Confocal Microscopy and Living Cell Studies

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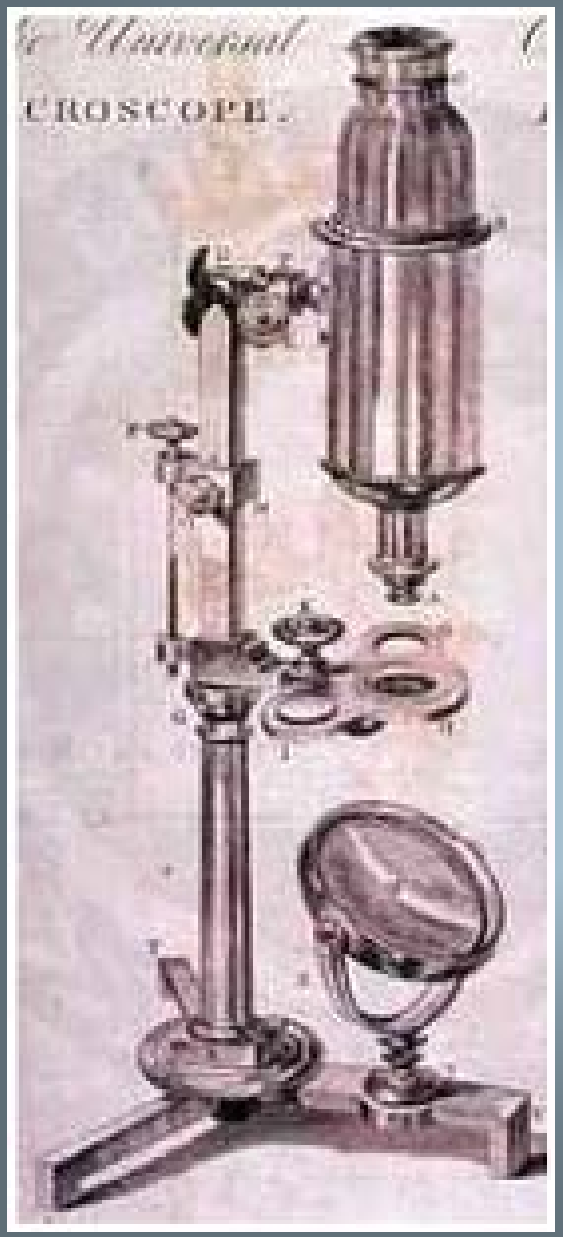
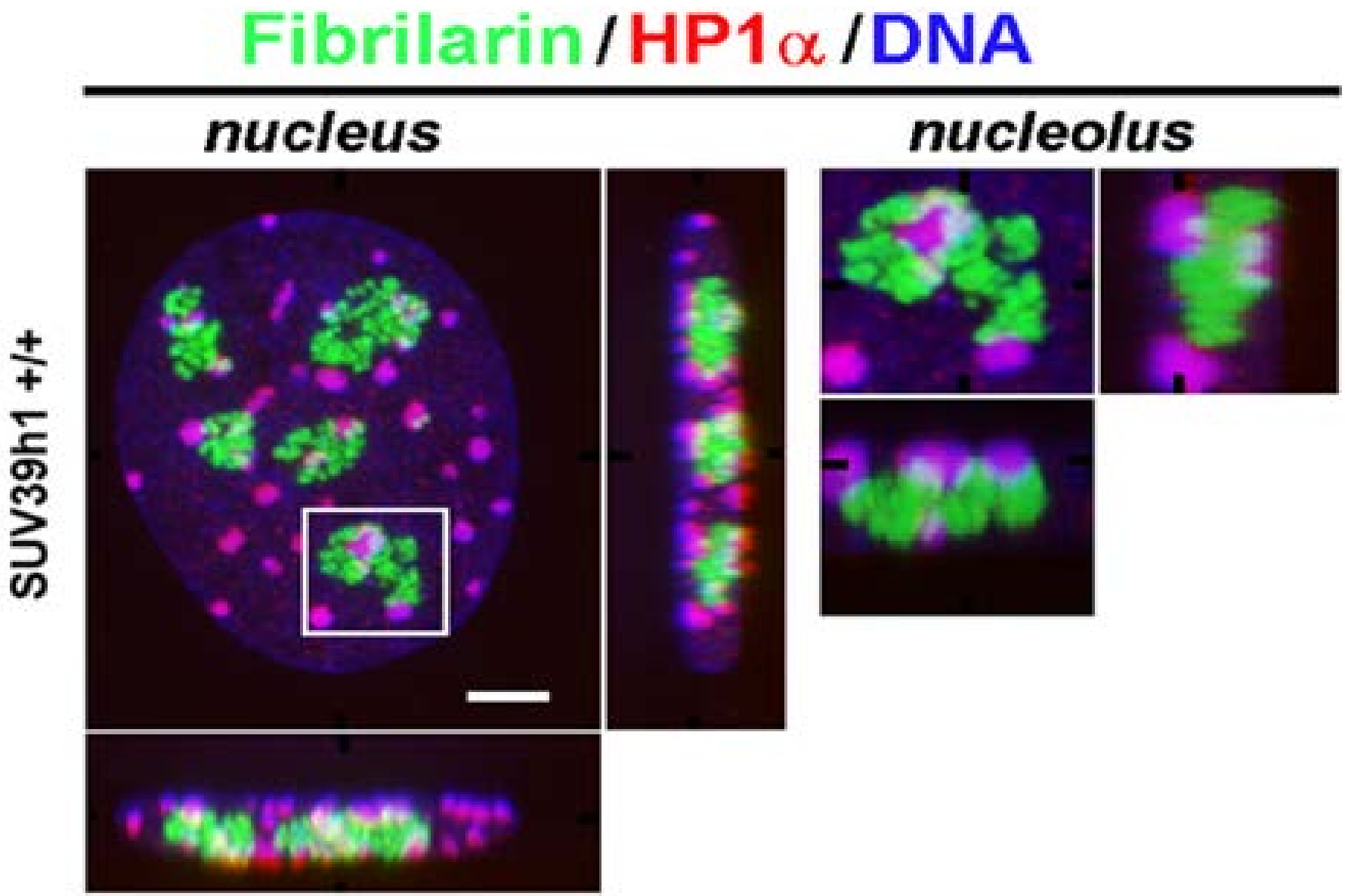
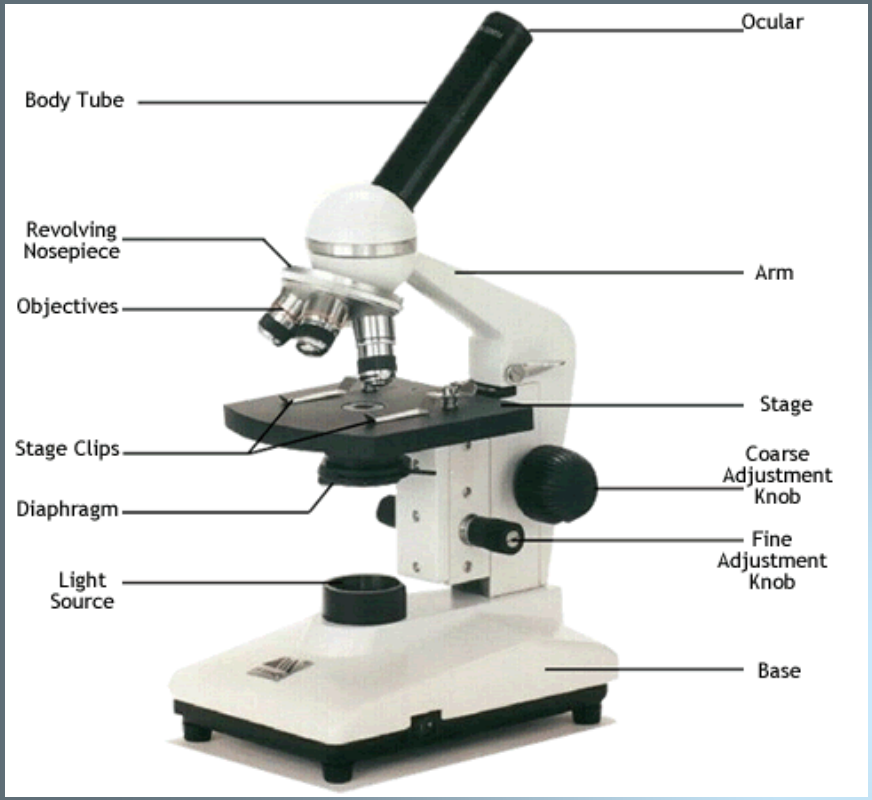


History of microscopy:

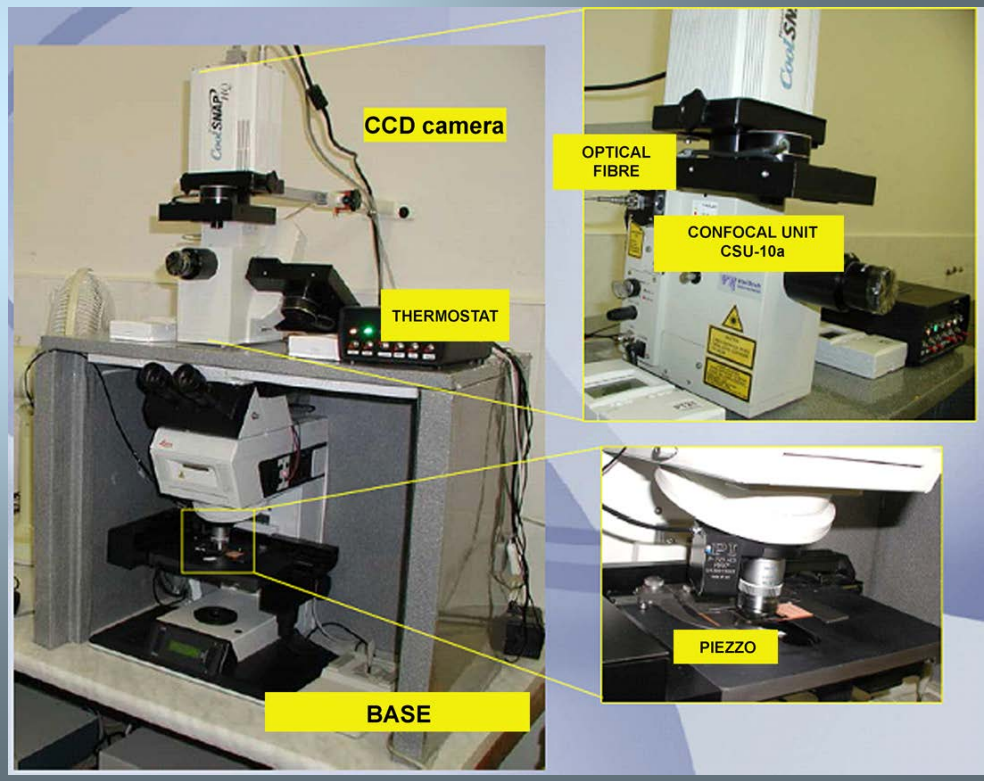
the light microscope (discovered by Robert Hook, 1665): an instrument that enables the human eye, by means of a lens or combinations of lenses, to observe enlarged images of tiny objects. It made visible the fascinating details of naked eye invisible worlds.

1957: Marvin Minsky patented the fist confocal microscope


Fibrilarin / HP1α / DNA
nucleus
nucleolus
SUV39h1 +/+



http://www.google.cz/imgres?imgurl=http://www.microscopehelp.com/



Brno laboratory at IBP

Visible Light Microscopy: Objectives: numerical aperture

- NA=ability of lens to gather light and resolve detail at a fixed distance from object.
  - Dependent on ability of lens to capture diffracted light rays.
- n=Refractive index is limiting (air=1.0, oil=1.51)
  - Do not mix mediums when using a lens
- Theoretical resolution depends on NA and the wavelength of light. NA=n·sin(μ)
  - Shorter wavelengths=higher resolution.
  - Resolution limit for green light (NA=1.4, 100X) is 0.2 μm.
    - R=0.61λ/NA

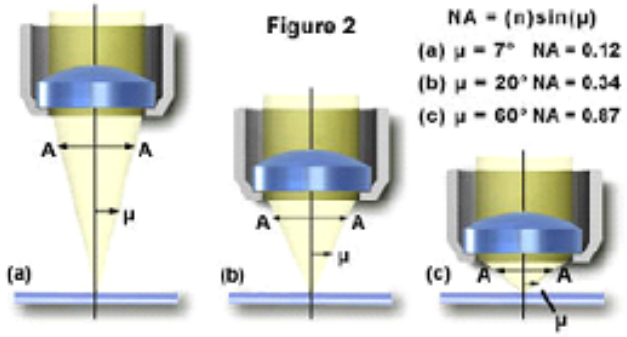
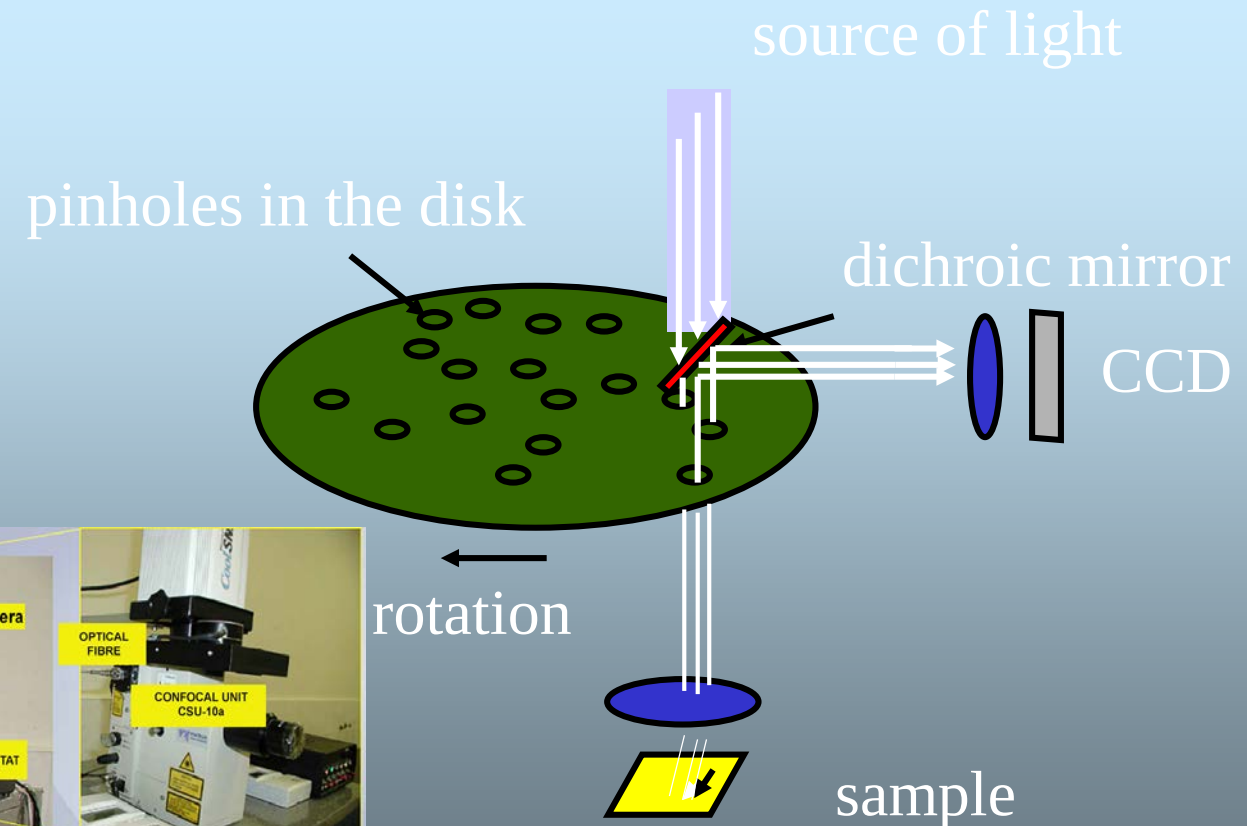
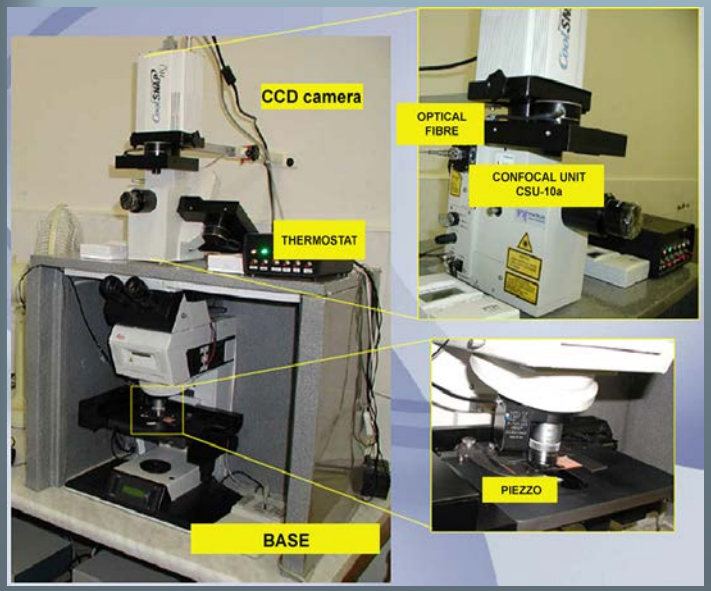


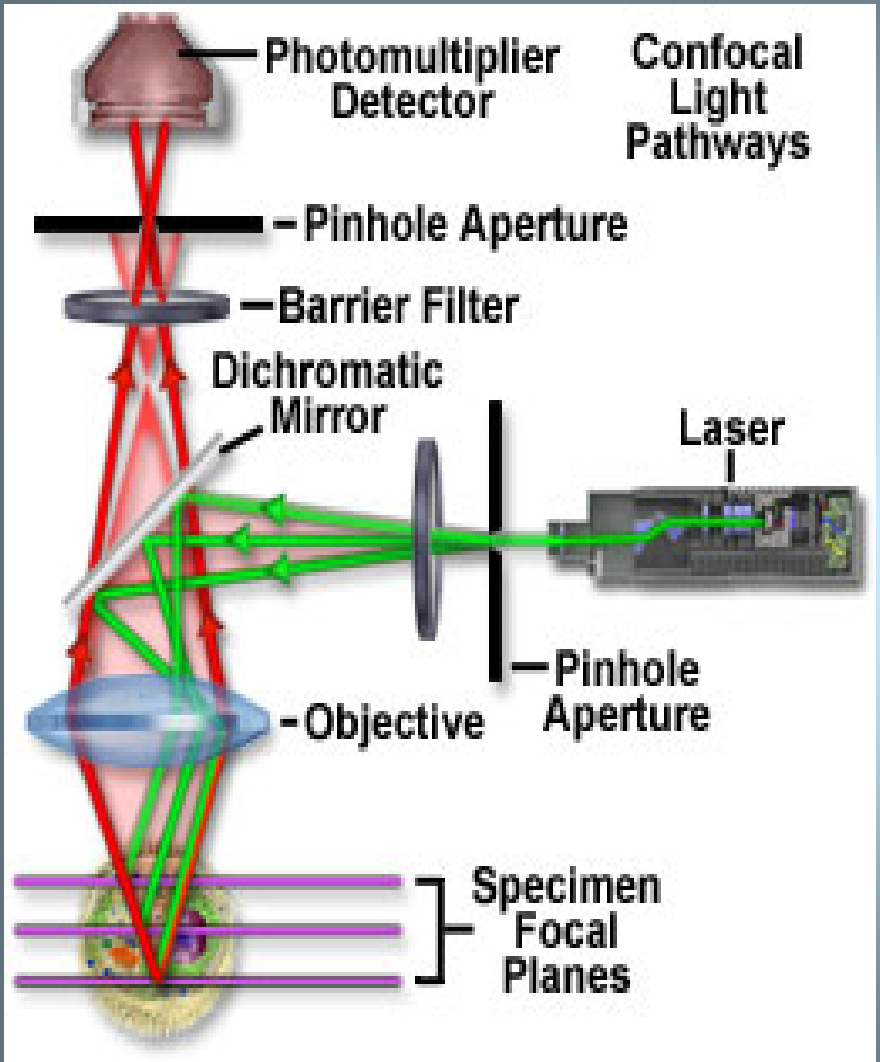
Figure 1

Tandem scanning microscopes based on Nipkow disk

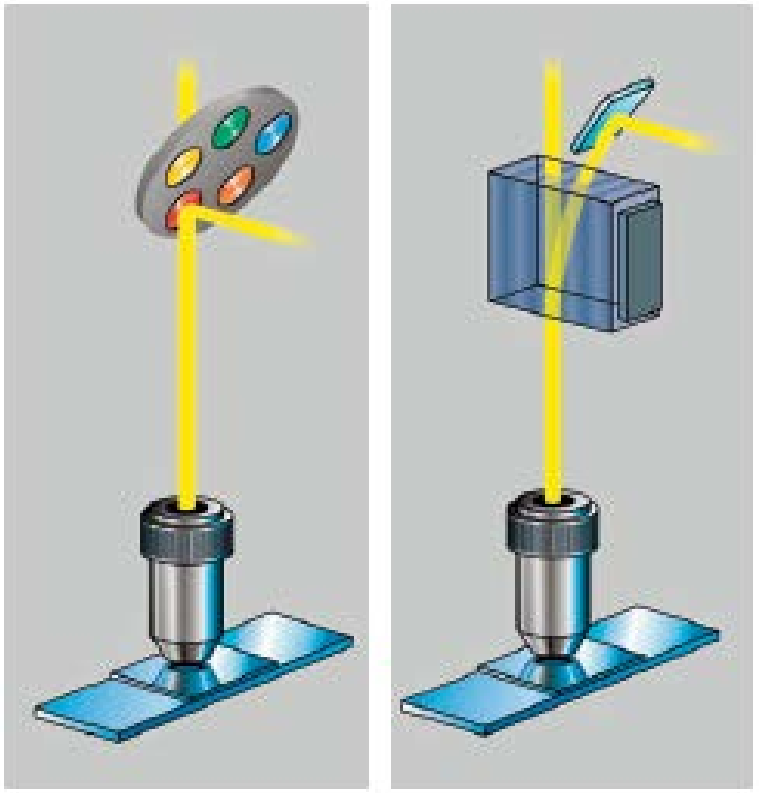




Confocal microscopy - principles

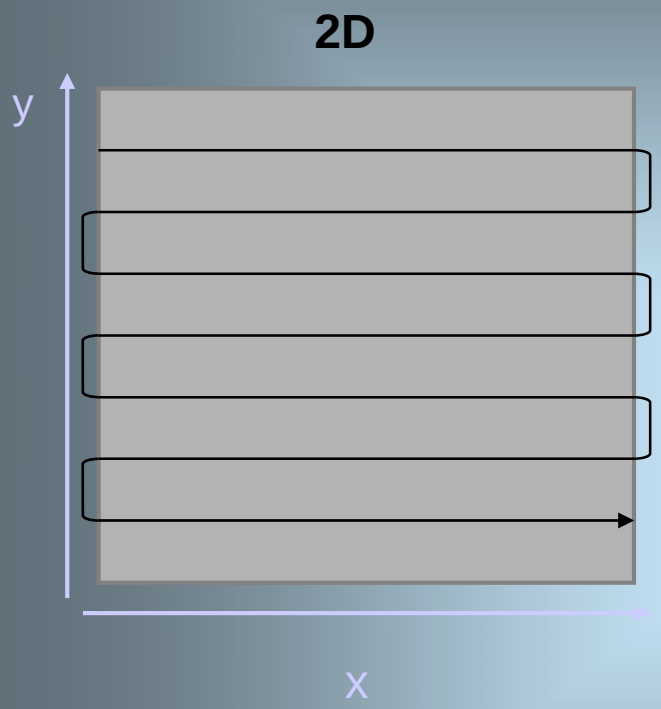


AOTF and AOBS

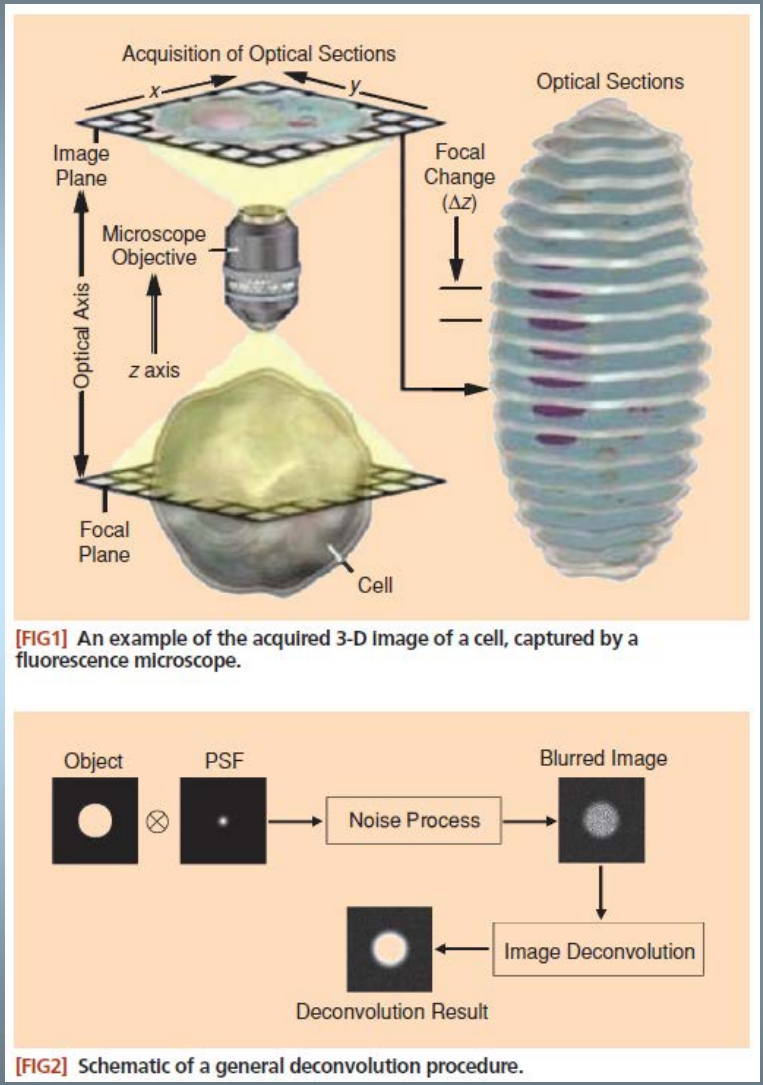


Left: conventional beam splitting by dichroic mirrors requires many optical elements with fixed properties.
Right: the AOBS® is electronically adaptable to all tasks.

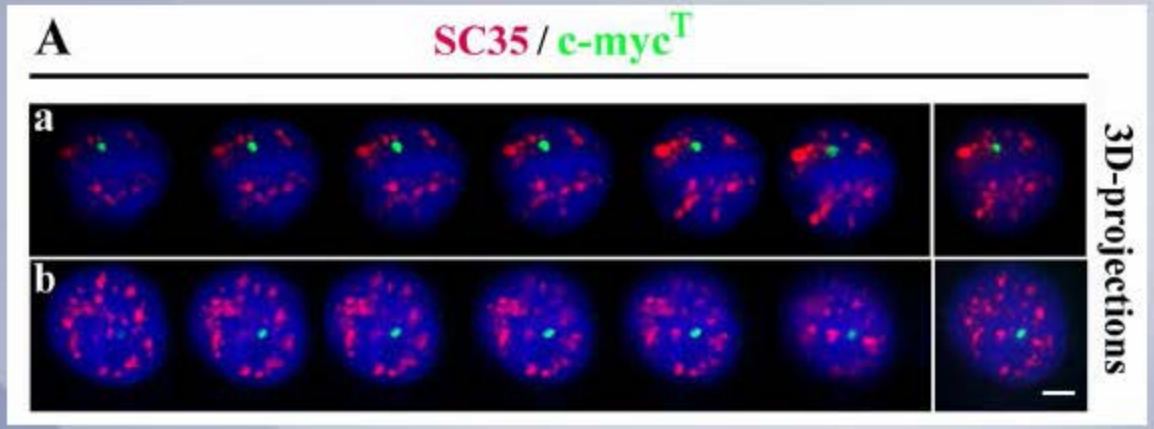
Scanning in 2D and 3D by confocal microscope

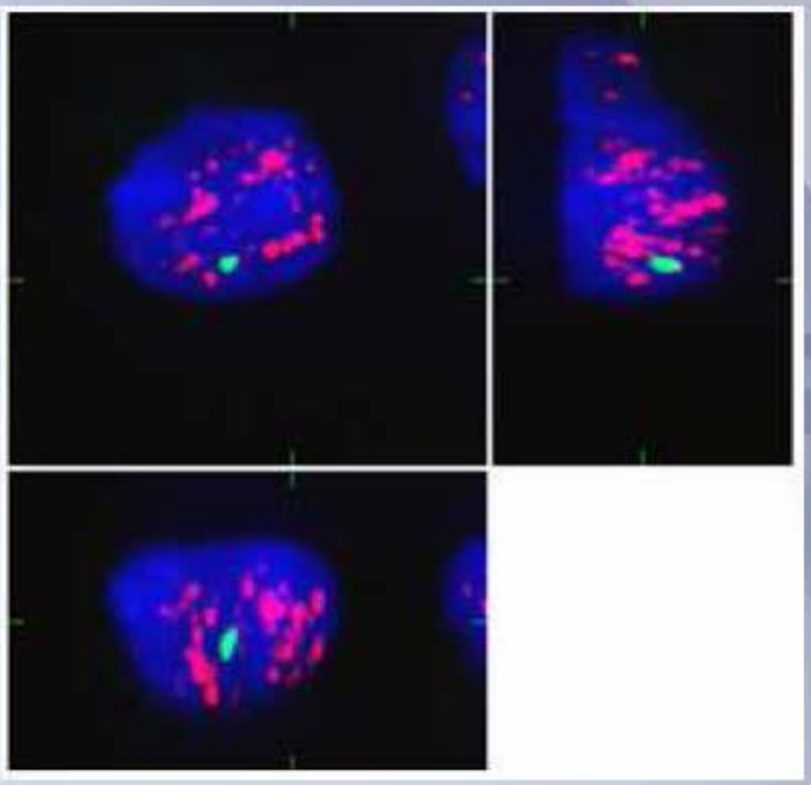


Laser beam moves firstly along *x* axis and then starts with new line in *y* axis.



3D-projection

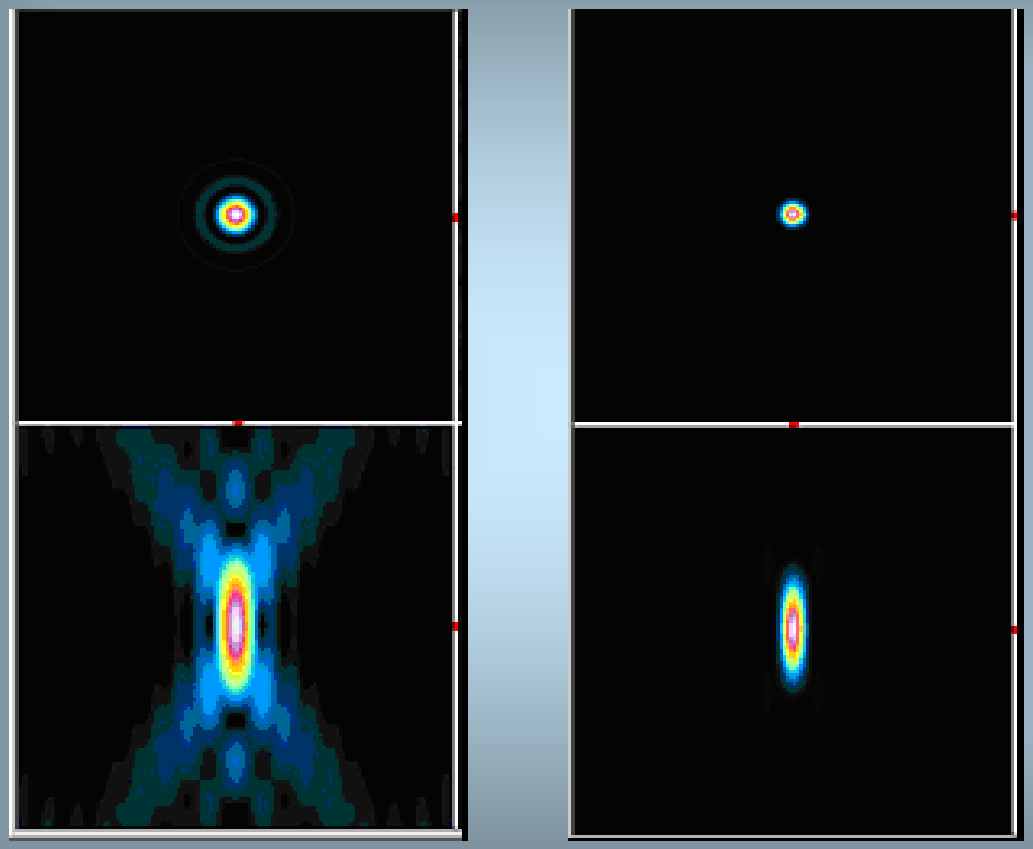




Optical resolution: conventional versus confocal

Conventional

Confocal



Res = 0.61*λ / NA

Res(xy) = 0.4*λ / NA

Res(xz) = 0.45*λ / n(1-cosα)

Formulas by Kino

4PI and STED resolution are much higher…

Deconvolution

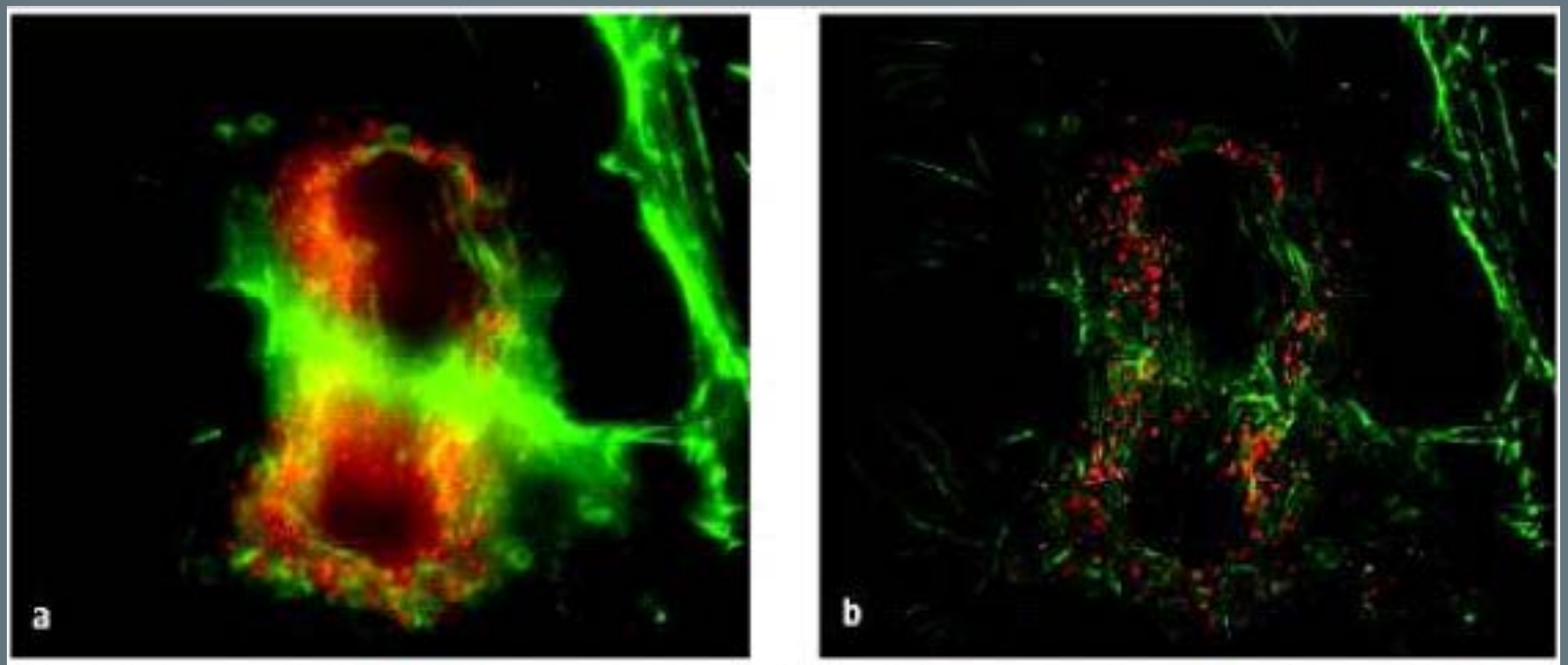


Fig. 3: Via deconvolution, artefacts can be computed out of fluorescence images. a). These artefacts are caused by the stray light from non-focused areas above and below the focus level. These phenomena, referred to as convolution, result in glare, distortion and blurriness. b). Deconvolution is a recognised mathematical procedure for eliminating such artefacts. The resulting image displayed is sharper with less noise and thus at higher resolution. This is also advantageous for more extensive analyses.

