(Paleo)Polyploidy – When Things Get Bigger



Whole-genome duplications



Current Biology 18

Examples of allopolyploid speciation



Hegarty and Hiscock 2008, Current Biology 18

Evolutionary significance of polyploidy



Leitch & Leitch (2008) Science 320

Whole-genome duplications of different age



neopolyploidy

mesopolyploidy

paleopolyploidy

time

Allopolyploidy, diversification, and the Miocene grassland expansion



- Most of the allopolyploidization events identified here occurred in the Late Miocene, simultaneous with or following the well documented expansion of the C4 grasslands.
- The dominant species of modern C4 grasslands are members of Andropogoneae, and most are allopolyploid. Many of these ecological dominants whose origin is dated to about 10.5 million years ago (mya) correlates closely with the date when C4 species came to dominate grasslands in Africa and Southern Asia (Pakistan), also estimated about 10-11 mya; the expansion in North America is dated about 7 mya.
- Allopolyploidy is thus correlated with ecological success.

Estep et al., PNAS (2014)

Model of the phylogenetic history of bread wheat (*Triticum aestivum*; AABBDD). Three rounds of hybridization/polyploidy.





Marcussen et al. (2014), Science

Whole-genome duplications in protozoa

• Aury *et al.* (2006) analyzed the unicellular eukaryote *Paramecium tetraurelia*

• most of 40,000 genes arose through at least 3 successive whole-genome duplications (WGDs)

• most recent duplication most likely caused an explosion of speciation events that gave rise to the *P. aurelia* complex (15 sibling species)

• some genes have been lost, some retained

 many retained (duplicated) genes do not generate functional innovations but are important because of the gene dosage effect





Whole-genome duplications in yeast

- genome comparison between two yeast species, *Saccharomyces cerevisiae* (n=16) and *Kluyveromyces waltii* (n=8)
- each region of K. waltii corresponding to two regions of S. cerevisiae
- the S. cerevisiae genome underwent a WGD after the two yeast species diverged

• in nearly every case (95%), accelerated evolution was confined to only one of the two paralogues (= one of the paralogues retained an ancestral function, the other was free to evolve more rapidly and acquired a derived function)

Whole-genome duplications in yeast



a) after divergence from *K. waltii*, the *Saccharomyces* lineage underwent a genome duplication event (2 copies of every gene and chromosome)

- b) duplicated genes were mutated and some lost
- c) two copies kept for only a small minority of duplicated genes
- d) the conserved order of duplicated genes (nos. 3-13) across different chromosomal segments
- e) comparison between genomes of *S. cerevisiae* and *K. waltii* reveals the duplicated nature of the *S. cerevisiae* genome



Kellis *et al.* 2004, Nature 428

Duplicated nature of the S. cerevisiae genome





S. *cerevisiae* chromosome 4 with sister regions in other chromosomes

Kellis et al. 2004, Nature 428



First evidence of a WGD in plants

Analysis of the genome sequence of the flowering plant *Arabidopsis thaliana*

The Arabidopsis Genome Initiative* AGI (2000)



What does the duplication in the Arabidopsis genome tell us about the ancestry of the species? As the majority of the Arabidopsis genome is represented in duplicated (but not triplicated) segments, it appears most likely that Arabidopsis, like maize, had a tetraploid ancestor. ...The diploid genetics of Arabidopsis and the extensive divergence of the duplicated segments have masked its evolutionary history.

Arabidopsis Species Are "Paleotetraploids" with 8 or 5 Chromosomes





AGI (2000) Nature, Hu et al. (2011) Nat Genet

The grapevine genome sequence suggests ancestral hexaploidization in major angiosperm phyla

The French-Italian Public Consortium for Grapevine Genome Characterization*

Nature 449, 2007



The formation of the palaeo-hexaploid ancestral genome occurred after divergence from monocots and before the radiation of the Eurosids. Star = a WGD (tetraploidization) event. The γ triplication may have been an ancient auto-hexaploidy formed from fusions of three identical genomes, or allo-hexaploidy formed from fusions of three somewhat diverged genomes.

Tang et al. 2008, Genome Research



WGD events in seed plants and angiosperms



Jiao et al. (2011) Nature; Clark and Donoghue (2017) Proc R Soc

Phylogenetic Tree of Sequenced Genomes with Whole Genome Duplications Marked



CoGePedia (http://genomevolution.org/wiki/)

Charles Darwin's abominable mystery solved?



