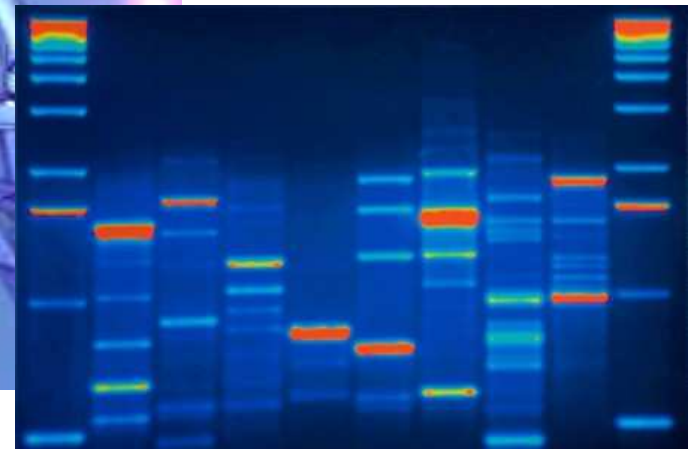
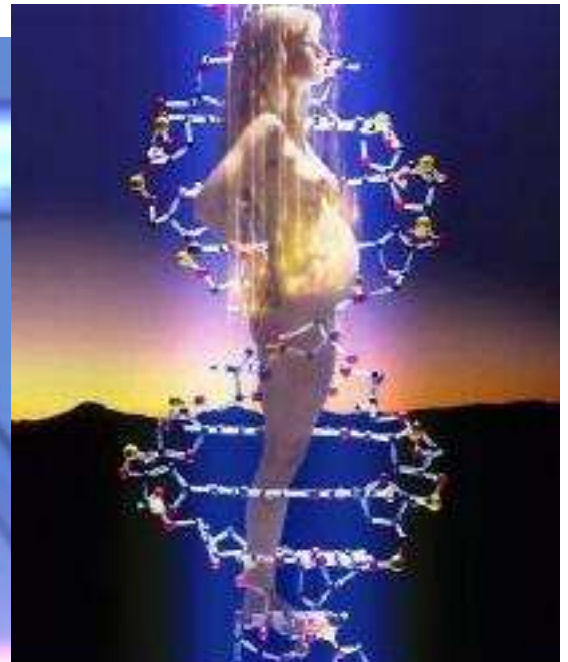
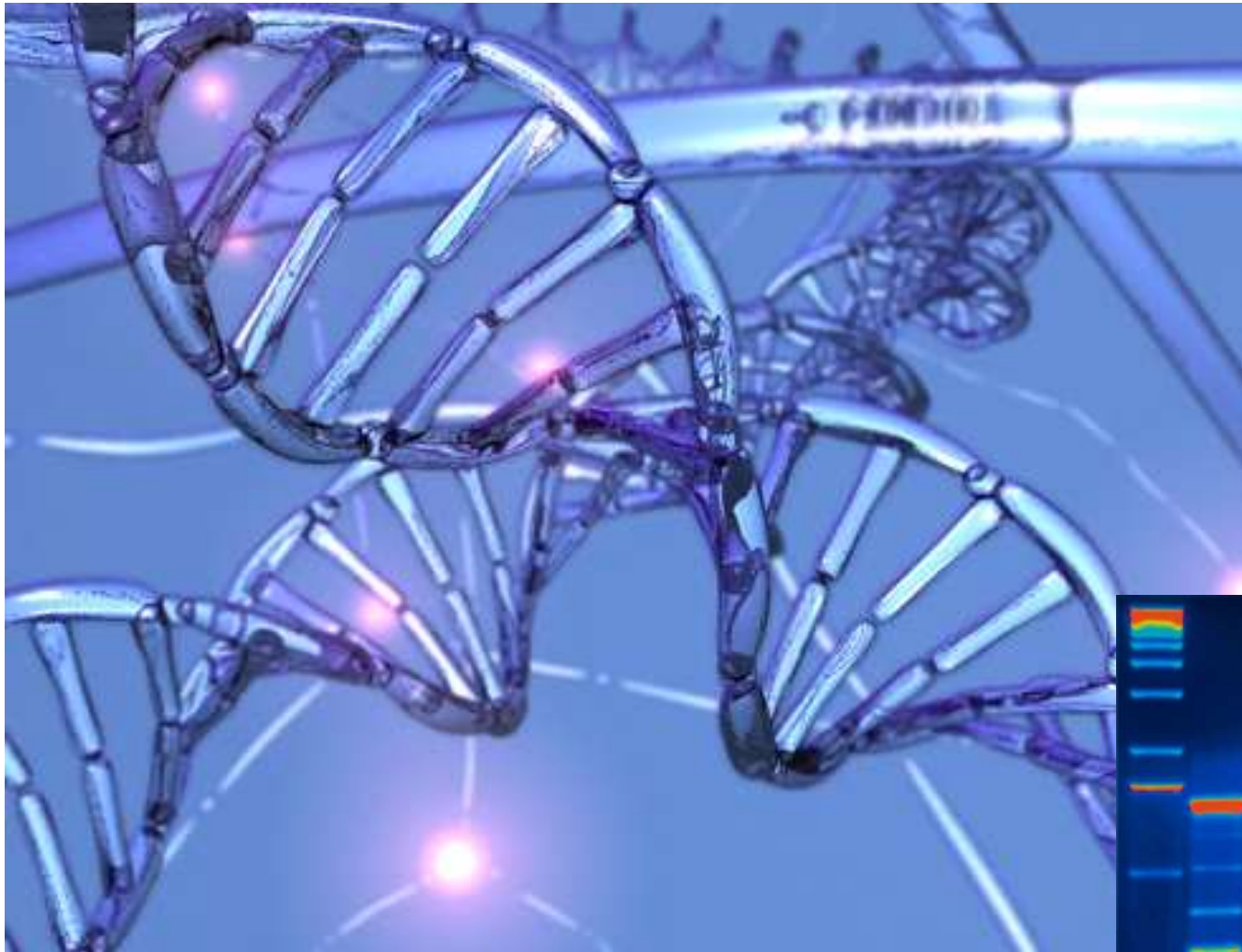
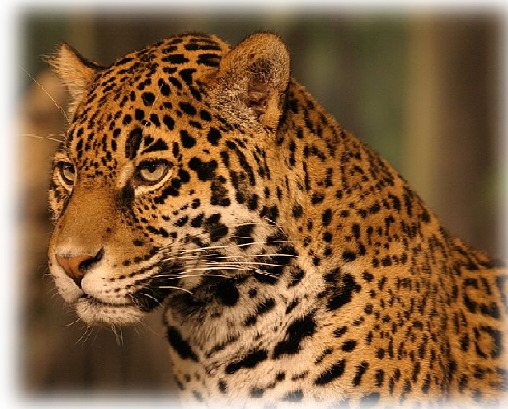