Assuming linearity, convolution of the object and the imaging system PSF is affected by noise and produces a blurred image. Deconvolution restores the original object to an improved resolution and higher signal-to-noise ratio (SNR) level.

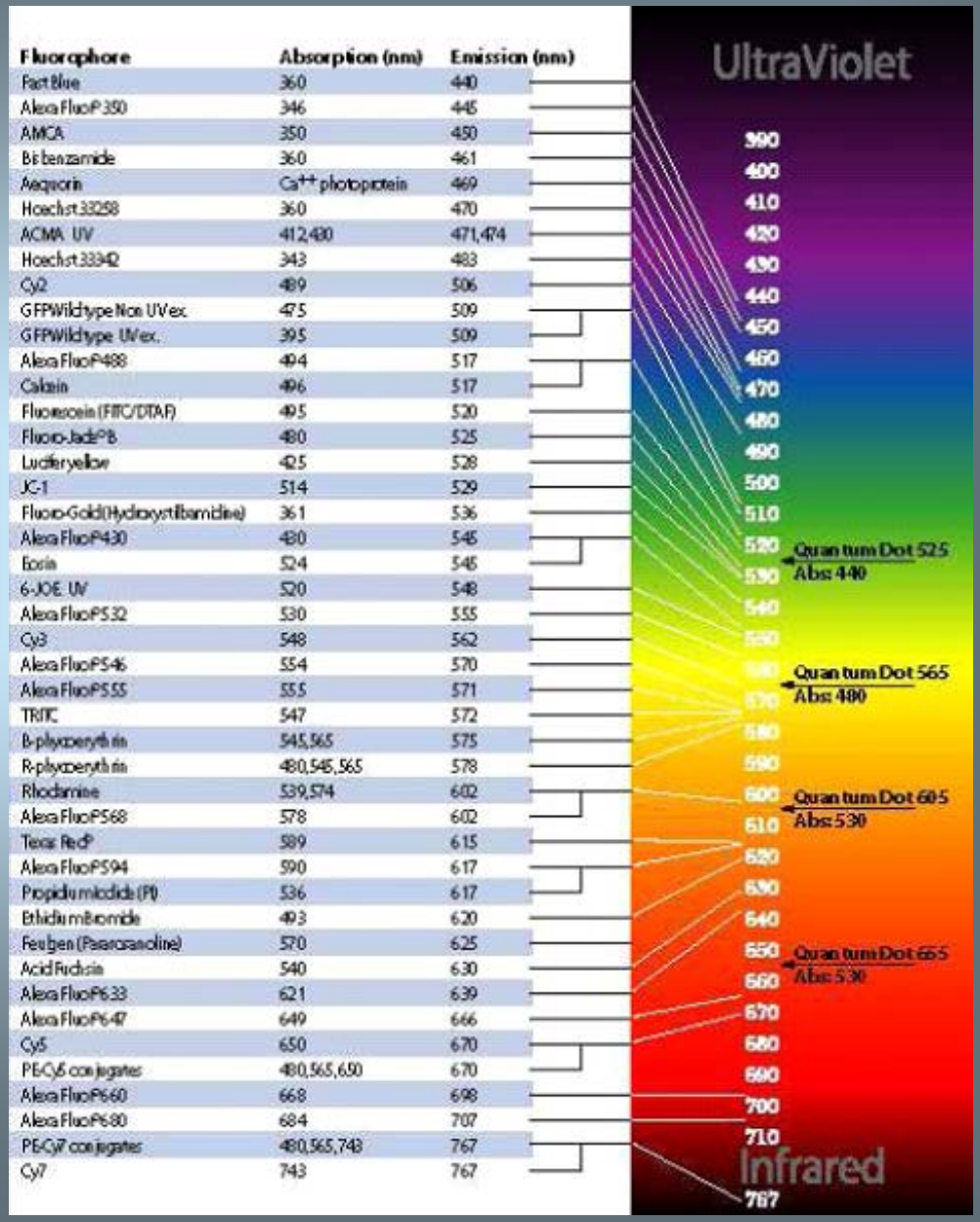
Types of fluorochromes

Fluorochromes are essentially dyes, which accept light energy (e.g. from a laser) at a given wavelength and re-emit it at a longer wavelength. These two processes are called excitation and emission.

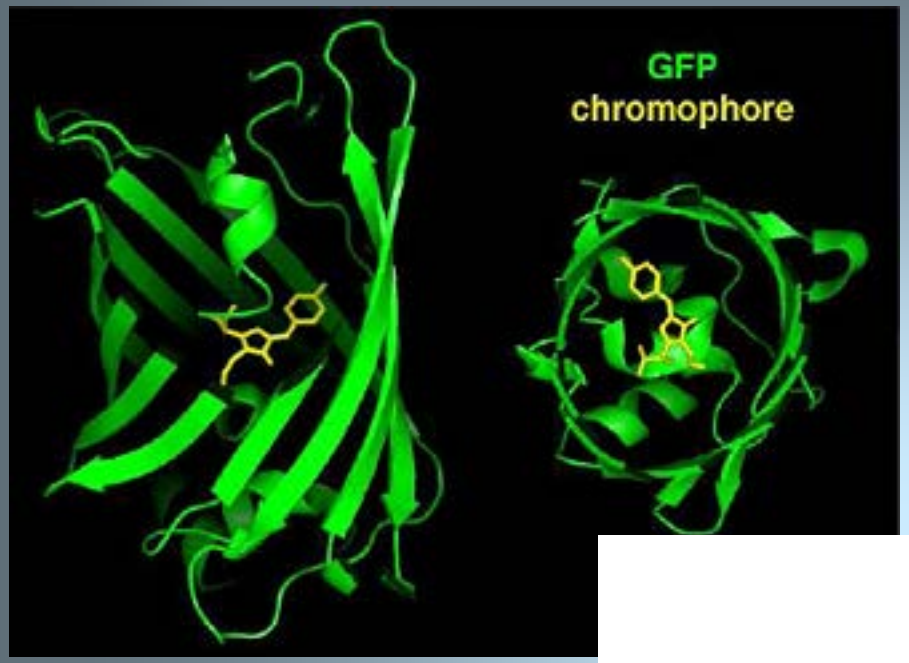
1. Fluorochromes conjugated with other molecule. Example represents quantum dots, used for for ultra-sensitive nonisotopic detection.

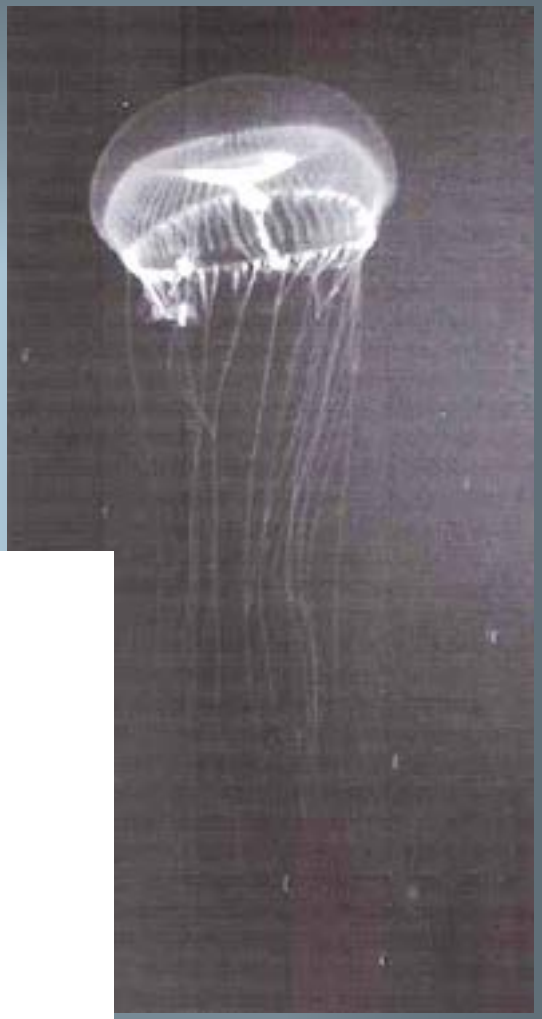
2. Fluorochormes that binds directly to some structure. For example, DAPI or PI binds to DNA

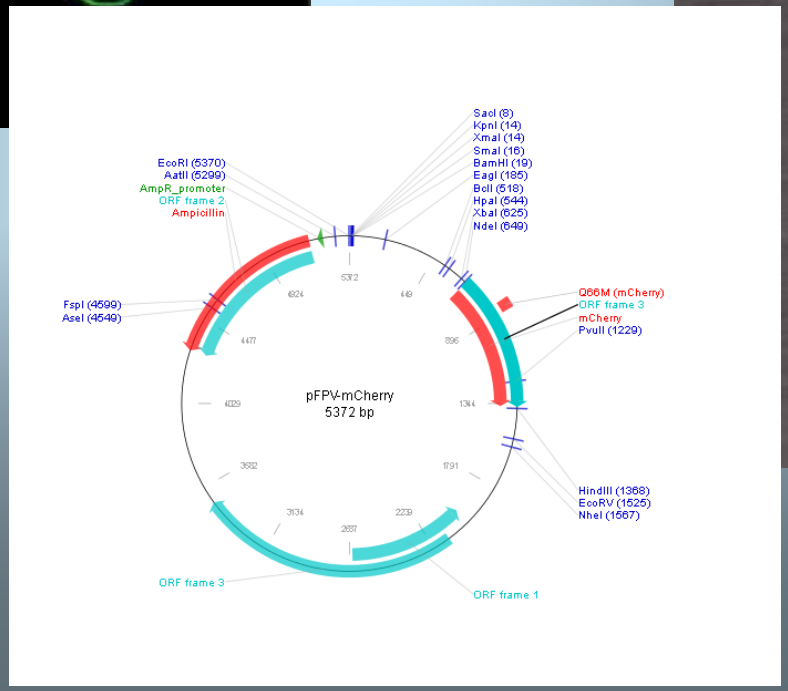
3. Fluorochromes produced by organism like *Aequorea victoria* (GFP) or octocoral *Dendronephthya sp.* (Dendra2)



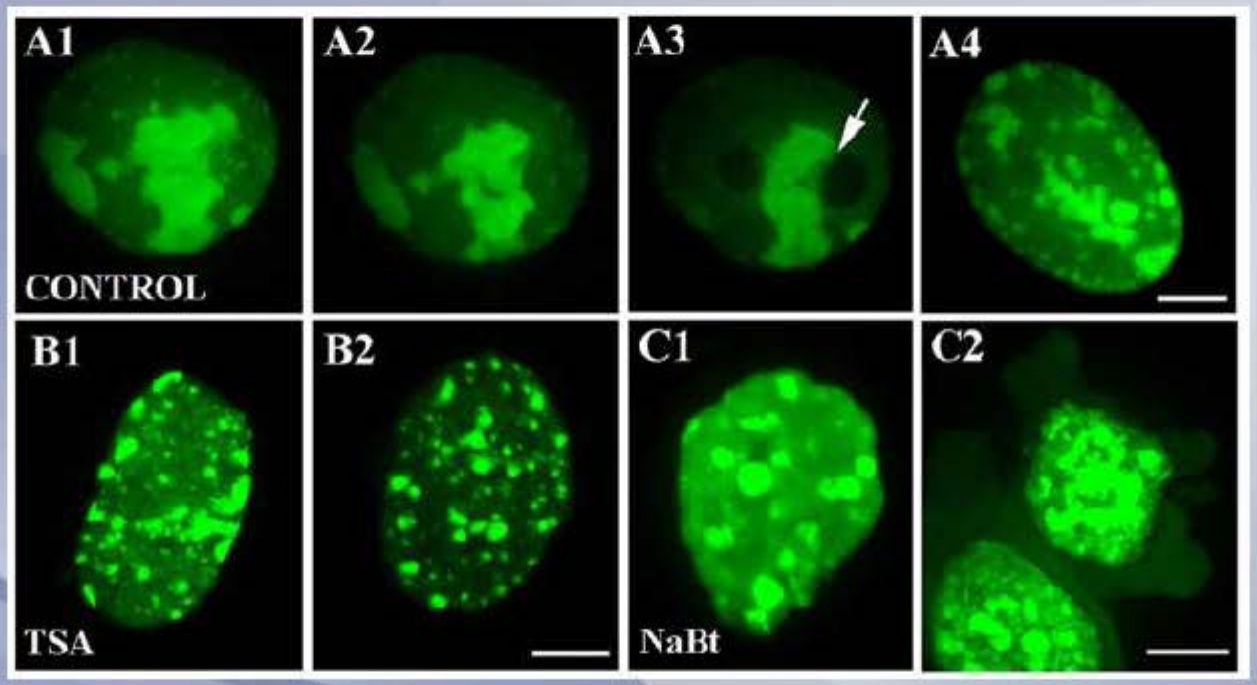
Living Cell Studies - GFP technology

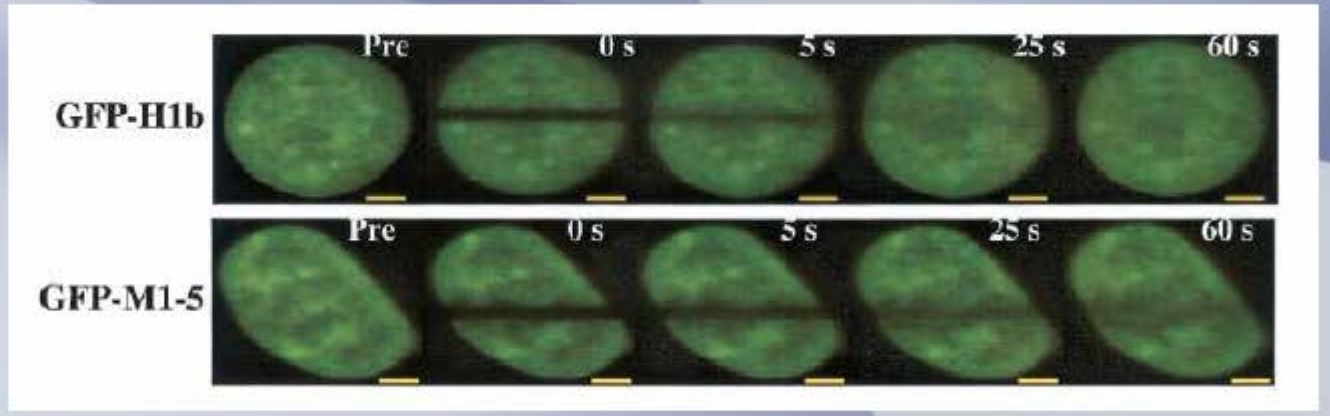


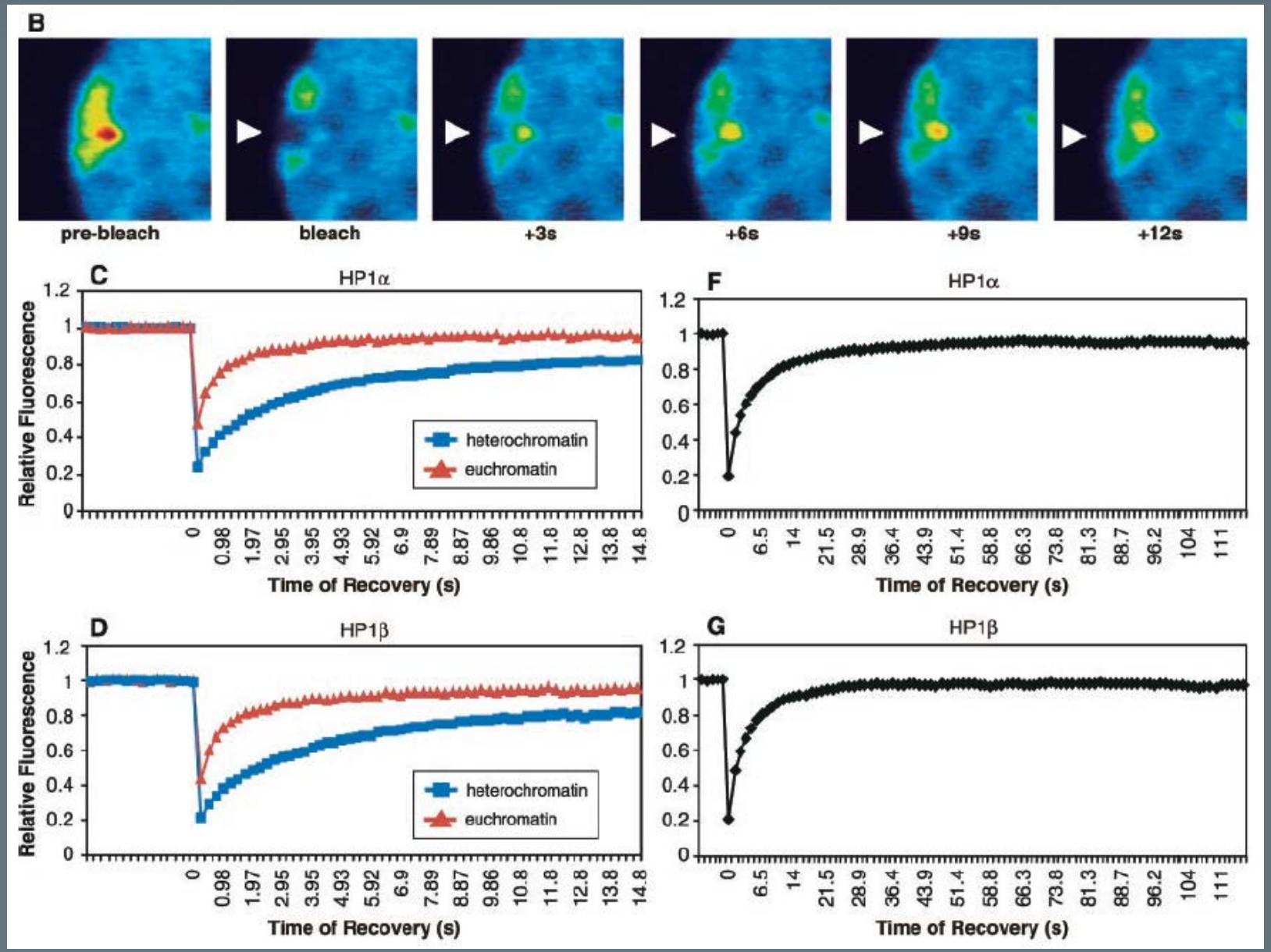




GFP-HP1 β

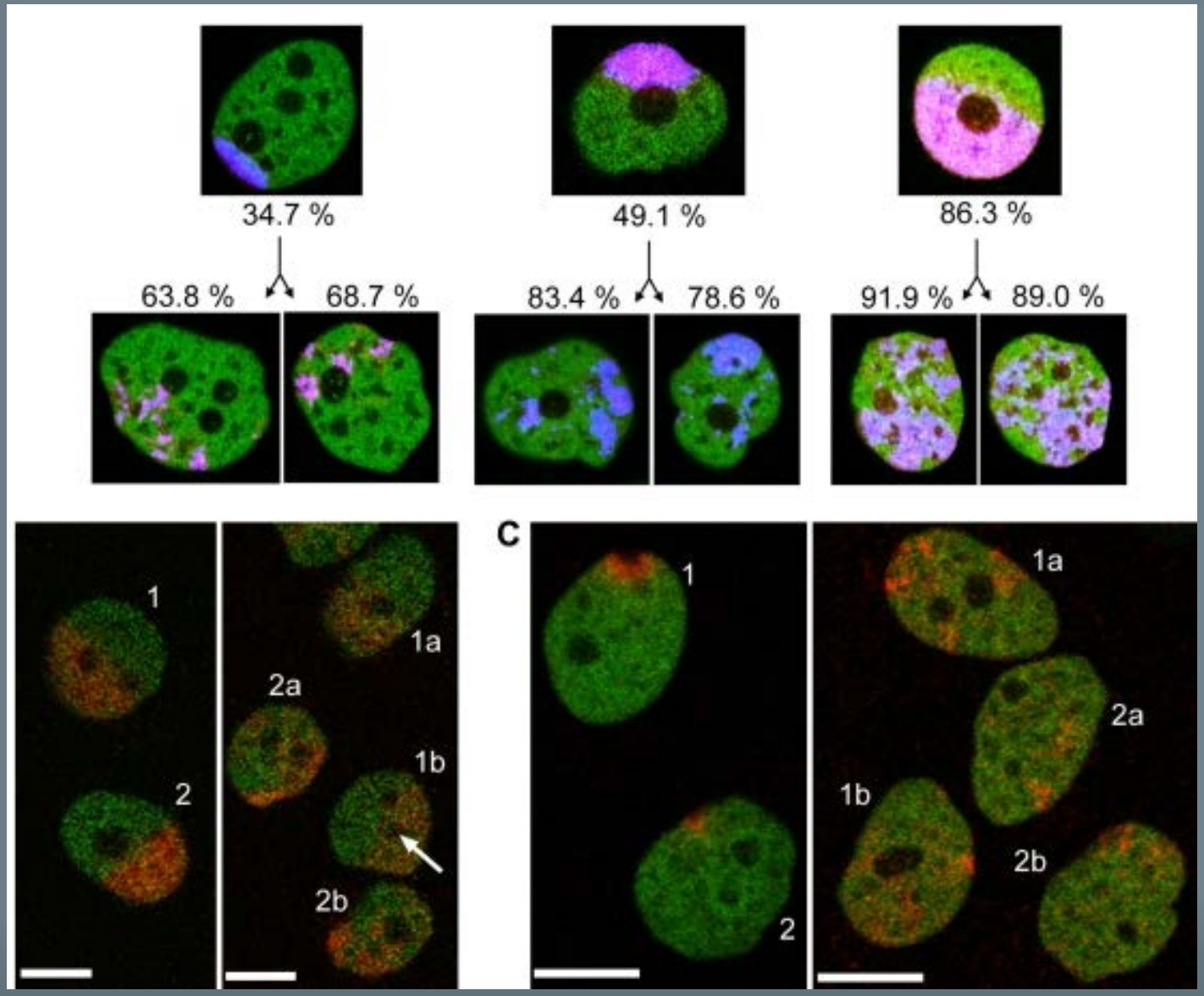

A1
A2
A3
A4
CONTROL
B1
B2
C1
C2
TSA
NaBt


GFP-H1b
Pre
0 s
5 s
25 s
60 s
GFP-M1-5
Pre
0 s
5 s
25 s
60 s

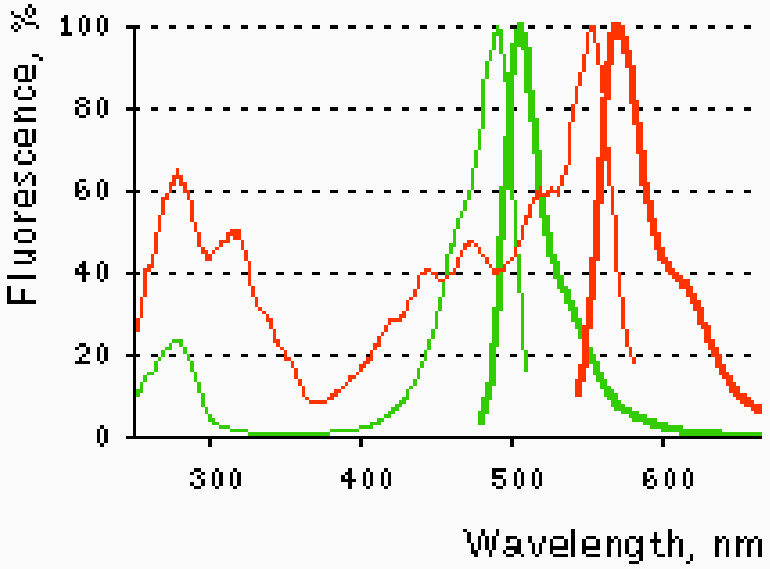


Cheutin et al. Science (2003)

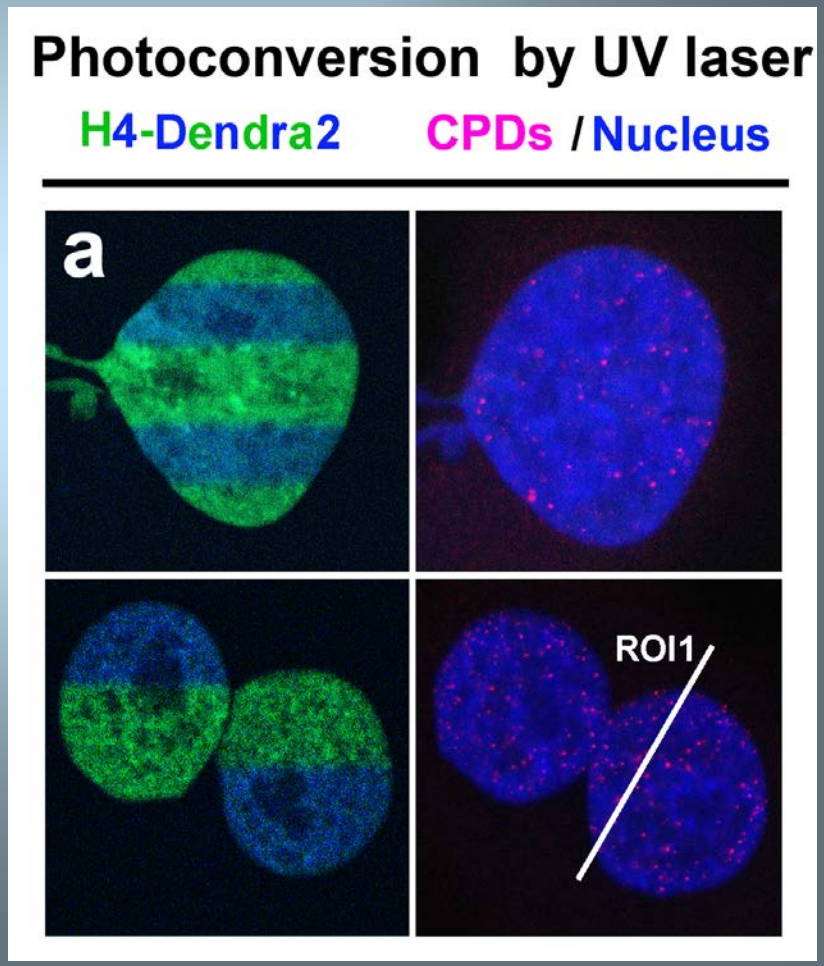
Dendra2 photo-conversion

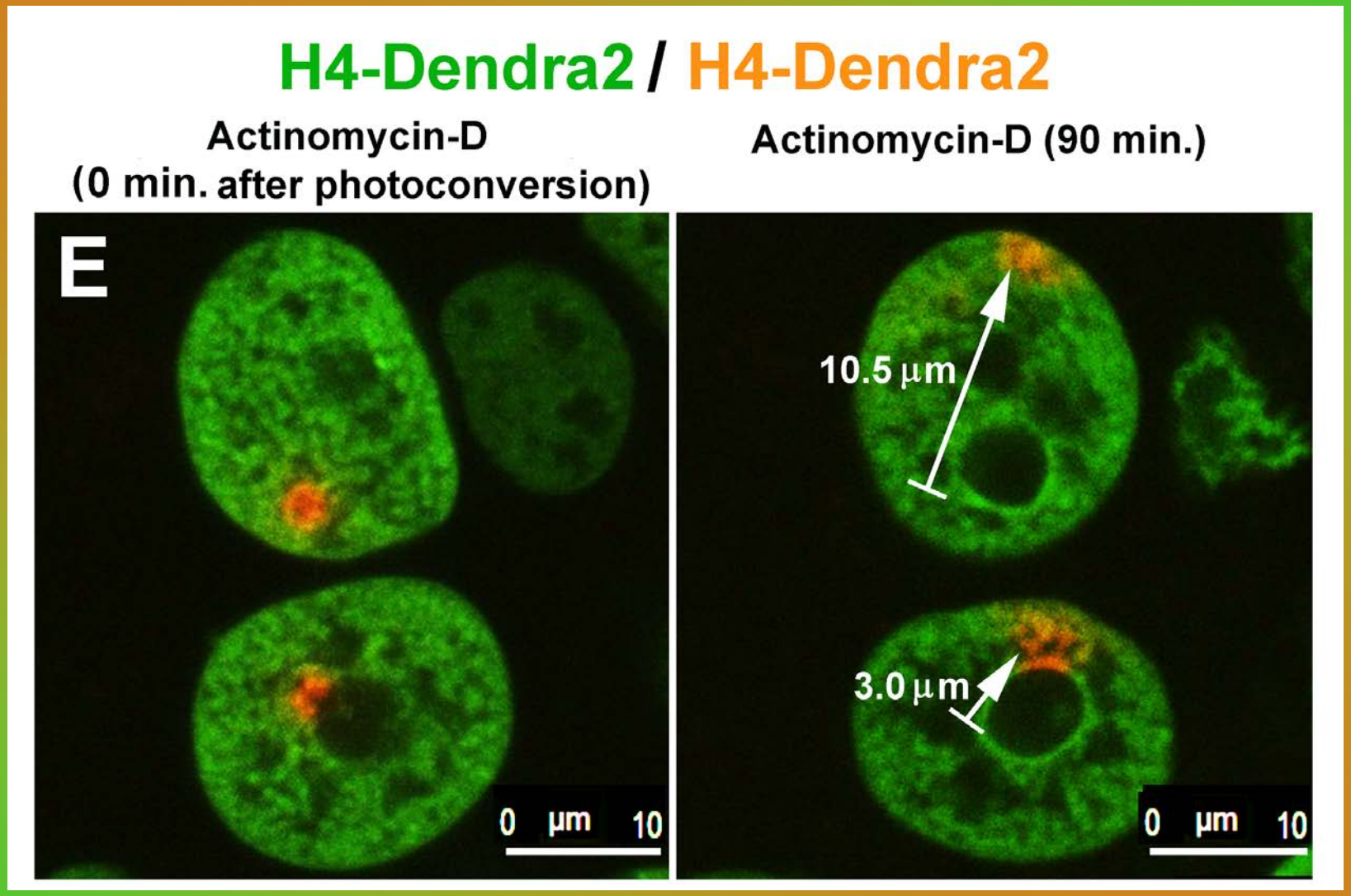


Dendra2 is an improved version of a green-to-red photoswitchable fluorescent protein Dendra, derived from octocoral *Dendronephthya* sp. (Gurskaya *et al.*, 2006).



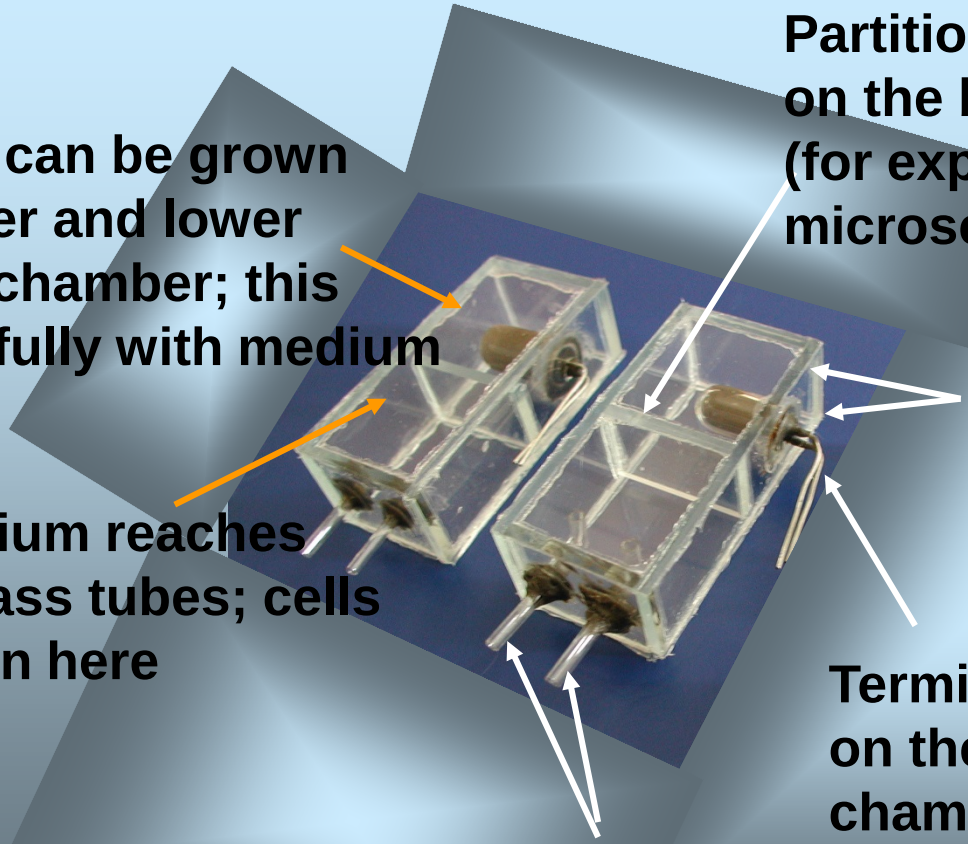
Normalized excitation (thin line) and emission (thick line) for non-activated (green) and activated (red) spectra.