Archaefructus liaoningensis (140 million year old fossil)



Afropollis (245 million year old angiosperm pollen)



"The rapid development as far as we can judge of all the higher plants within recent geological times is an abominable mystery." (Charles Darwin in a letter to Sir Joseph Hooker, 1879)



Theres is evidence of ancient polyploidy throughout the major angiosperm lineages. It means that a genome-scale duplication event probably occurred PRIOR to the rapid diversification of flowering plants

Plants with double genomes might have had a better chance to survive the Cretaceous–Tertiary extinction event

Jeffrey A. Fawcett^{a,b,1}, Steven Maere^{a,b,1}, and Yves Van de Peer^{a,b,2}

PNAS 106 (2009)

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Could WGD event(s) help plants to survive the mass extinction (one or more catastrophic events such as a massive asteroid impact) at the Cretaceous-Tertiary boundary ?

Analysis of 41 plant genomes supports a wave of successful genome duplications in association with the Cretaceous–Paleogene boundary

Kevin Vanneste,^{1,2} Guy Baele,³ Steven Maere,^{1,2} and Yves Van de Peer^{1,2,4}

Genome Res (2014)

Possible establishment of polyploid plants following the K/Pg mass extinction (66 million y. ago)

WGDs clustered around theCretaceous-Tertiary (KT) boundary

the KT extinction event - the most recent mass extinction (one or more catastrophic events such as a massive asteroid impact and/or increased volcanic activity)

 the KT extinction event extinction of 60% of plant species, as well as a majority of animals, including dinosaurs





Lohaus and Van de Peer (2016) Curr Opin Pl Biol

Whole-genome duplication in land plants

Cambrian– Carbonifer	Permia	in	Triassic	Jurassic	Cretaceous	Paleogene- Quaternary	
Plants	ous		Angiosperms				 Cucumis melo Cucumis sativus Citrullus lanatus Juglans regia Malus domestica Pyrus bretschneideri Pyrus persica Prunus mume Fragaria vesca Cilycine max Calganus cajan Medicago fruncatula Cicer arietinum Lotus Japonicus Ricinus communis Mainibat esculenta Jatropha curcas Linum usitatissimum Populus trichocarpa Salix suchowensis Brassica rapa Theilungila parvula Arabidopsis hyrata Casiyptus grandis Vitis vinifera Solanum luberosum Petunia hybrida Ipomea nil Solanum luberosum Patuna sativa Daucus carota Neliengo fumosa Brachypodium distachyon Hordeum vulgare Oryza sativa Zarapha curmina Solanum locolor Seatria talica Anabidopsis prata Patunia hybrida Ipomoea nil Solanum luberosum Petunia carota Malus bo nucljera Aquilegia formosa Brachypodium distachyon Hordeum vulgare Oryza sativa Zea mays Sorghum bicolor Setaria talica Ananas comosus Musa acuminata Phoenix dactylifera Phoenix app. Pinus spp. Pinus
Moss							Ceratodon purpureus Physcomitrella patens
					_		

Van de Peer et al. (2017) Nat Review

Orchids probably share a common WGD driving the early divergence of the family



- largest vascular plant family
- c. 28,000 species
- 736 genera



Multiple WGDs across the Asteraceae family tree



- the second largest family of vascular plants
- some 24,700 species
- enormous karyological variation ->180 different mitotic chromosome counts
- chromosome numbers from n = 2 to
 c. n = 216



Huang et al. (2016) MBE

Whole-genome duplications, diploidization, and the consequences



Consequences of WGD events: the Solanaceae-specific genome triplication (49 million y. ago) contributed to the evolution of the tomato fruit



Gene and genome duplications, key innovations and coevolution



- WGDs (core Brassicales, Brassicaceae)
- chemical arms race
- plants glucosinolates
- butterflies countertactic (detoxification)
- repeated escalation of key innovations (glucosinolate synthesis) → diversification in Brassicales plants and Pierinae butterflies over 80 million years



Edger et al. (2015) PNAS

Gene and genome duplications, key innovations and coevolution





Whole-genome duplication and diploidization

Whole-genome duplication and diploidization



The WGD Radiation Lag-Time Model



Cytogenomic features of post-polyploid genome diploidization



- recurrent WGDs
- different age of WGDs
- different diploidization rates

Cytogenetic evidence for biased subgenome fractionation during post-polyploid diploidization.



