

# CG920 Genomics

## Lesson 12

### Systems Biology Tools

### Model organisms, PCR and PCR Primer Design

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INVESTICE DO ROZVOJE VZDĚLÁVÁNÍ

Tato prezentace je spolufinancována  
Evropským sociálním fondem  
a státním rozpočtem České republiky

# Literature

## ■ Literature sources for Chapter 12:

- Wilt, F.H., and Hake, S. (2004). [Principles of Developmental Biology](#). (New York ; London: W. W. Norton)
- Roscoe B. Jackson Memorial Laboratory., and Green, E.L. (1966). [Biology of the laboratory mouse](#). (New York: Blakiston Division) <http://www.informatics.jax.org/greenbook/index.shtml>
- Eden, E., Navon, R., Steinfeld, I., Lipson, D., and Yakhini, Z. (2009). GOrilla: a tool for discovery and visualization of enriched GO terms in ranked gene lists. *BMC Bioinformatics* 10, 48.
- The Arabidopsis Genome Initiative. (2000). Analysis of the genome sequence of the flowering plant *Arabidopsis thaliana*. *Nature* 408, 796-815.
- Gregory, S.G., Sekhon, M., Schein, J., Zhao, S., Osoegawa, K., Scott, C.E., Evans, R.S., BurrIDGE, P.W., Cox, T.V., Fox, C.A., Hutton, R.D., Mullenger, I.R., Phillips, K.J., Smith, J., Stalker, J., Threadgold, G.J., Birney, E., Wylie, K., Chinwalla, A., Wallis, J., Hillier, L., Carter, J., Gaige, T., Jaeger, S., Kremitzki, C., Layman, D., Maas, J., McGrane, R., Mead, K., Walker, R., Jones, S., Smith, M., Asano, J., Bosdet, I., Chan, S., Chittaranjan, S., Chiu, R., Fjell, C., Fuhrmann, D., Girn, N., Gray, C., Guin, R., Hsiao, L., Krzywinski, M., Kutsche, R., Lee, S.S., Mathewson, C., McLeavy, C., Messervier, S., Ness, S., Pandoh, P., Prabhu, A.L., Saeedi, P., Smailus, D., Spence, L., Stott, J., Taylor, S., Terpstra, W., Tsai, M., Vardy, J., Wye, N., Yang, G., Shatsman, S., Ayodeji, B., Geer, K., Tsegaye, G., Shvartsbeyn, A., Gebregeorgis, E., Krol, M., Russell, D., Overton, L., Malek, J.A., Holmes, M., Heaney, M., Shetty, J., Feldblyum, T., Nierman, W.C., Catanese, J.J., Hubbard, T., Waterston, R.H., Rogers, J., de Jong, P.J., Fraser, C.M., Marra, M., McPherson, J.D., and Bentley, D.R. (2002). A physical map of the mouse genome. *Nature* 418, 743-750.
- Benitez, M. and Hejatko, J. Dynamics of cell-fate determination and patterning in the vascular bundles of *Arabidopsis thaliana* (submitted)

# Outline

- Tools of **systems biology**
  - **Gene ontology** analysis
  - **Molecular Regulatory Networks Modeling**
- Model organisms
  - *Mus musculus*
  - *Arabidopsis thaliana*
- Selected **methods of molecular biology**
  - Preparation of transgenic organisms
  - PCR
  - Design and preparation of primers (Dr. Hana Konečná)

# Outline

- Tools of **systems biology**
    - **Gene ontology** analysis
- 

# Results of –omics Studies vs Biologically Relevant Conclusions

- Results of **–omics studies** are represented by **huge amount of data**, e.g. differential gene expression. But how to get any **biologically relevant conclusions**?

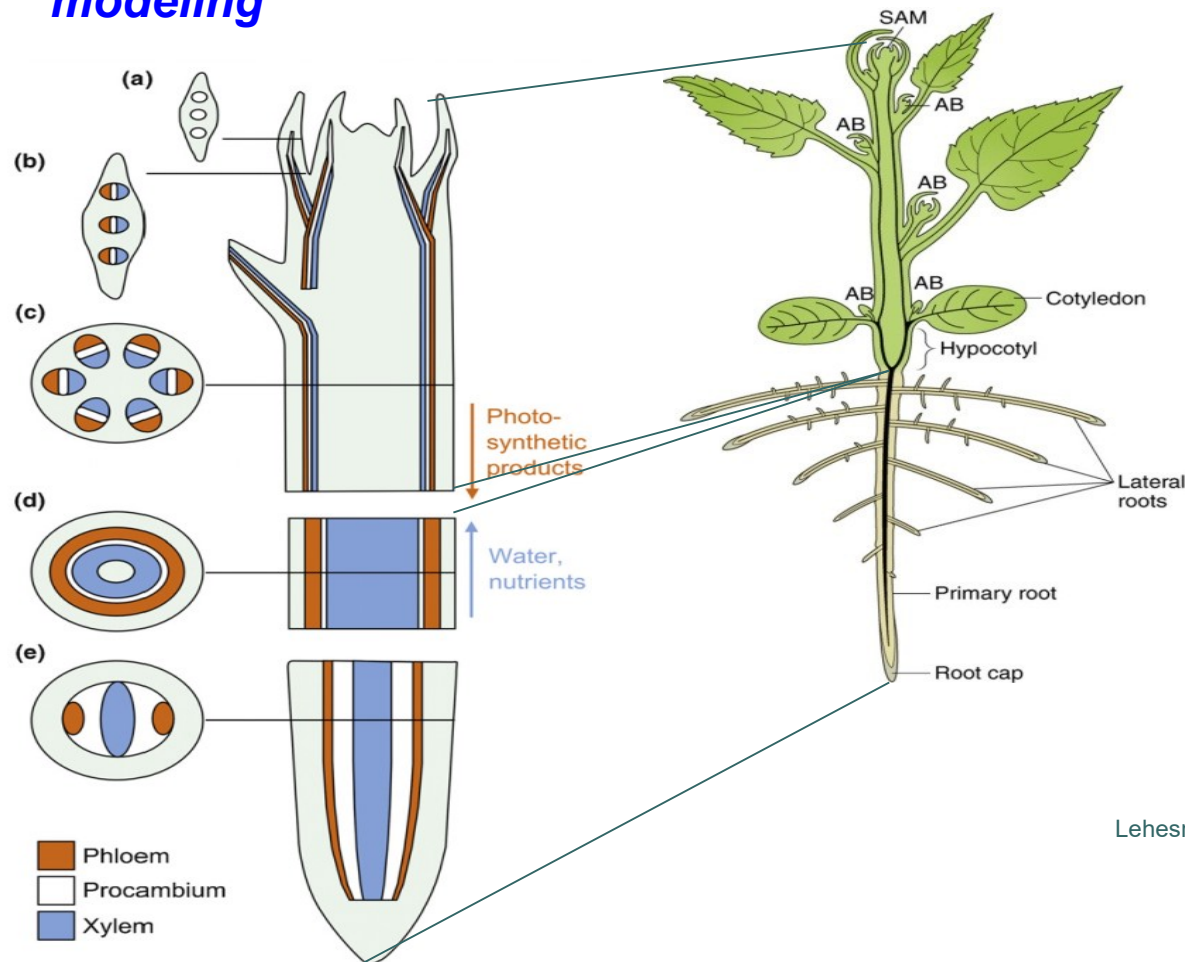
Didi et al., unpublished

gene	locus	sample_1	sample_2	status	value_1	value_2	log2(fold_change)	test_stat	p_value	q_value	significant
AT1G07795	1:2414285-2414967	WT	MT	OK	0	1,1804	1.79769e+308	1.79769e+308	6.88885e-05	0,00039180	1 yes
HRS1	1:4556891-4558708	WT	MT	OK	0	0,696583	1.79769e+308	1.79769e+308	6.61994e-06	4.67708e-05	yes
ATMLO14	1:9227472-9232296	WT	MT	OK	0	0,514609	1.79769e+308	1.79769e+308	9.74219e-05	0,00053505	5 yes
NRT1.6	1:9400663-9403789	WT	MT	OK	0	0,877865	1.79769e+308	1.79769e+308	3.2692e-08	3.50131e-07	yes
AT1G27570	1:9575425-9582376	WT	MT	OK	0	2,0829	1.79769e+308	1.79769e+308	9.76039e-06	6.647e-05	yes
AT1G60095	1:22159735-22162419	WT	MT	OK	0	0,688588	1.79769e+308	1.79769e+308	9.95901e-08	9.84992e-07	yes
AT1G03020	1:698206-698515	WT	MT	OK	0	1,78859	1.79769e+308	1.79769e+308	0,00913915	0,0277958	yes
AT1G13609	1:4662720-4663471	WT	MT	OK	0	3,55814	1.79769e+308	1.79769e+308	0,00021683	0,00108079	yes
AT1G21550	1:7553100-7553876	WT	MT	OK	0	0,562868	1.79769e+308	1.79769e+308	0,00115582	0,00471497	yes
AT1G22120	1:7806308-7809632	WT	MT	OK	0	0,617354	1.79769e+308	1.79769e+308	2.48392e-06	1.91089e-05	yes
AT1G31370	1:11238297-11239363	WT	MT	OK	0	1,46254	1.79769e+308	1.79769e+308	4.83523e-05	0,00028514	3 yes
APUM10	1:13253397-13255570	WT	MT	OK	0	0,581031	1.79769e+308	1.79769e+308	7.87855e-06	5.46603e-05	yes
AT1G48700	1:18010728-18012871	WT	MT	OK	0	0,556525	1.79769e+308	1.79769e+308	6.53917e-05	0,00037473	6 yes
AT1G59077	1:21746209-21833195	WT	MT	OK	0	138,886	1.79769e+308	1.79769e+308	0,00122789	0,00496816	yes
AT1G60050	1:22121549-22123702	WT	MT	OK	0	0,370087	1.79769e+308	1.79769e+308	0,00117953	0,0048001	yes
AT4G15242	4:8705786-8706997	WT	MT	OK	0,00930712	17,9056	10,9098	-4,40523	1.05673e-05	7.13983e-05	yes
AT5G33251	5:12499071-12500433	WT	MT	OK	0,0498375	52,2837	10,0349	-9,8119	0	0	yes
AT4G12520	4:7421055-7421738	WT	MT	OK	0,0195111	15,8516	9,66612	-3,90043	9.60217e-05	0,000528904	yes
AT1G60020	1:22100651-22105276	WT	MT	OK	0,0118377	7,18823	9,24611	-7,50382	6.19504e-14	1.4988e-12	yes
AT5G15360	5:4987235-4989182	WT	MT	OK	0,0988273	56,4834	9,1587	-10,4392	0	0	yes

Example of an output of transcriptional profiling study using Illumina sequencing performed in our lab. Shown is just a tiny fragment of the complete list, comprising about 7K genes revealing differential expression in the studied mutant

# Molecular Regulatory Networks Modeling

- **Vascular tissue** as a developmental model for **GO analysis** and **MRN modeling**

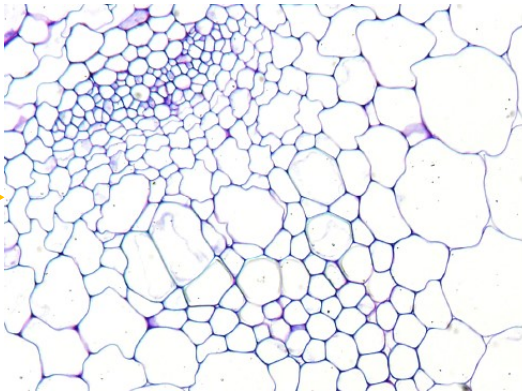


Lehesranta et al., *Trends in Plant Sci* (2010)

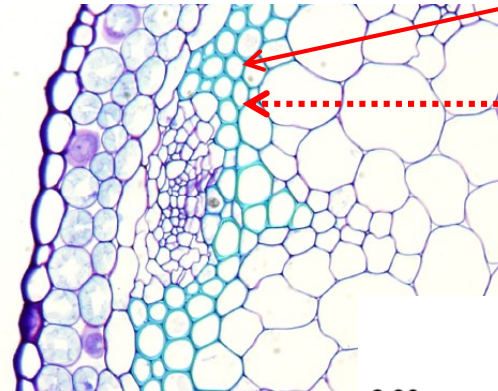
# Hormonal Control Over Vascular Tissue Development

- Plant **Hormones Regulate Lignin Deposition** in Plant Cell Walls and **Xylem Water Conductivity**