ORIGIN OF GENETIC VARIATION



Consequence of the H-W principle:

if the assumptions of the H-W population hold true, polymorphism can be maintained solely by random mating and Mendelian inheritance



BUT!

real populations usually differ from the model:

population size finite

mating may be nonrandom

migration

selection

emergence of new alleles by mutation

MAIN MICROEVOLUTIONARY MECHANISMS:

mutation (incl. transposition)

recombination

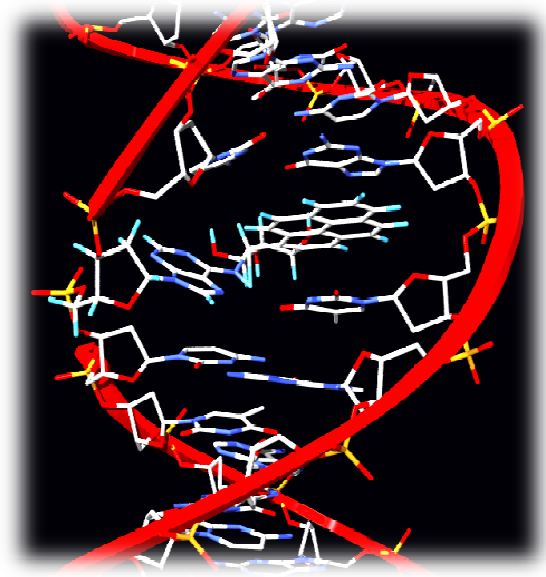
migration (gene flow)