H4-Dendra2 / H4-Dendra2
Actinomycin-D
(0 min. after photoconversion)
Actinomycin-D (90 min.)
E
10.5 μm
3.0 μm
0 μm 10
0 μm 10


Leica TCS SP-5 X

Incubation chambers for living cell studies



Partition with opening on the bottom of the chamber (for experiments under upright microscope)

Part I – cells can be grown on both upper and lower sides of the chamber; this part is filled fully with medium

Cover glass 24x50 mm

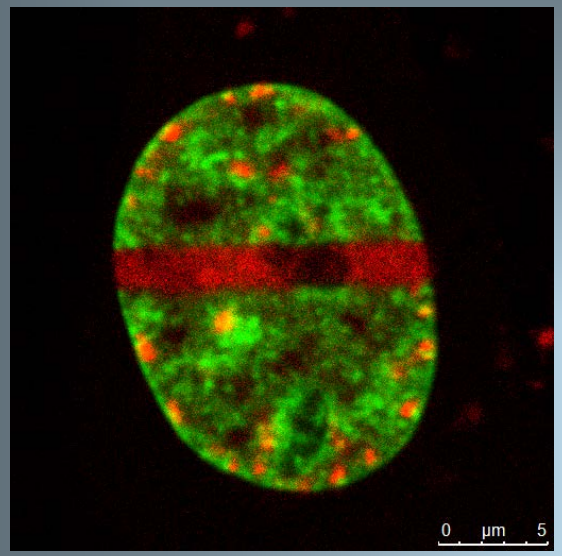
Part II – medium reaches under the glass tubes; cells are not grown here

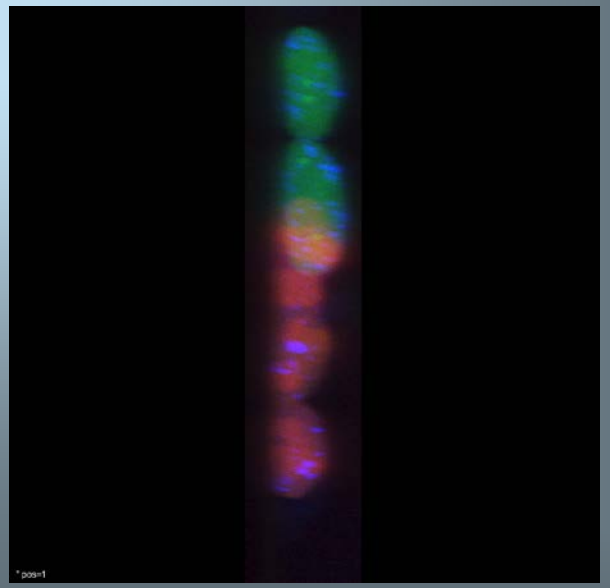
Termistors in short tubes on the side of the chamber

Glass tubes bent under the upper glass (to enable CO_2 to flow above medium in the part II of the chamber)

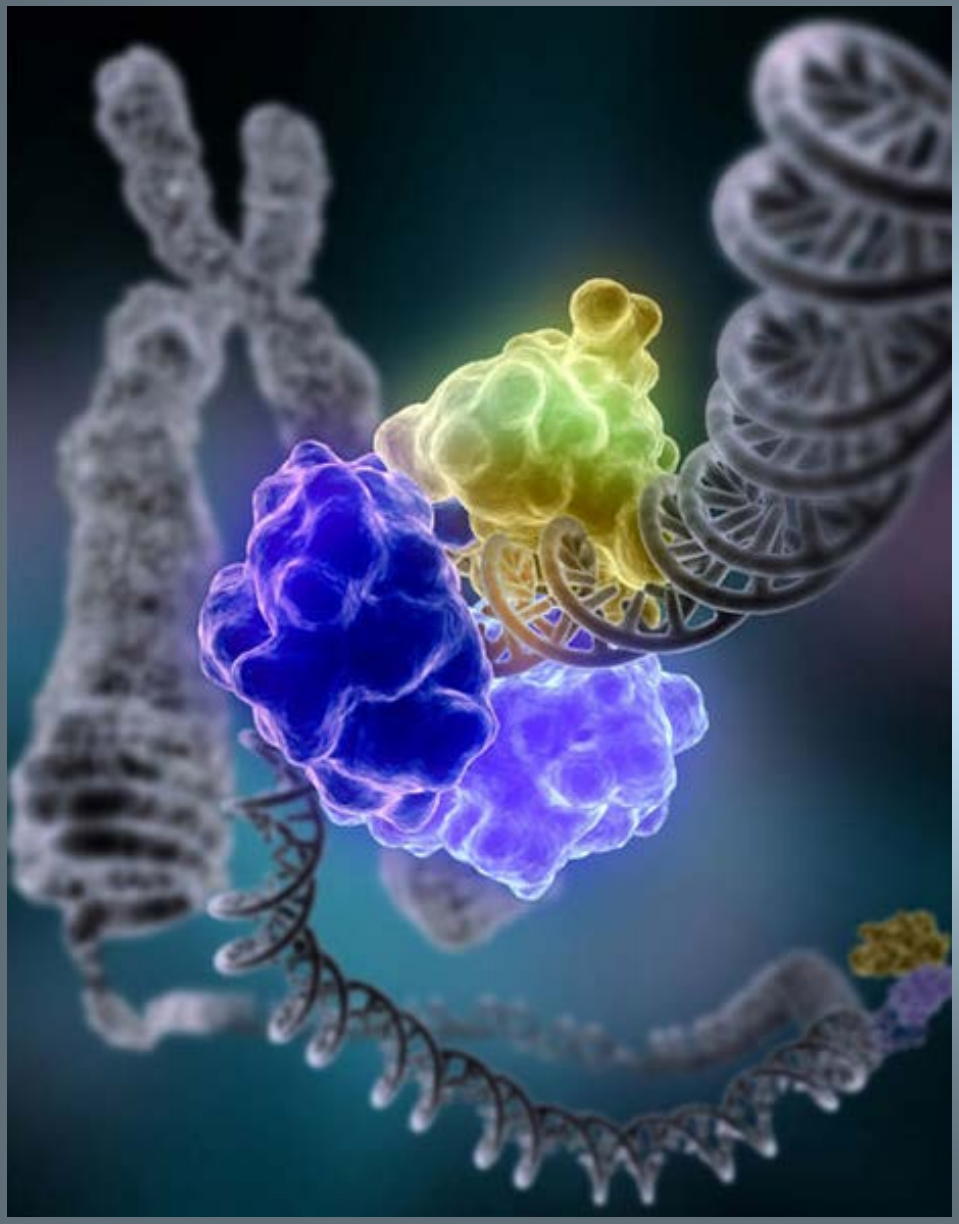
Application of UV–laser (355 nm) in experimental studies on DNA repair

Advanced confocal microscopy





DNA repair



Single-strand damage

- Base excision repair (**BER**), which repairs damage to a single base caused by oxidation, alkylation, hydrolysis, or deamination.
- Nucleotide excision repair (**NER**), which recognizes bulky, helix-distorting lesions such as pyrimidine dimers and 6,4 photoproducts.
- Mismatch repair (**MMR**), which corrects errors of DNA replication and recombination that result in mispaired (but undamaged) nucleotides.

Double-strand breaks

- non-homologous end joining (**NHEJ**)
- microhomology-mediated end joining (**MMEJ**)
- homologous recombination (**HR**)

DNA repair

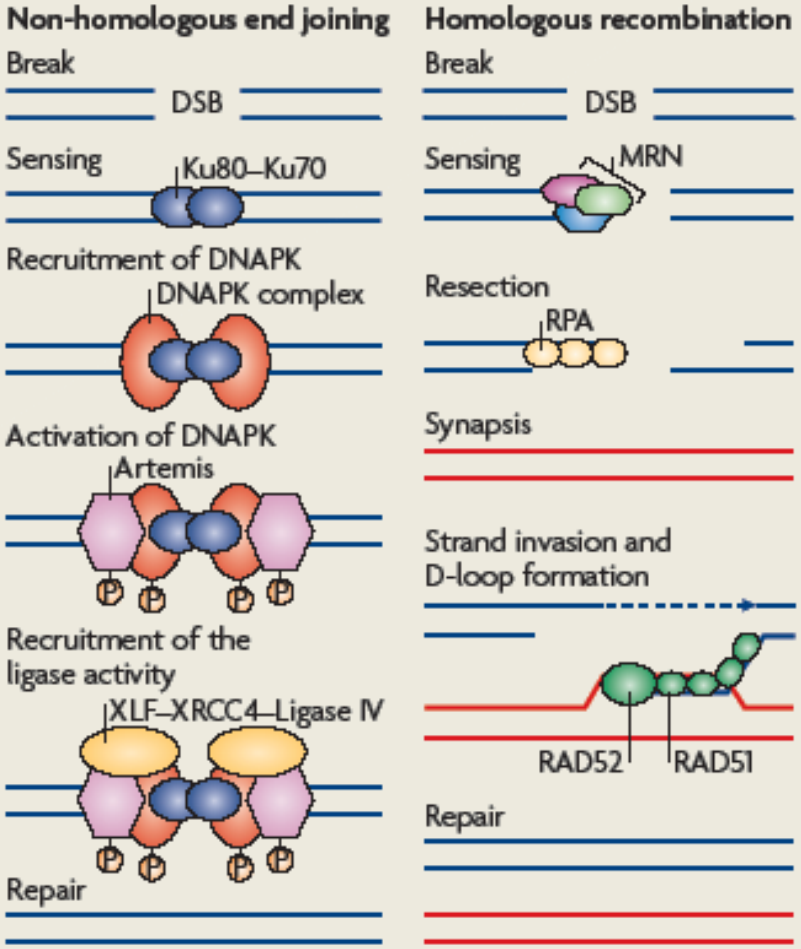
Box 1 | **The two main types of double-stranded DNA-break repair**

Non-homologous end joining

A DNA lesion (a double-stranded DNA break (DSB)) is sensed by the Ku80–Ku70 heterodimer, which in turn recruits the DNA-dependent protein kinase catalytic subunit DNAPKcs, resulting in assembly of the DNAPK complex and activation of its kinase activity (see the figure; left panel). Increasing evidence suggests that DNAPK functions as a regulatory component of non-homologous end joining (NHEJ), potentially facilitating and regulating the processing of DNA ends. DNAPK also increases the recruitment of XRCC4, DNA ligase IV, XLF and Artemis, which carry out the final rejoining reaction.

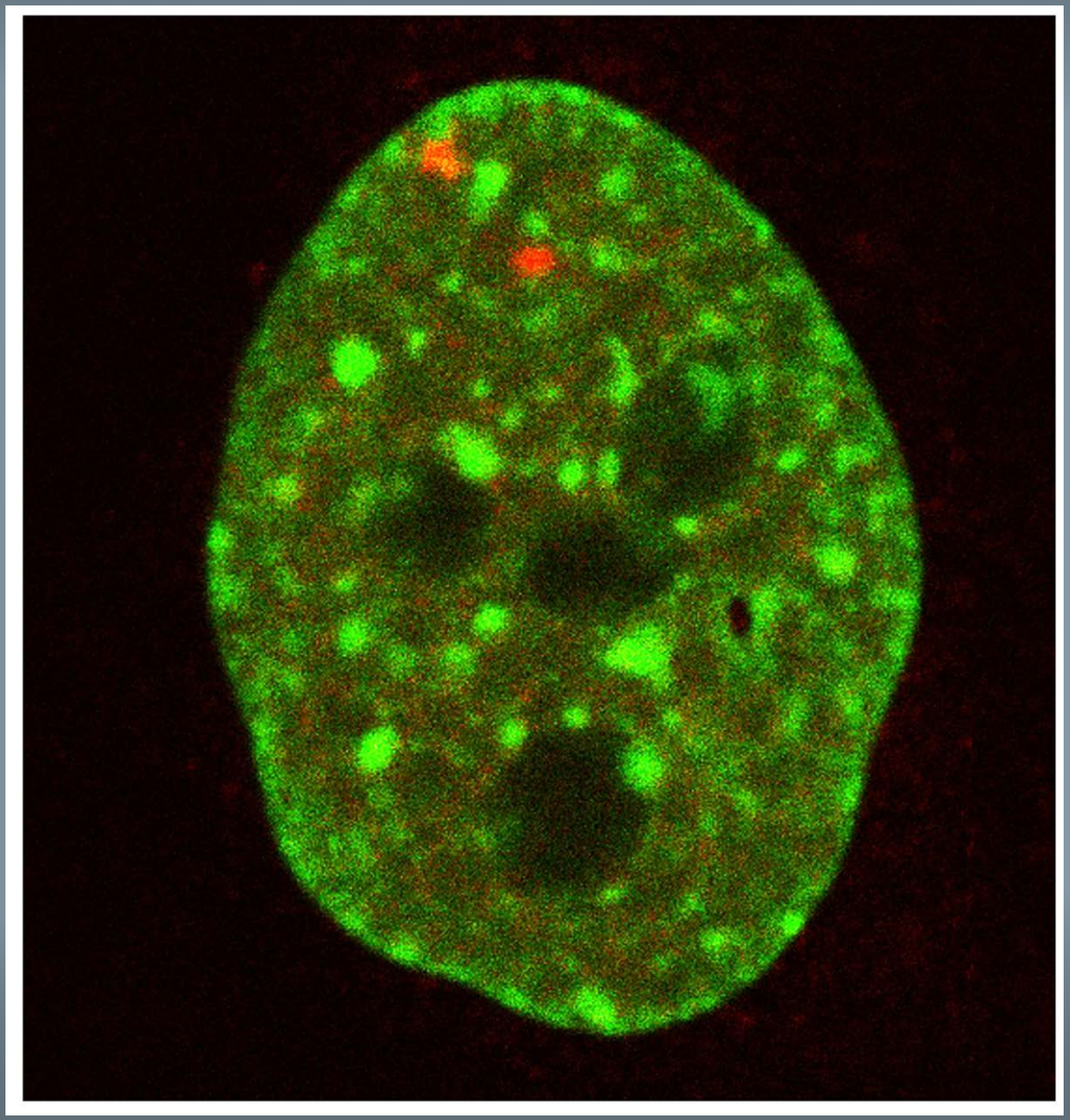
Homologous recombination repair

A DNA lesion is recognized by the MRN (MRE11–RAD50–NBS1) complex, which is recruited to the DSB to generate single-stranded DNA by resection (see the figure; right panel). The single-stranded ends are bound by replication protein A (RPA), RAD51 and RAD52 and can subsequently invade the homologous template, creating a D-loop and a Holliday junction, to prime DNA synthesis and to copy and ultimately restore genetic information that was disrupted by the DSB.

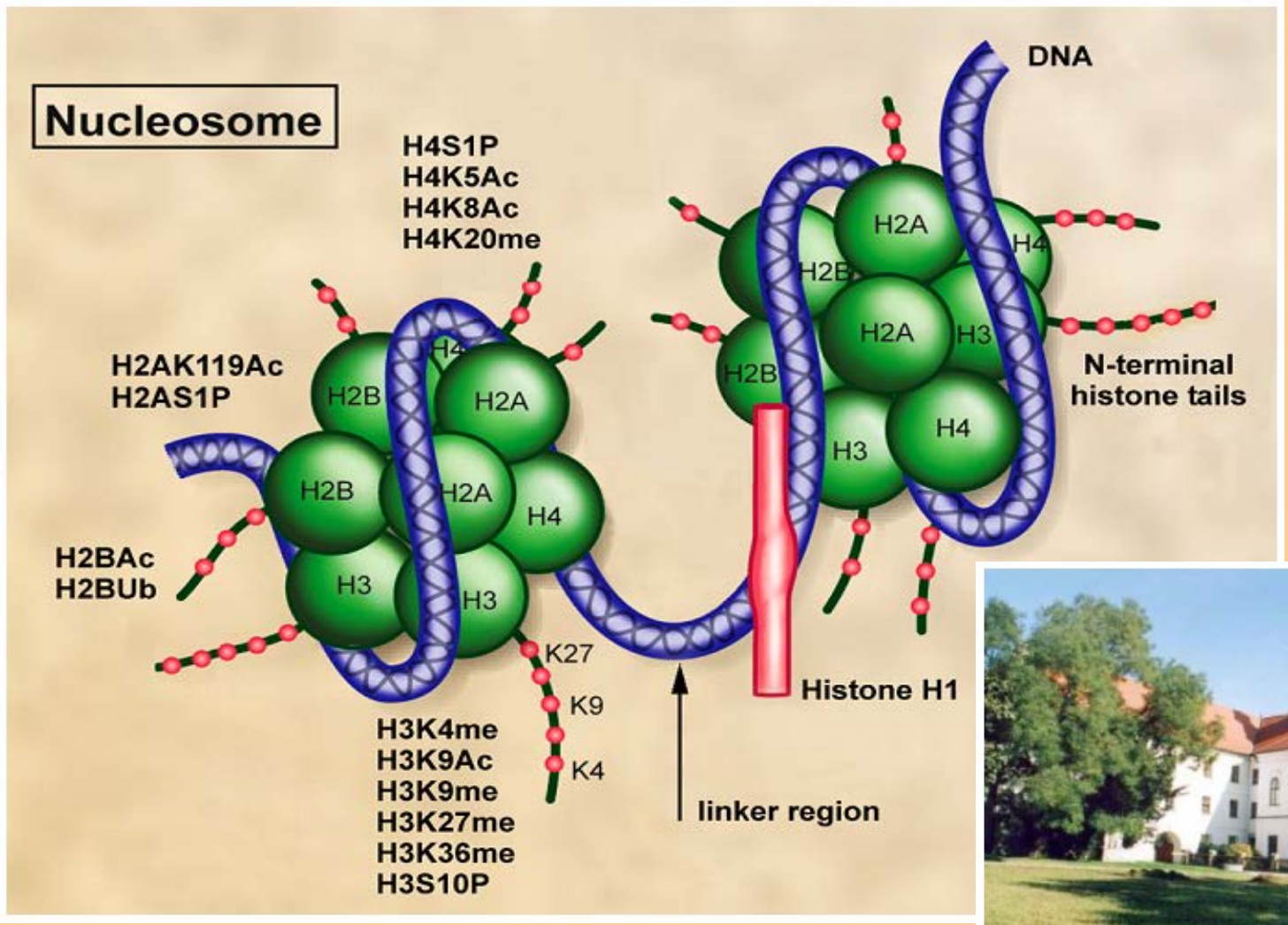


Misteli and Soutogou (2009)

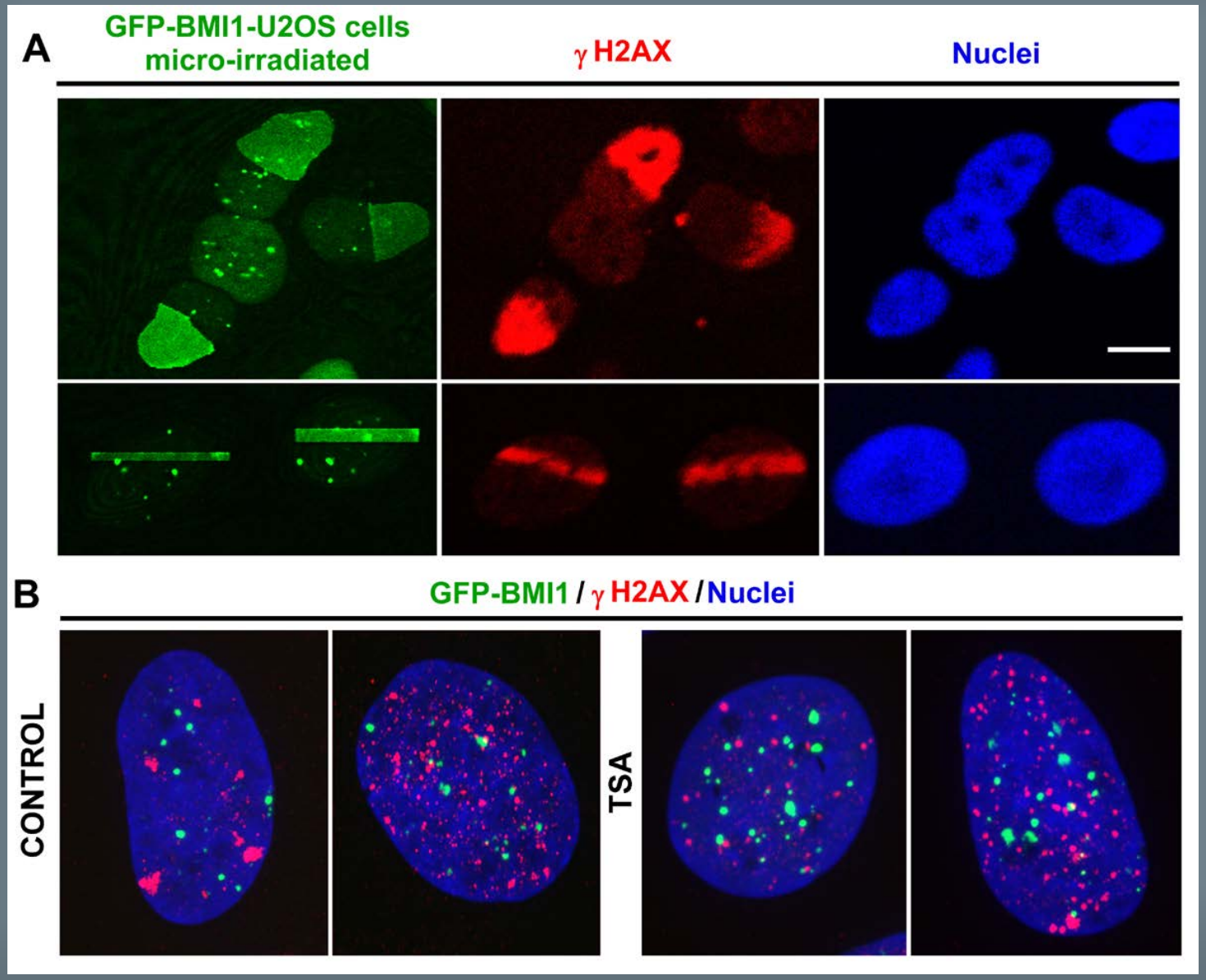
DNA repair foci



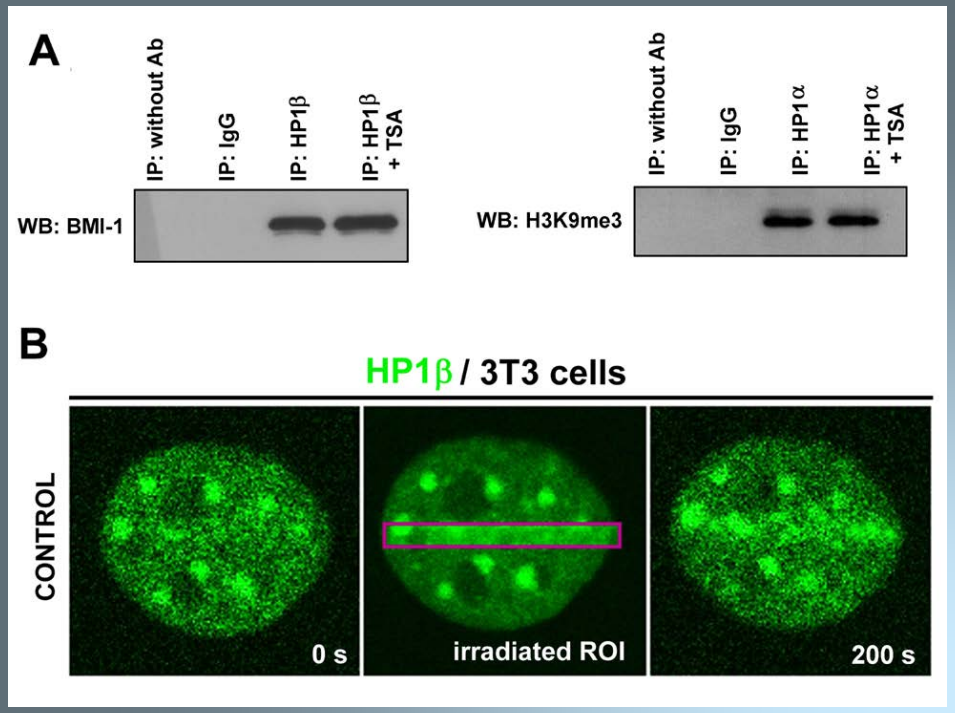
Histone signature
Brno nomenclature for histone modifications
(Turner, 2005)