**WT**

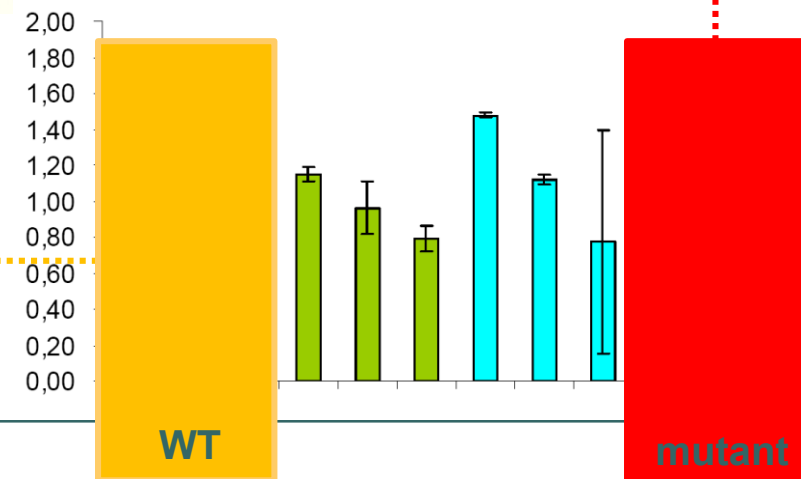
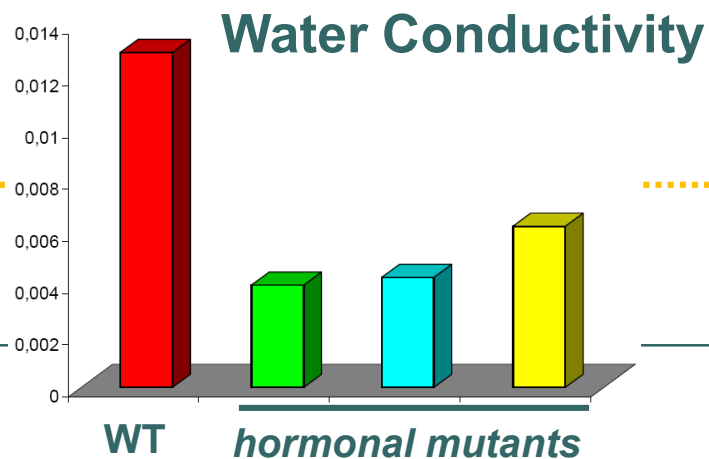


*hormonal mutant*



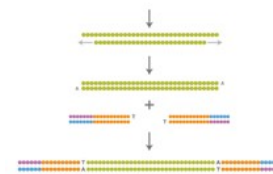
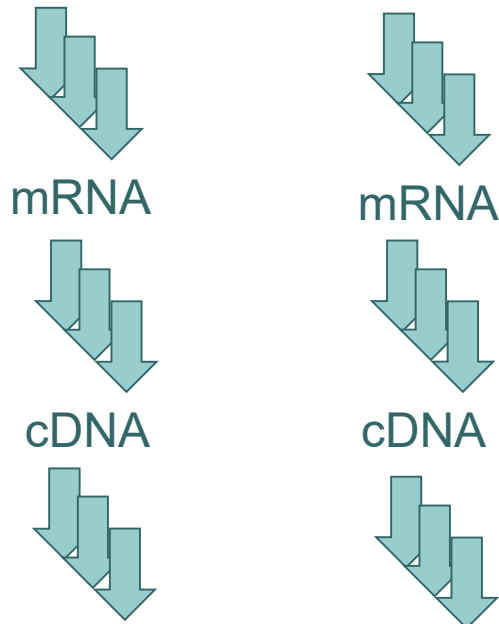
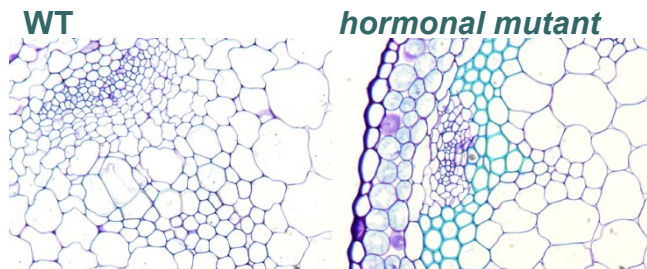
**lignified cell walls**

Acid-insoluble lignins



# Hormonal Control Over Vascular Tissue Development

- *Transcriptional profiling* via *RNA sequencing*



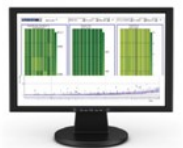
Library Preparation  
~2 h [15 min hands-on (Nextera)]  
< 6 h [< 3 h hands-on (TruSeq)]



Cluster Generation  
~5 h (<10 min hands-on)



Sequencing by Synthesis  
~1.5 to 11 days



CASAVA  
2 days (30 min hands-on)

Sequencing by Illumina and  
**number of transcripts** determination



# Results of –omics Studies vs Biologically Relevant Conclusions

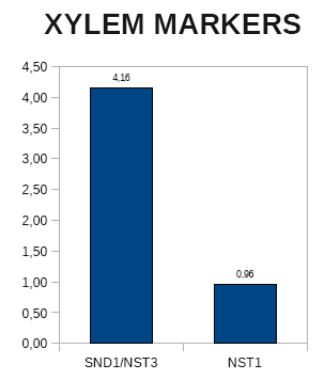
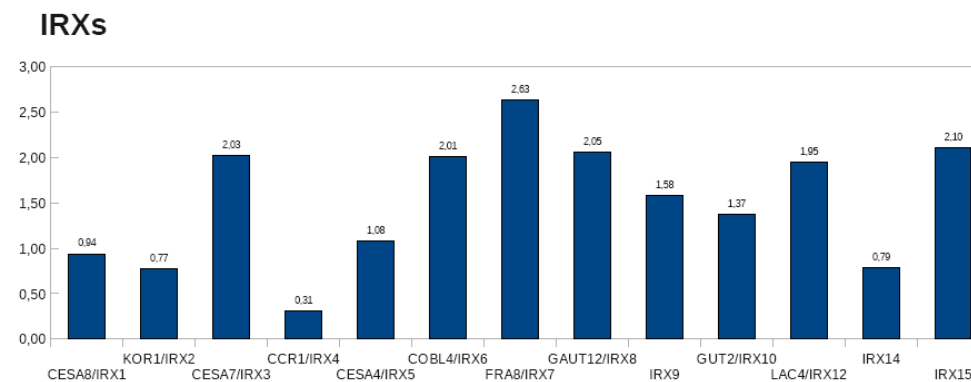
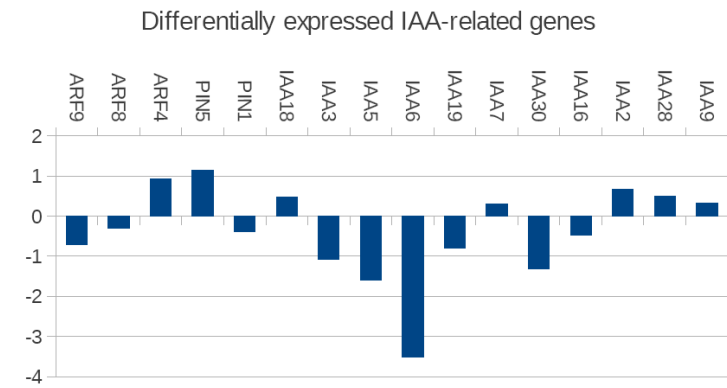
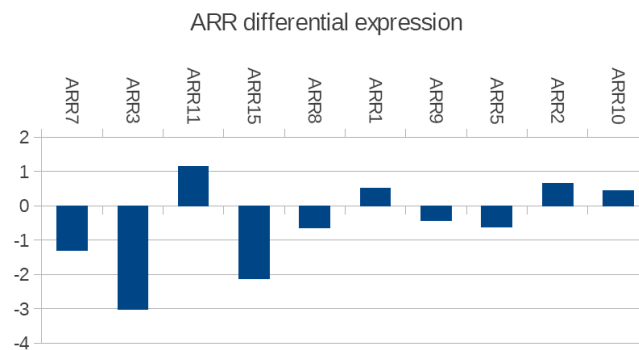
- Transcriptional profiling yielded more than **7K differentially regulated genes...**

Dii et al., unpublished

gene	locus	sample_1	sample_2	status	value_1	value_2	log2(fold_change)	test_stat	p_value	q_value	significant
AT1G07795	1:2414285-2414967	WT	MT	OK	0	1,1804	1.79769e+308	1.79769e+308	6.88885e-05	0,00039180	1 yes
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AT5G15360	5:4987235-4989182	WT	MT	OK	0,0988273	56,4834	9,1587	-10,4392	0	0	yes

# Gene Ontology Analysis

- One of the possible approaches is to study **gene ontology**, i.e. previously demonstrated **association** of genes to **biological processes**



# Gene Ontology Analysis

- Several tools allow **statistical evaluation** of **enrichment** for genes **associated with specific processes**

One of such recent and very useful tools is Gorilla software, freely available at <http://cbl-gorilla.cs.technion.ac.il/>.

Eden et al., *BMC Bioinformatics* (2009)

**GORILLA**  
Gene Ontology enRICHment anaLysis and visualiZation tool

Gorilla is a tool for identifying and visualizing enriched GO terms in ranked lists of genes. It can be run in one of two modes:

1. Searching for enriched GO terms that appear densely at the top of a ranked list of genes or
2. Searching for enriched GO terms in a target list of genes compared to a background list of genes.

For further details see [References](#).

[Running example](#) [Usage instructions](#) [GORilla News\(Updated December 3rd 2012\)](#) [References](#)

**Step 1: Choose organism**  
Arabidopsis thaliana

**Step 2: Choose running mode**  
 Single ranked list of genes  Two unranked lists of genes (target and background lists)

**Step 3: Paste a ranked list of gene/protein names**  
Names should be separated by an <ENTER>. The preferred format is gene symbol. Other supported formats are: gene and protein RefSeq, Uniprot, Unigene and Ensembl. Use [WebGestalt](#) for conversion from other identifier formats.

Or upload a file: D:\Results\2012\Mariane [Procházet]

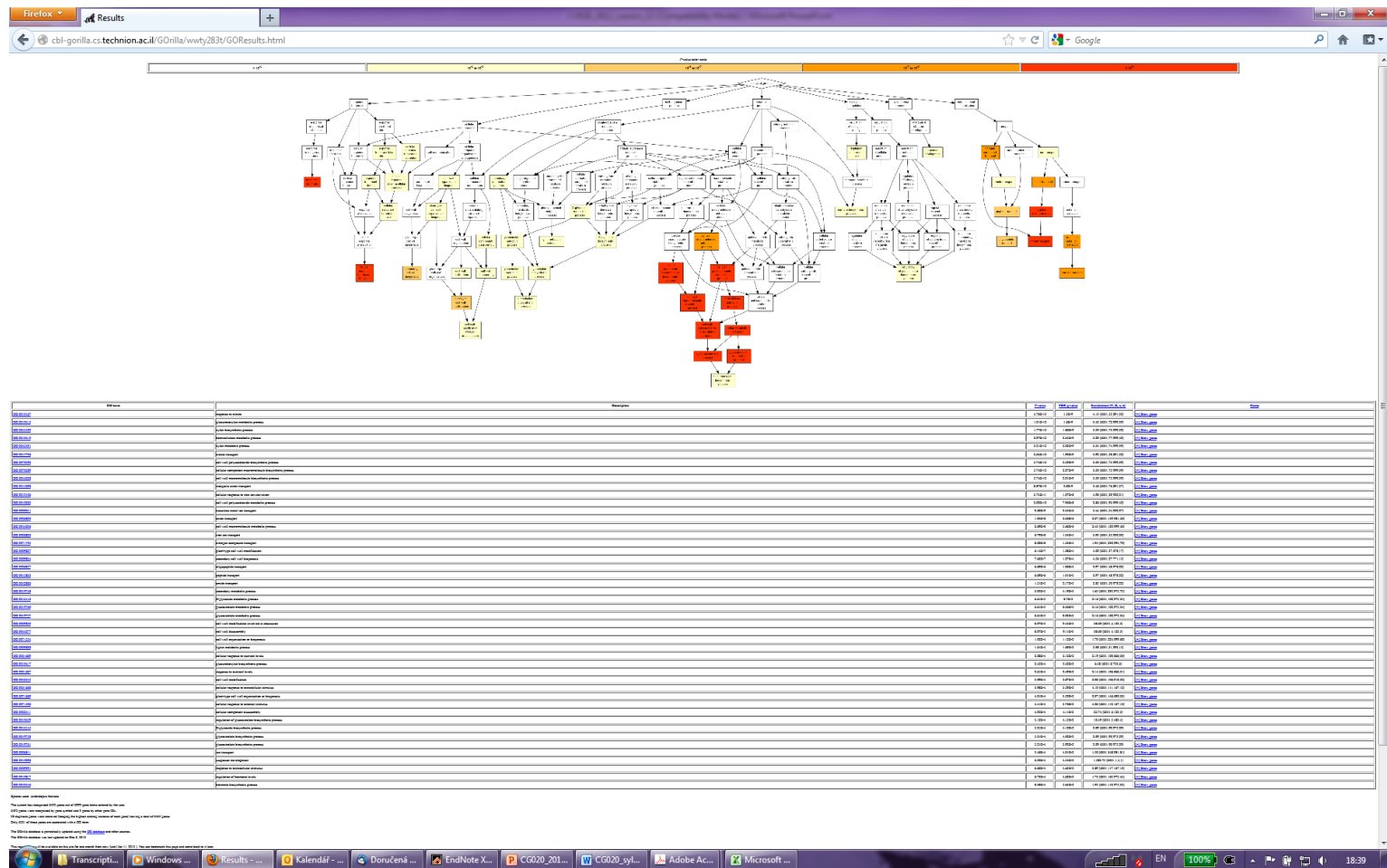
**Step 4: Choose an ontology**  
 Process  Function  Component  All

