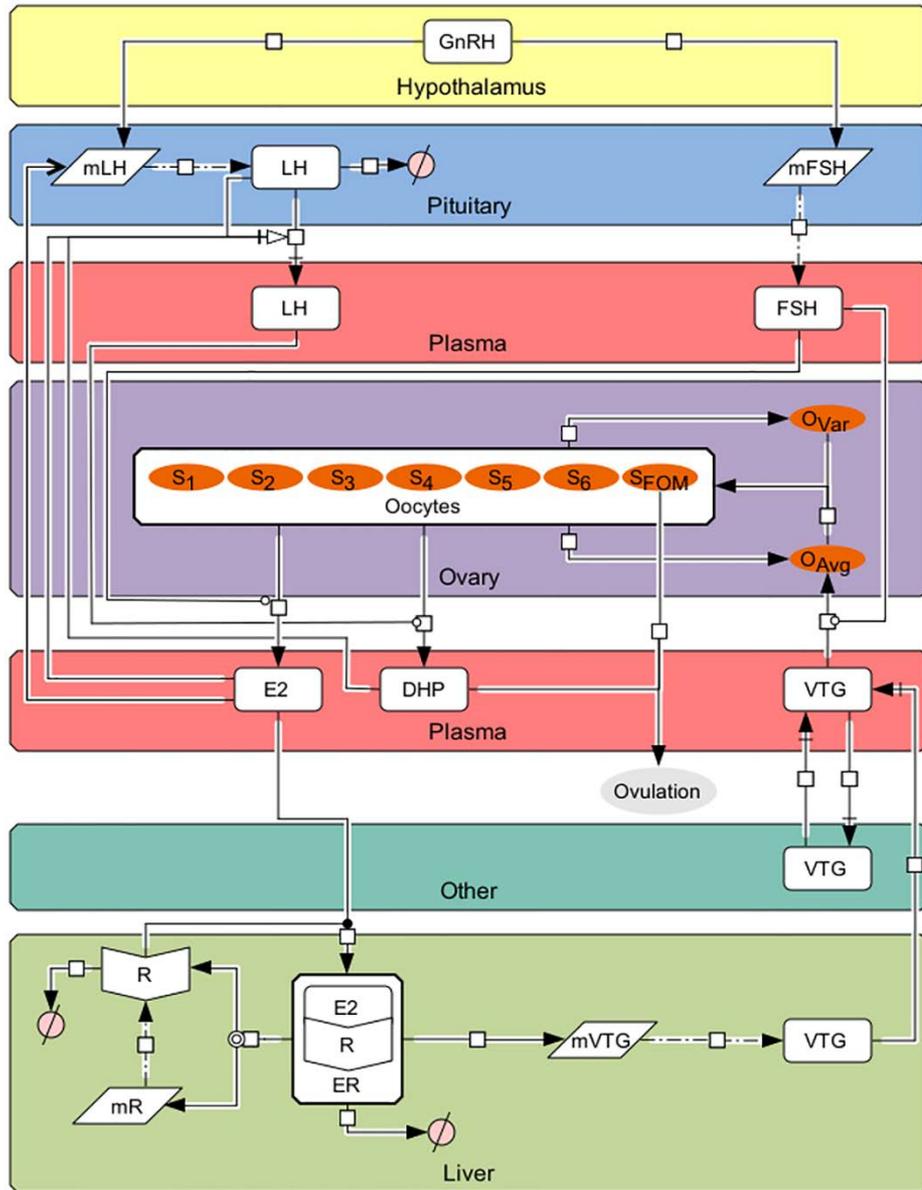


Fig 1. The HPOL signaling network in rainbow trout as formulated in our model.

Arrows and symbols on graph follow CellDesigner vs. 4.4 notation ([www.cellDesigner.org](http://www.cellDesigner.org)). GnRH is secreted from the hypothalamus into the pituitary stimulating the production of mFSH and mLH, which then leads to formation of FSH and LH, respectively. FSH, which is being continuously secreted from the pituitary, travels to the ovaries to stimulate production of E2. E2 then travels to the liver to bind with E2 receptors (R; translated from mR) to form ER. ER then stimulates the production of mVTG, which produces VTG<sub>L</sub>. Secreted VTG then travels from the liver to the ovaries via the plasma (VTG<sub>p</sub>) where it is absorbed by follicles in stages 3 through 6 (the proportion of follicles in these stages are denoted by S<sub>j</sub>, j = 3, 4, 5, and 6) during vitellogenesis, the rate of which is affected by FSH<sub>p</sub> to promote oocyte growth (O<sub>Avg</sub>). Oocyte growth then progresses the oocytes through the stages using a Weibull distribution created from O<sub>Avg</sub> together with O<sub>Var</sub>. In the later stages LH<sub>p</sub> stimulates the oocytes to produce DHP. Finally, oocytes undergo final maturation (S<sub>FOM</sub>) and combined with DHP, determine when the fish ovulates

# Kvantitativní mechanistické modelování

Fig 3. HPOL **model predictions** for (A) pituitary levels of FSH $_{\beta}$  subunit mRNA, (B) pituitary levels of LH $_{\beta}$  subunit mRNA, (C) Hepatic levels of E2 receptor mRNA and (D) Hepatic levels of VTG mRNA  
**Observed data** (dark grey circles; mean  $\pm$  TG mRn = 3)