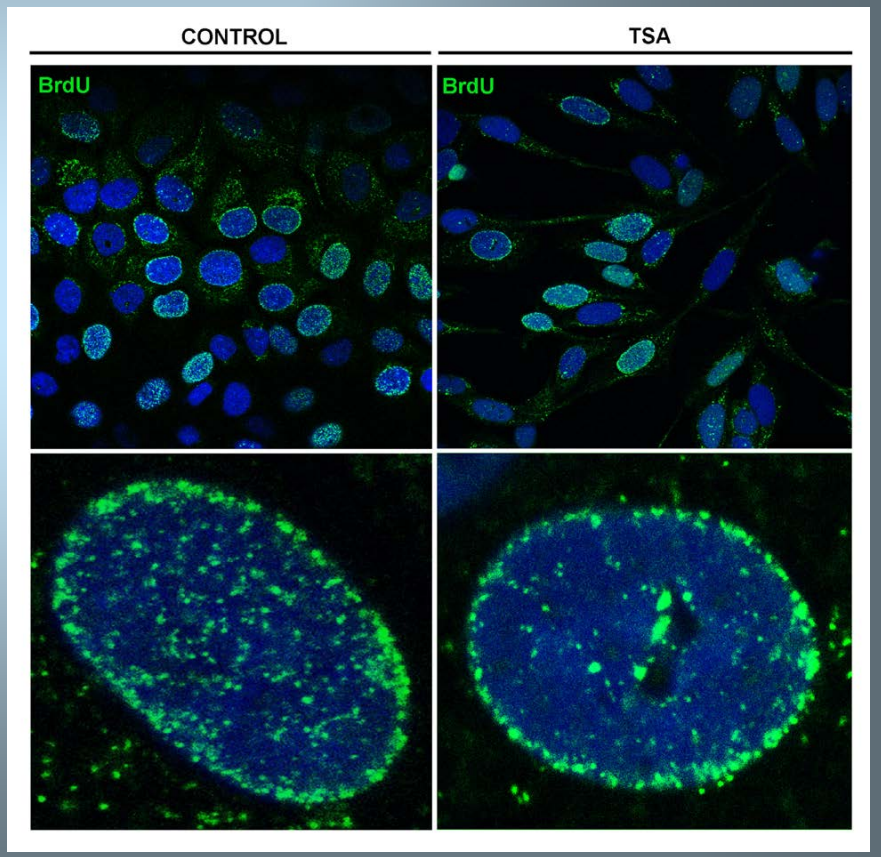


Využití UV laseru 355 nm ke studiu DNA reparace

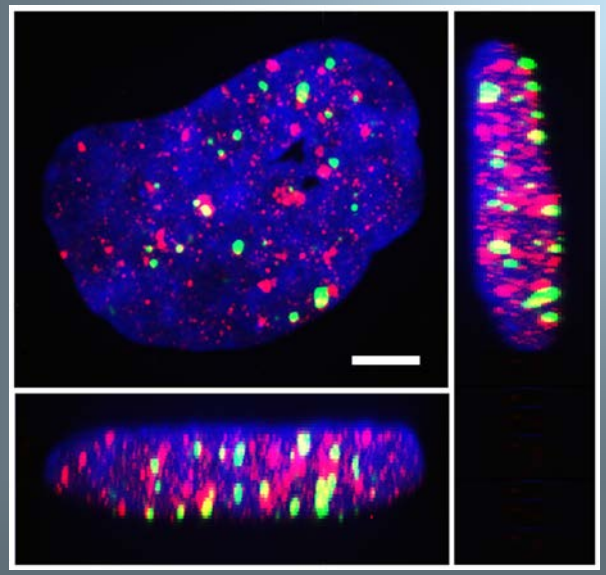


Experiments of Gabriela Galiová

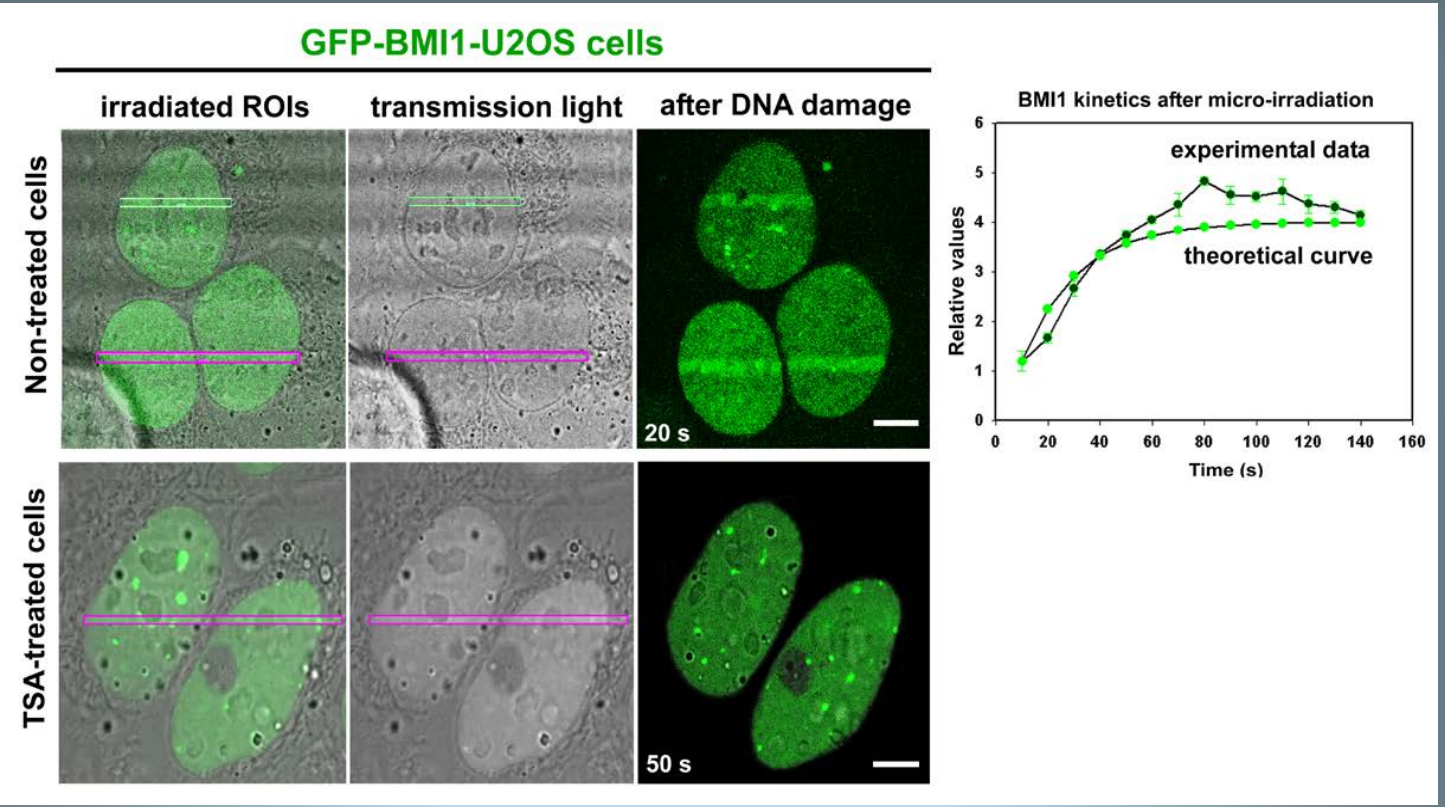


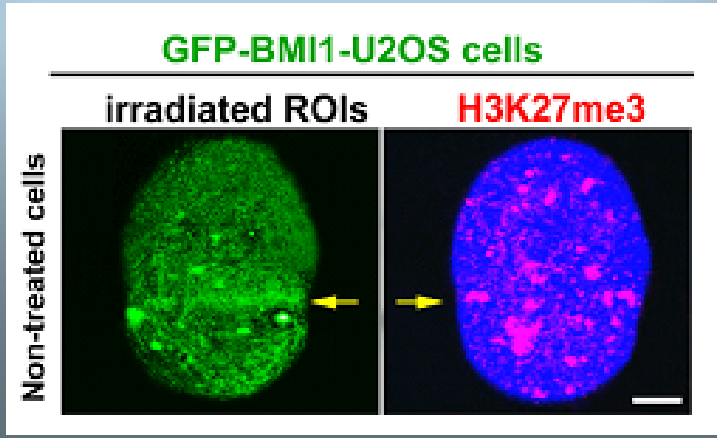


HP1β / BMI1

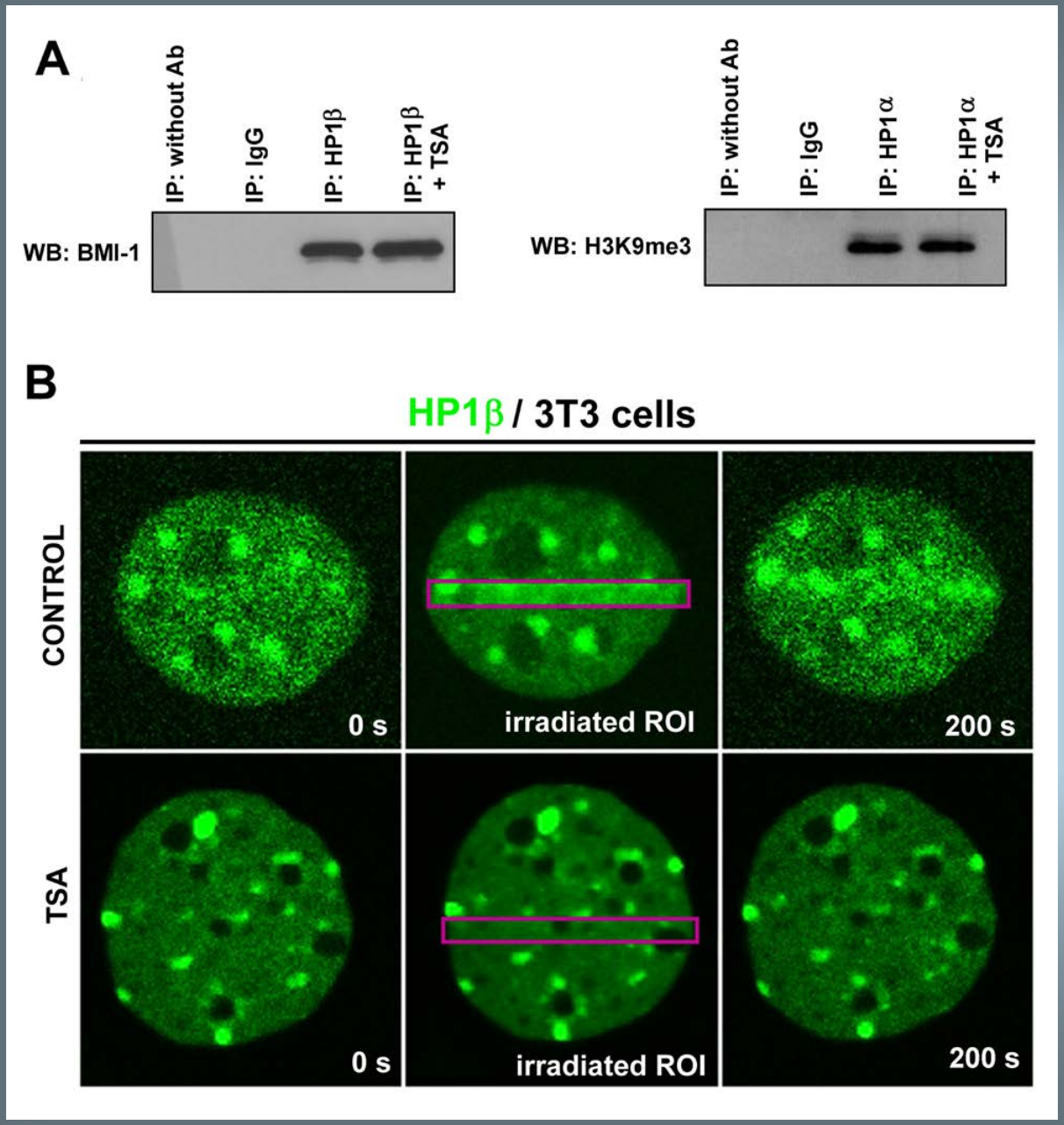


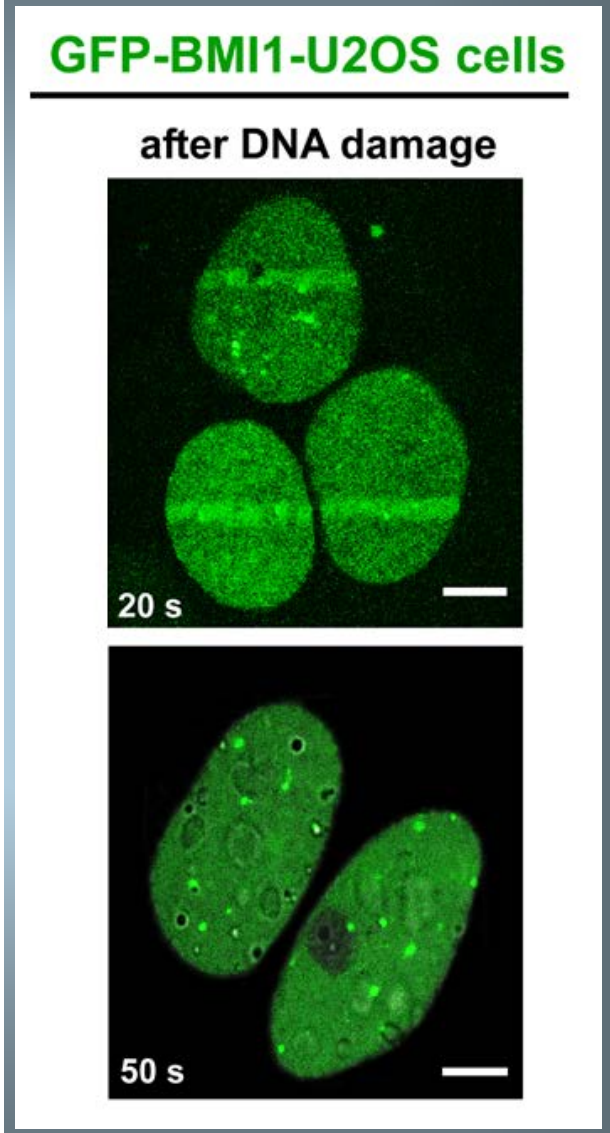
Experiments of Gabriela Šustáčková, Eva Bártová and Lenka Stixová





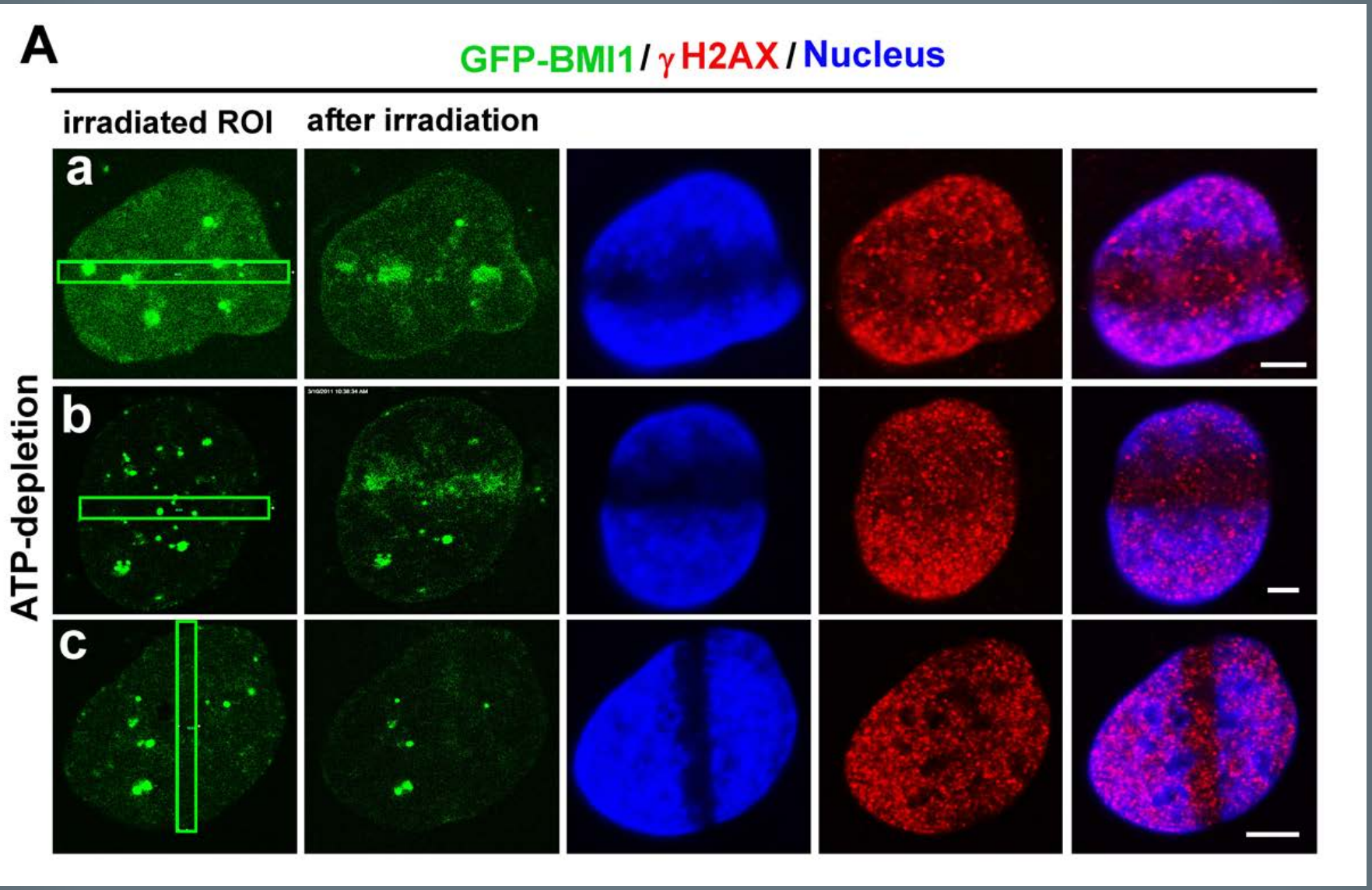
Experiments of Gabriela Galiová



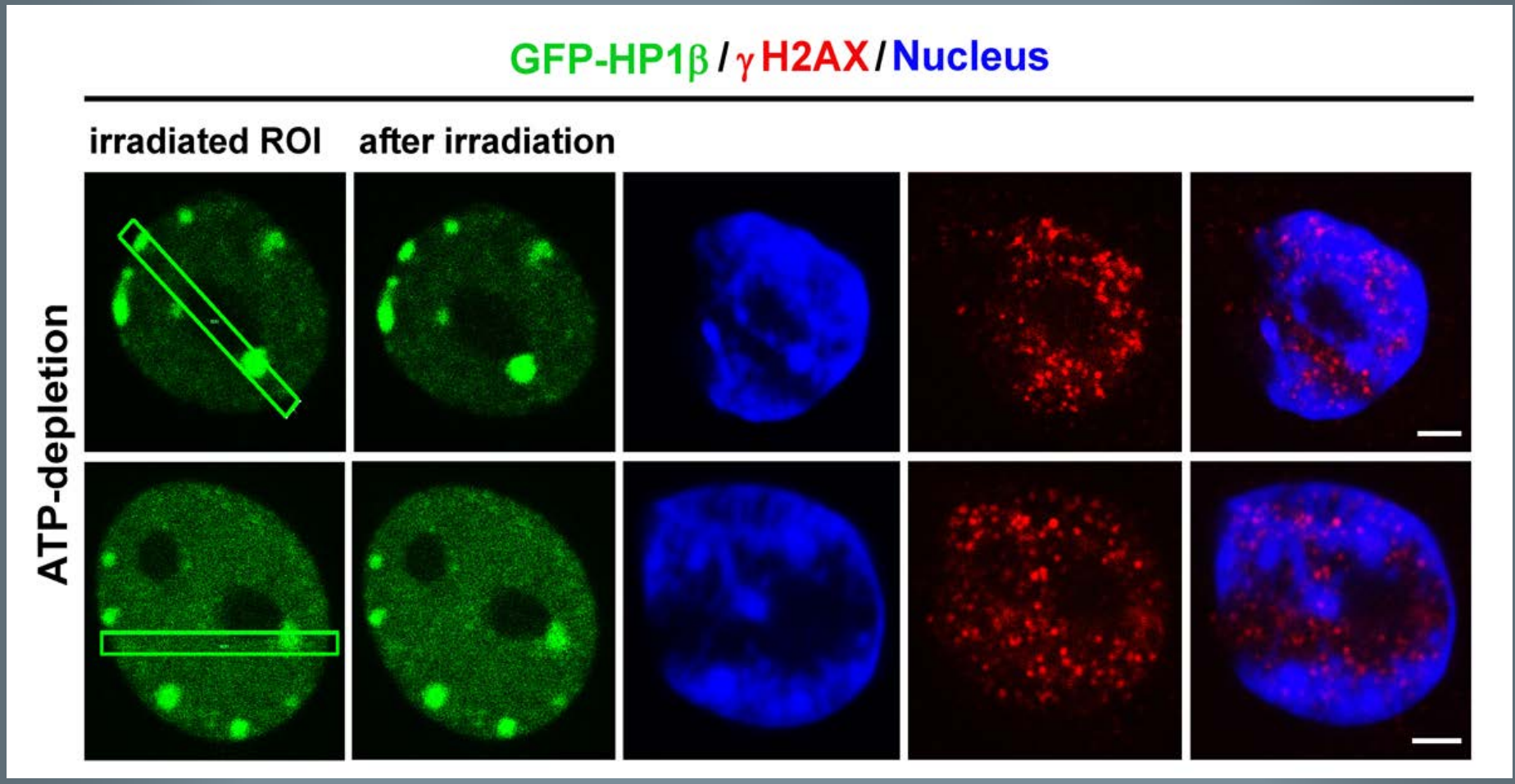


Experiments of Gabriela Galiová and Lenka Stixová

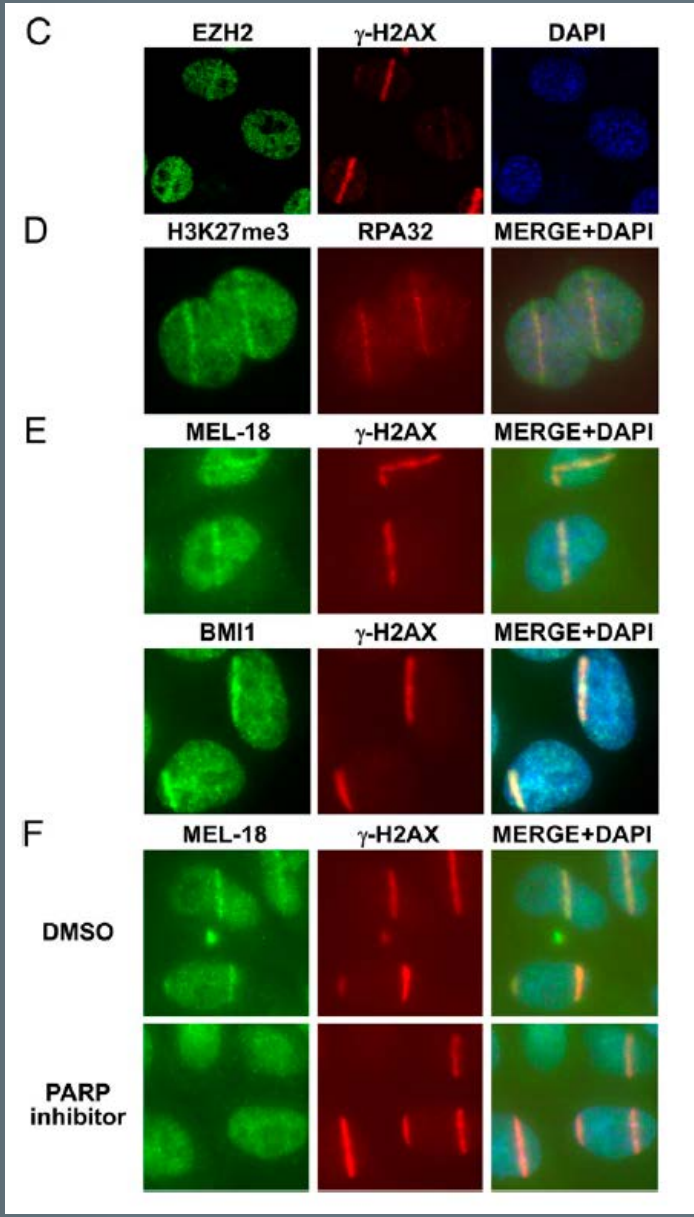
ATP depletion



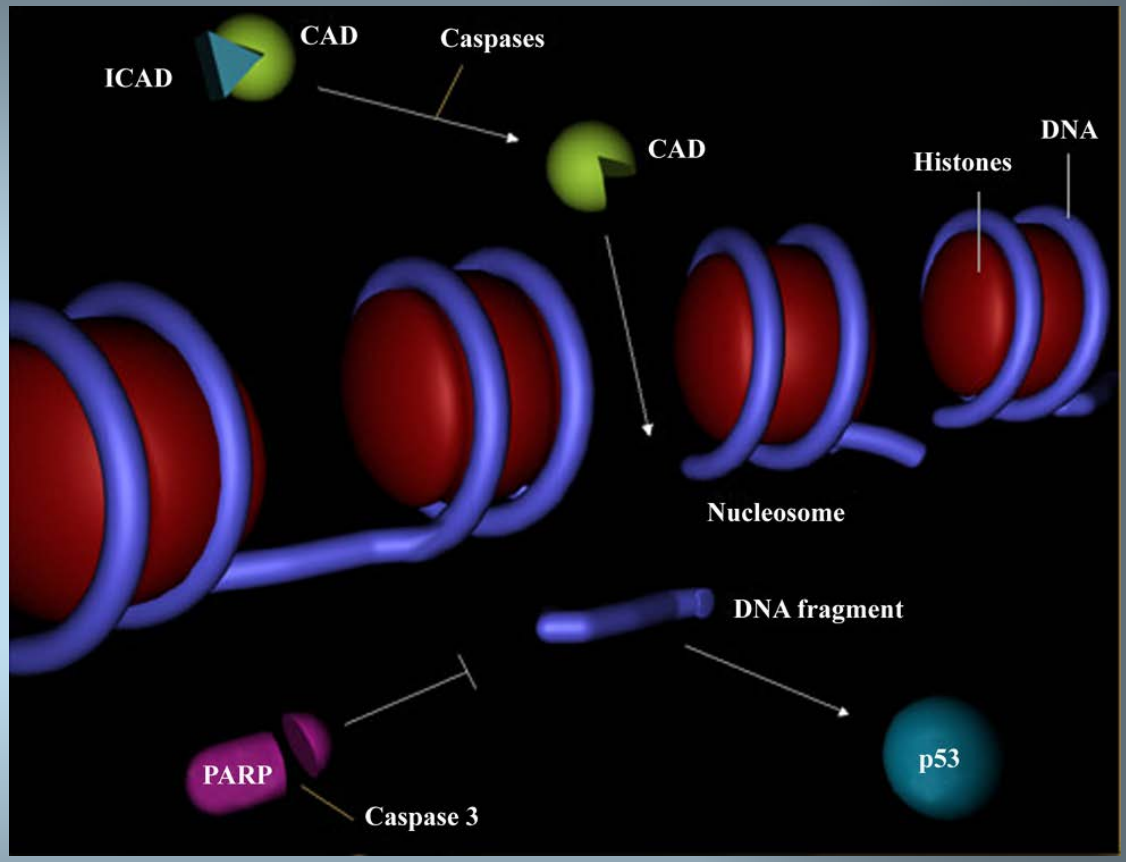
Šustáčková et al., JCP, 2011



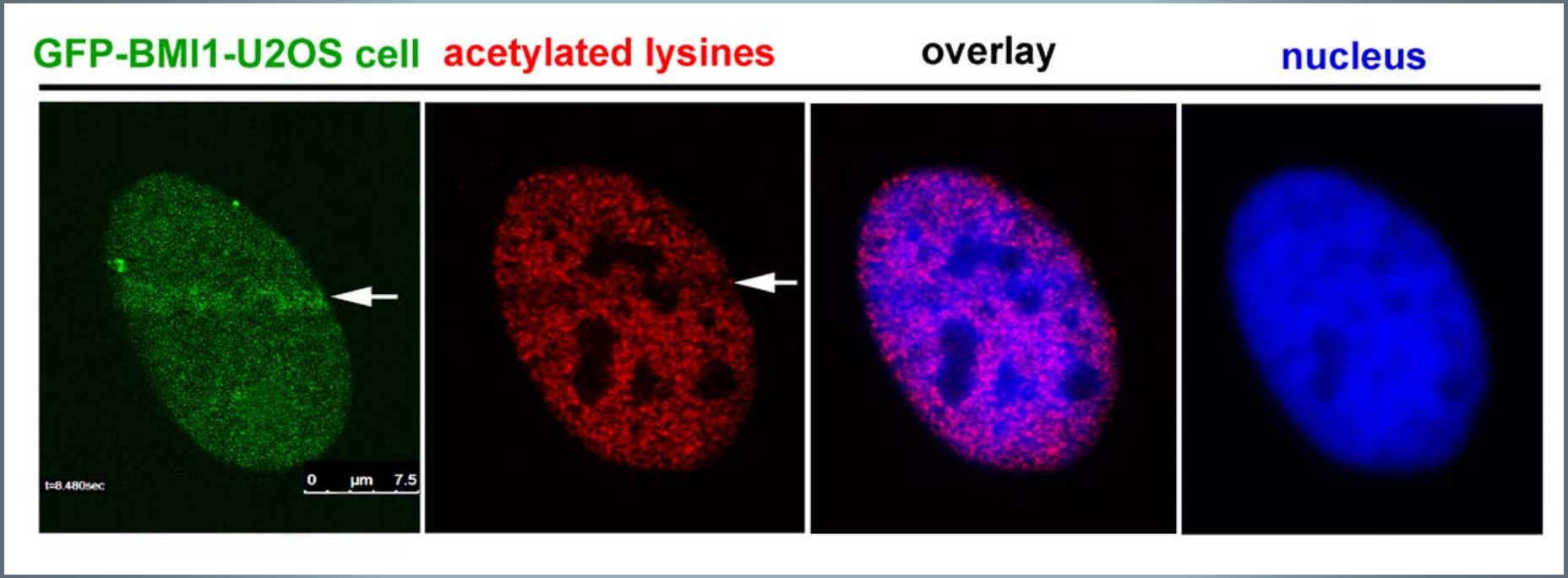
Experiments of Gabriela Šustáčková



Chou et al., PNAS (2010)

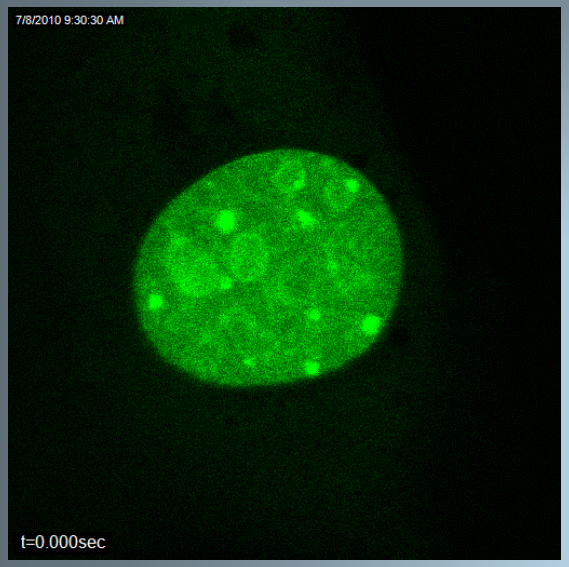


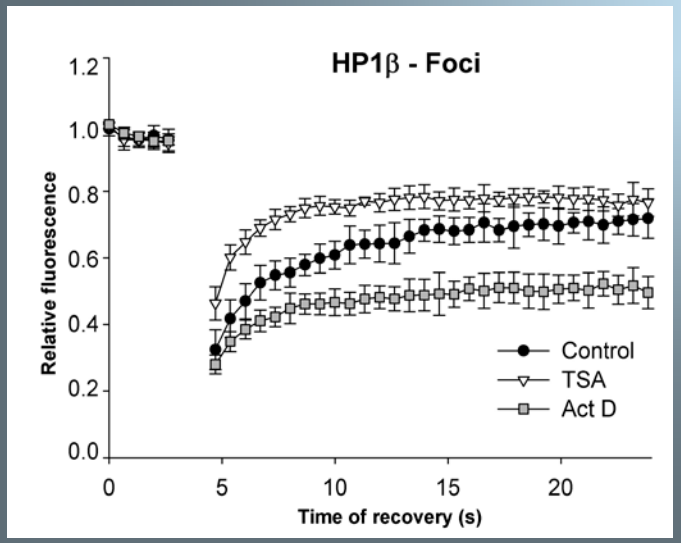
DNA repair

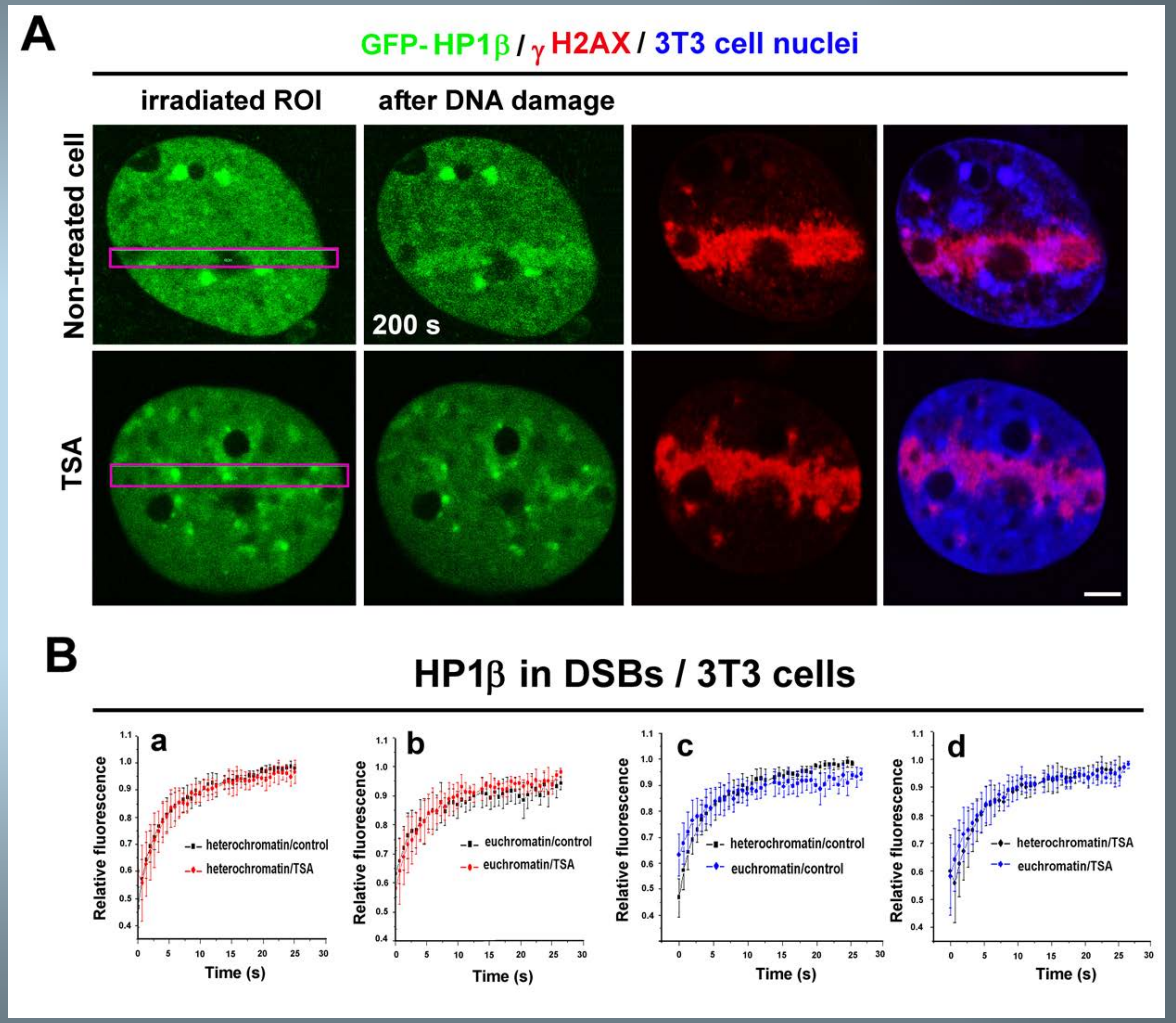


Experiments of Gabriela Šustáčková and Soňa Legartová

FRAP in UV-damaged chromatin with accumulated BMI1 and HP1β

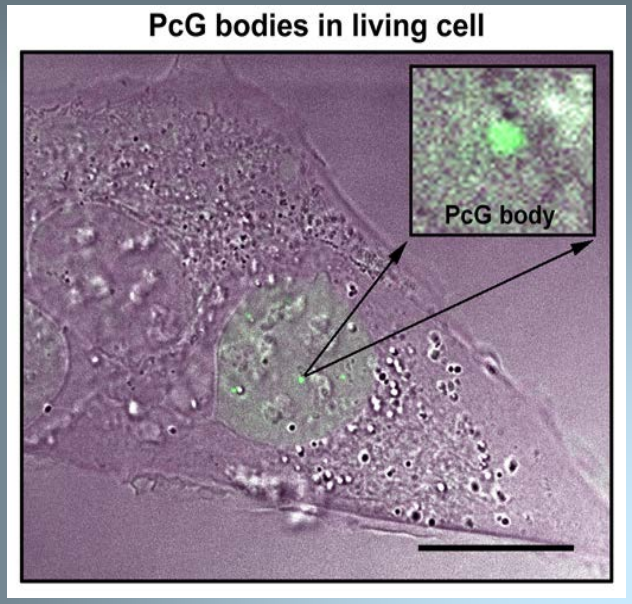


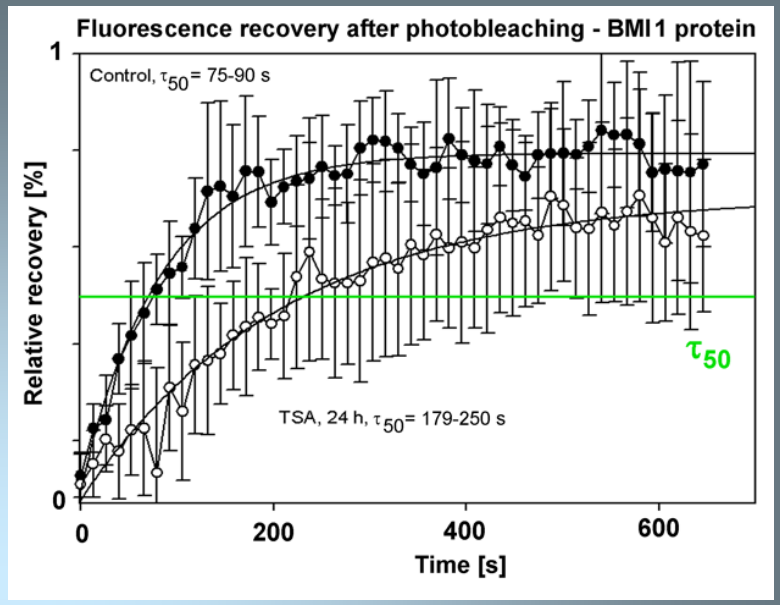


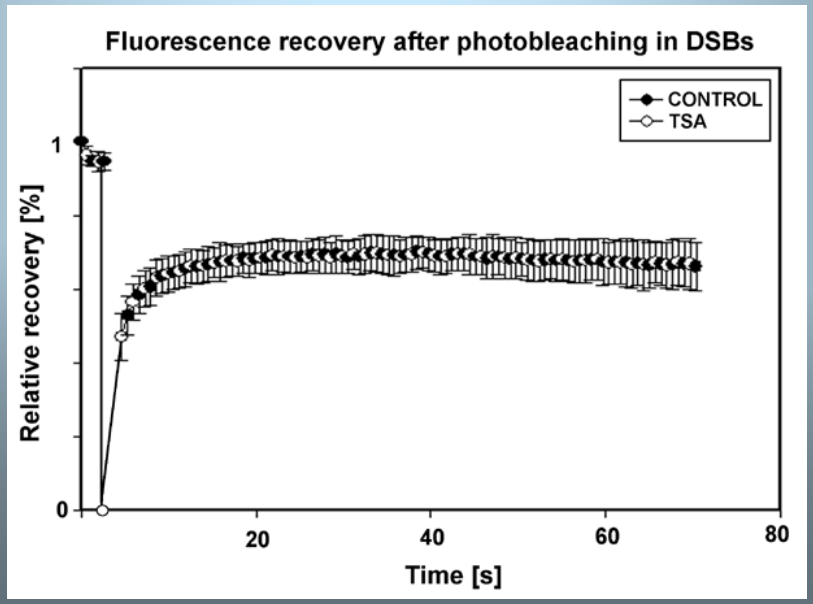


Experiments of Gabriela Šustáčková and Darya Orlova

FRAP – BMI1



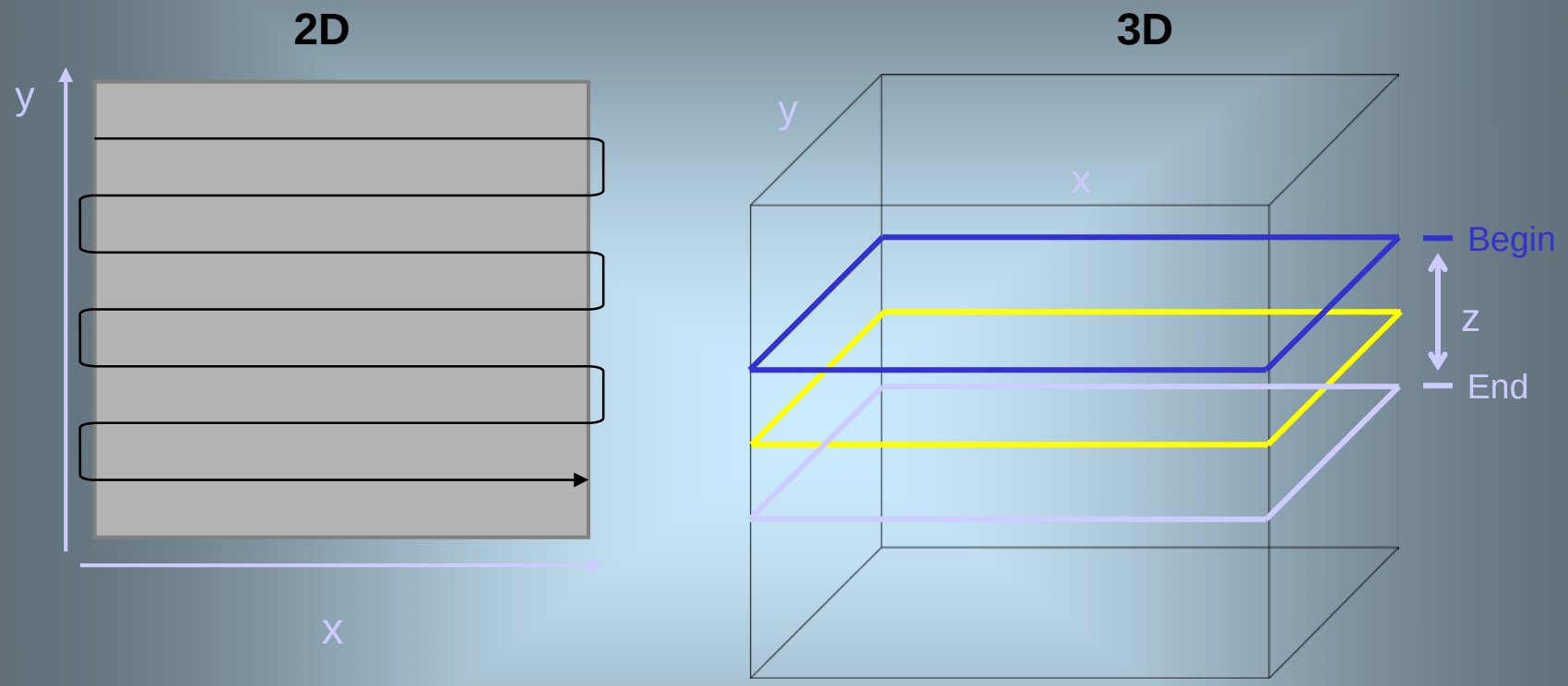




Experiments of Stanislav Kozubek

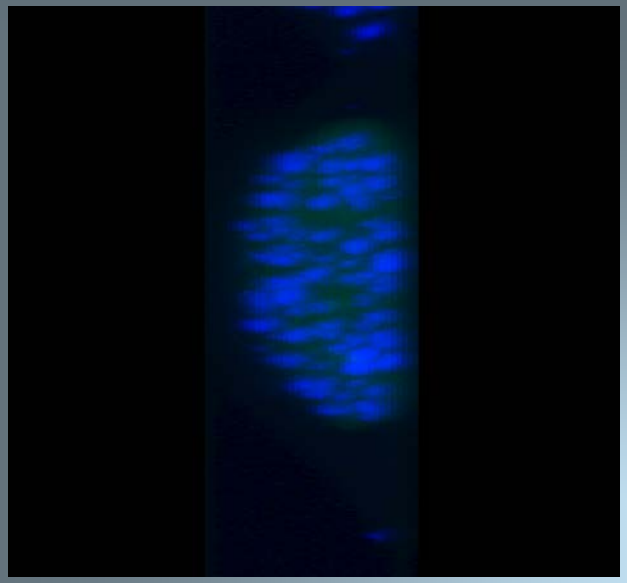
Additional molecular-biology methods with application for confocal microscopy