nonrandom mating

natural selection

random genetic drift (incl. bottleneck, founder effect)

(molecular drive)



MUTATION



spontaneous × induced

in germ cells × somatic

according to their deleterious/beneficial effect:

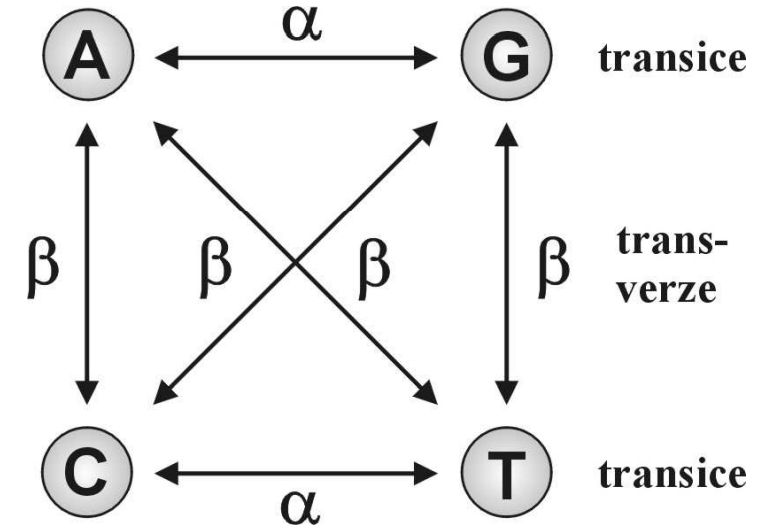
beneficial (positive)

deleterious (lethal, negative)

neutral

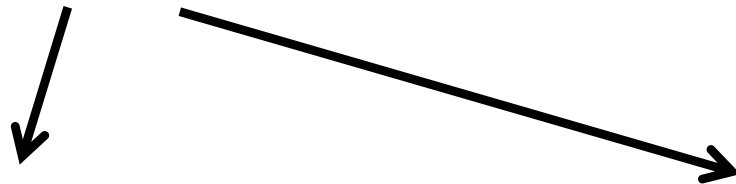
According to effect

point (gene)
chromosomal
genome



Point mutations:

substitutions (transitions, transversions)



synonymous

GTC \rightarrow GTA
Val \rightarrow Val

GTC \rightarrow TTC
Val \rightarrow Phe
AAG \rightarrow TAG
Lys \rightarrow ochre (stop)

nonsynonymous

missense
nonsense

insertions ACGGT → AC**A**GGT
deletions A**C**GGT → AGGT } **indels** → shift of reading frame

back mutations: generally 10-times lower frequencies

recurrent mutations → **mutation pressure**:

eg. when allele frequency $A = 0,5$; $2N = 2000$, i.e. : $N_A = 1000$

after 1st generation → $N = 1001$ ⇒ increase to 0,5005

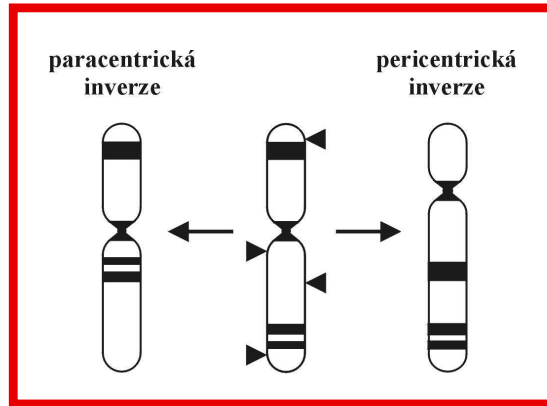
after 100 generations → 0,55 ...