Two Evolutionarily Distinct Classes of Paleopolyploidy

Olivier Garsmeur,^{†,1} James C. Schnable,^{†,2} Ana Almeida,² Cyril Jourda,¹ Angélique D'Hont,^{*†,1} and Michael Freeling^{*,†,2}

Species	WGD Class	Substitution Rate (Ks)	Bias Ratio between	Fractionation Pattern	Genome Dominance	Expression Data from	
			Duplicate Regions				
Medicago	I	0.87	1.23	Biased	No data		
Sorghum	1	0.95	1.24	Biased (Schnable et al. 2012)	Yes	Dugas et al. (2011)	
Arabidopsis	1	0.76	1.17	Biased (Thomas et al. 2006)	Yes	Gan et al. (2011)	
Brassica	1	0.34	1.47	Biased (Wang et al. 2011)	Yes (Cheng et al. 2012)		
Maize	1	0.17	1.46	Biased (Woodhouse et al. 2010)	Yes (Schnable et al. 2011)		
Poplar	н	0.23	1.05	Unbiased	No data		
Soybean	н	0.15	1.03	Unbiased	No	Schmidt et al. (2011)	
Banana	II	0.39	1.06	Unbiased	No	D'Hont et al. (2012) and supplementary table S4, Supplementary Material online	

Table 2. Fractionation Pattern and Genome Dominance in Eight Species.





Evolution of the Ancestral Crucifer Genome –

ANCIENT POLYPLOIDS



Brassicas Are Ancient Hexaploids (Mesopolyploids)



Parkin et al. (2005) Genetics

Diplotaxis erucoides 2n = 14



Morisia monanthos 2n = 14 Brassica oleracea 2n = 18



Moricandia arvensis 2n = 28



Lysak et al. (2005) Genome Res, (2007) Plant Physiol

Diploidization in *Brassica* is marked by the asymmetrical evolution of polyploid genomes



The genome of the mesopolyploid crop species Brassica rapa

The Brassica rapa Genome Sequencing Project Consortium



The density of orthologous genes in **three subgenomes** (LF, MF1 and MF2) of *B. rapa* compared to *A. thaliana*.

Three *B. rapa* Subgenomes Contain Genome Block Associations Unique to the tPCK Ancestral Genome



Cheng et al. (2013) Plant Cell, Mandakova and Lysak (2008) Plant Cell

Whole-Genome Triplication Spurred Genome and Taxonomic Diversity in *Brassica* and Tribe Brassiceae



ORIGINAL ARTICLE

Multispeed genome diploidization and diversification after an ancient allopolyploidization

Terezie Mandáková¹ | Milan Pouch¹ | Klára Harmanová¹ | Shing Hei Zhan² | Itay Mayrose³ | Martin A. Lysak¹

Australia: 15 genera, 47 species New Zealand: *Pachycladon*, 11 species





The allopolyploid origin evidenced by single-copy nuclear gene phylogenies





Polyploid Origin of Pachycladon (n=10)



Mandakova, Heenan and Lysak (2010) BMC Evol Biol



Does Ancient Polyploidy Explain the Rapid Species Radiation in *Heliophila* ?

c. 90 (-100) endemic spp.



















Whole-Genome Triplication in the Southern African Tribe Heliophileae



		clade	meso- neo-mesopolyploid gen		genomes
			6x (10x) ₩GD	12x (20x) ₩GD	24x (40x)
	4	В	2 <i>n</i> = 16	2 <i>n</i> = 32	2 <i>n</i> = 64
6x (10x)	species	А	2 <i>n</i> = 18	2 <i>n</i> = 36	
		А	2 <i>n</i> = 20	2 <i>n</i> = 40	2 <i>n</i> = 80
	number	B+C	2 <i>n</i> = 22	2 <i>n</i> = 44	2 <i>n</i> = 88
	reduction	С	2 <i>n</i> = 26		

Mandakova et al. (2012) Taxon

Lineage-Specific Mesopolyploid WGDs in Brassicaceae





Mandáková et al. (2017) Plant J

Existing species and genomic diversity of many Brassicaceae clades result from post-polyploid diploidizations



Two hypotetraploid *Cardamine* species:

recent and starting diploidizations via descending dysploidy



Speciation and diversification driven by post-polyploid diploidization *via* descending dysploidy



'Many more, if not all, higher plant species, considered as diploids because of their genetic and cytogenetic behaviour, are actually ancient polyploids' (Paterson *et al.* 2005).