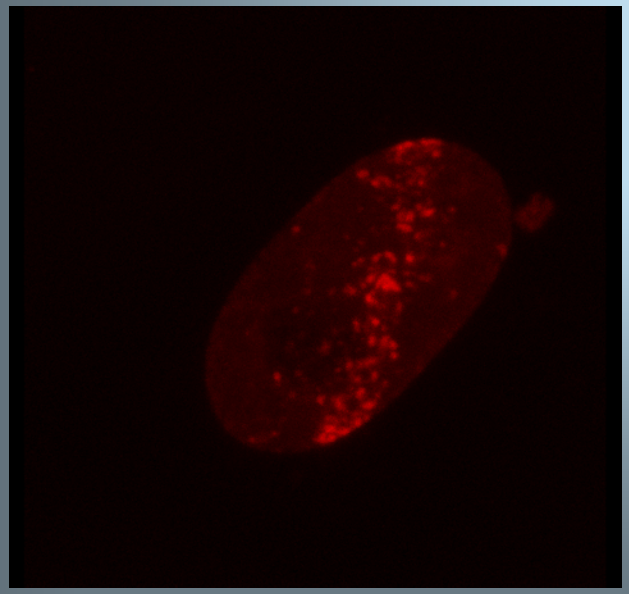
Scanning in 2D and 3D by confocal microscope

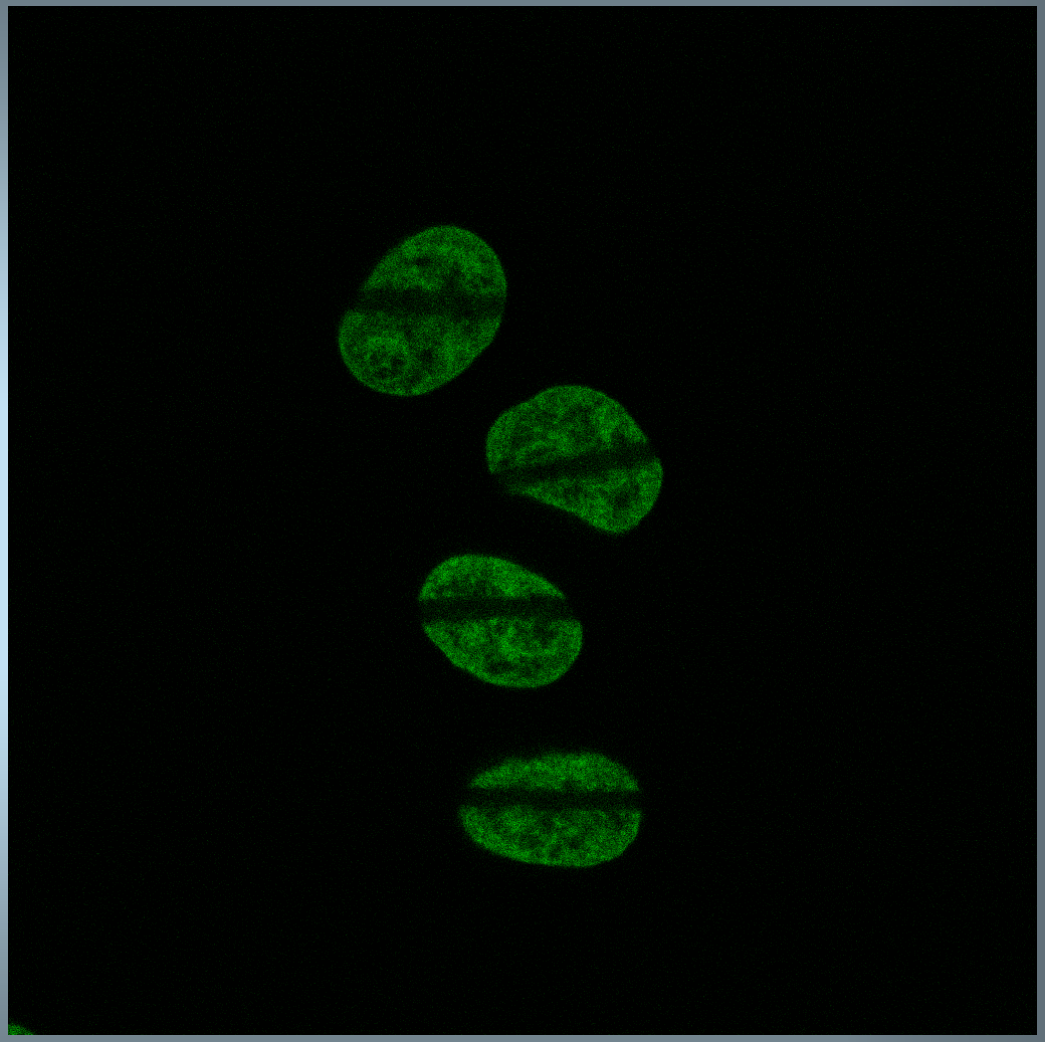


Laser beam moves firstly along *x* axis and then starts with new line in *y* axis.

Finishing scanning of one thin optical slice in *xy* plane, the scanning plane is moved in *z* axis to other slice







Subdiffraction Multicolor Imaging of the Nuclear Periphery with 3D Structured Illumination Microscopy

Lothar Schermelleh,[1*] Peter M. Carlton,[2*] Sebastian Haase,[2,4] Lin Shao,[2] Lukman Winoto,[2] Peter Kner,[2] Brian Burke,[3] M. Cristina Cardoso,[4] David A. Agard,[2] Mats G. L. Gustafsson,[5] Heinrich Leonhardt,[1*†] John W. Sedat[2*†]

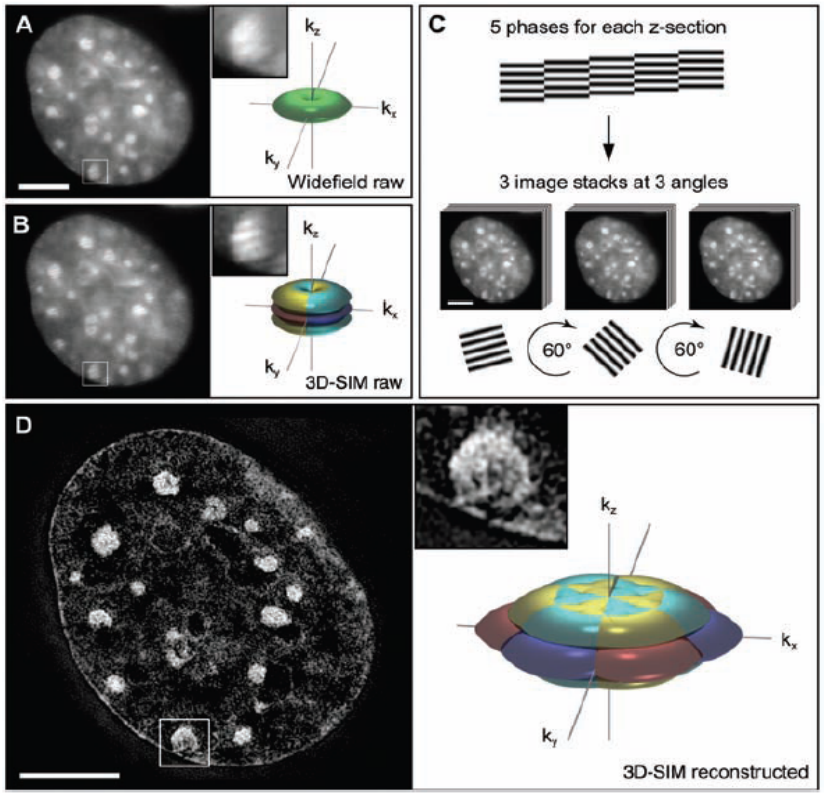
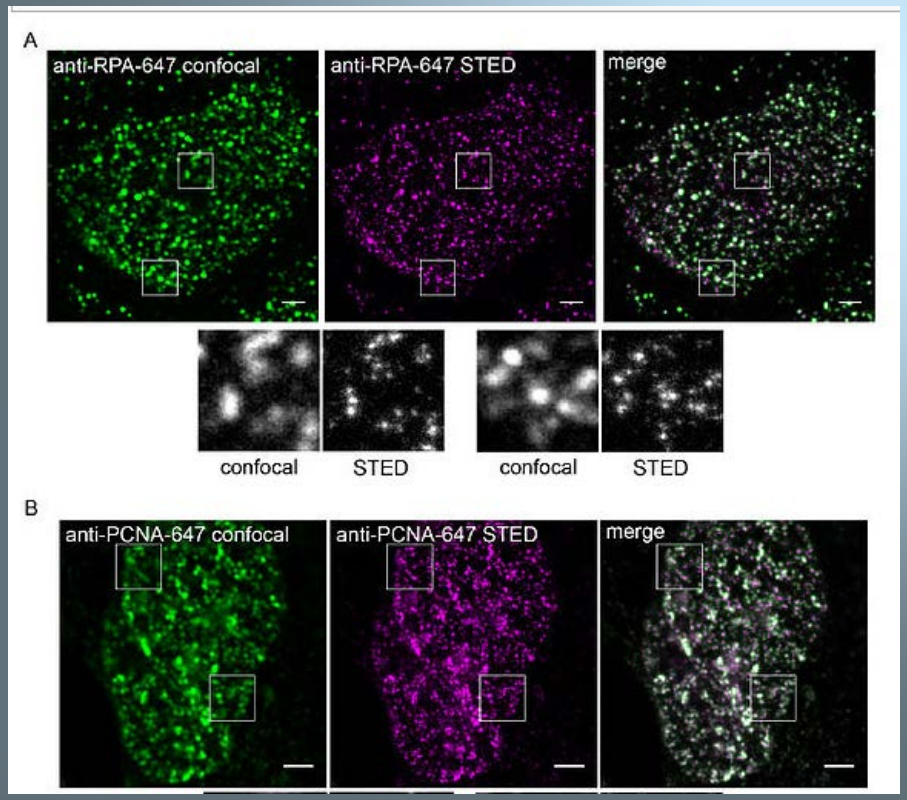


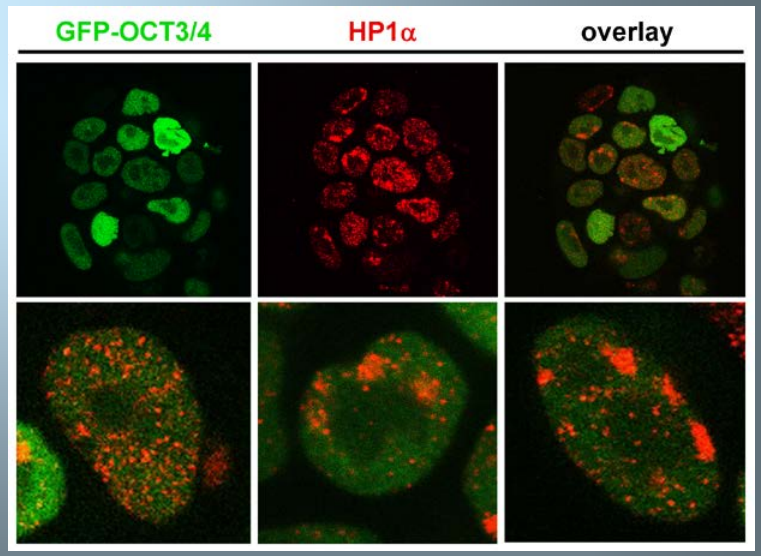
Fig. 1. Subdiffraction resolution imaging with 3D-SIM. (**A** and **B**) Cross section through a DAPI-stained C2C12 cell nucleus acquired with conventional wide-field illumination (A) and with structured illumination (B), showing the striped interference pattern (inset). The renderings to the right illustrate the respective support of detection in frequency space. The axes k_x, k_y, and k_z indicate spatial frequencies along the x, y, and z directions. The surfaces of the renderings represent the corresponding resolution limit. The depression of the frequency support ("missing cone") in z direction in (A) indicates the restriction in axial resolution of conventional wide-field microscopy. With 3D-SIM, the axial support is extended but remains within the resolution limit. (**C**) Five phases of the sine wave pattern are recorded at each z position, allowing the shifted components to be separated and returned to their proper location in frequency space. Three image stacks are recorded with the diffraction grating sequentially rotated into three positions 60° apart, resulting in nearly rotationally symmetric support over a larger region of frequency space. (**D**) The same cross section of the reconstructed 3D-SIM image shows enhanced image details compared with the original image (insets). The increase in resolution is shown in frequency space on the right, with the coverage extending two times farther from the origin. Scale bars indicate 5 μm.

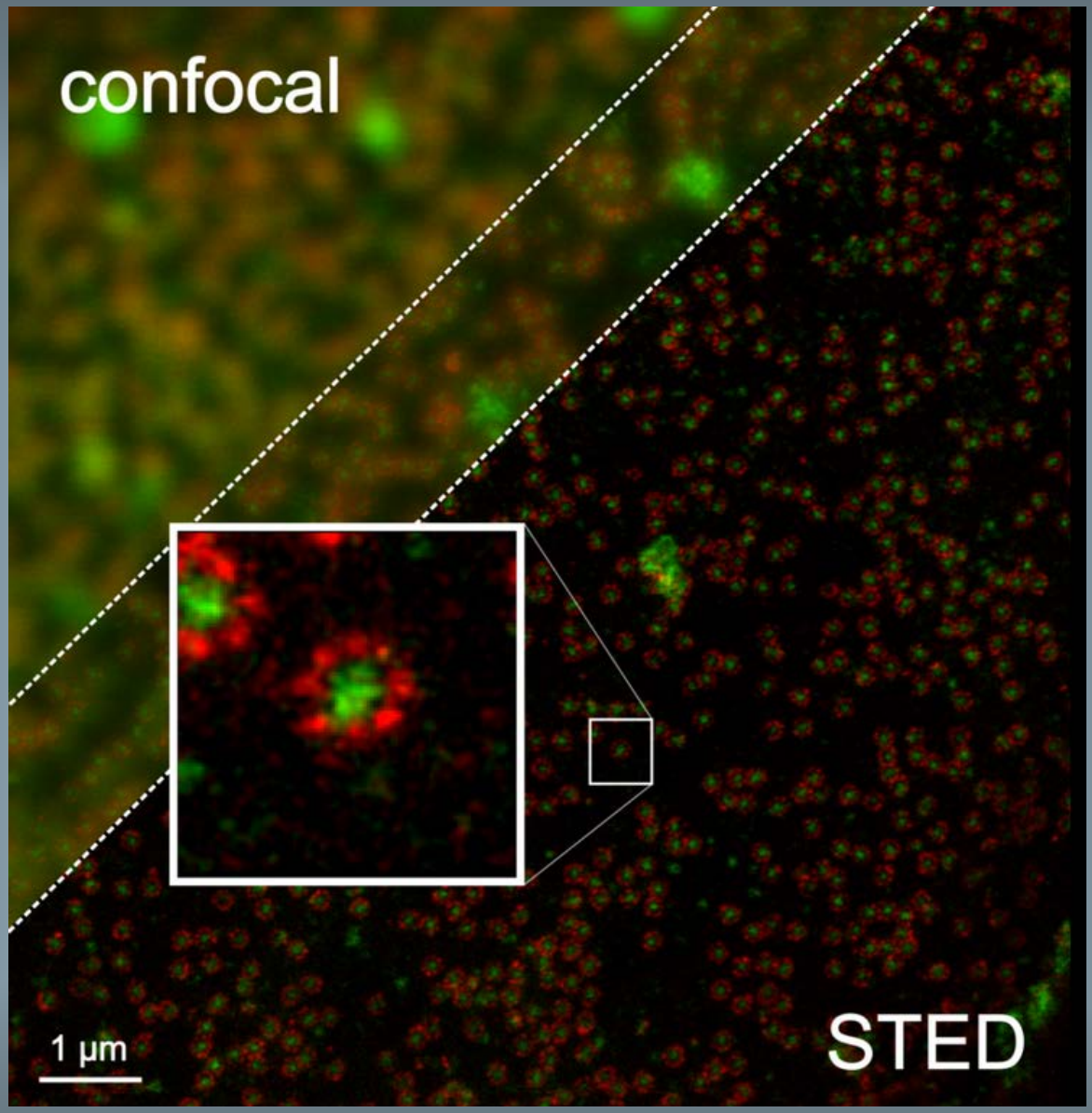
Stimulated Emission Depletion microscopy, or STED microscopy, is a fluorescence microscopy technique that uses the non-linear de-excitation of fluorescent dyes to overcome the resolution limit imposed by diffraction with standard confocal laser scanning microscopes and conventional far-field optical microscopes.

STED

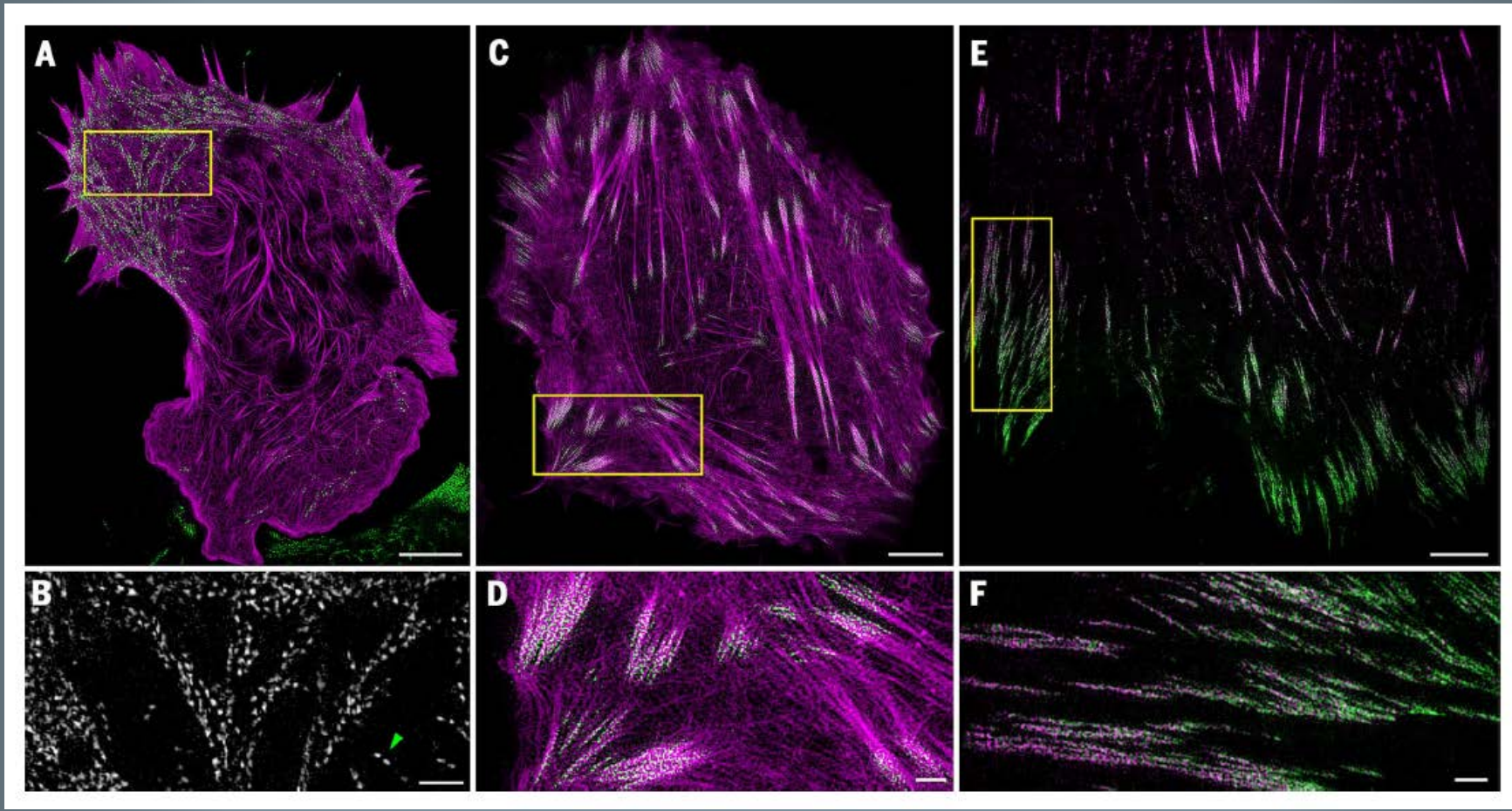


SP-5 LSCM




confocal
1 µm
STED

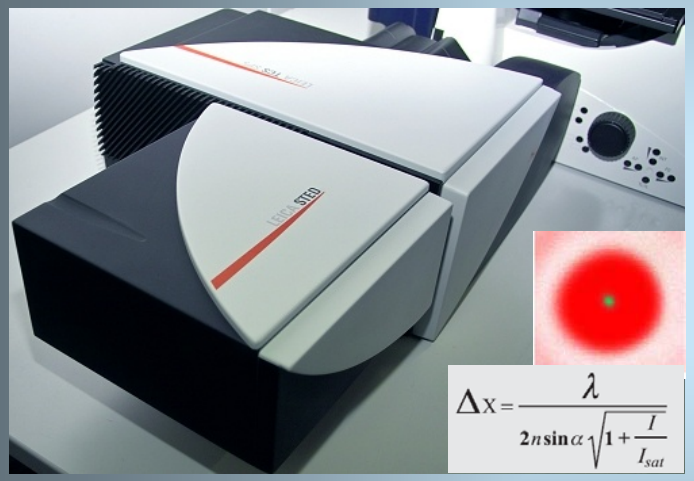
High-speed live-cell imaging

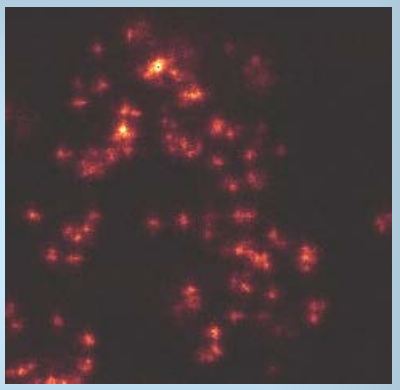


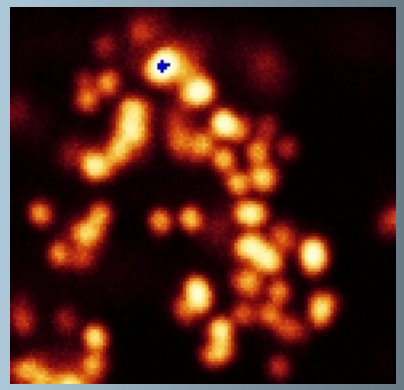
Li et al., Science (2015)

Leica STED – STimulated Emission Depletion
SUPERRESOLUTION (subdiffraction) in xy plane

Hell, S. W. and J. Wichmann (1994). *Opt. Lett.*
"Breaking the diffraction resolution limit by stimulated emission"







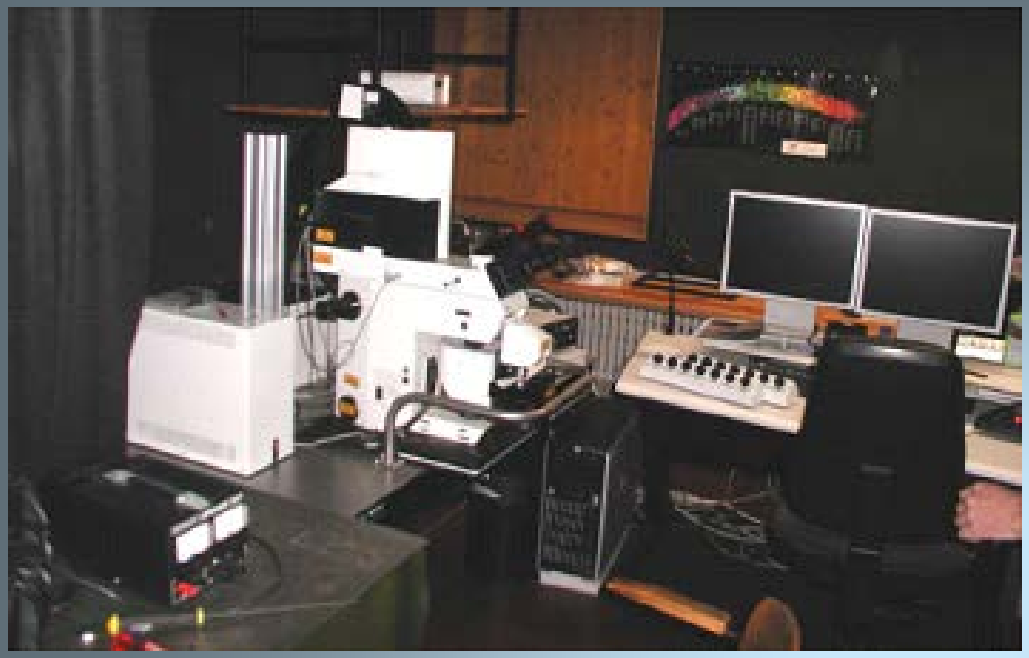
neurobiology
membrane biology
membrane rafts
intracellular transport

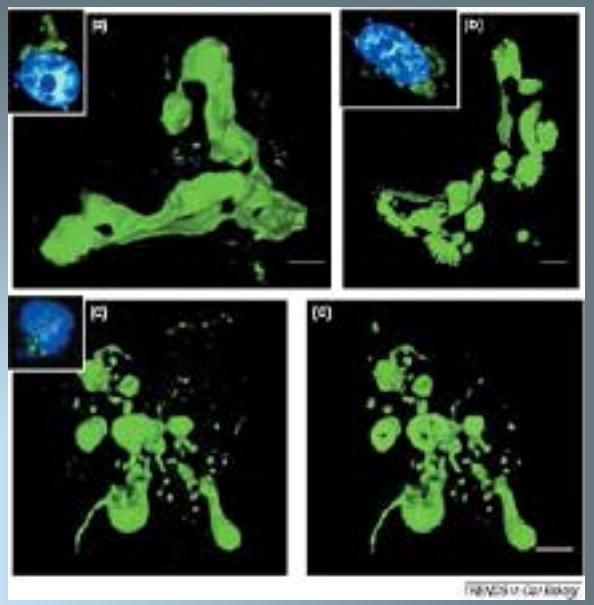
Willig KI et al. *Nature* 2006
Sieber JJ et al. *Biophy J* 2006
Kittel RJ et al. *Science* 2006
Fitzner D et al. *EMBO J* 2006

Kellner RR et al. *Neurosience* 2007
Lin W et al. *PNAS* 2007
Seebach J *Cardiovas. Res.* 2007
Sieber JJ *Science* 2007

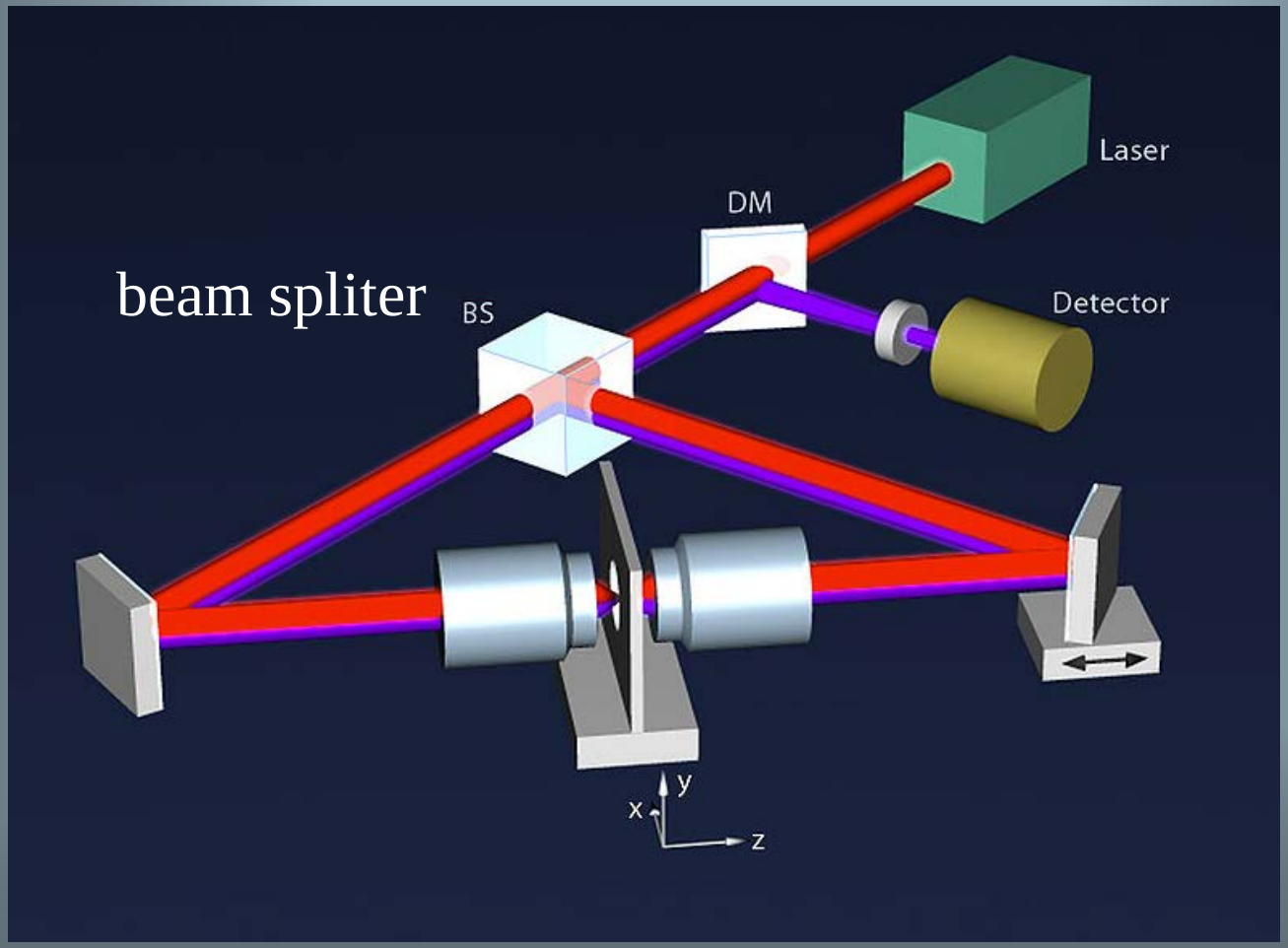
Unique

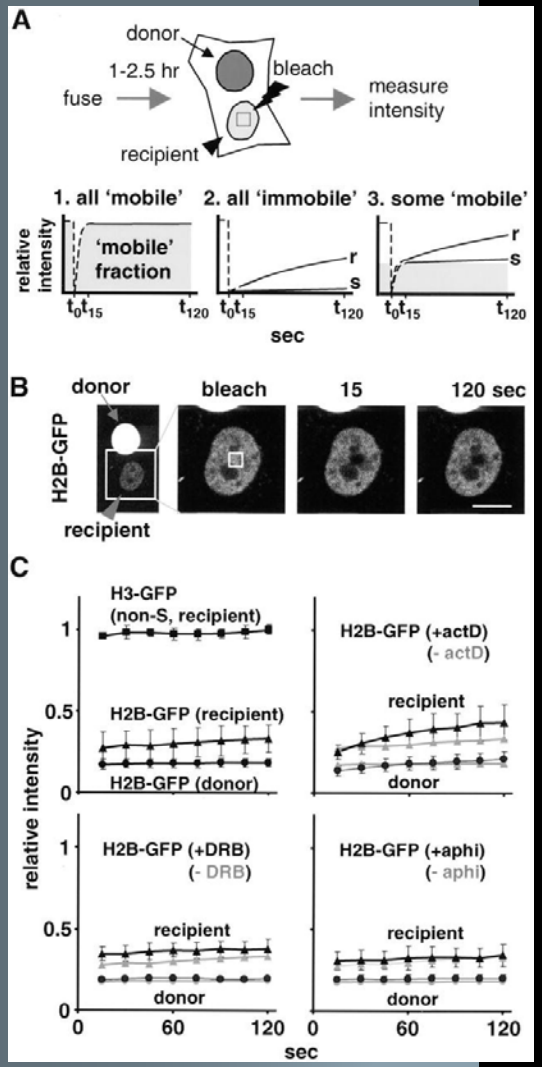
4-pi microscopy (CC lab in Heidelberg)

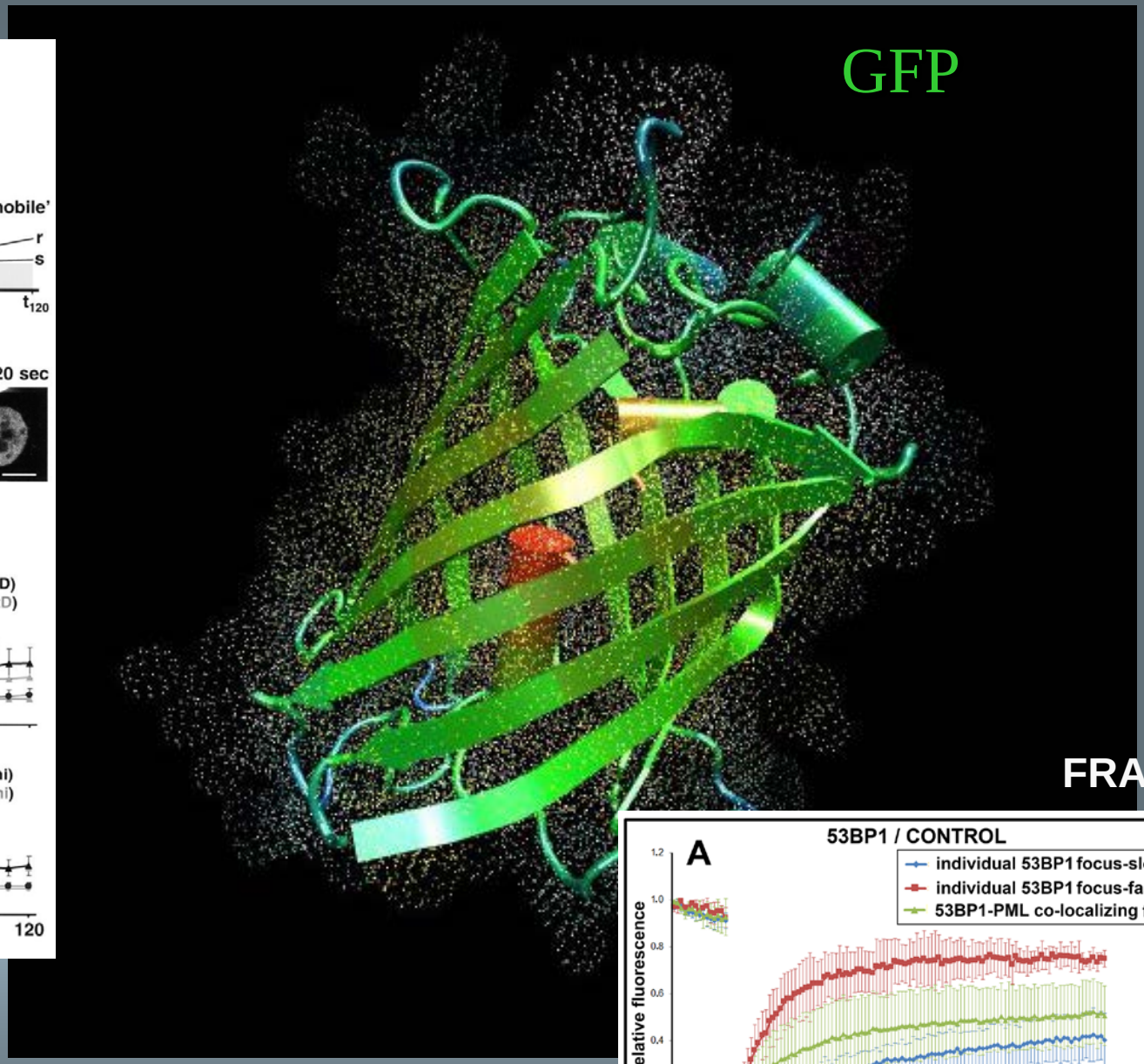




4pi: improved axial resolution. The typical value of 500-700 nm can be improved to 100-150 nm which corresponds to an almost spherical focal spot with 5-7 times less volume than that of standard confocal microscopy.