**Search Enriched GO terms**

Reset form

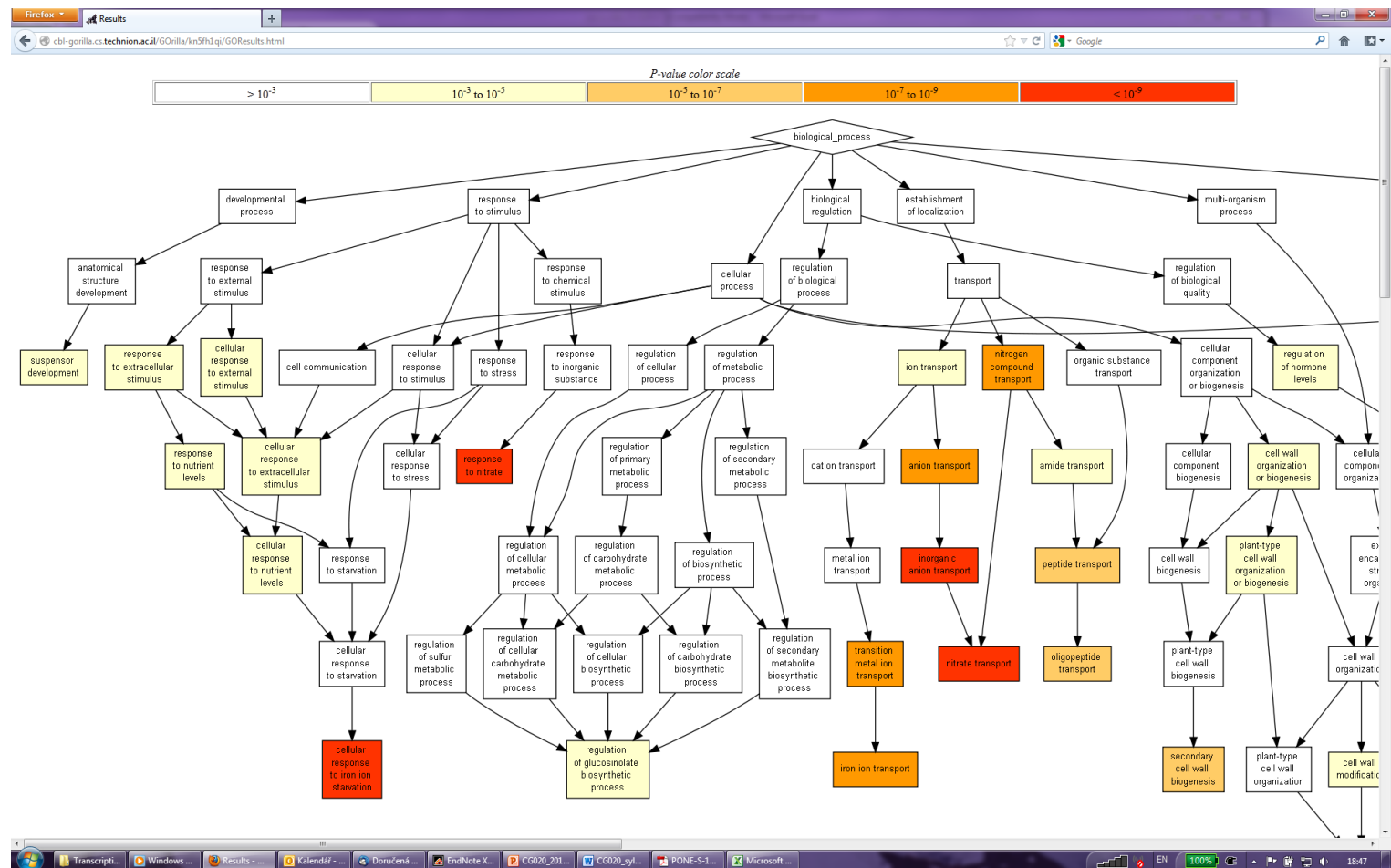
# Gene Ontology Analysis

- Several tools allow **statistical evaluation** of **enrichment** for genes **associated with specific processes**



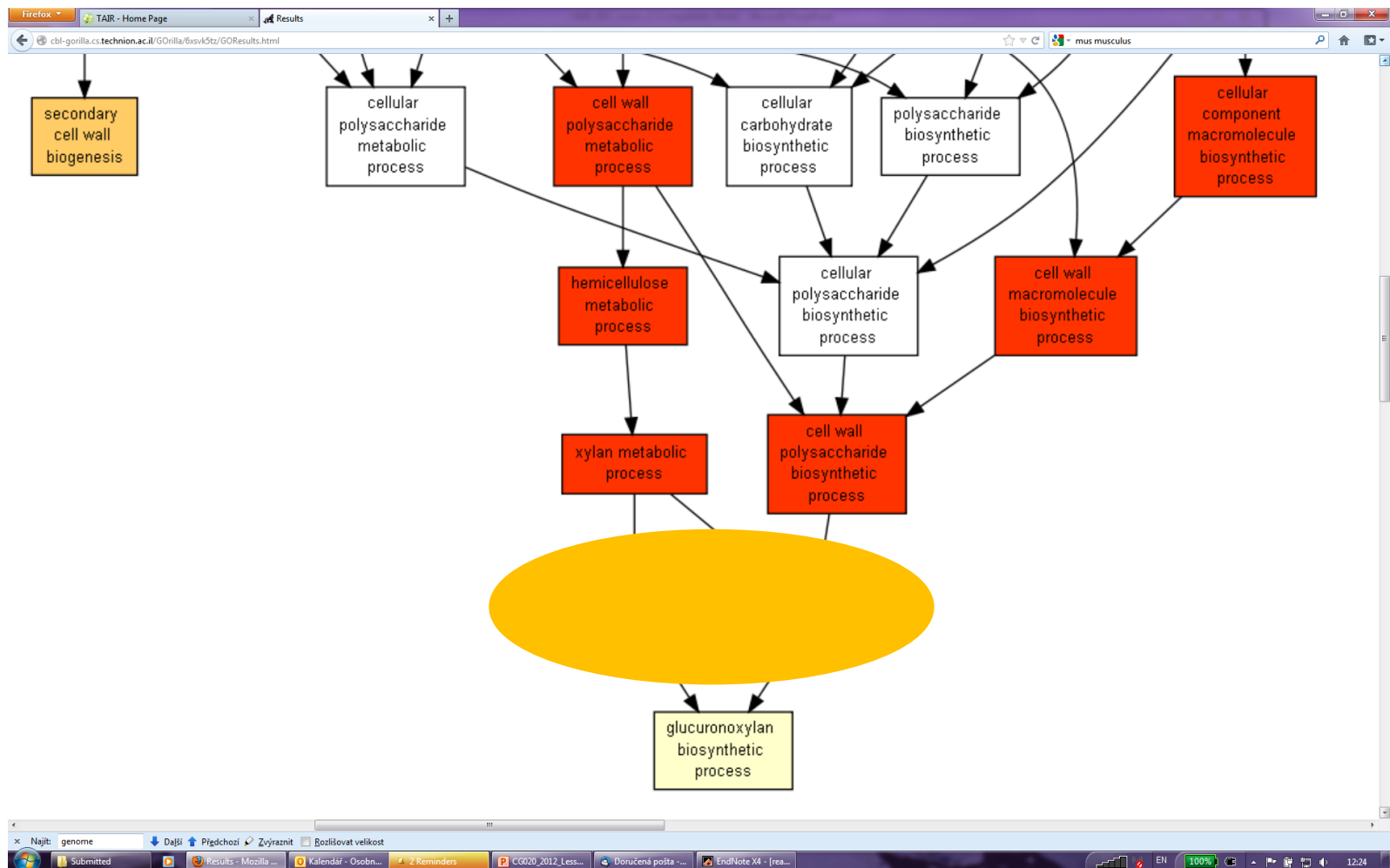
# Gene Ontology Analysis

- Several tools allow **statistical evaluation** of **enrichment** for genes **associated with specific processes**



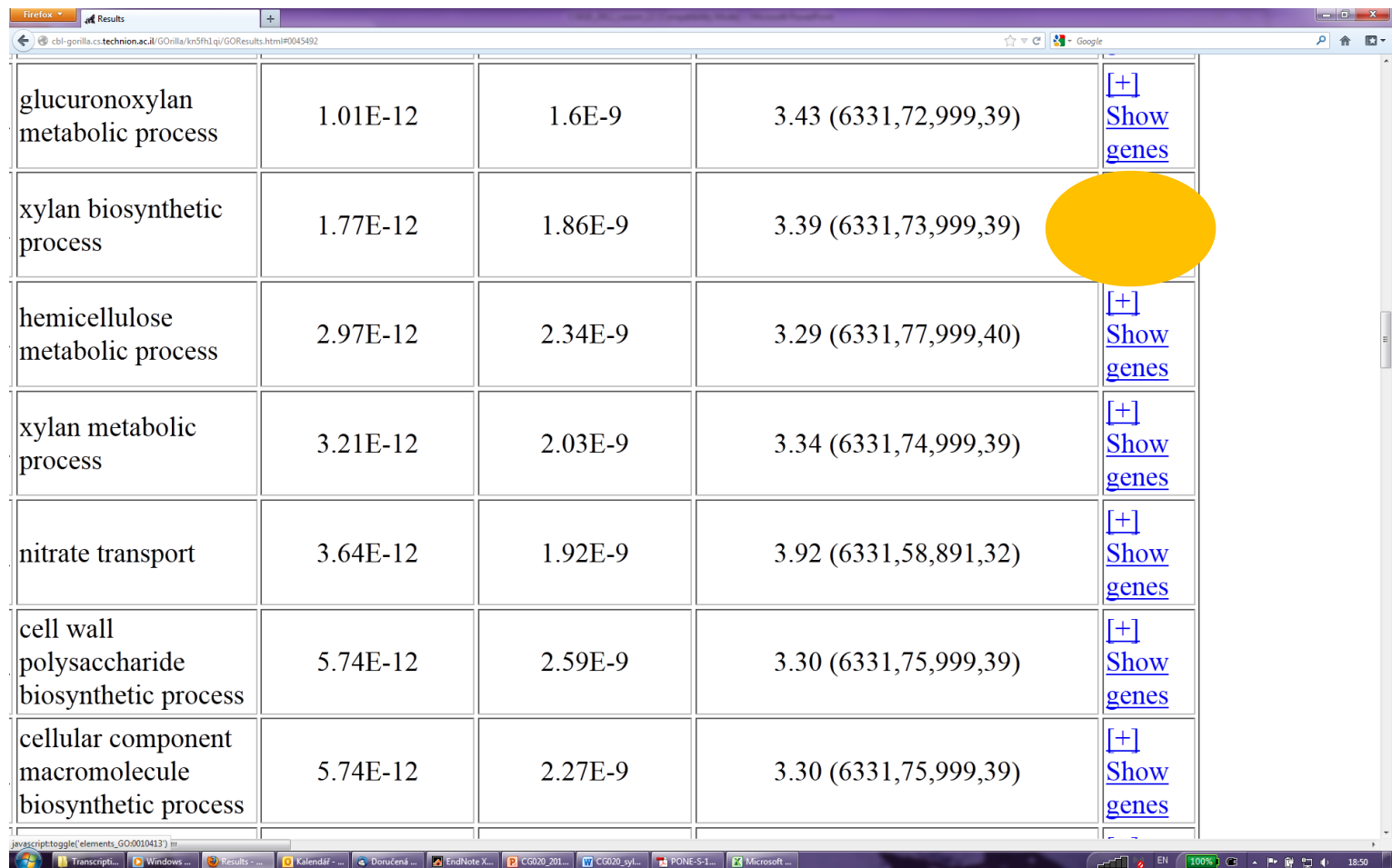
# Gene Ontology Analysis

- Several tools allow **statistical evaluation** of **enrichment** for genes **associated with specific processes**



# Gene Ontology Analysis

- Several tools allow **statistical evaluation** of **enrichment** for genes **associated with specific processes**



glucuronoxylan metabolic process	1.01E-12	1.6E-9	3.43 (6331,72,999,39)	[+] <a href="#">Show genes</a>
xylan biosynthetic process	1.77E-12	1.86E-9	3.39 (6331,73,999,39)	[+] <a href="#">Show genes</a>
hemicellulose metabolic process	2.97E-12	2.34E-9	3.29 (6331,77,999,40)	[+] <a href="#">Show genes</a>
xylan metabolic process	3.21E-12	2.03E-9	3.34 (6331,74,999,39)	[+] <a href="#">Show genes</a>
nitrate transport	3.64E-12	1.92E-9	3.92 (6331,58,891,32)	[+] <a href="#">Show genes</a>
cell wall polysaccharide biosynthetic process	5.74E-12	2.59E-9	3.30 (6331,75,999,39)	[+] <a href="#">Show genes</a>
cellular component macromolecule biosynthetic process	5.74E-12	2.27E-9	3.30 (6331,75,999,39)	[+] <a href="#">Show genes</a>

# Gene Ontology Analysis

- Several tools allow **statistical evaluation** of **enrichment** for genes **associated with specific processes**