⇒ change of allele frequencies by mutations very slow

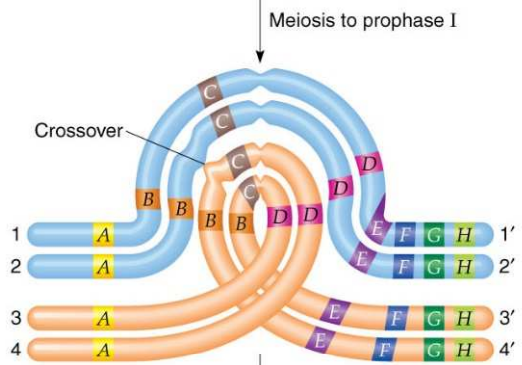
Chromosomal mutations (chr. rearrangements)

inversions

pericentric
paracentric



Normal chromosomes
Inversion chromosome



Normal product
(all genes present)

1 A B C D E F G H 1' Viable

Deletion/duplication
product (EFGH deleted;
A duplicated)

2 A B C D A 4 Inviabile

Inversion product
(all genes present)

3 A D C B E F G H 3' Viable

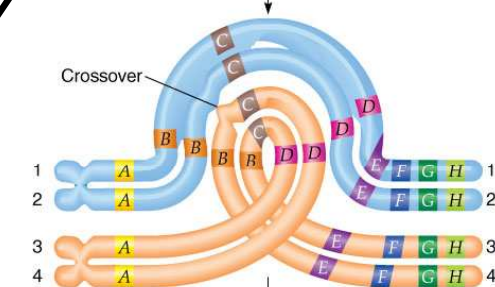
Deletion/duplication
product (A deleted;
EFGH duplicated)

4' H G F E B C D E F G H 2' Inviabile

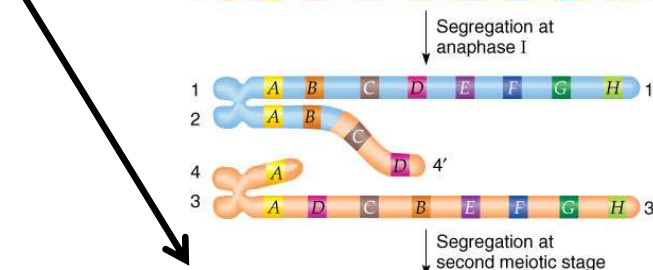
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crossing-over
ale žádná
rekombinace!

Normal chromosome
Inversion chromosome
Meiosis to prophase I



Random break
in dicentric
bridge
Acentric fragment
lost



Segregation at
anaphase I

Segregation at
second meiotic stage

Normal product 1 A B C D E F G H 1'

Deletion products 2 A B C D 4'

4 A

Inversion product
(all genes present) 3 A D C B E F G H 3'

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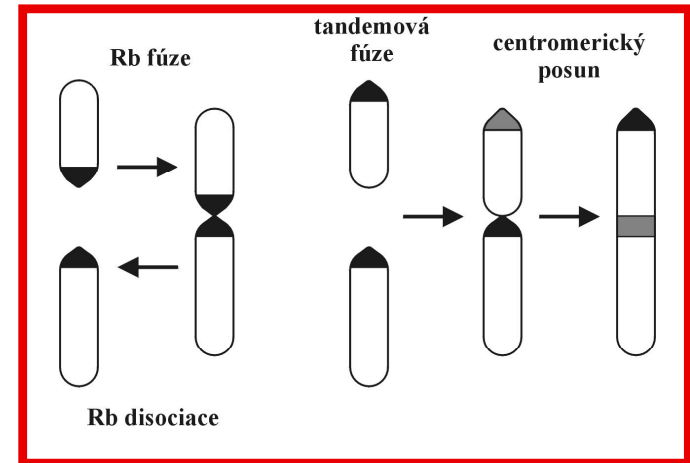
translocations

fusions and dissociations

(Robertsonian translocations)



house mouse



AA AA AA AA AA AA AA AA AA AA

AA AA AA AA AA AA AA AA AA AA

XX XX XX XX XX XX XX XX XX XX

Rapid chromosomal evolution in house mice on the island of Madeira

One population of mice introduced to island in 1400s

Two populations evolved different sets of Robertsonian translocations, hybrid offspring are sterile