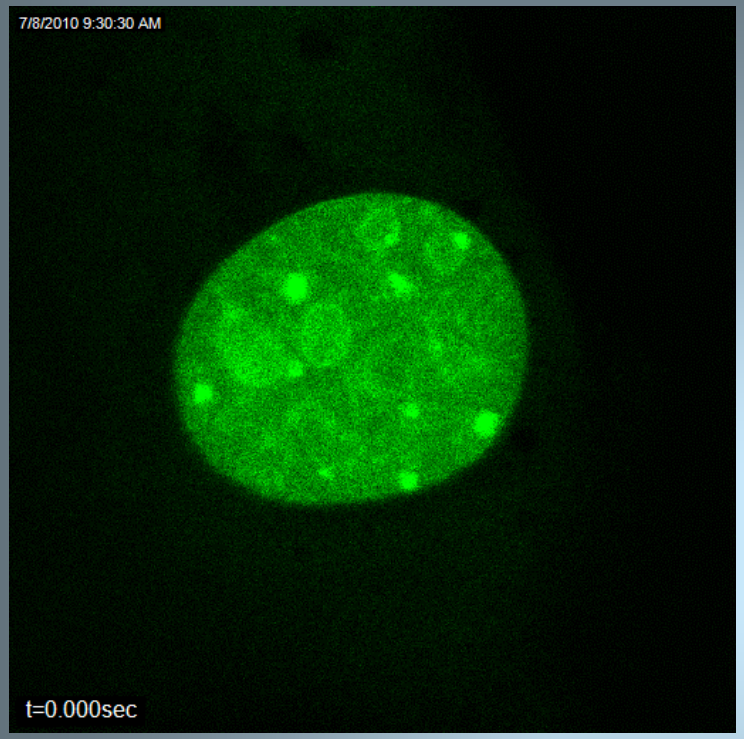


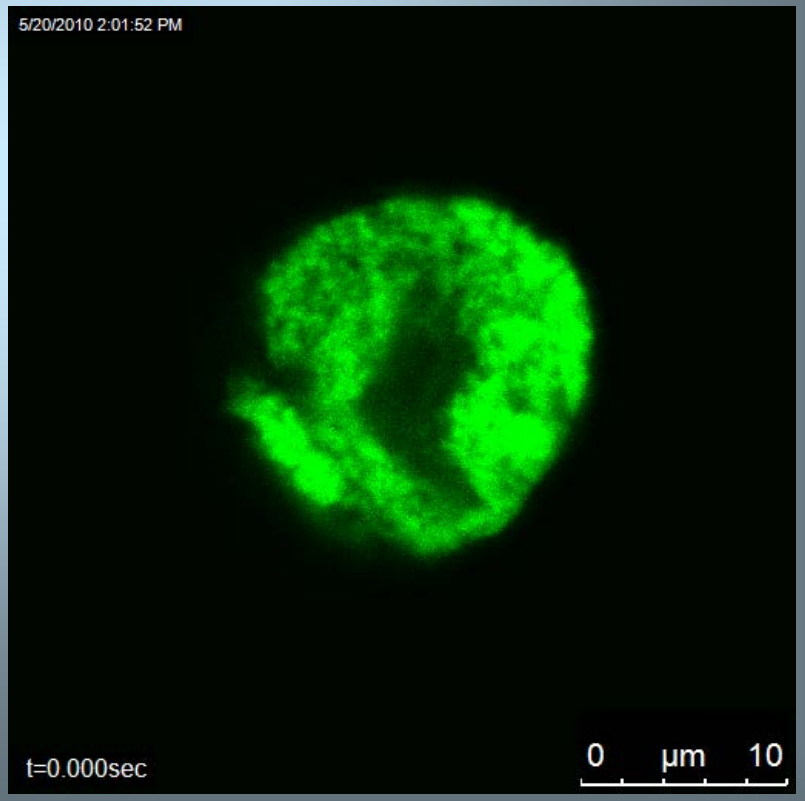


JCB

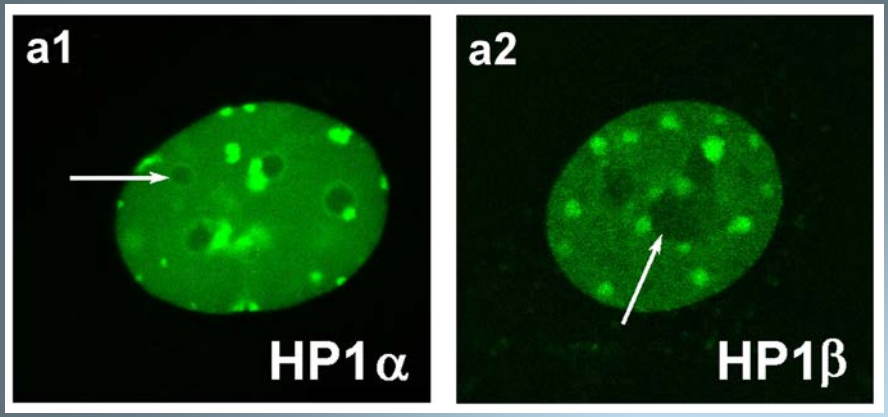
Kimura and Cook (2001)

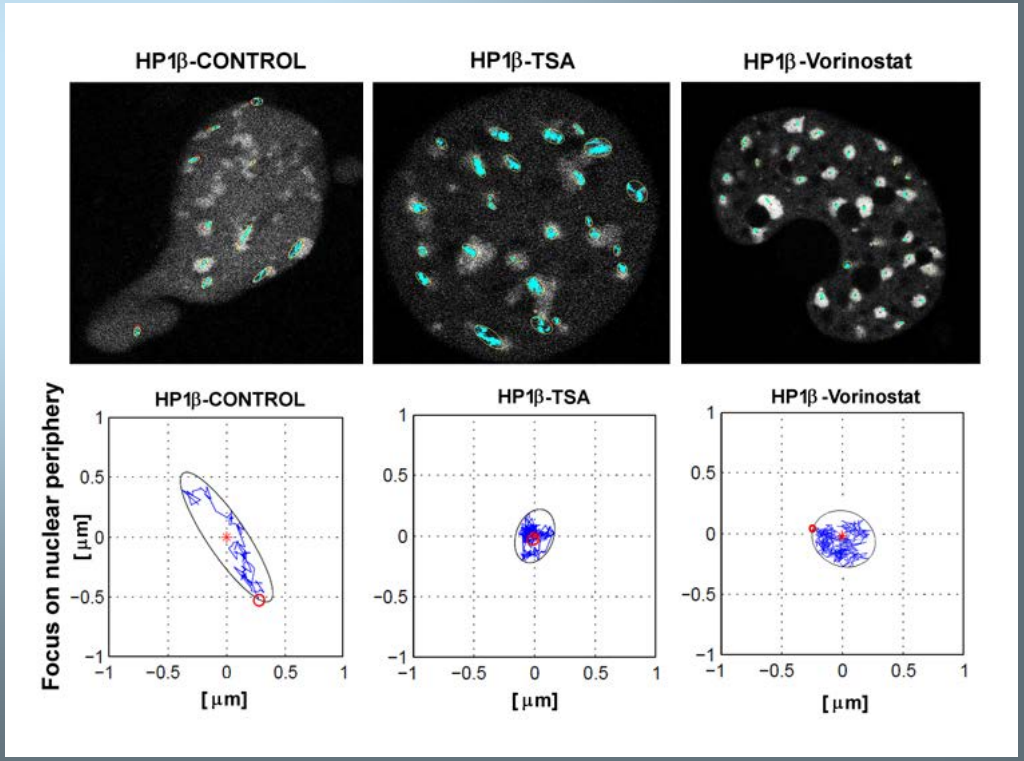
FRAP results on Jmjd2b-tagged by GFP



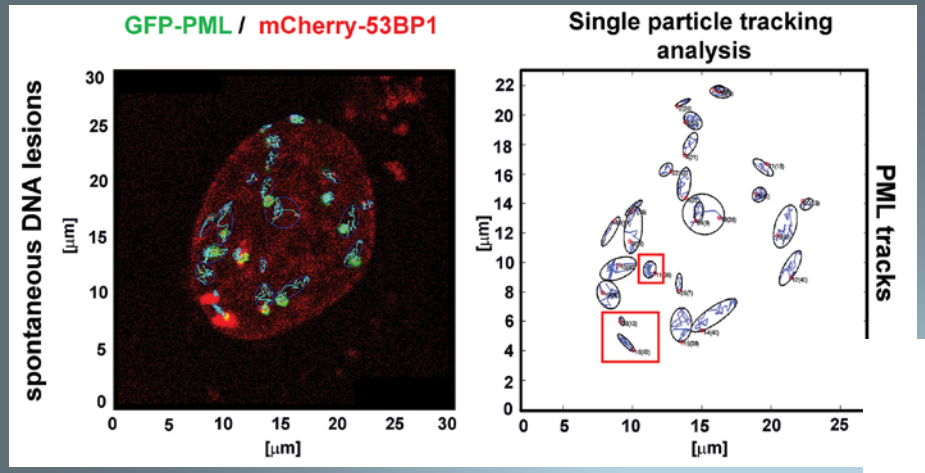


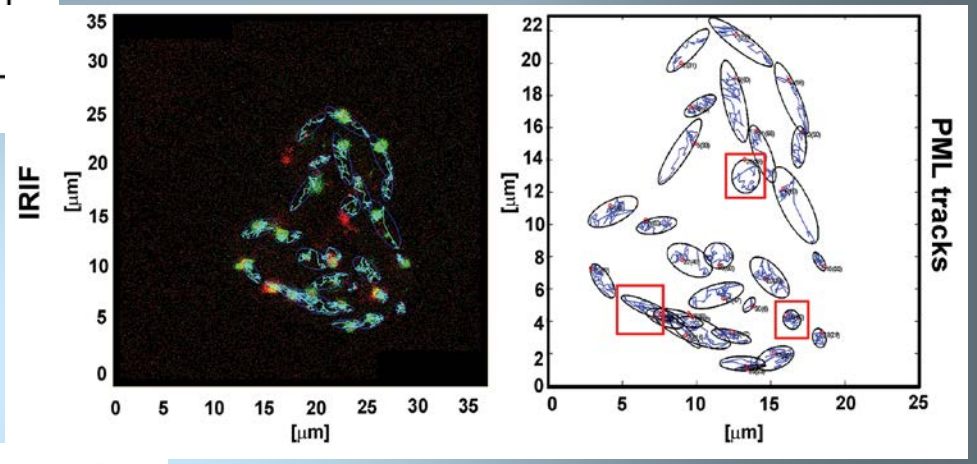
Single particle tracking analysis

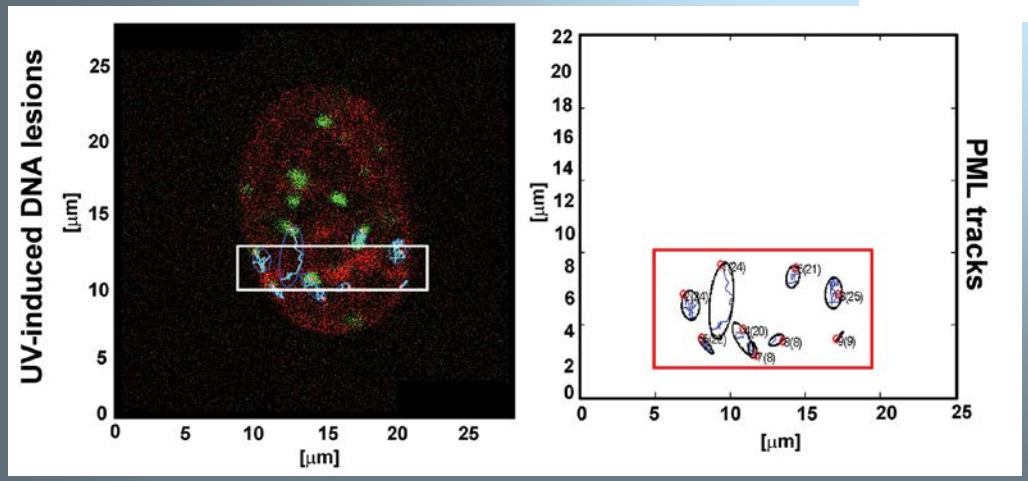




Experiments of Lenka Stixová and Pavel Matula

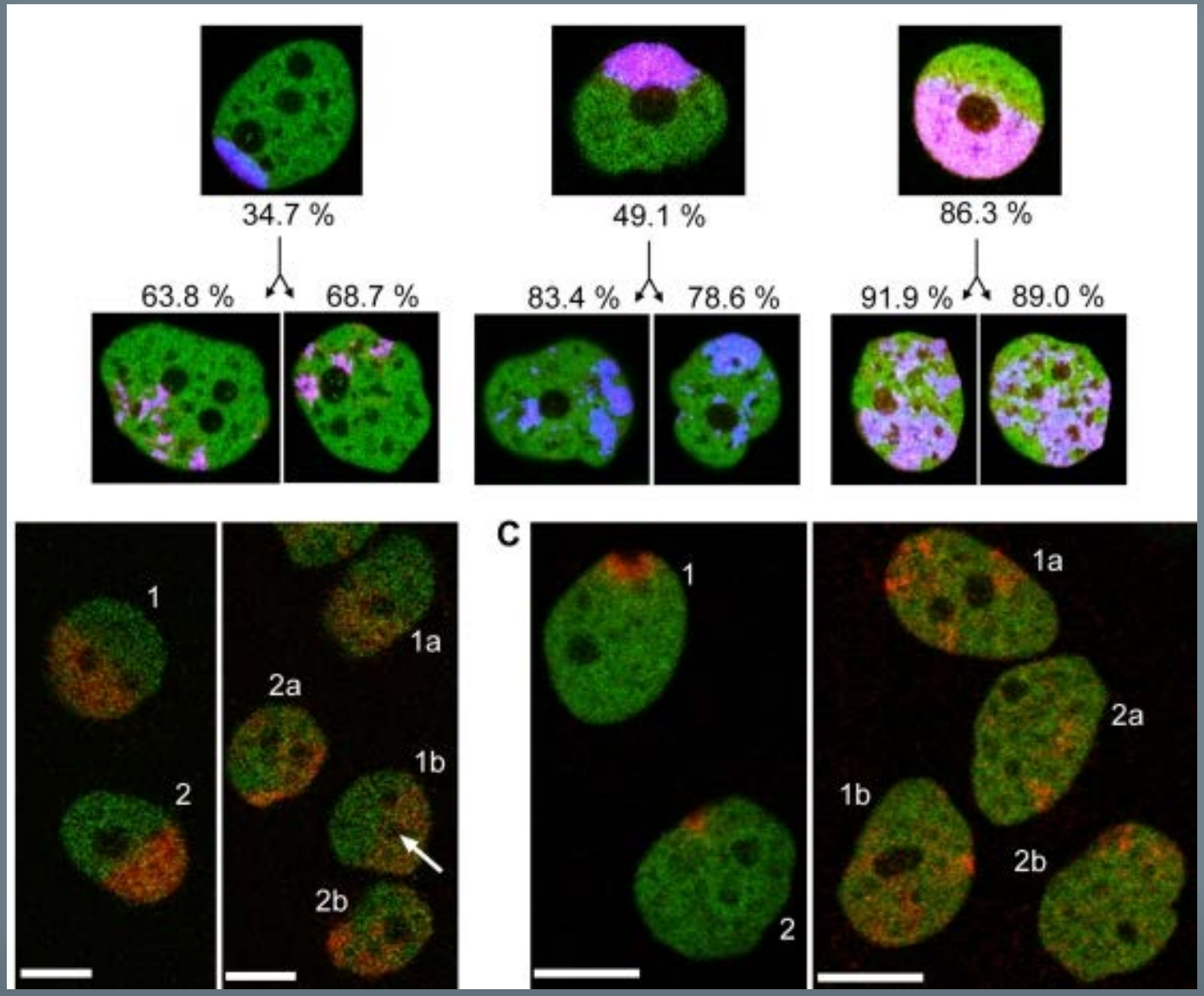




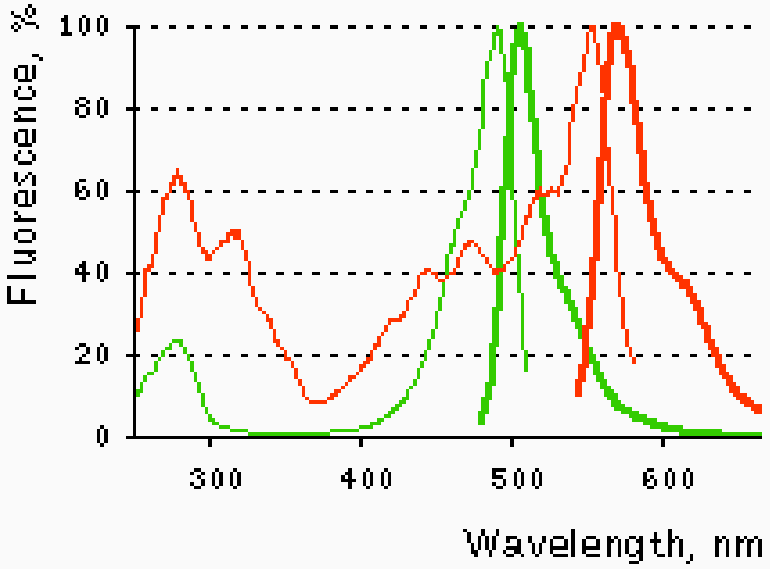


Experiments of Veronika Foltanková and Dmitry Sorokin

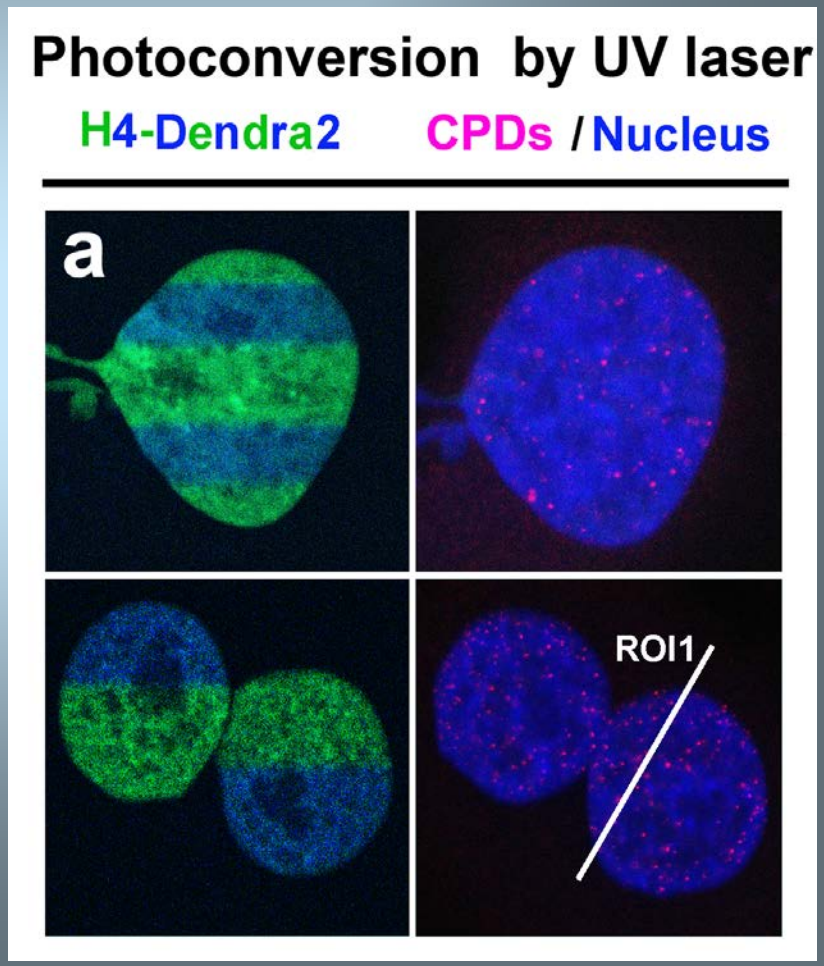
Dendra2 photo-conversion



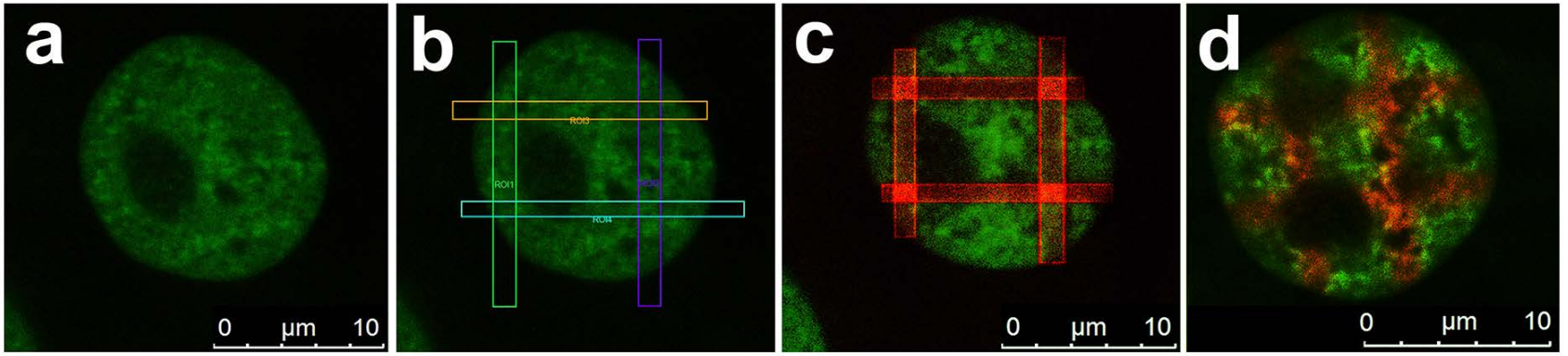
Dendra2 is an improved version of a green-to-red photoswitchable fluorescent protein Dendra, derived from octocoral *Dendronephthya* sp. (Gurskaya *et al.*, 2006).



Normalized excitation (thin line) and emission (thick line) for non-activated (green) and activated (red) spectra.



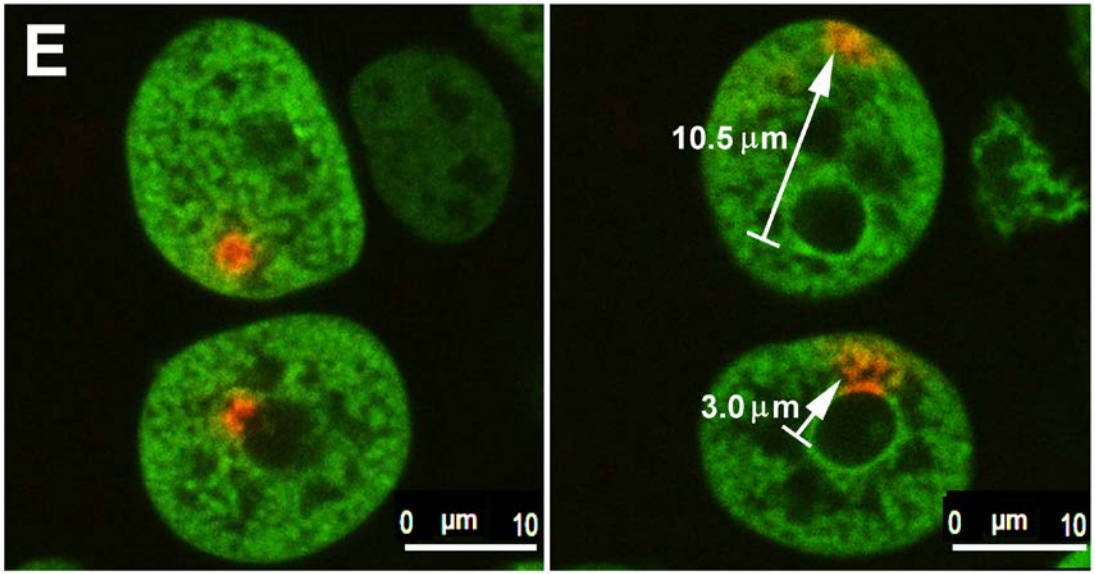
Dendra2-H4 photoconversion



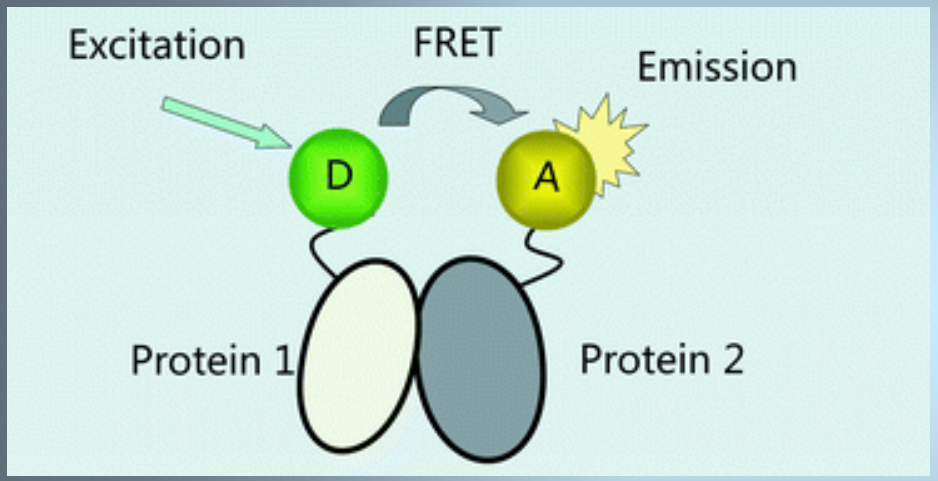
H4-Dendra2 / H4-Dendra2

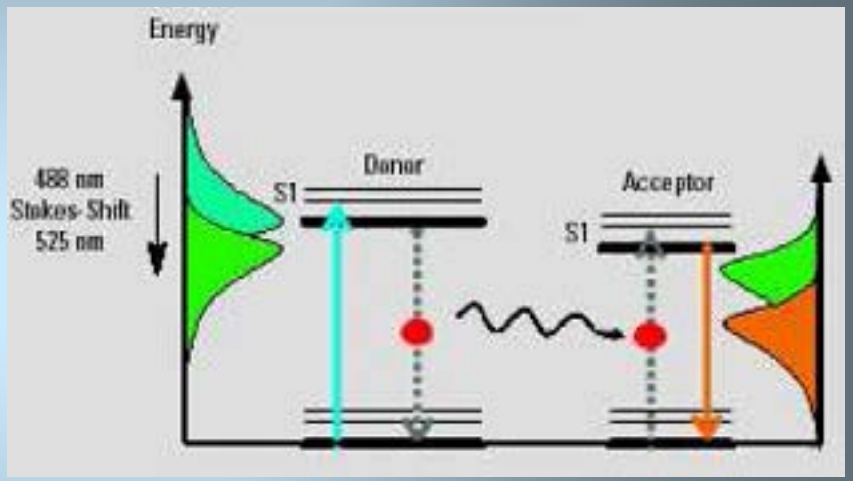
Actinomycin-D
(0 min. after photoconversion)

Actinomycin-D (90 min.)

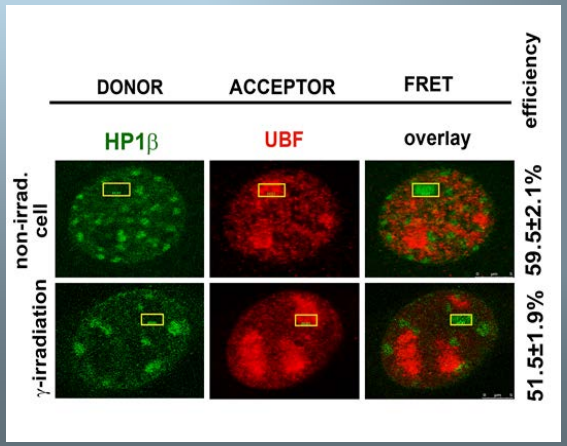


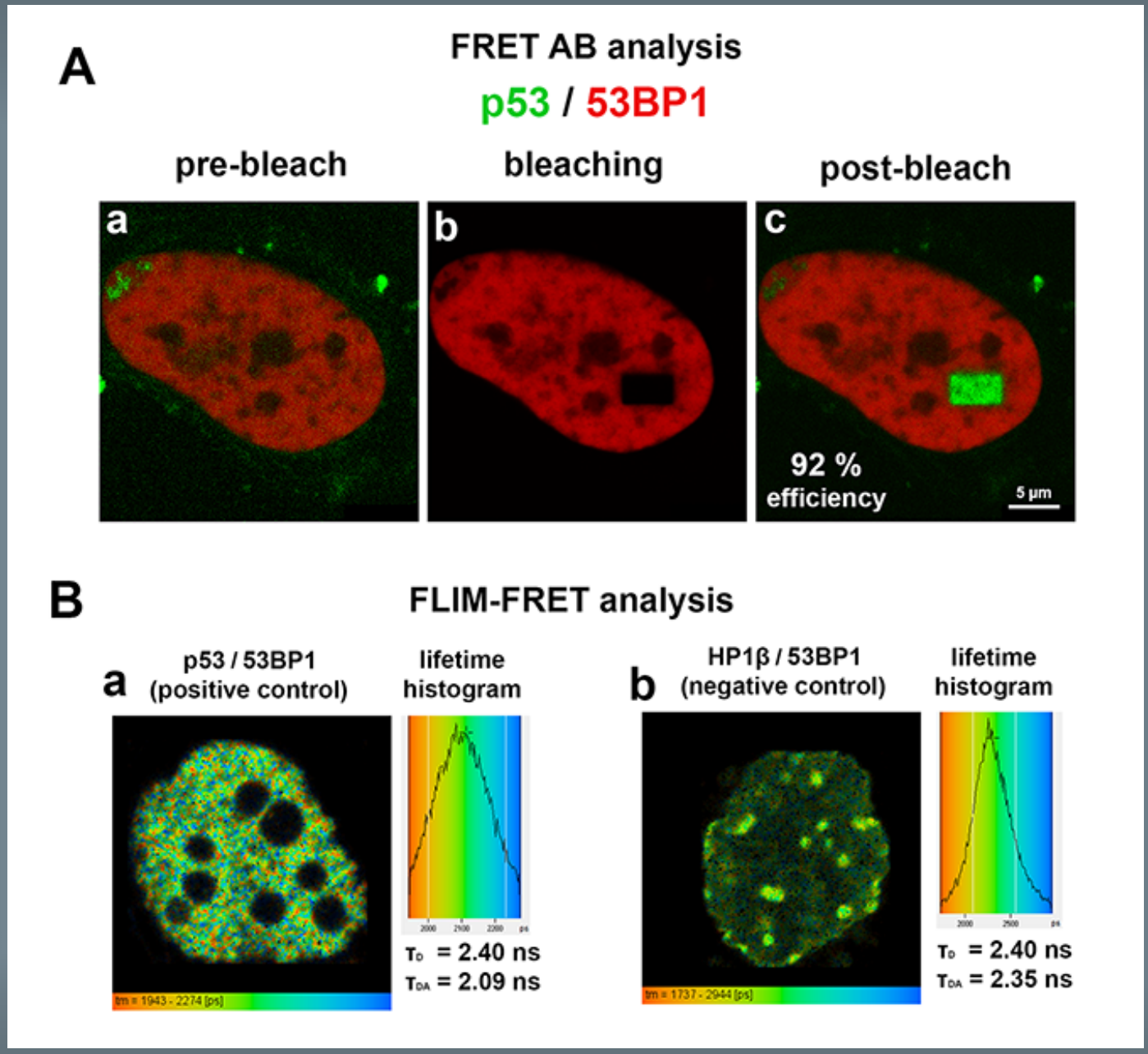
FRET (Fluorescence Resonance Energy Transfer) is a technique for measuring interactions between two proteins in vivo. In this technique, two different fluorescent molecules (fluorophores) are genetically fused the two proteins of interest.