Description	P-value	FDR q-value	Enrichment (N, B, n, b)	Genes
response to nitrate	4.76E-13	1.5E-9	4.13 (6331,55,891,32)	[+] Show genes
glucuronoxytan metabolic process	1.01E-12	1.6E-9	3.43 (6331,72,999,39)	[+] Show genes
xylan biosynthetic process	1.77E-12	1.86E-9	3.39 (6331,73,999,39)	[-] Hide genes GUT2 - putative glycosyltransferase PG5IP3 - plant glycogenin-like starch initiation protein 3 FRA8 - exostosin-like protein GAUT12 - alpha-1,4-galacturonyltransferase AT4G22460 - bifunctional inhibitor/lipid-transfer protein/seed storage 2s albumin-like protein AT5G42180 - peroxidase 64 AT3G10910 - ring-h2 finger protein at72 LAC17 - laccase 17 KNAT7 - homeobox protein knotted-1-like 7 NAC012 - nac domain-containing protein 12 IRX9 - nucleotide-diphospho-sugar transferases-like protein AT1G70500 - pectin lyase-like protein CESA4 - cellulose synthase a catalytic subunit 4 [udp-forming] AT1G08340 - rho gtpase activating protein with pak-box/p21-rho-binding domain CTL2 - chitinase-like protein 2 IRX6 - cobra-like protein 4 MYB63 - myb domain protein 63 PG5IP1 - plant glycogenin-like starch initiation protein 1 AT5G46340 - putative o-acetyltransferase AT3G21710 - hypothetical protein AT2G03200 - aspartyl protease-like protein AT1G09440 - protein kinase family protein AT5G40020 - pathogenesis-related thaumatin-like protein AT3G23090 - targeting protein for xk1p2-like protein AT5G67210 - hypothetical protein AT3G56230 - btb/poz domain-containing protein AT2G31930 - hypothetical protein JP630 - putative polygalacturonase non-catalytic subunit jp630 MAP70-5 - microtubule-associated proteins 70-5 AT3G50220 - hypothetical protein AGL44 - protein agamous-like 44 IRX12 - laccase-4 NAC073 - nac domain containing protein 73 IRX3 - cellulose synthase a catalytic subunit 7 [udp-forming] AT4G27435 - hypothetical protein MYB46 - transcription factor myb46 AT1G72220 - ring-h2 finger protein at154 FRD3 - mate efflux family protein AT1G33800 - hypothetical protein
hemicellulose metabolic process	2.07E-12	2.24E-9	3.20 (6331,77,999,40)	[+] Show genes

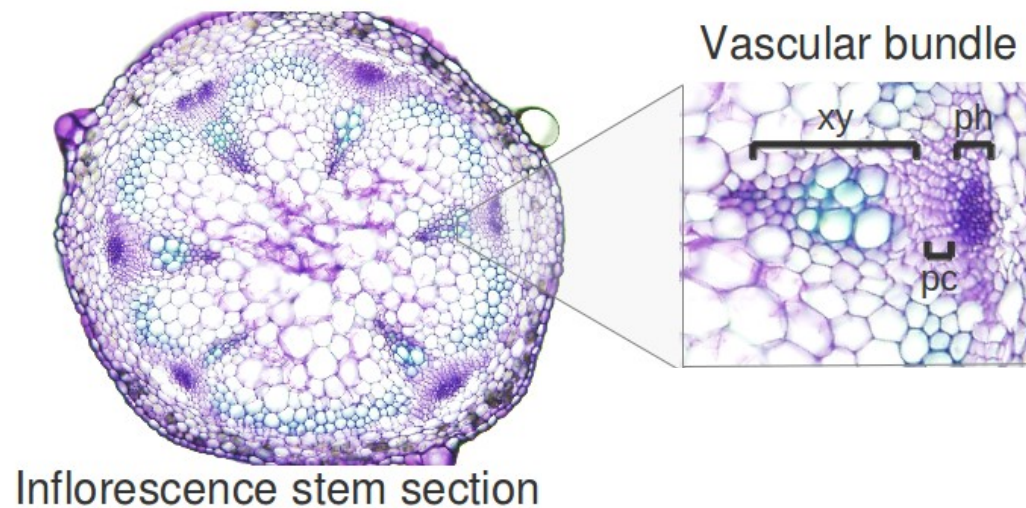


# Outline

- Tools of systems biology
  - Gene ontology analysis
  - **Molecular Regulatory Networks Modeling**

# Molecular Regulatory Networks Modeling

- **Vascular tissue** as a developmental model for **MRN modeling**



# Molecular Regulatory Networks Modeling

- **Literature search** for published data and creating small database

Interaction	Evidence	References
A-ARRs $\neg$ CK signaling	Double and higher order type-A ARR mutants show increased sensitivity to CK.	[27]
	Spatial patterns of A-type ARR gene expression and CK response are consistent with partially redundant function of these genes in CK signaling.	[27]
	A-type ARRs decreases B-type ARR6-LUC.	[13]
	Note: In certain contexts, however, some A-ARRs appear to have effects antagonistic to other A-ARRs.	[27]
AHP6 $\neg$ AHP	ahp6 partially recovers the mutant phenotype of the CK receptor WOL.	[9]
	Using an in vitro phosphotransfer system, it was shown that, unlike the AHPs, native AHP6 was unable to accept a phosphoryl group. Nevertheless, AHP6 is able to inhibit phosphotransfer from other AHPs to ARRs.	[9]

# Molecular Regulatory Networks Modeling

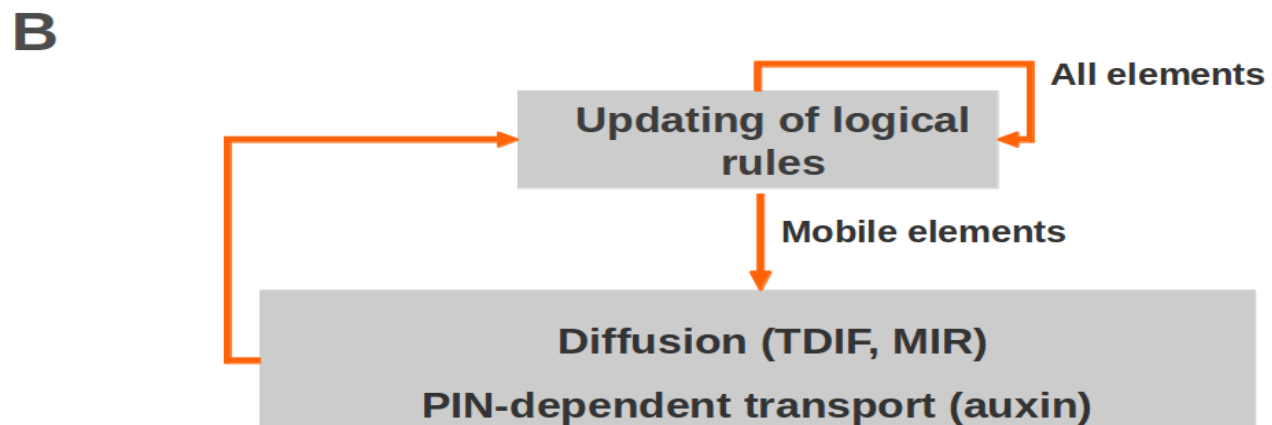
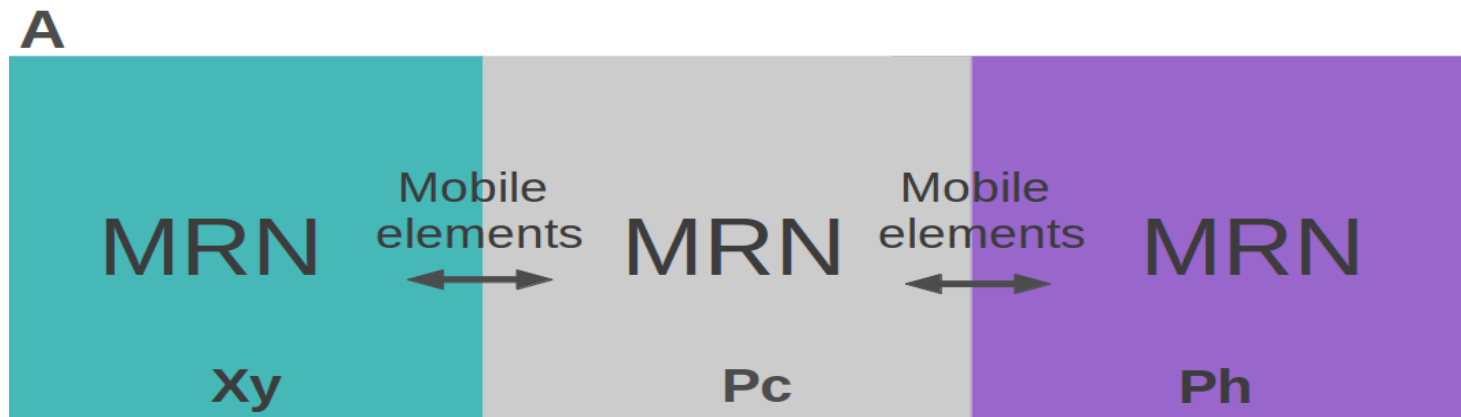
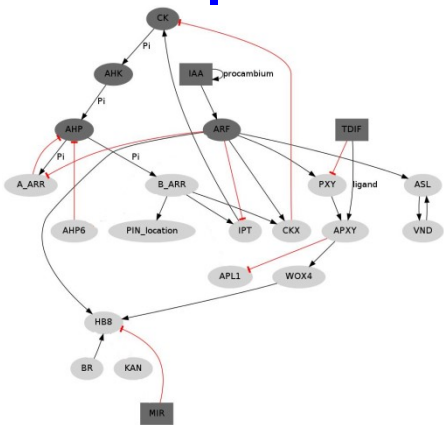
- Formulating *logical rules* defining the *model dynamics*

Network node	Dynamical rule
CK	2 If ipt=1 and ckx=0 1 If ipt=1 and ckx=1 0 else
CKX	1 If barr>0 or arf=2 0 else
AHKs	ahk=ck
AHPs	2 If ahk=2 and ahp6=0 and aarr=0 1 If ahk=2 and (ahp6+aarr<2) 1 If ahk=1 and ahp6<1 0 else
B-Type ARRs	1 If ahp>0 0 else
A-Type ARRs	1 If arf<2 and ahp>0 0 else

Benitez and Hejatko, *submitted*

# Molecular Regulatory Networks Modeling

- Specifying *mobile elements* and their model behaviour



According to experimental evidence for the system under study, the hormone IAA, the peptide TDIF, and the microRNA MIR165/6 are able to move among the cells. In the case of TDIF and MIR165/6, the mobility is defined as diffusion and is given by the following equation:

$$g(t+1)T[i] = H(g(t)[i] + D(g(t)[i+1] + g(t)[i-1] - N(g(t)[i])) - b) \quad (2),$$

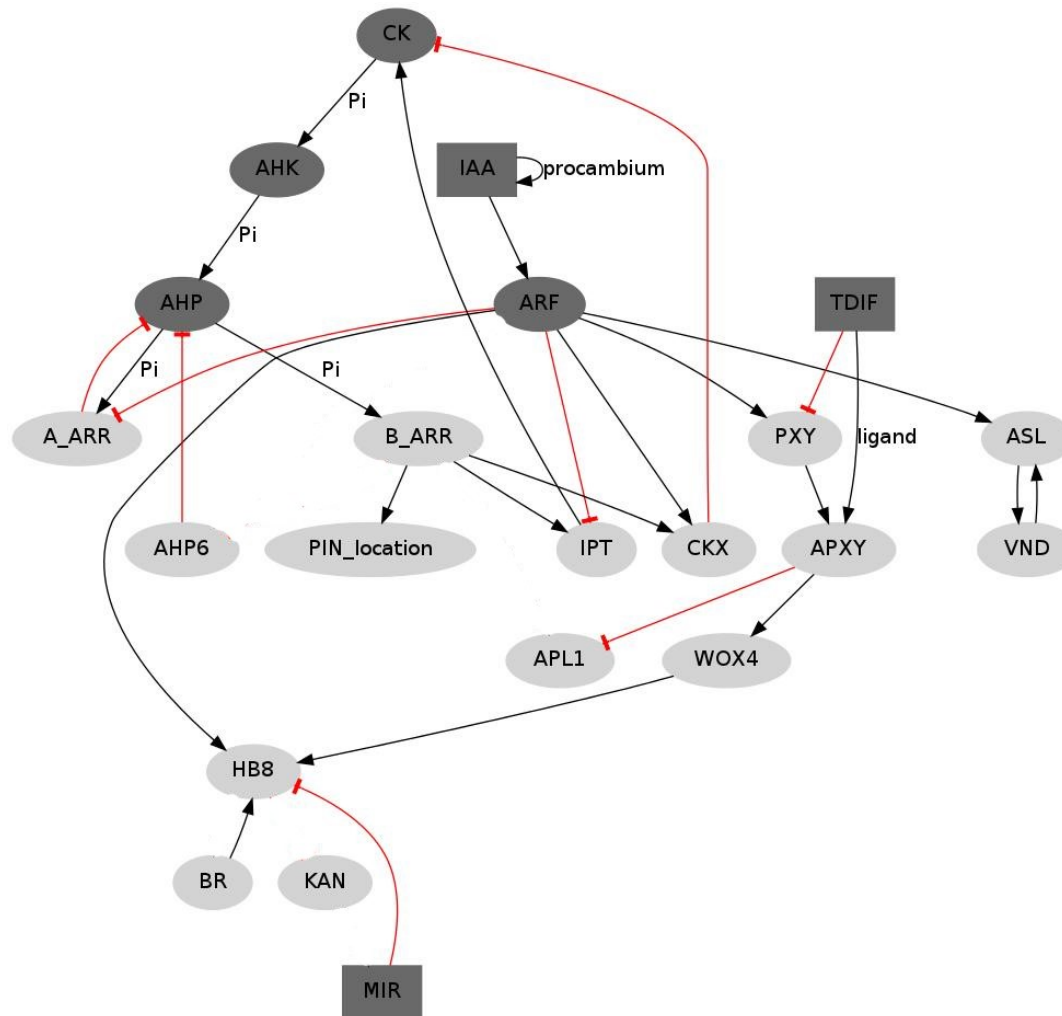
where  $g(t)T[i]$  is the total amount of TDIF or MIR165 in cell ( $i$ ).  $D$  is a parameter that determines the proportion of  $g$  that can move from any cell to neighboring ones and is correlated to the diffusion rate of  $g$ .  $b$  is a constant corresponding to a degradation term.  $H$  is a step function that converts the continuous values of  $g$  into a discrete variable that may attain values of 0, 1 or 2.  $N$  stands for the number of neighbors in each cell. Boundary conditions are zero-flux. In the case of IAA, the mobility is defined as active transport dependent on the radial localization of the PIN efflux transporters and is defined by the equation:

$$iaa(t+1)T[i] = H(iaa(t)[i] + Diaa(pin(t)[i+1])(iaa(t)[i+1]) + Diaa(pin(t)[i-1])(iaa(t)[i-1]) - N(Diaa)(pin(t)[i])(iaa(t)[i]) - b_{iaa}) \quad (3),$$

where  $Diaa$  is a parameter that determines the proportion of IAA that can be transported among cells. The transport depends on the presence of IAA and PIN in the cells and  $b_{iaa}$  corresponds to a degradation term. As in equation 2,  $H$  is a step function that converts the continuous values to discrete ones and  $N$  stands for the number of neighbors in each cell. Boundary conditions for IAA motion are also zero-flux.

# Molecular Regulatory Networks Modeling

- Preparing the *first version* of the model and its *testing*



The proposed model considers data that we identified and evaluated through an extensive search (up to January 2012). It takes into account molecular interactions, hormonal and expression patterns, and cell-to-cell communication processes that have been reported to affect vascular patterning in the bundles of Arabidopsis. The model components and interactions are graphically presented in the figure above. In the network model, nodes stand for molecular elements regulating one another's activities. Most of the nodes can take only 1 or 0 values (light gray nodes in the figure), corresponding to "present" or "not present," respectively. Since the formation of gradients of hormones and diffusible elements may have important consequences in pattern formation, mobile elements TDIF and MIR, as well as members of the CK and IAA signaling systems, can take 0, 1 or 2 values (dark gray nodes in the figure above) Benitez and Hejatko, submitted.



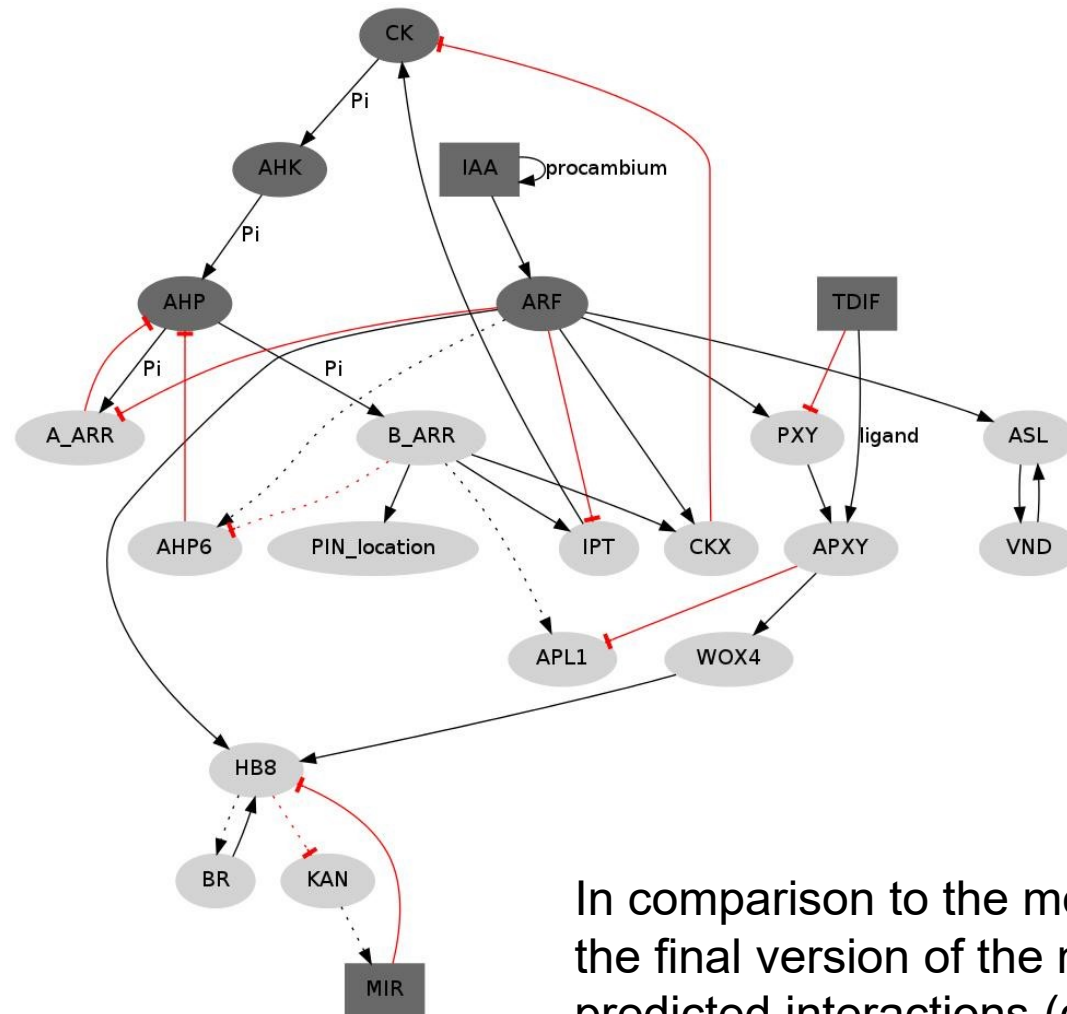
# Molecular Regulatory Networks Modeling

- Specifying of missing interactions via *informed predictions*

Interaction	Evidence	References
CK → PIN7 radial localization	<p>Predicted interaction (could be direct or indirect)</p> <p>Informed by the following data:</p> <p>During the specification of root vascular cells in <i>Arabidopsis thaliana</i>, CK regulates the radial localization of PIN7.</p> <p>Expression of PIN7:GFP and PIN7::GUS is upregulated by CK with no significant influence of ethylene.</p> <p>In the root, CK signaling is required for the CK regulation of PIN1, PIN3, and PIN7. Their expression is altered in <i>wol</i>, <i>cre1</i>, <i>ahk3</i> and <i>ahp6</i> mutants.</p>	<p>[18]</p> <p>[18,20]</p> <p>[19]</p>
CK→ APL	<p>Predicted interaction (could be direct or indirect)</p> <p>Consistent with the fact that APL overexpression prevents or delays xylem cell differentiation, as does CKs.</p> <p>Partially supported by microarray data and phloem-specific expression patterns of CK response factors.</p>	<p>[21]</p> <p>(TAIR, ExpressionSet:1005823559, [22])</p>

# Molecular Regulatory Networks Modeling

- Preparing the *next version* of the model and its *testing*



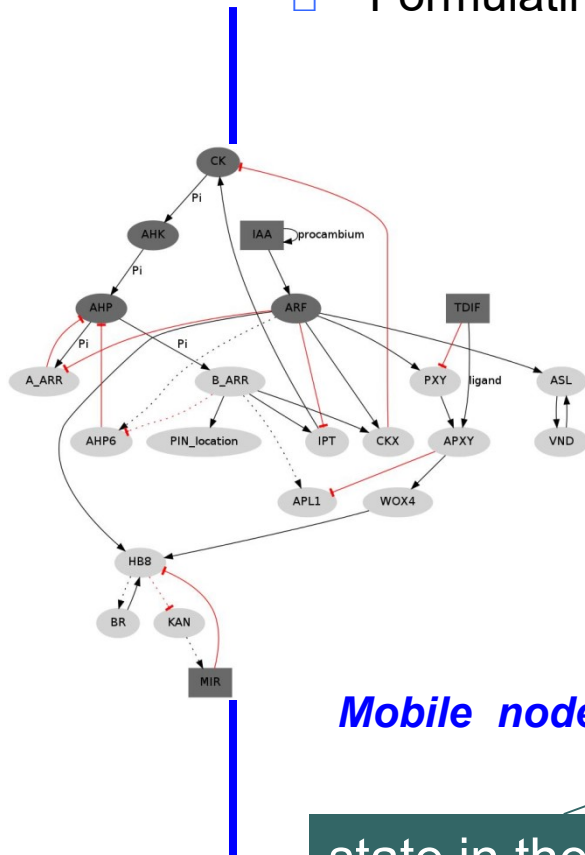
Benitez and Hejatko, *PlosONE*, 2013

In comparison to the model shown on slide 23, the final version of the model contains the predicted interactions (dashed lines).



# Molecular Regulatory Networks Modeling

- Formulating **equations** describing the **relationships** in the model



logical rule function

state in the time  $t$

$$\text{Static nodes: } g_n(t+1) = F_n(g_{n1}(t), g_{n2}(t), \dots, g_{nk}(t))$$

state in the time  $t+1$

Amount of TDIF or MIR165 in cell  $i$

$$\text{Mobile nodes: } g_{(t+1)T[i]} = H(g_{(t)[i]} + D(g_{(t)[i+1]} + g_{(t)[i-1]} - N(g_{(t)[i]})) - b)$$

state in the time  $t+1$

constant corresponding to a degradation term

proportion of movable element

# Molecular Regulatory Networks Modeling

- **Good model** should be able to **simulate reality**

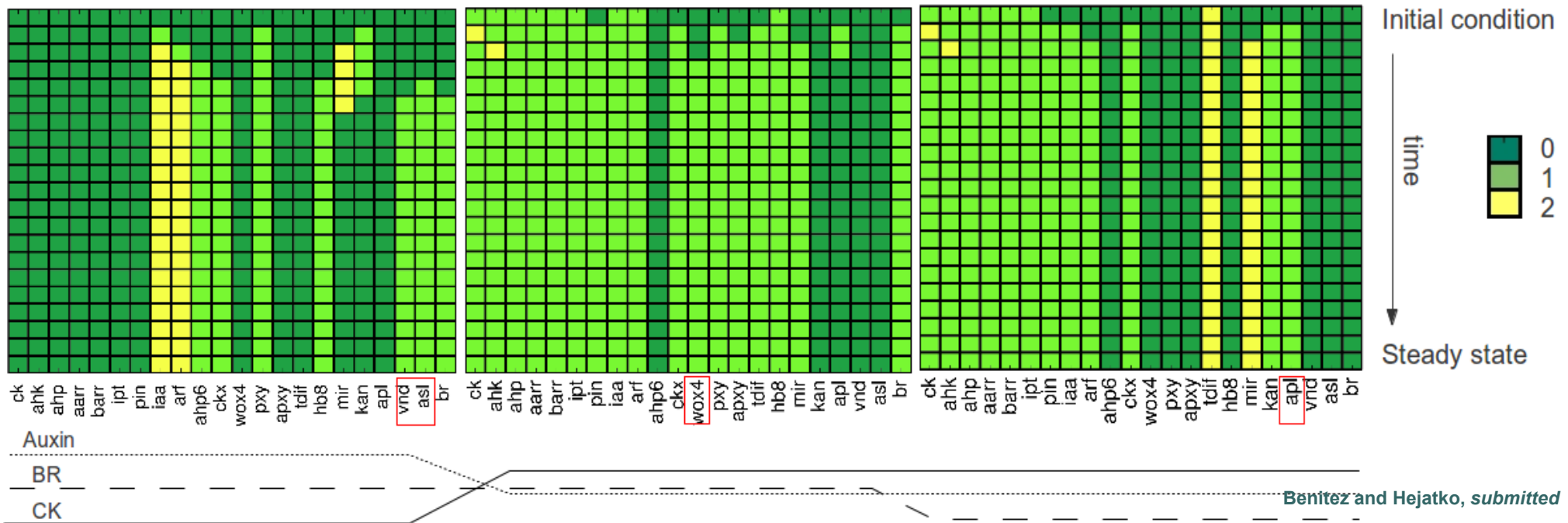
**Static nodes:**  $g_n(t+1) = F_n(g_{n1}(t), g_{n2}(t), \dots, g_{nk}(t))$

**Mobile nodes:**  $g_{(t+1)T[i]} = H(g_{(t)[i]} + D(g_{(t)[i+1]} + g_{(t)[i-1]} - N(g_{(t)[i]})) - b)$

Xylem

Procambium

Phloem



The initial conditions specify the initial state of some of the network elements (figure above) and are the following :

I) In the procambial position (central compartment), CK is initially available and there is an initial and sustained IAA input or self-upregulation. This condition is supported by several lines of evidence. Also *HB8*, a marker of early vascular development that has been found in procambial cells, is assumed to be initially present at this position. These conditions are not fixed, however. After the initial configuration, all the members of the CK and IAA signaling pathways, as well as *HB8*, can change their states according to the logical rules.