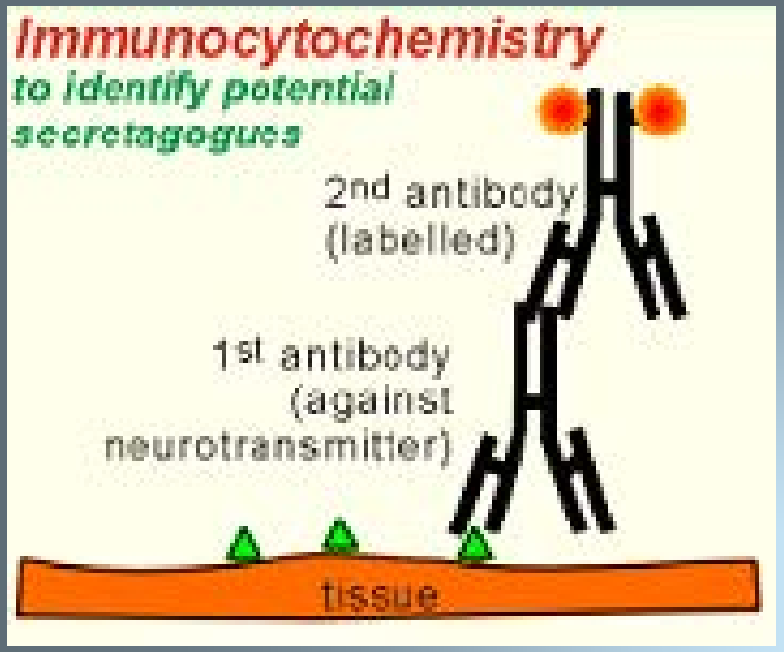




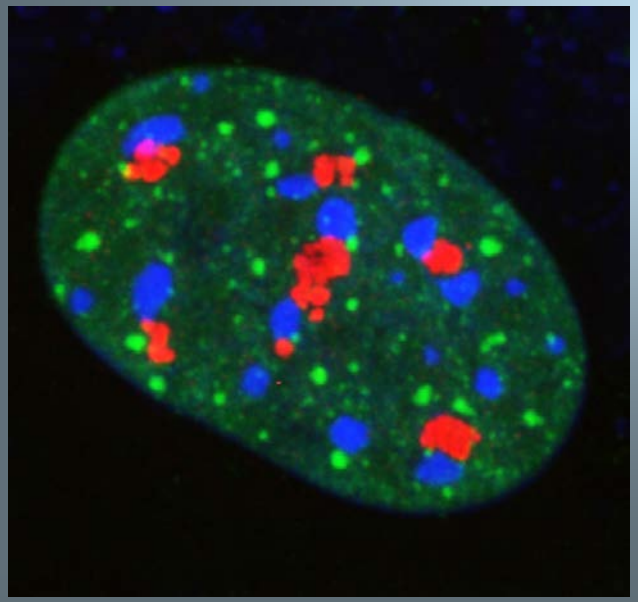
http://www.rsc.org/publishing/journals/

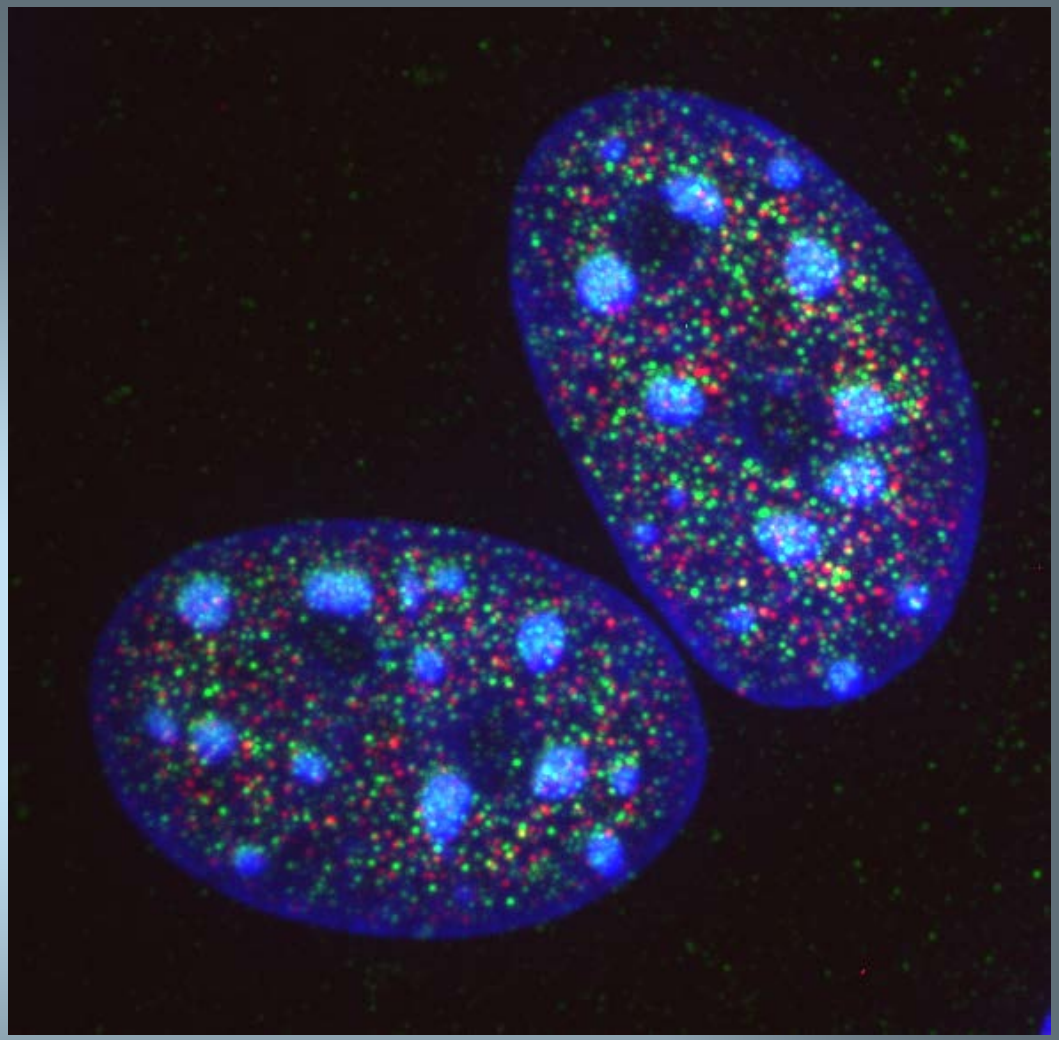



A
FRET AB analysis
p53 / 53BP1
pre-bleach
bleaching
post-bleach
a
b
c
92 %
efficiency
5 µm
B
FLIM-FRET analysis
a
p53 / 53BP1
(positive control)
lifetime
histogram
τD = 2.40 ns
τDA = 2.09 ns
b
HP1β / 53BP1
(negative control)
lifetime
histogram
τD = 2.40 ns
τDA = 2.35 ns



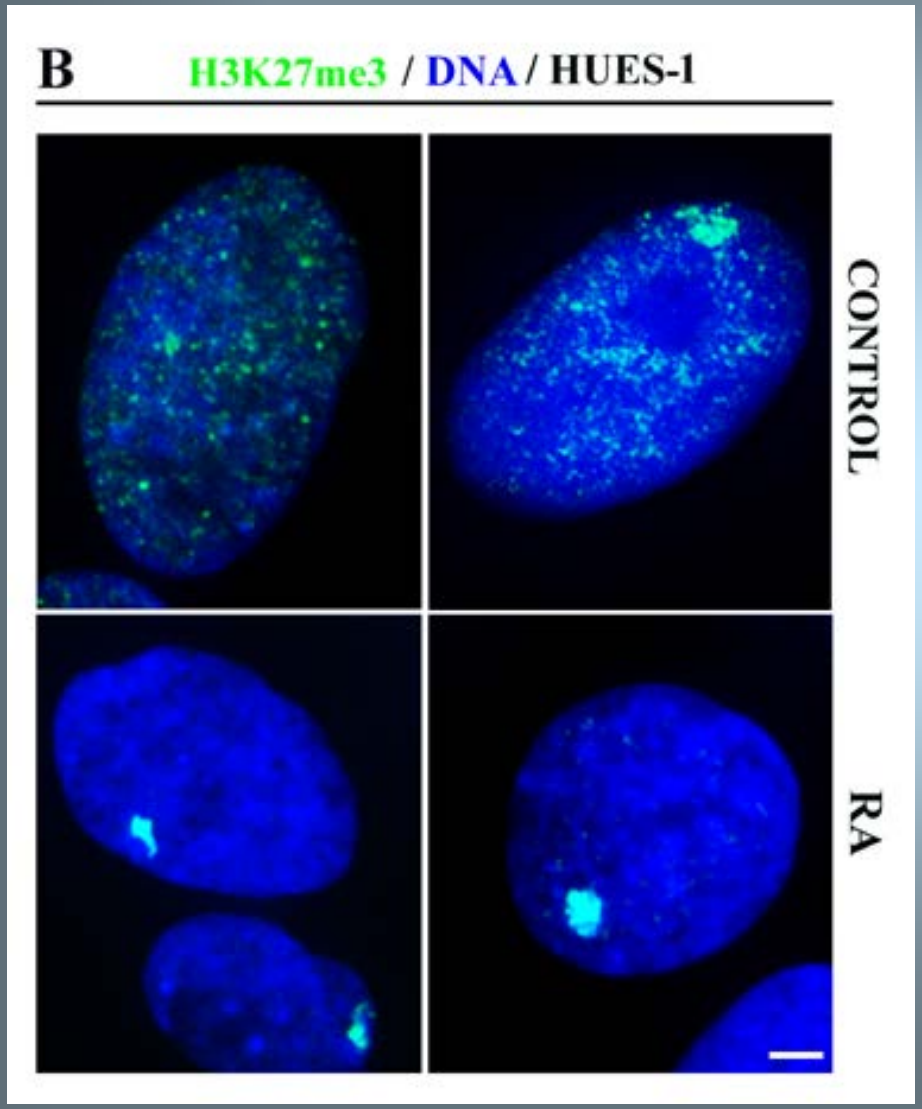
http://www.celanphy.science.ru.nl/Bruce%20web/construction.htm

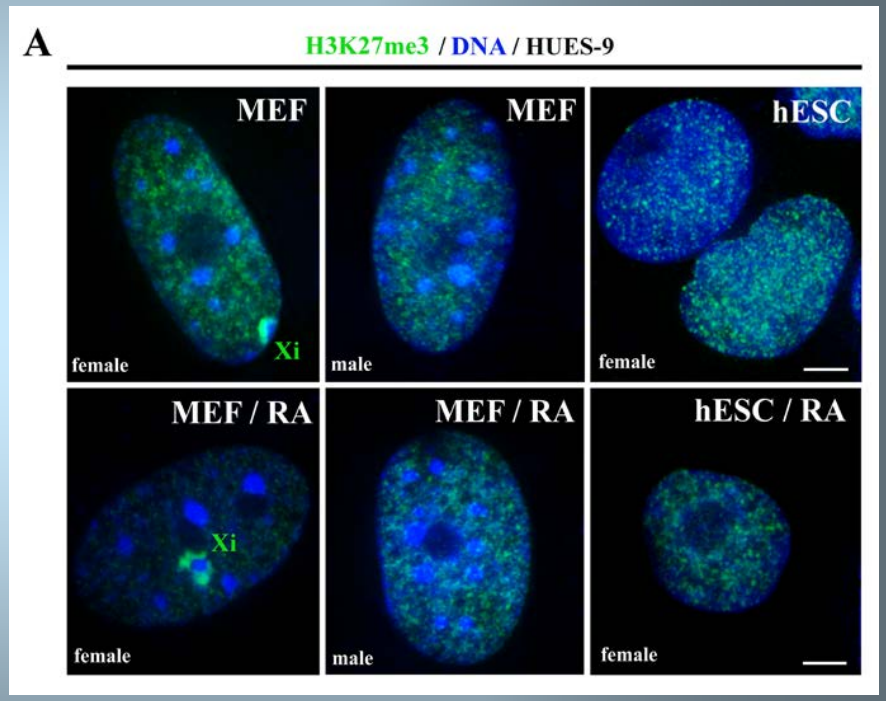


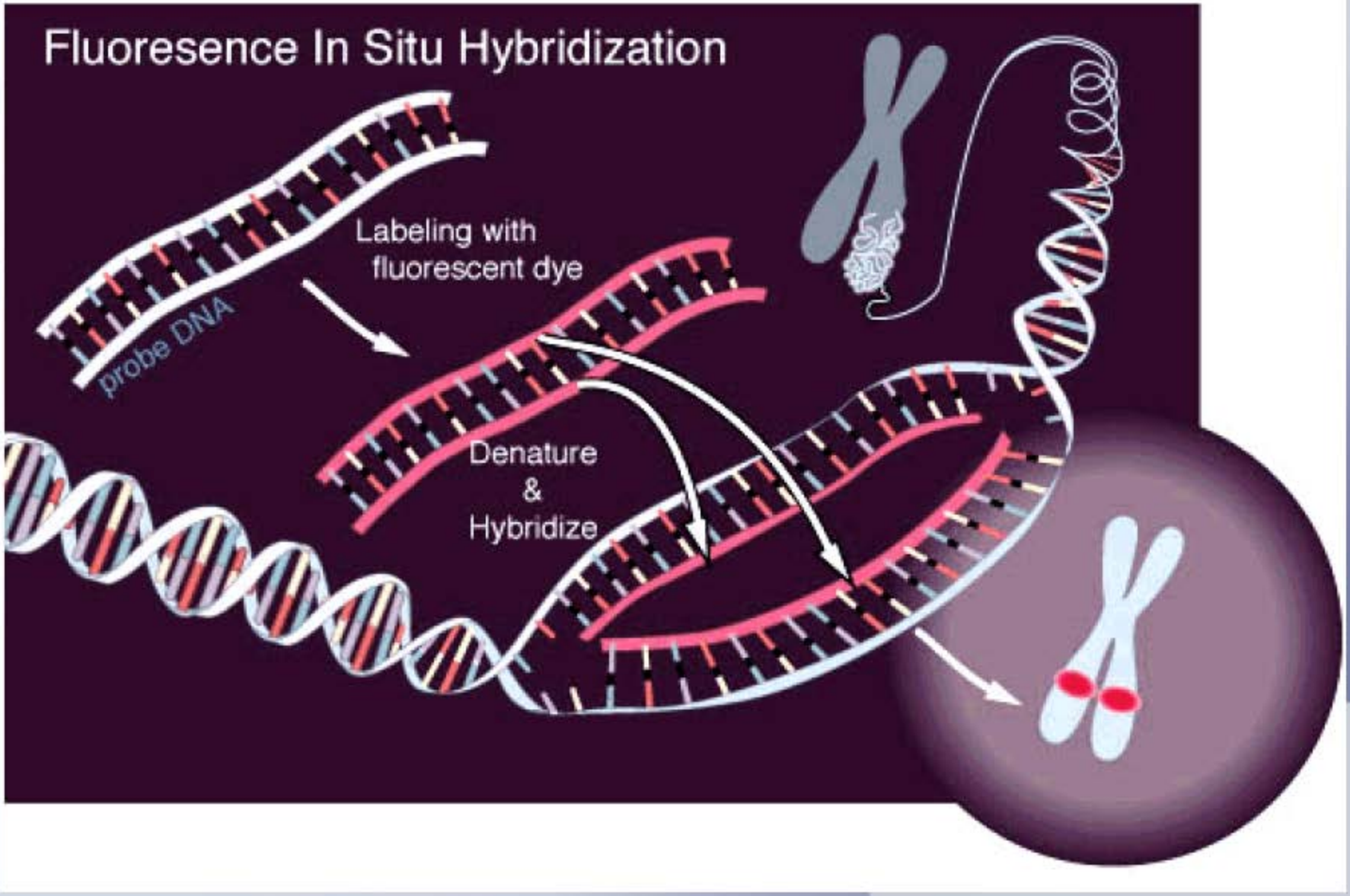


EB group, IBP, Brno

Inactivation of X chromosome in hESCs in comparison to MEFs

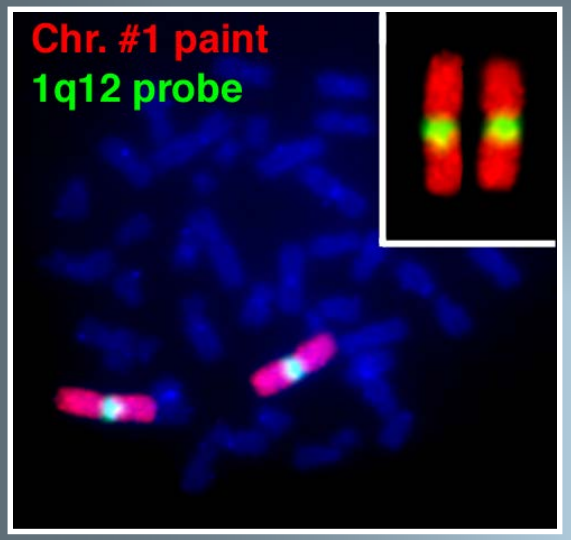


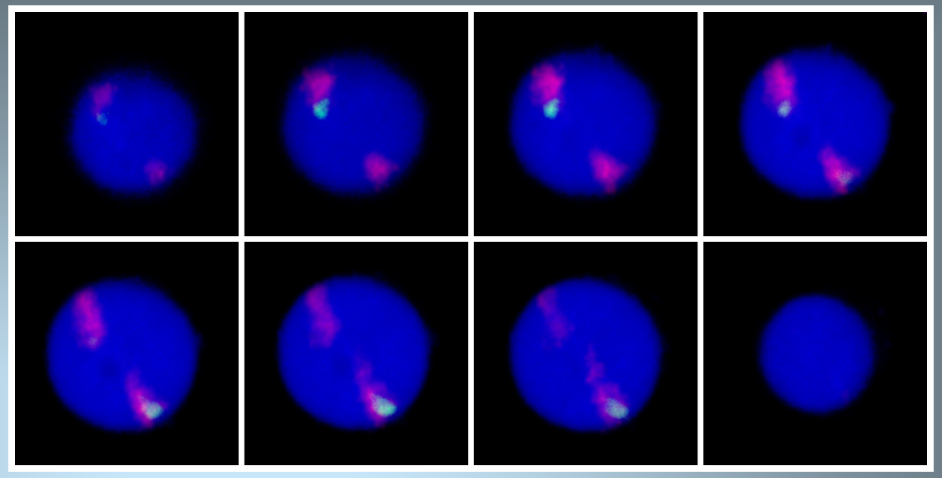




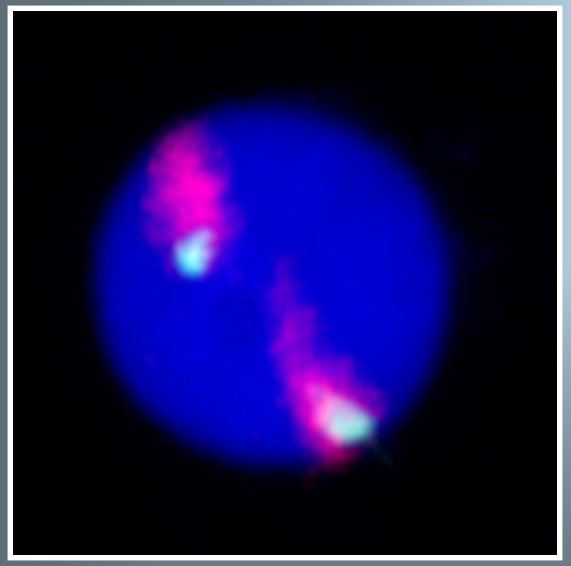
http://www.accessexcellence.org/RC/VL/GG/fish.html

3D-FISH a konfokální mikroskopie

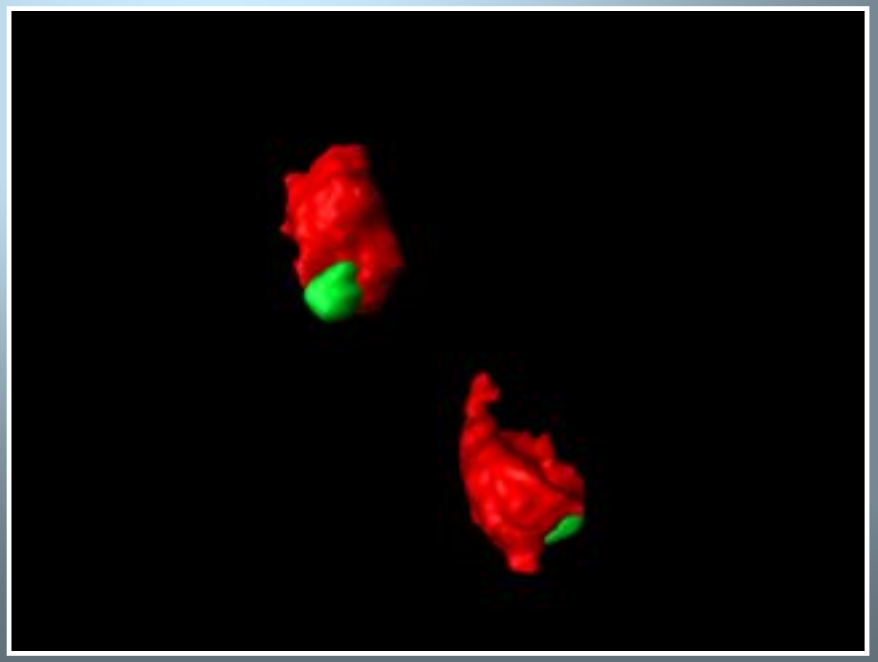




Galerie optických řezů

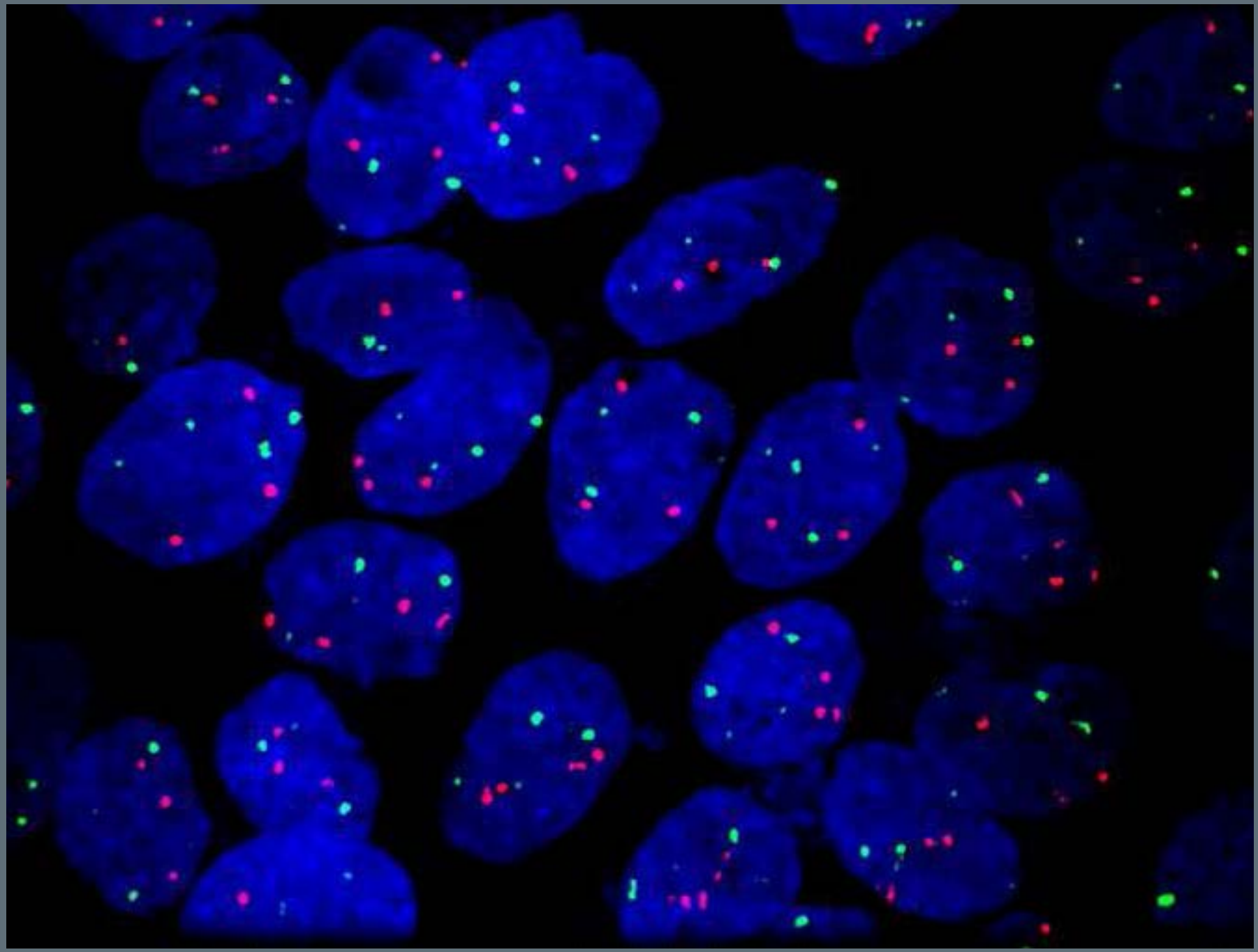


Maximální obraz
Všech řezů



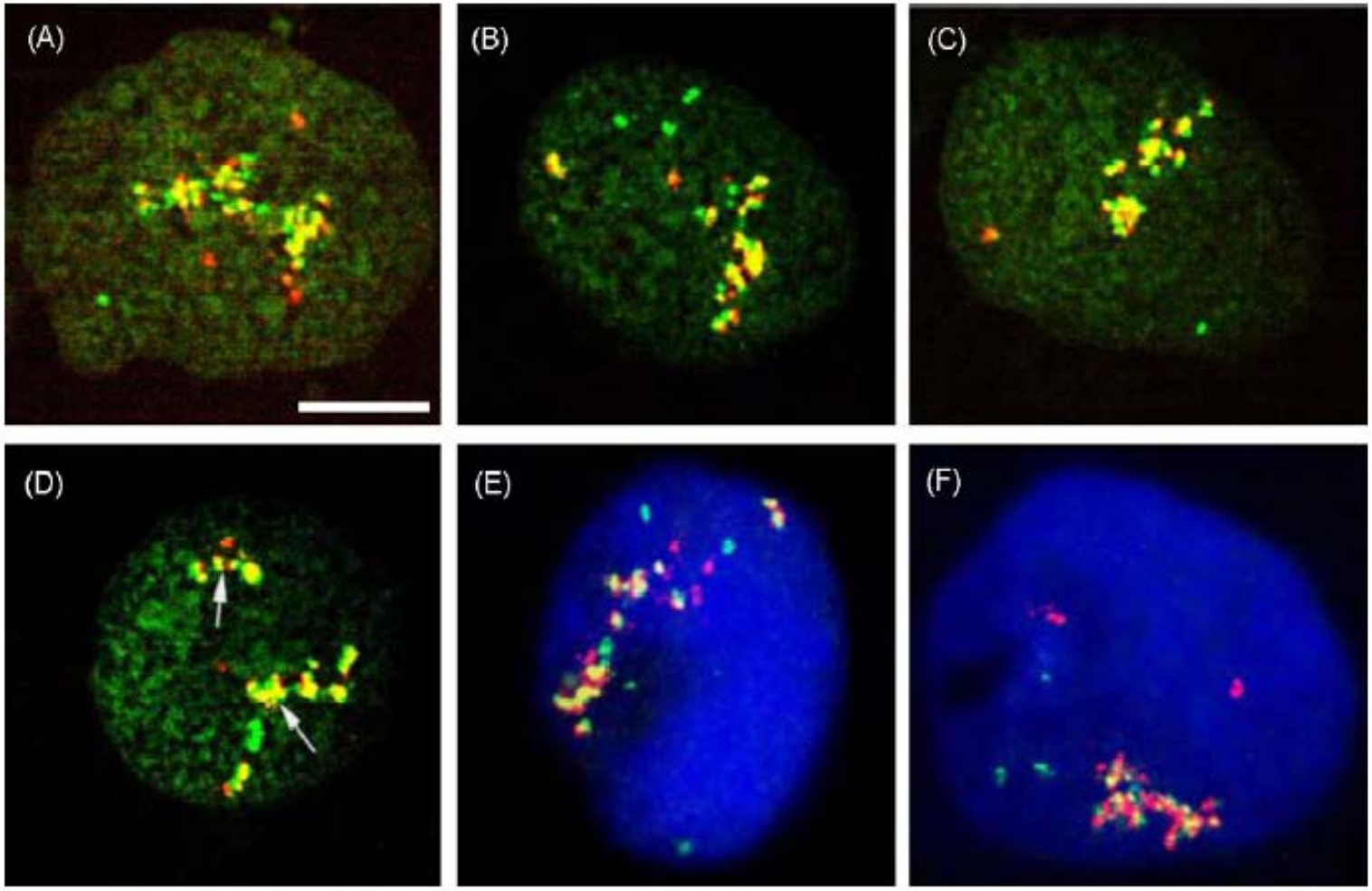
3D reconstrukce CT

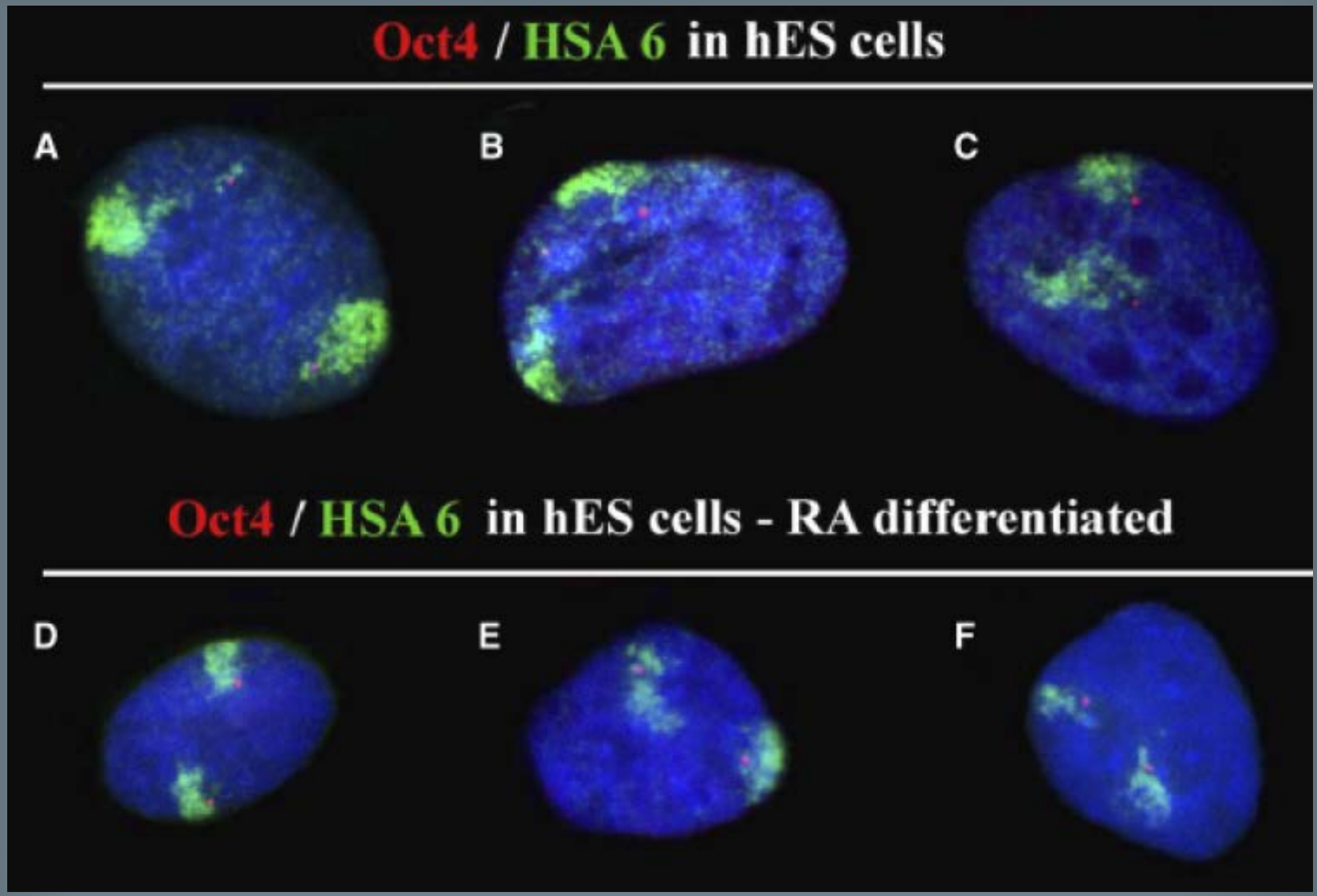
Weierich et al., (2003) in press



K562 cells t(9;22)