II) In the xylem and phloem positions, it is assumed that no element is initially active except for the CK signaling pathway and TDIF, both in the phloem position. The level of expression for a given node is represented by a discrete variable  $g$  and its value at a time  $t+1$  depends on the state of other components of the network ( $g_1, g_2, \dots, g_N$ ) at a previous time unit. The state of every gene  $g$  therefore changes according to:

$$g_n(t+1) = F_n(g_{n1}(t), g_{n2}(t), \dots, g_{nk}(t)) \quad (1).$$

In this equation,  $gn1, gn2, \dots, gnk$  are the regulators of gene  $gn$  and  $F_n$  is a discrete function known as a logical rule (logical rules are grounded in available experimental data, for example see slide 20). Given the logical rules, it is possible to follow the dynamics of the network for any given initial configuration of the nodes expression state. One of the most important traits of dynamic models is the existence of steady states in which the entire network enters into a self-sustained configuration of the nodes state. It is thought that in developmental systems such self-sustained states correspond to particular cell types.

According to experimental evidence for the system under study, the hormone IAA, the peptide TDIF, and the microRNA MIR165/6 are able to move among the cells. In the case of TDIF and MIR165/6, the mobility is defined as diffusion and is given by the following equation:

$$g(t+1)T[i] = H(g(t)[i] + D(g(t)[i+1] + g(t)[i-1] - N(g(t)[i])) - b) \quad (2),$$

where  $g(t)T[i]$  is the total amount of TDIF or MIR165 in cell  $(i)$ .  $D$  is a parameter that determines the proportion of  $g$  that can move from any cell to neighboring ones and is correlated to the diffusion rate of  $g$ .  $b$  is a constant corresponding to a degradation term.  $H$  is a step function that converts the continuous values of  $g$  into a discrete variable that may attain values of 0, 1 or 2.  $N$  stands for the number of neighbors in each cell.

Boundary conditions are zero-flux. In the case of IAA, the mobility is defined as active transport dependent on the radial localization of the PIN efflux transporters and is defined by the equation:

$$iaa(t+1)T[i]=Hiaa(iaa(t)[i]+Diaa(pin(t)[i+1])(iaa(t)[i+1])+Diaa(pin(t)[i-1])(iaa(t)[i-1]) - N(Diaa)(pin(t)[i])(iaa(t)[i]) - biaa) \quad (3),$$

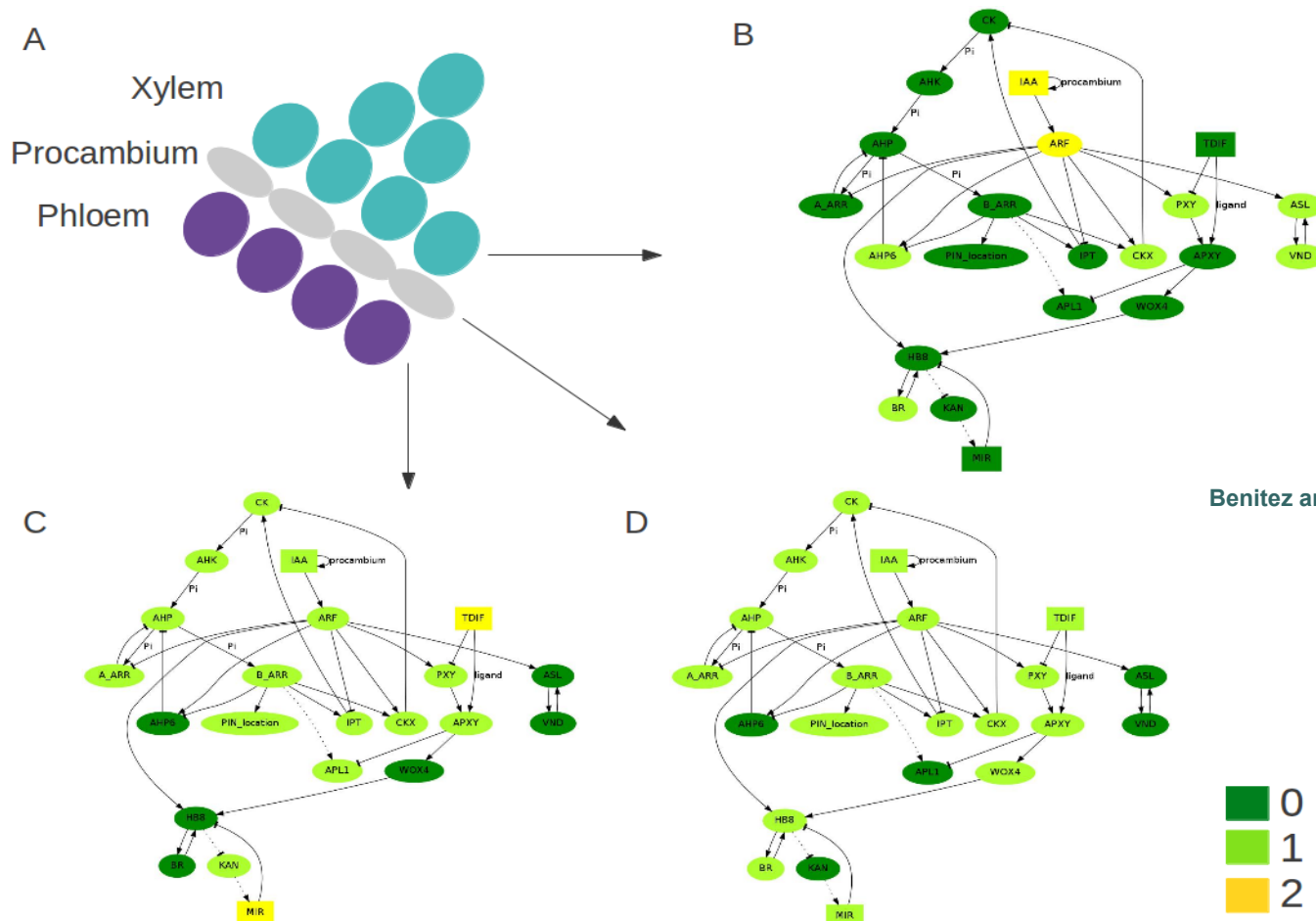
where *Diaa* is a parameter that determines the proportion of IAA that can be transported among cells. The transport depends on the presence of IAA and PIN in the cells and *biaa* corresponds to a degradation term. As in equation 2, *H* is a step function that converts the continuous values to discrete ones and *N* stands for the number of neighbors in each cell. Boundary conditions for IAA motion are also zero-flux.

Using the logical rules, equations 1–3, and a broad range of parameter values (not shown here), it is possible fully to reproduce the results and analyses reported in the following sections (see the figure above for the simulation time course).



# Molecular Regulatory Networks Modeling

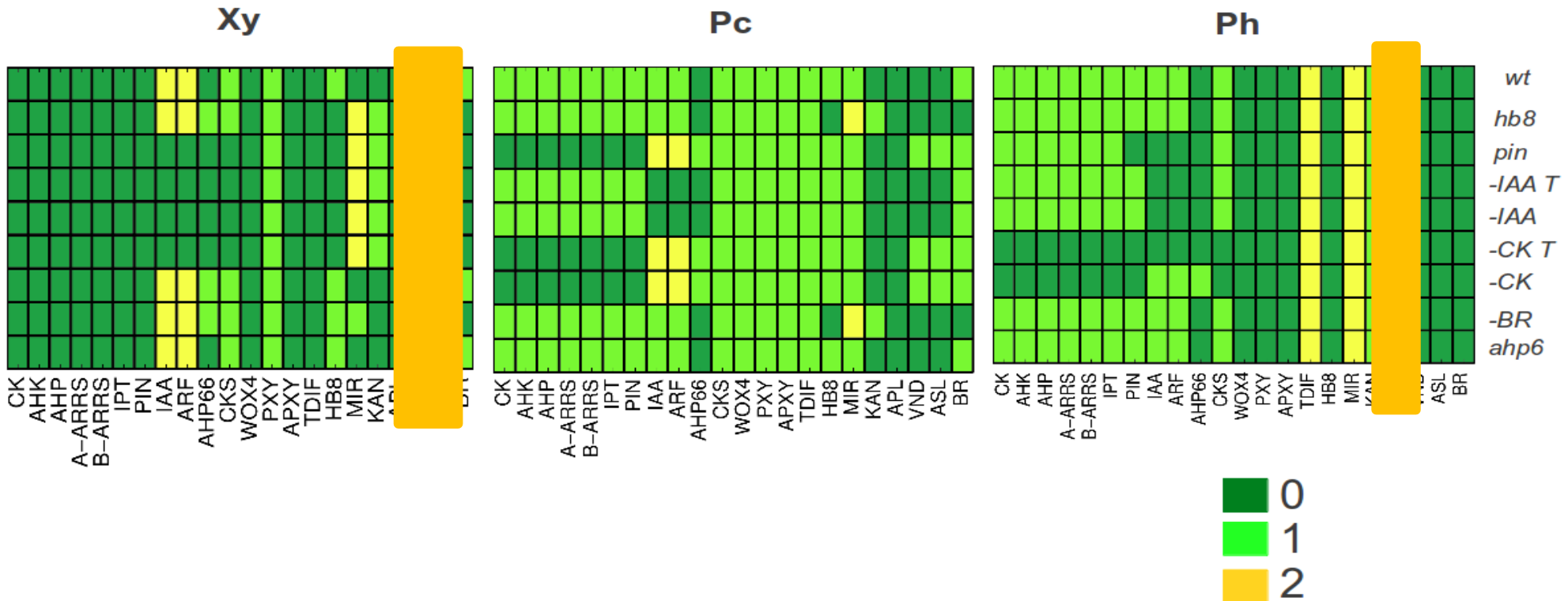
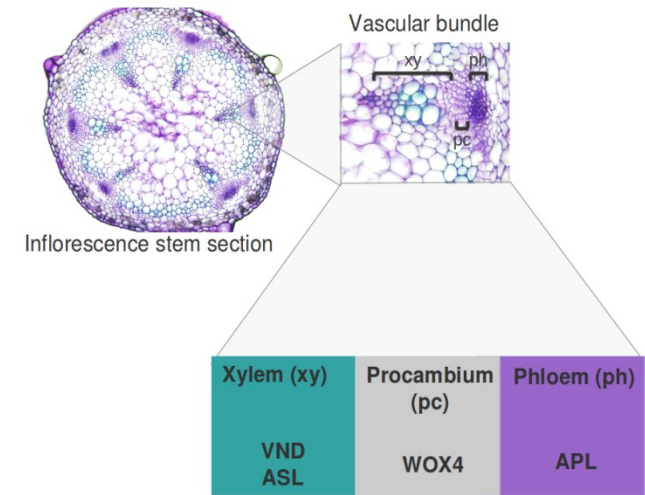
- The **good model** should be able to **simulate reality**



Another representation of the distinct expression profiles in the individual vascular bundle compartments (phloem, procambium and xylem).

# Molecular Regulatory Networks Modeling

- Simulation of *mutants*





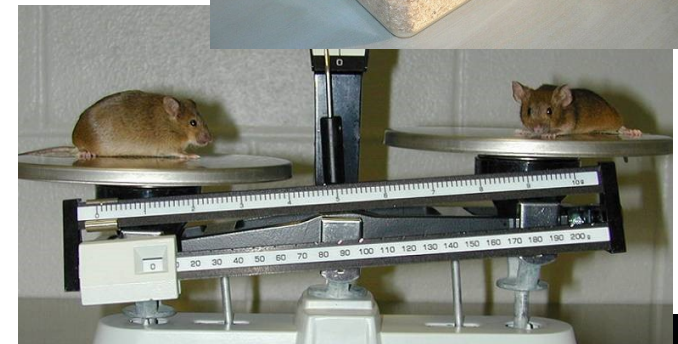
# Outline

- Tools of systems biology
  - Gene ontology analysis
  - Molecular Regulatory Networks Modeling
- Model organisms
  - *Mus musculus*

# *Mus musculus*

## house mouse

- Low requirements for area
- Relatively large number of offspring (3-14, 6-8 on average)
- Genome size is close to the size of human genome (about 3000 Mbp), the number of genes as well (about 24K)
- 20 chromosomes (19+1)
- Suitable for a wide range of physiological experiments (anatomical and physiological similarity to human)
- Possibility to obtain (quite easily) KO mutants and transgenic lines



More info about mouse at

<http://www.informatics.jax.org/greenbook/index.shtml>.

# Mus musculus

house mouse

- Genome known since 2002  
(<http://www.ncbi.nlm.nih.gov/projects/genome/assembly/grc/mouse/>)

Genome Reference Consortium

Mouse Genome Overview

Information concerning the continuing improvement of the mouse genome.

The GRC is working hard to provide the best possible reference assembly for mouse. We do this by both generating multiple representations ([alternate loci](#)) for regions that are too complex to be represented by a single path. Additionally, we are releasing regional fixes known as [patches](#). This allows users who are interested in a specific locus to get an improved representation without affecting users who need chromosome coordinate stability.

**Getting Data**  
 GRCm38.p1 (Latest minor release from the GRC): [FTP](#)  
 GRCm38 (Latest Major release from the GRC): [FTP](#)  
 Alignments of MGSCv37 to GRCm38: [GRC FTP](#)  
 Information on regions under review: [FTP](#)  
 Annotated clone assembly problems: [FTP](#)

**Next assembly update**  
 The next assembly update (patch release 2) will be a minor update (only patches) and will happen in March 2013.

Regions containing alternate-loci  
 Regions containing fix patches

An ideogram representation of the latest mouse assembly (not showing unplaced or unlocalized sequences).

GRCm38.p1 GRCm38 MGSCv37

**GRCm38.p1**  
 Release date: 23 Aug 2012  
 Release type: minor

**GRC Blog**

[The GRC and the 10th International Zebrafish Genetics and Development Meeting \(June 20-24, 2012 - Madison, Wisconsin\)](#) 26 Jul 2012

[Hidden assembly problems exposed](#) 06 Jul 2012

The human reference genome GRCh37 represents th... [see all](#)

**Recently Resolved Mouse Issues**

**Mouse (MG-4136)** Nov16, 2012  
 Inversion found in the assembly has been corrected in component AL611930.25.

**Mouse (MG-4212)** Nov16, 2012  
 There is an assembly gap between AC132090.3 and AC239617.4, originally described in MG-3584, and then updated in MG-4143. This duplicate Jira issue is being used to manage an update at this gap that is being considered for patch release. [see all](#)

**References**

**Whole Genome Papers**  
[The Mouse Genome WGS Assembly](#)  
[The Mouse Genome: Clone based assembly](#)

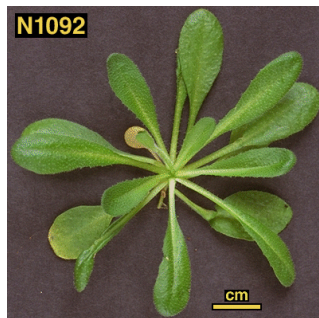
# Outline

- Tools of systems biology
  - Gene ontology analysis
  - Molecular Regulatory Networks Modeling
- Model organisms
  - *Mus musculus*
  - *Arabidopsis thaliana*

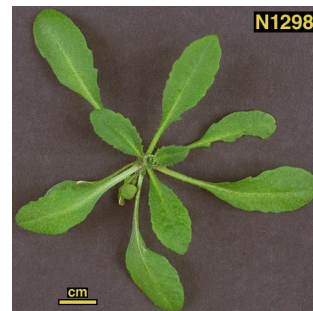
# *Arabidopsis thaliana*

mouse-ear cress

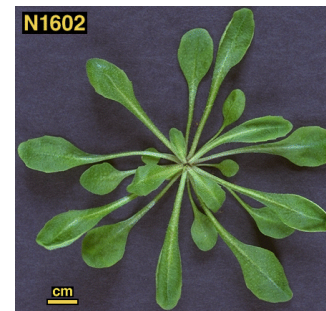
- Low requirements for cultivation area
- High number of seeds (20.000 per plant and more)
- Small and compact genome, (125 MBp, about 25.000 genes, average size 3 kb)
- 5 chromosomes
- Suitable for wide range of physiological experiments
- High natural variability (approximately 750 ecotypes (Nottingham Arabidopsis Seed Stock Centre))



Columbia 0



Landsberg 0



Wassilewskija 0



# Arabidopsis thaliana

mouse-ear cress

- Genome known since 2000 (<http://www.arabidopsis.org/>)

The screenshot shows the TAIR website homepage. The browser window title is "TAIR - Home Page" and the address bar shows "www.arabidopsis.org". The page layout includes a top navigation bar with links for Home, Help, Contact, About Us, and Login/Register. Below this is a secondary navigation bar with tabs for Search, Browse, Tools, Portals, Download, Submit, News, and ABRC Stocks. The main content area is titled "The Arabidopsis Information Resource" and contains several paragraphs of text describing the database and its resources. On the right side, there are sections for "Breaking News" with links to subscribe to news feeds, follow on Twitter, and join a Facebook group. Below that, there are announcements for "New Set of Confirmed T-DNA Lines Available" and "New from ABRC Education and Outreach!". At the bottom of the main content area, there is a large banner for a new online submission form, which includes a form with fields for Article #, Locus, Gene, and GeneFunction Method. The banner text says "Click here to try our new online submission form and submit the molecular function (e.g. protein kinase), biological process (e.g. seed development), localization (e.g. plasma membrane) or interacting partner of your favorite gene". The bottom of the browser window shows the Windows taskbar with various open applications and the system tray.

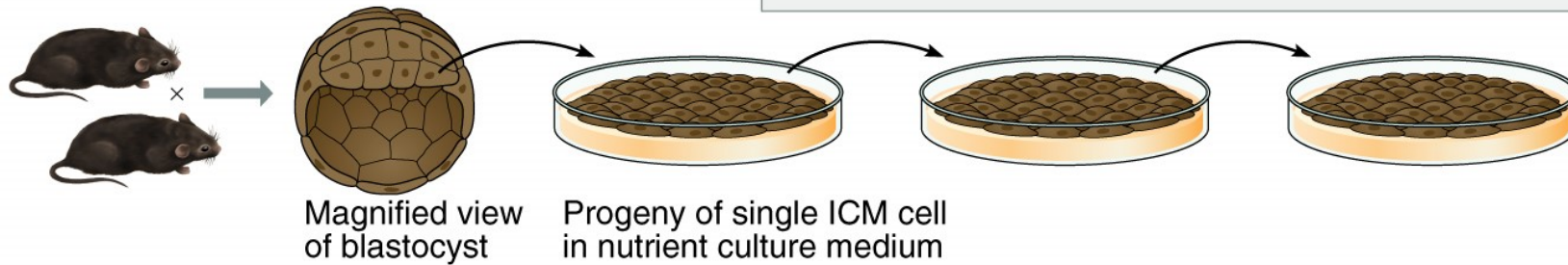


# Outline

- Tools of systems biology
  - Gene ontology analysis
  - Molecular Regulatory Networks Modeling
- Model organisms
  - *Mus musculus*
  - *Arabidopsis thaliana*
- Selected **methods of molecular biology**
  - Preparation of transgenic organisms

1 Isolate single cells from a blastocyst of black mouse parents.

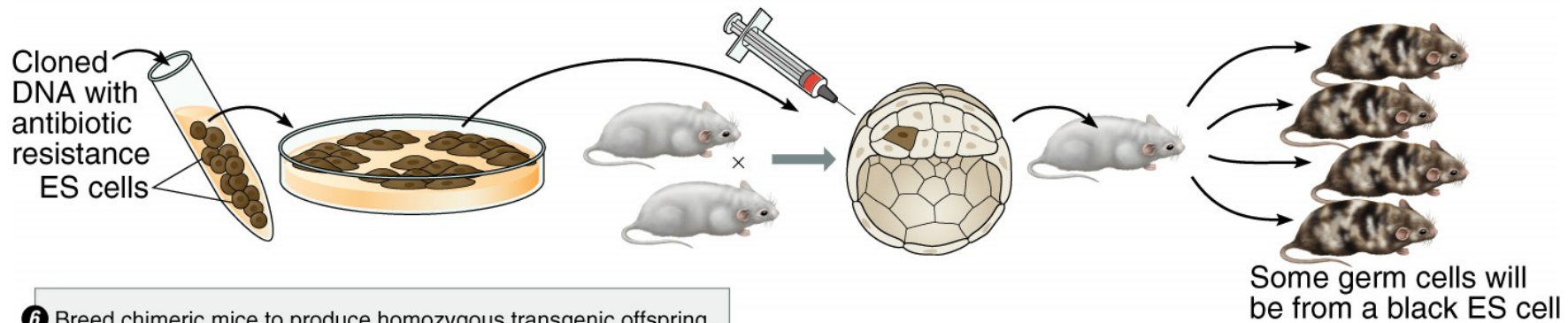
2 Pick a single cell from the first culture and grow a clone of this cell in cultures for 15 mitotic generations. Repeat every 10 days for a year. These are ES (embryonic stem) cells.



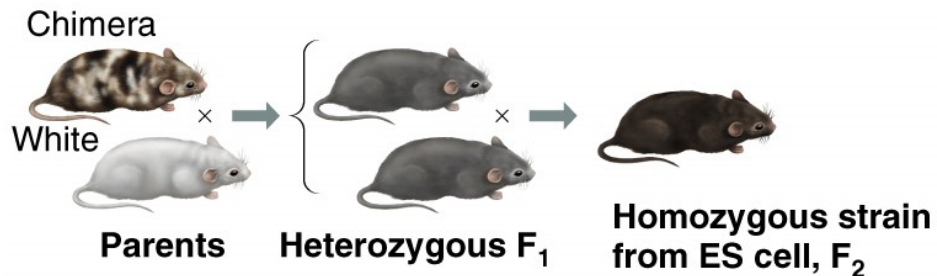
3 Transform stem cells with a cloned gene. Include an antibiotic resistance marker in the cloned gene. Culture ES cells in presence of antibiotic to select transformants.

4 Inject transformed ES cells into blastocysts from white mice. Implant into surrogate mother.

5 Resultant pups will be chimeras of ES cells from black parent and white parent. Black ES cells contain transgene.



6 Breed chimeric mice to produce homozygous transgenic offspring.



Individual ICM cells of the embryo could be isolated and later re-introgressed into the new embryo. These ICM cells are called **embryonic stem (ES) cells**. It is very important technique that allows production of transgenic mice.

The isolated ES cells are transformed via foreign DNA construct and it is injected within the embryo. The transformed cell becomes a part of the embryo and might result into formation of different tissue types, among them the spermatogonia or oogonia. i.e. the tissue that provides progenitor for sperm or egg cells in the resulting chimera. Thus, the progeny of those chimeras will inherit the modified cell with certain probability and these individuals will carry the transgene in every cell of their body. Thus, the transgenic mice will be produced.

This is very important mainly with regard of the knockout mutant (K.O.) production. In the modified ES, the genes might be specifically eliminated via DNA recombination. In that way, function of many of the mice genes was identified.

E.g. the gene *NODAL* is expressed in the anterior portion of the primitive streak that is equivalent to the Hensen's node. *nodal/nodal* embryos are lethal, they do not undergo gastrulation and form almost no mesoderm.

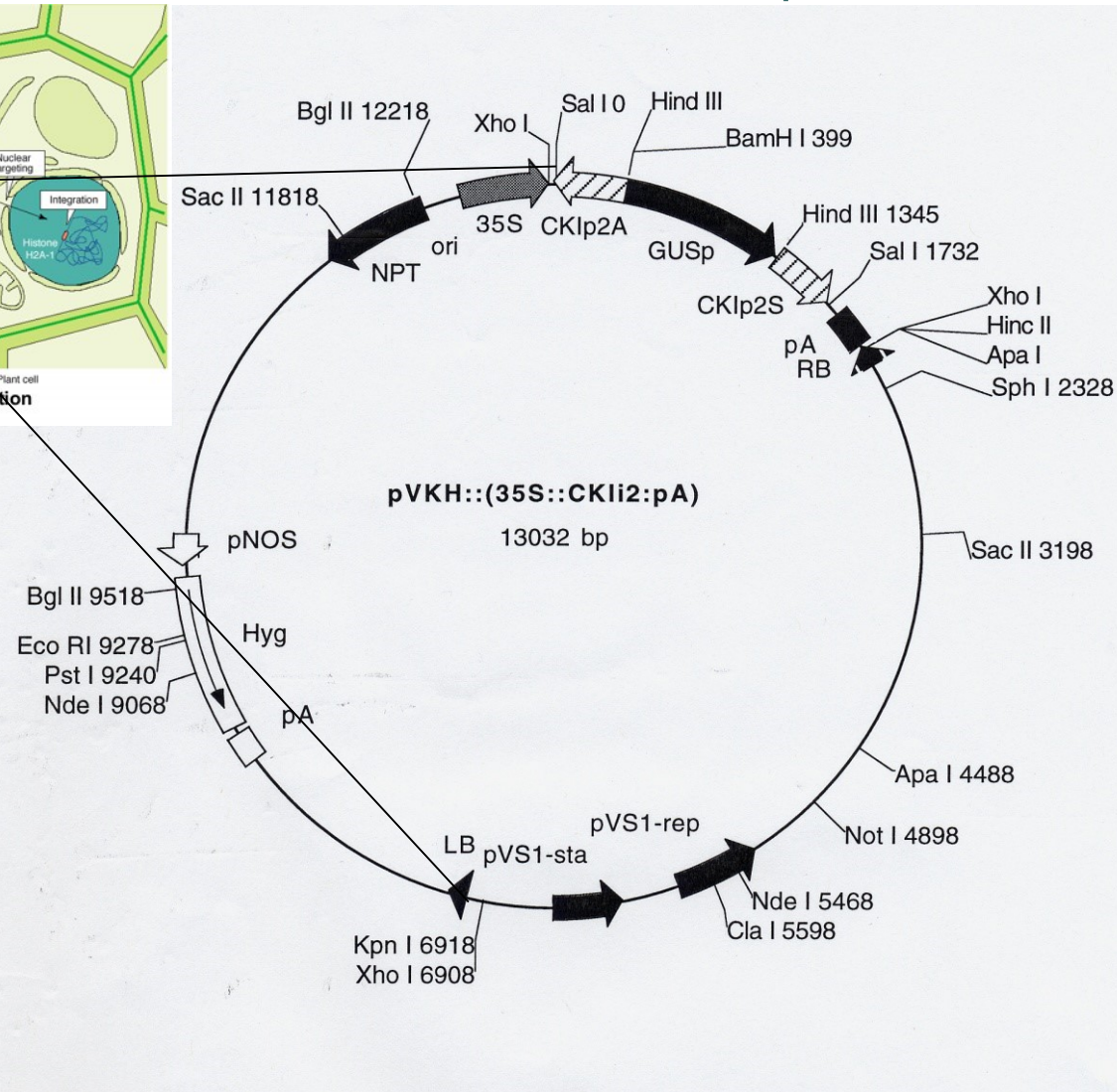
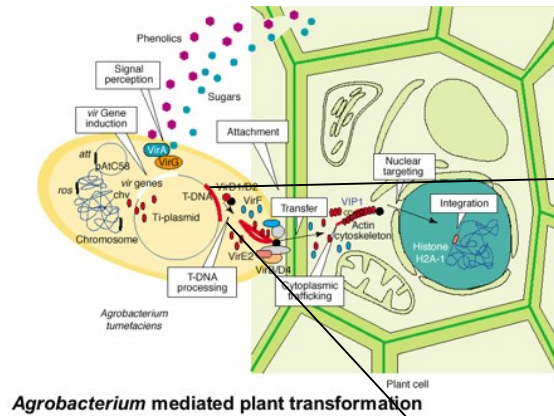
## Transformation of *Arabidopsis* by *Agrobacterium tumefaciens*