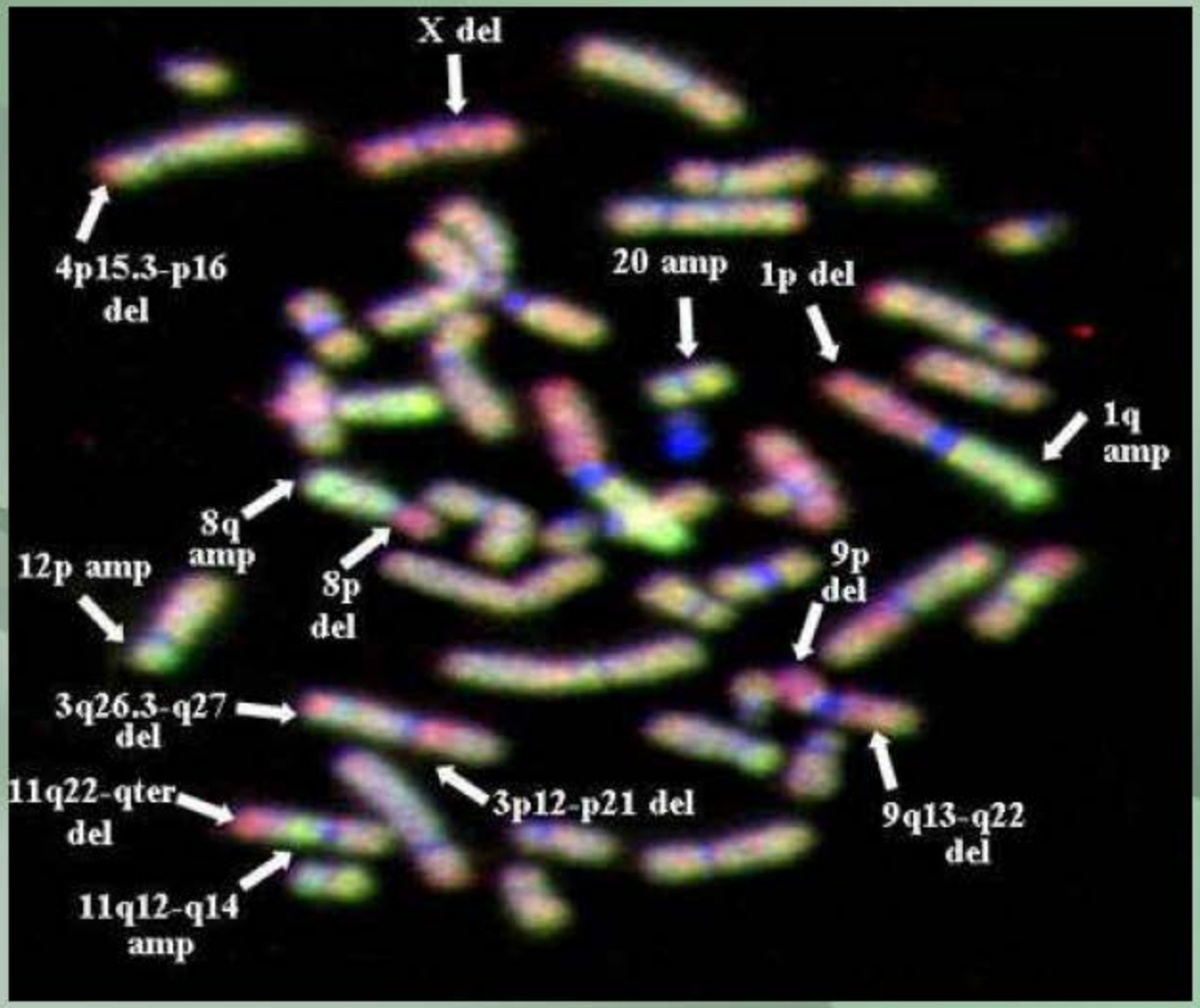
E. Bártová et al. / Leukemia Research 29 (2005) 901–913



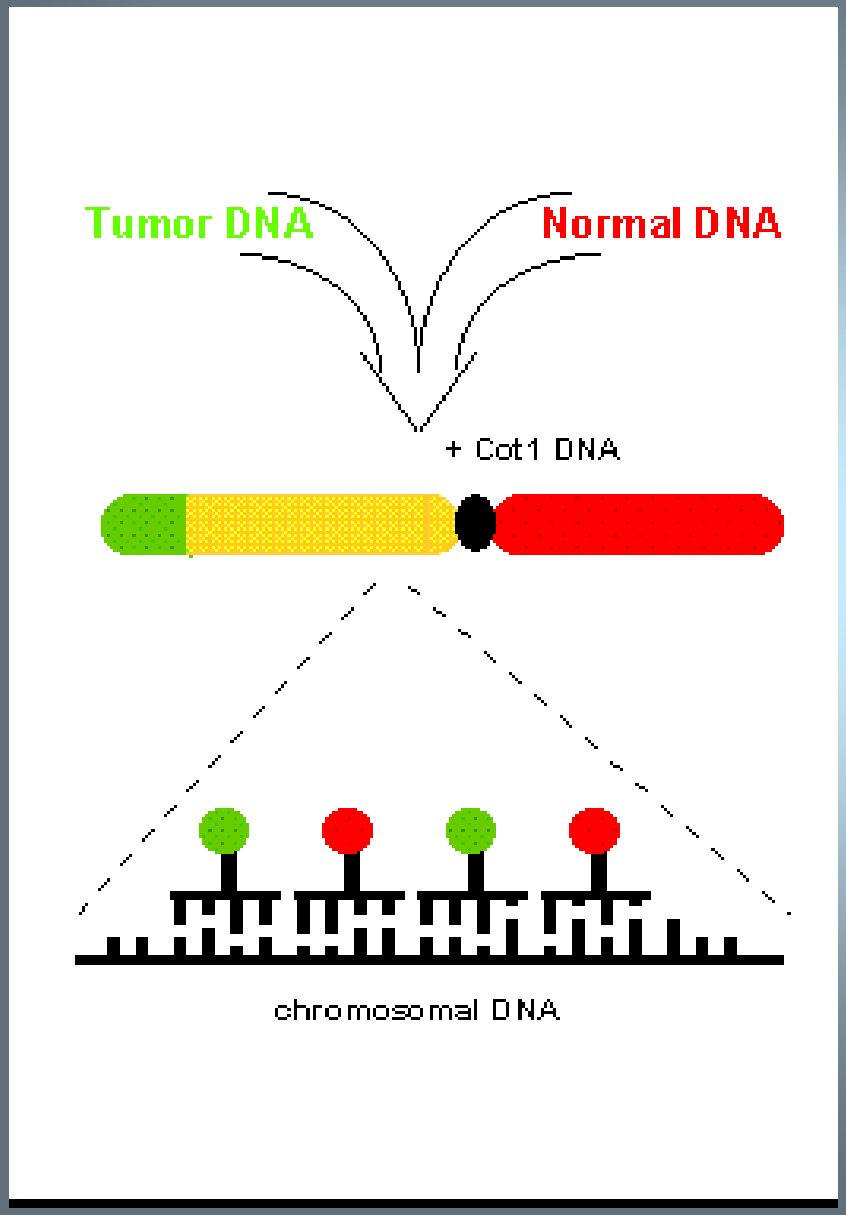


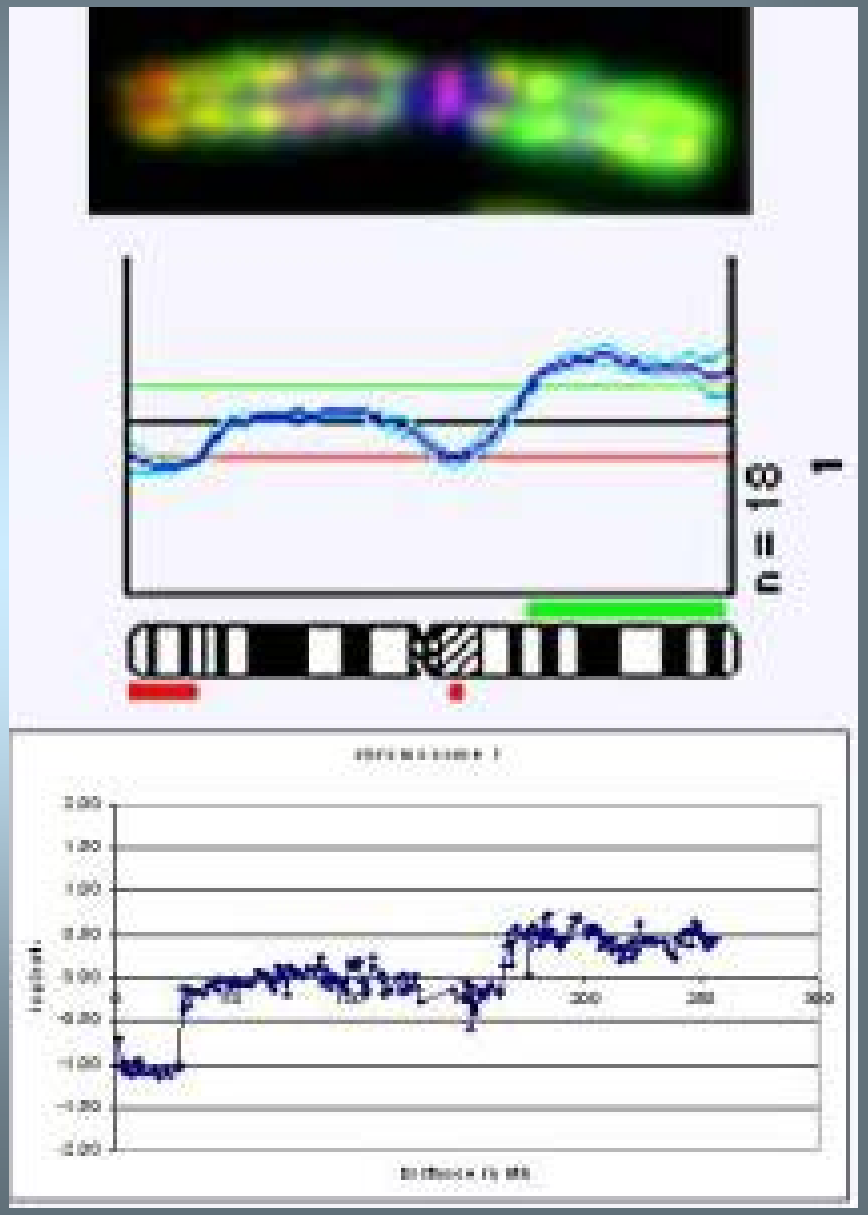
Bártová et al., Differentiation (2008)

Comparative genome hybridization

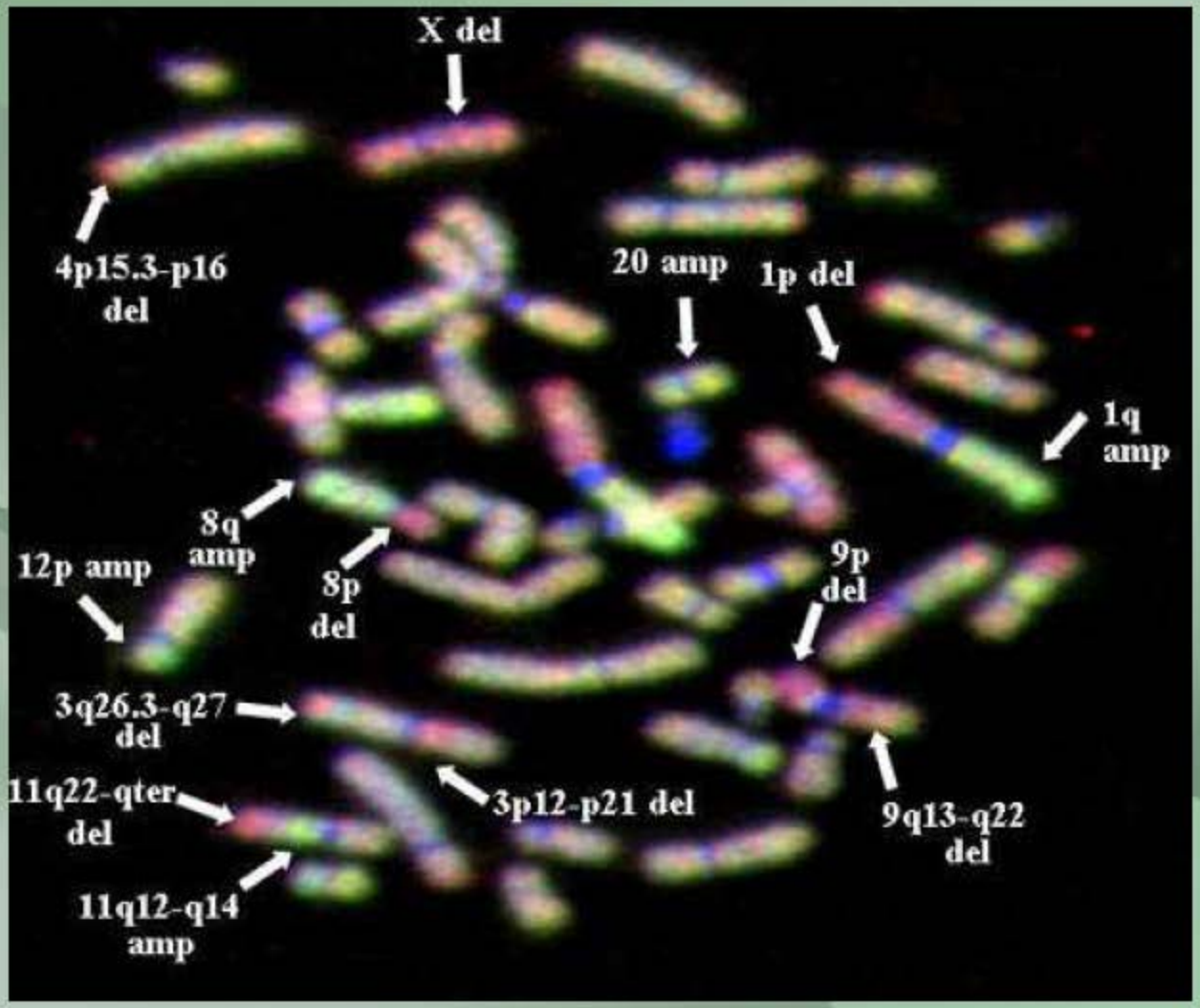


CGH on metaphase spreads





Comparative genome hybridization



Advanced microscopic techniques

Electron Microscopes are: **scientific instruments that use a beam of highly energetic electrons to examine objects on a very fine scale. This examination can yield the following information:**

Topography

The surface features of an object or "how it looks", its texture; direct relation between these features and materials properties (hardness, reflectivity...etc.)

Morphology

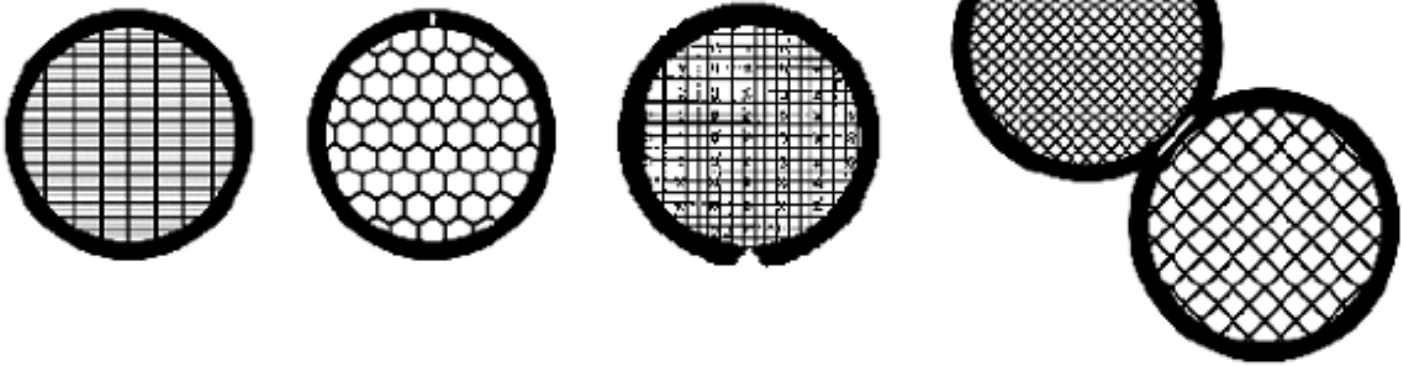
The shape and size of the particles making up the object; direct relation between these structures and materials properties (ductility, strength, reactivity...etc.)

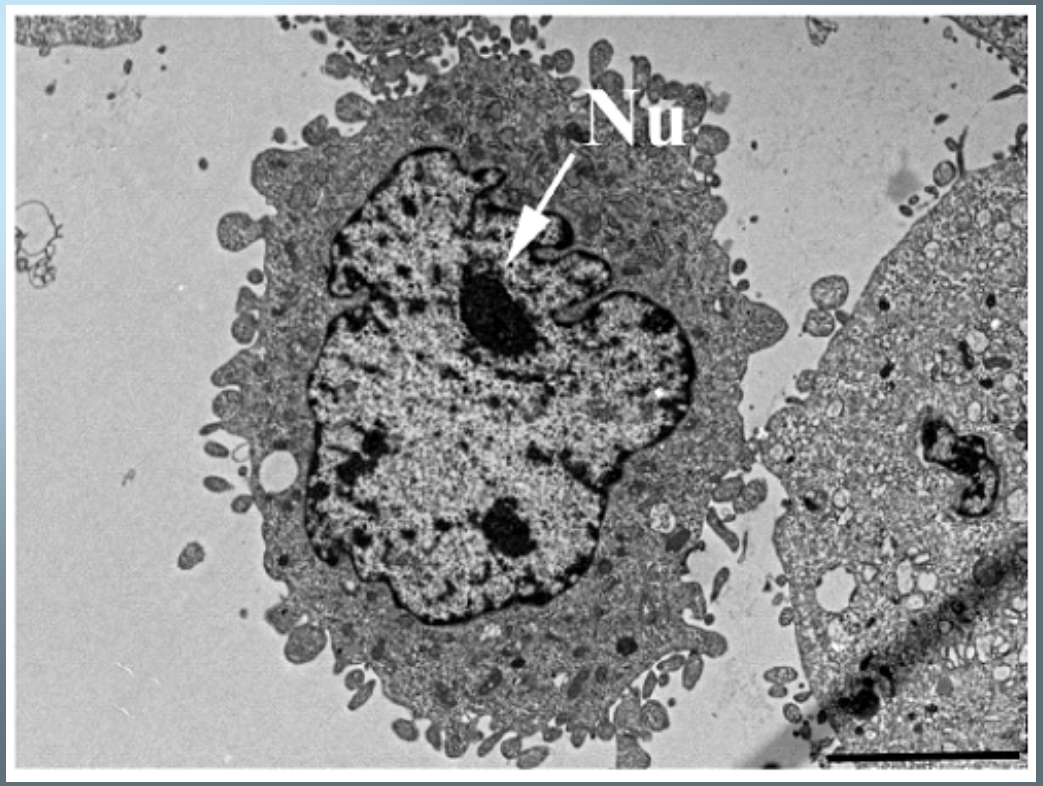
Composition

The elements and compounds that the object is composed of and the relative amounts of them; direct relationship between composition and materials properties (melting point, reactivity, hardness...etc.)

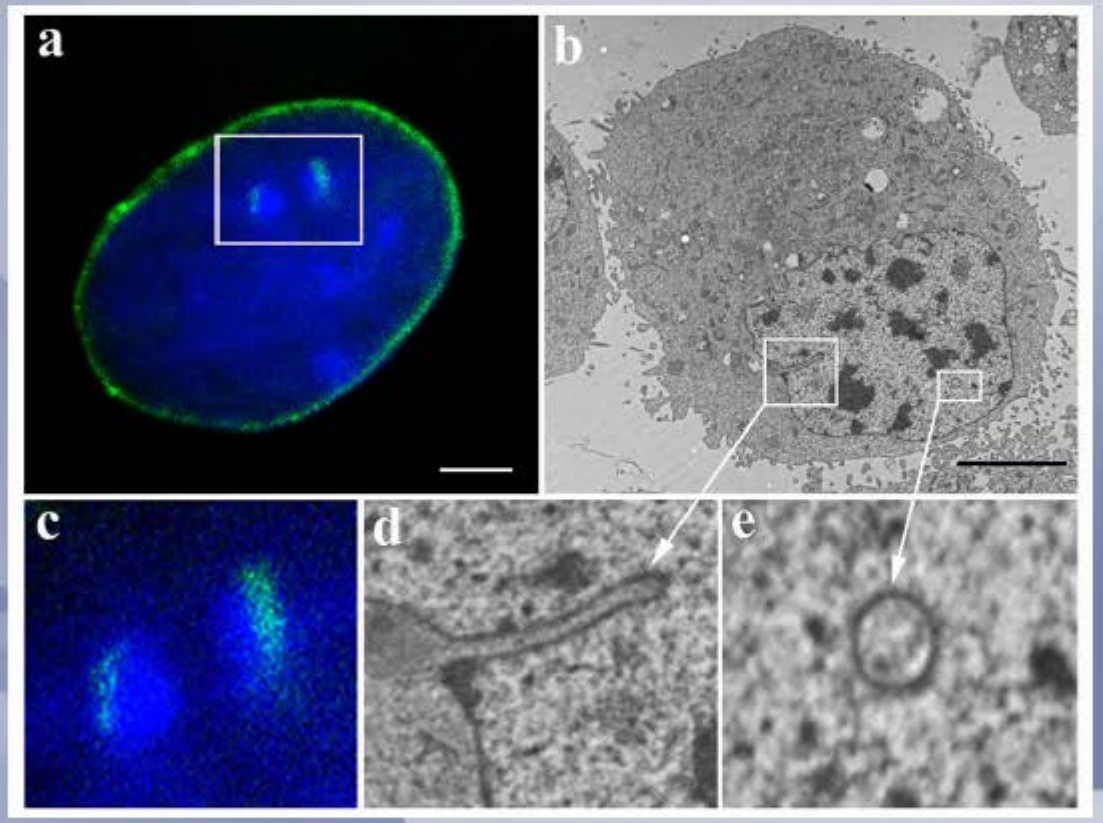
Crystallographic Information

How the atoms are arranged in the object; direct relation between these arrangements and materials properties (conductivity, electrical properties, strength, etc.)



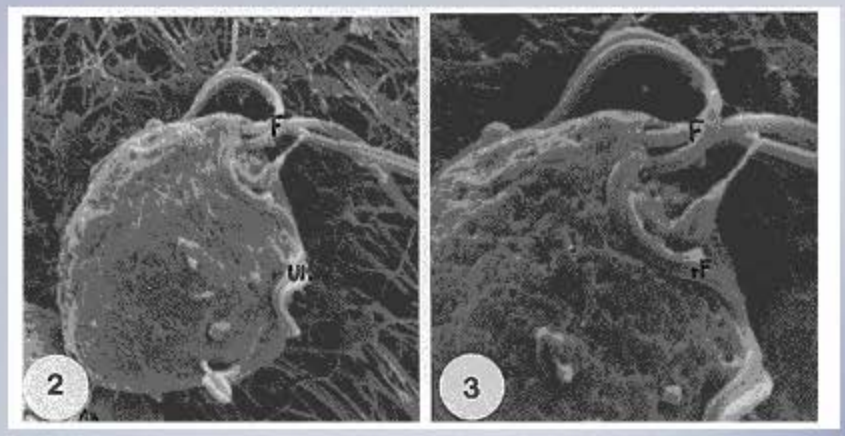

Nu

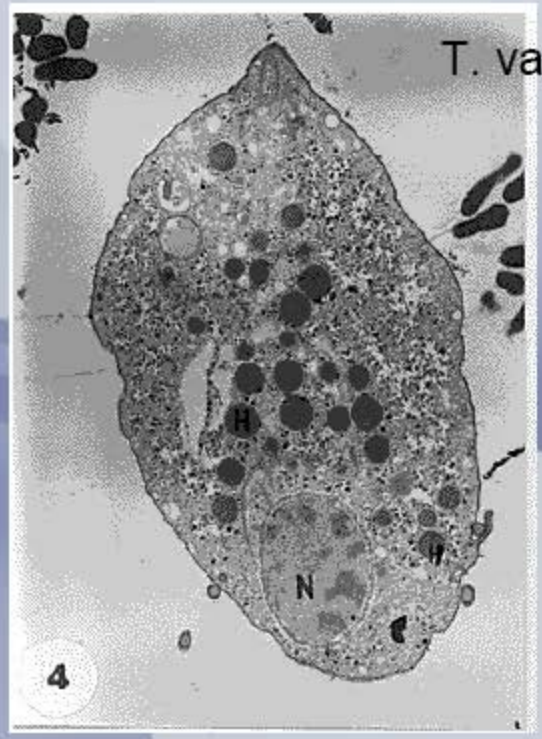
Transmission Electron Microscopes (TEM)



Galiová et al. EJCB (2008)

Scanning Electron Microscopes





X-RAY MICROSCOPY

Soft X-ray microscopes can be used to study hydrated cells up to 10 μm thick and produce images of 30 nm resolutions. X-ray microscopy, that has the more pronounced properties of laser scanning confocal microscopy (LSCM), has been a long-standing goal for experimental science (Seres et al., 2005). Since the cells are imaged in the X-ray transmissive "water window", where organic material absorbs approximately an order of magnitude more strongly than water, chemical contrast enhancement agents are not required to view the distribution of cellular structures (Meyer-Ilse et al., 2001). In such experiments, cells must be rapidly frozen to be studied on a cryostage, showing information which is closely similar to 4D-living cell observation by LSCM.

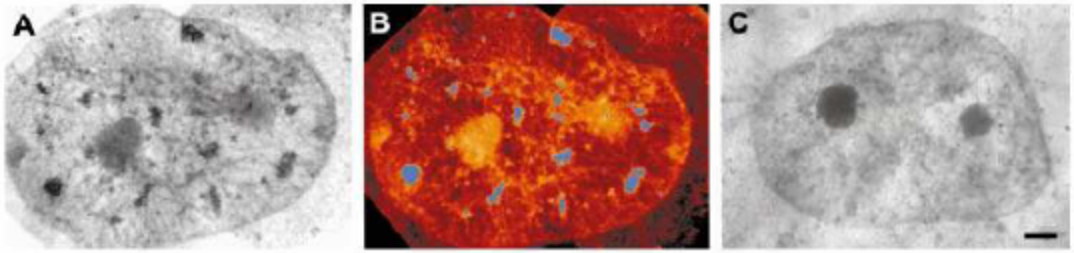
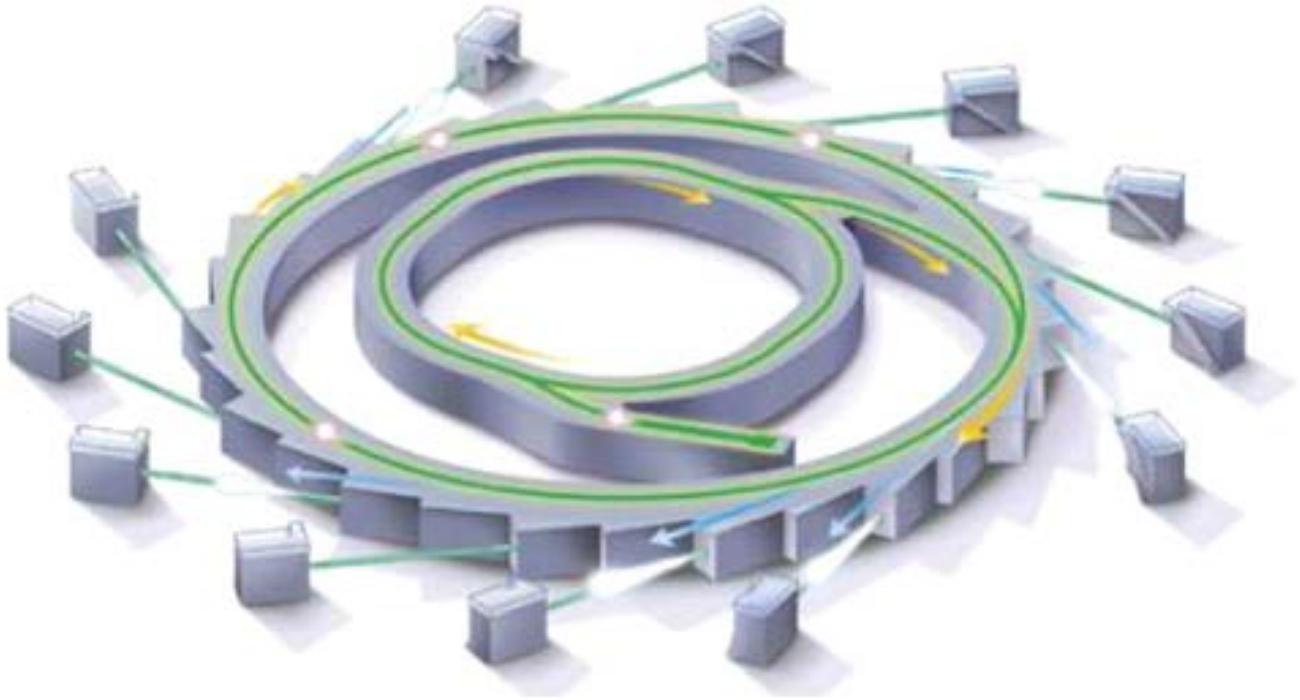


Fig. 4. Nuclei of human mammary epithelial cells (T4) labelled for RNA splicing factor (SRm300). (A) X-ray micrograph of a single nucleus after silver enhancement. This image is a montage compiled from two individual X-ray microscope images. (B) Same nucleus after colour coding to emphasize the label. (C) Control; single nucleus that was exposed to secondary antibodies and silver enhancement but not primary antibodies. This image is a montage compiled from two individual X-ray microscope images. Magnification = 2400×, 0.034 NA with 20 nm pixel size at 517 eV (λ = 2.4 nm).

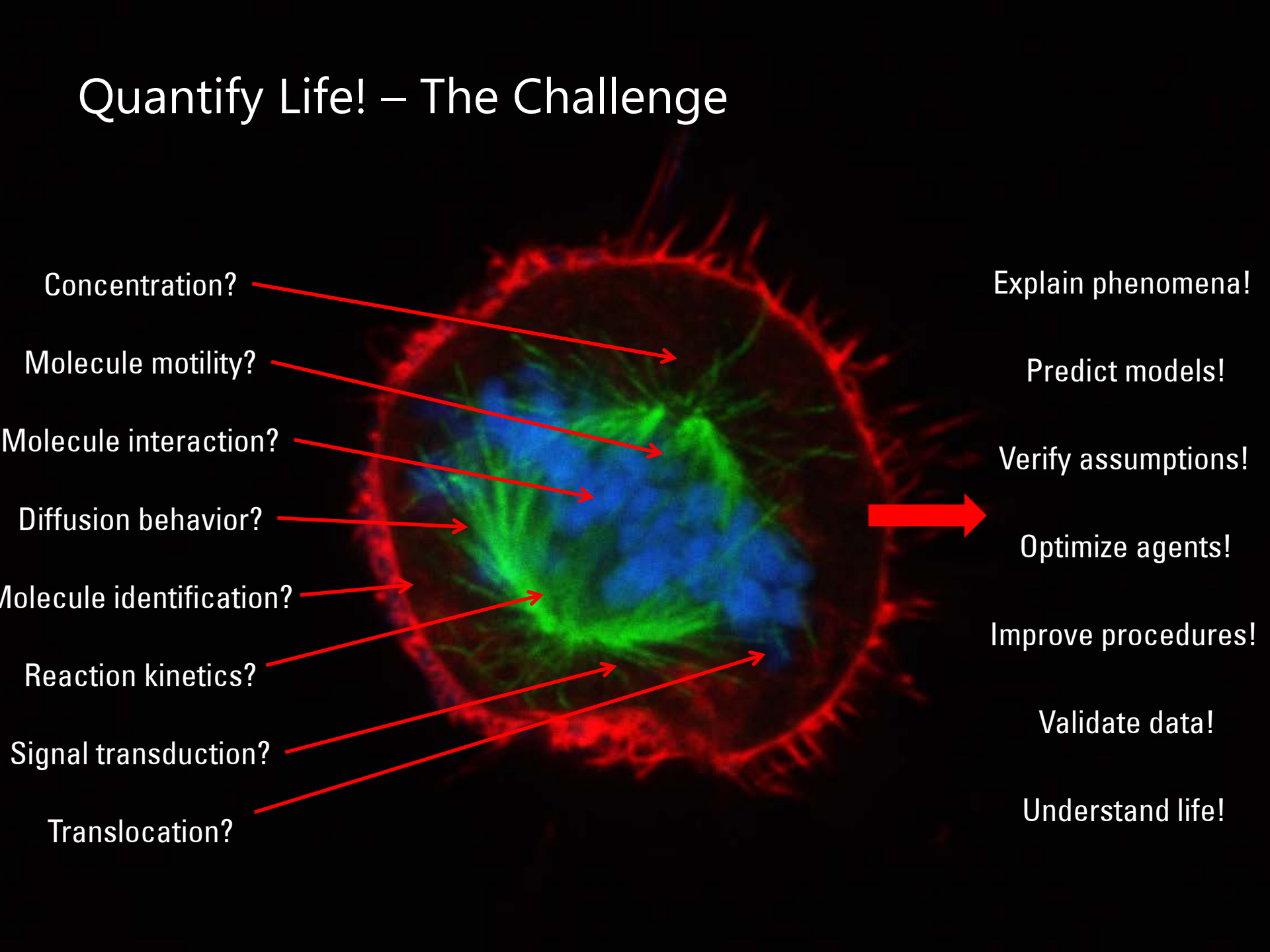
(Meyer-Ilse et al., 2000)

WHAT IS A SYNCHROTRON

A synchrotron is a device that accelerates electrons to almost the speed of light. As the electrons are deflected through magnetic fields they create extremely bright light. The light is channelled down beamlines to experimental workstations where it is used for research.



Quantify Life! – The Challenge



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