**Crown gall of raspberry caused by *Agrobacterium tumefaciens*.**

# Transformation of *Arabidopsis* by *Agrobacterium tumefaciens*

## Transfer of bacterial DNA into plant cells

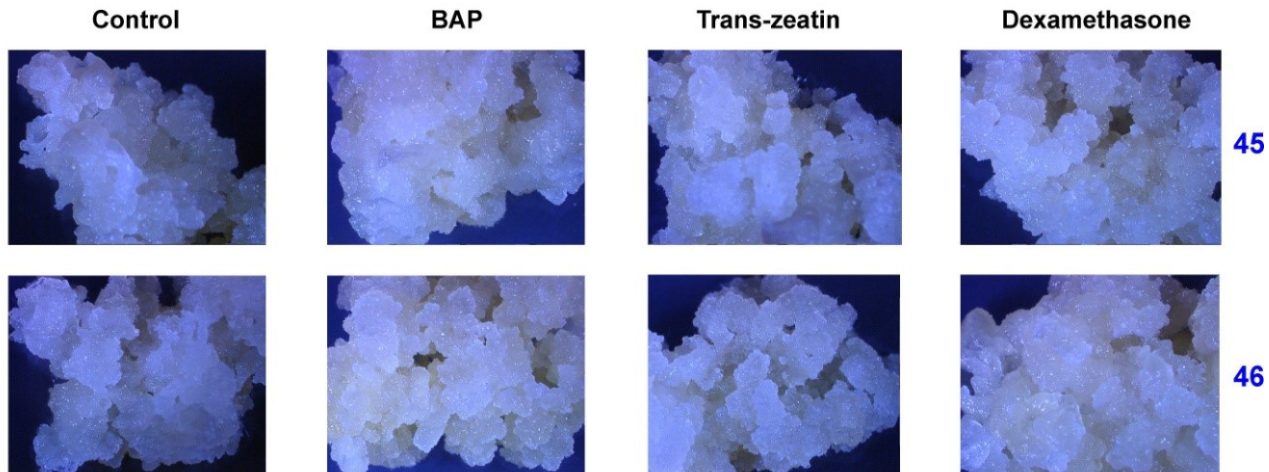


# Transformation by cocultivation of leaf discs

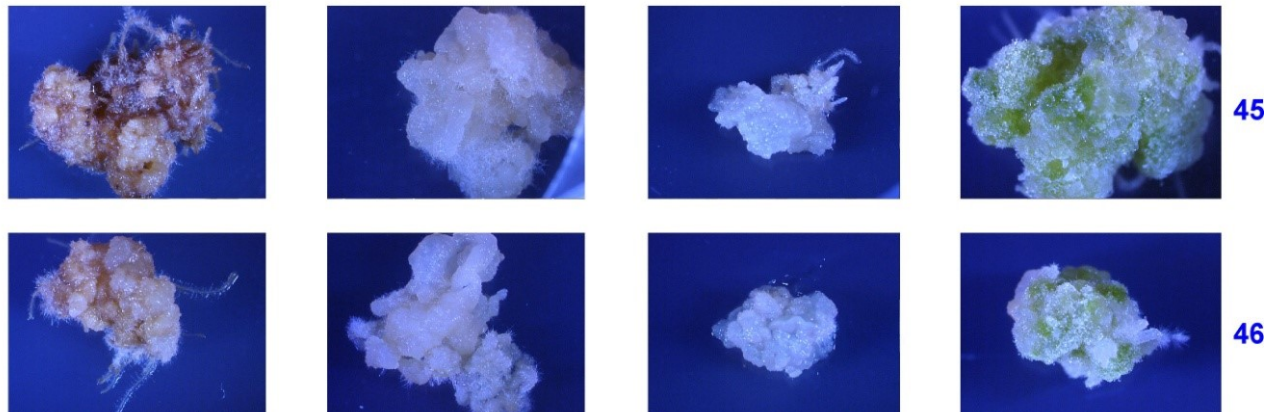


# Transformation by cocultivation of calluses

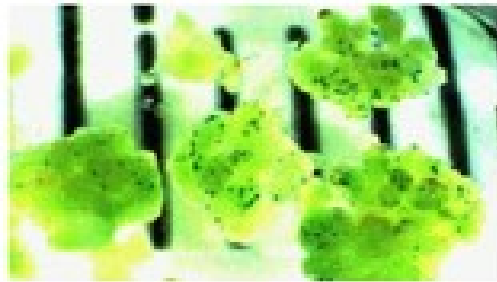
2 mg/l IAA, 0,5 mg/l 2,4-D, 0,5 mg/l 9-ipR



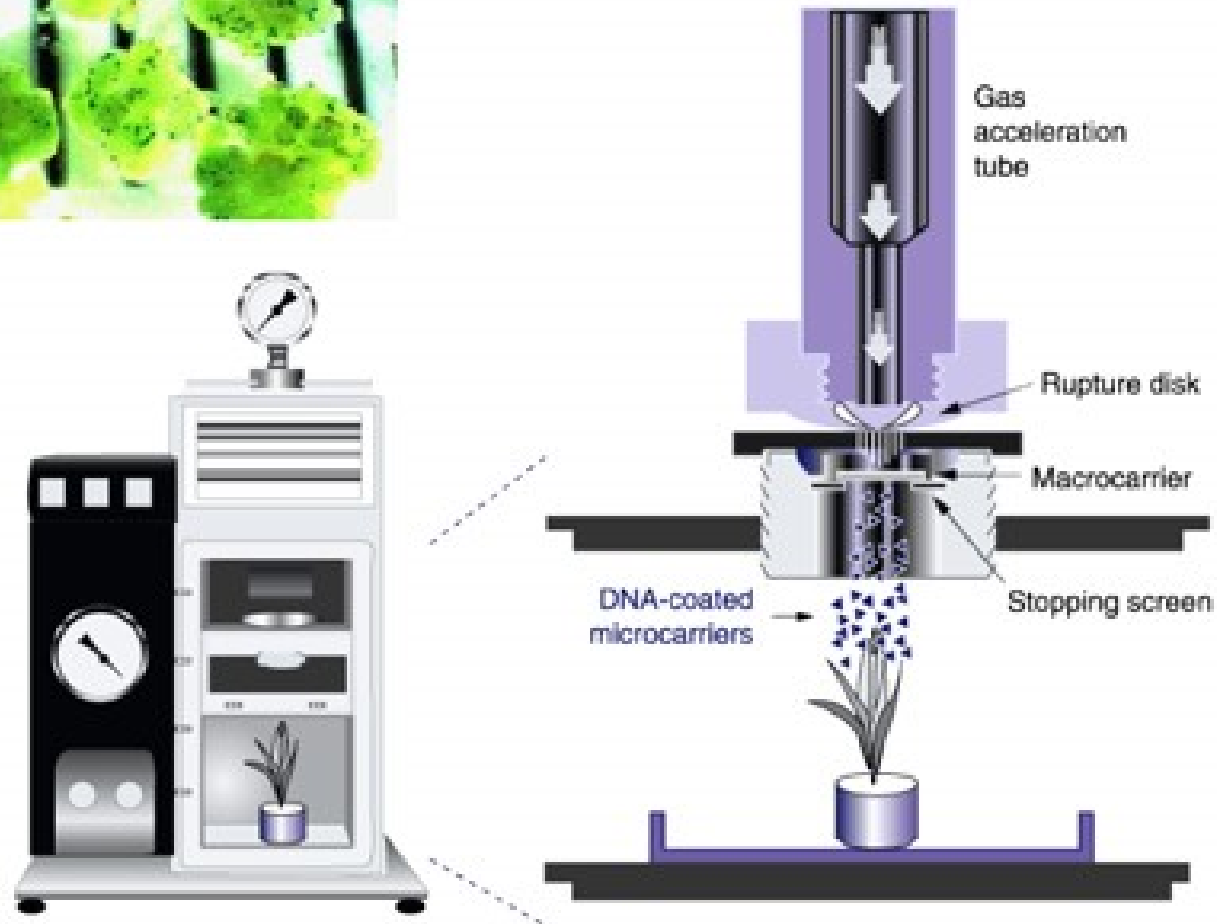
No hormones



## Transformation by biolistic delivery of DNA



### *Biolistic delivery of DNA*





## Transformation of inflorescence



When plants have primary bolts 5-15 cm they are ready to infiltrate. Clipping of primary bolts is not necessary.



After infiltration, pots are placed on their sides to allow for drainage and are covered with plastic wrap. Plants are returned, in this state, to the growth chamber for 24 hours. After 24 hours, they are turned upright into a fresh flat.



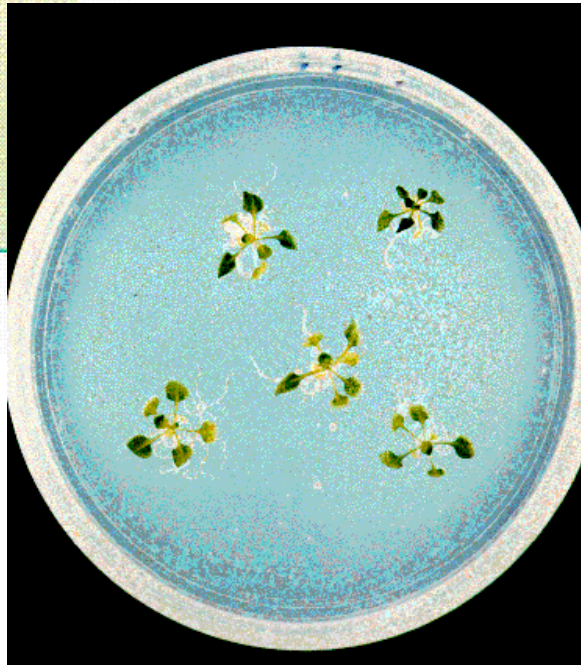
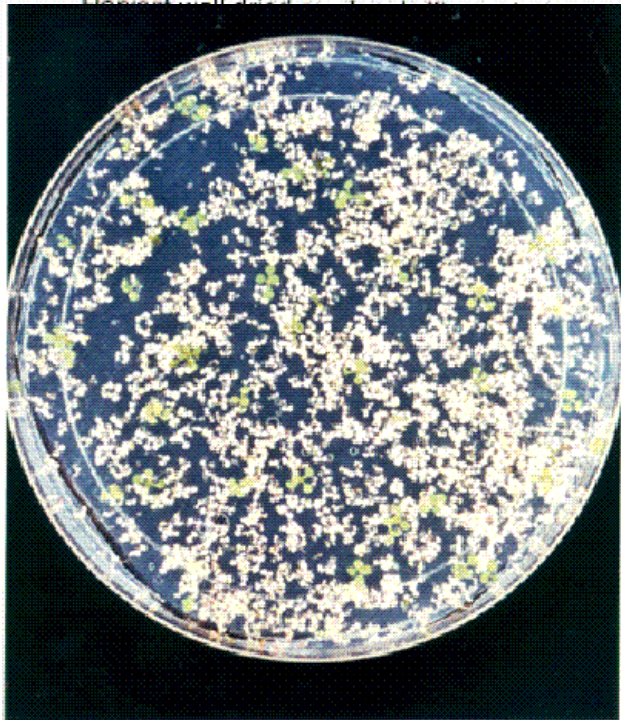
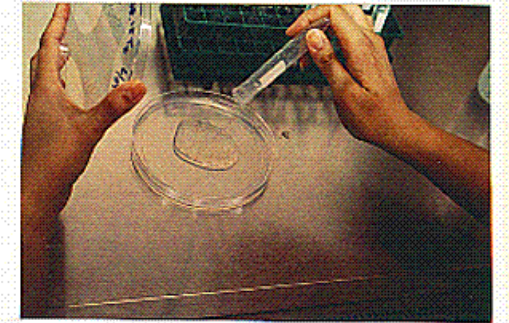
Plants are allowed to grow to maturity. They are staked to avoid seed loss and facilitate plant harvesting.

NOTE: Leaves degenerate within 2 weeks of infiltration. This is normal and does not affect seed set.

# Transformation of inflorescence



Sterilize seed in bleach solution.



Plant transformed seedlings in soil.

<http://www.bch.msu.edu/pamgreen/green.htm>  
Transformed seedlings are grown and have true leaves on selective medium (a 40mg/l kanamycin plate is shown).

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  - Molecular Regulatory Networks Modeling
- Model organisms
  - *Mus musculus*
  - *Arabidopsis thaliana*
- Selected **methods of molecular biology**
  - Preparation of transgenic organisms
  - PCR
  - Design and preparation of primers (Dr. Hana Konečná)