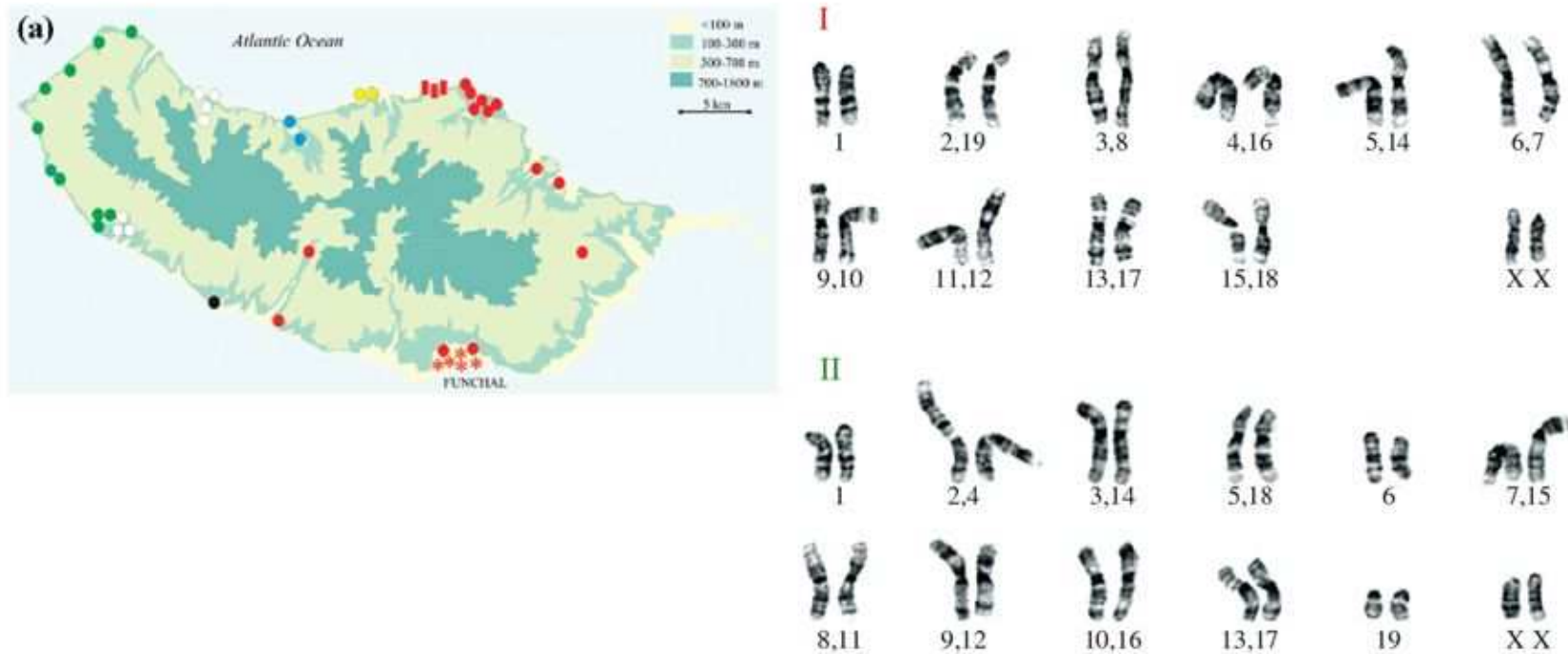


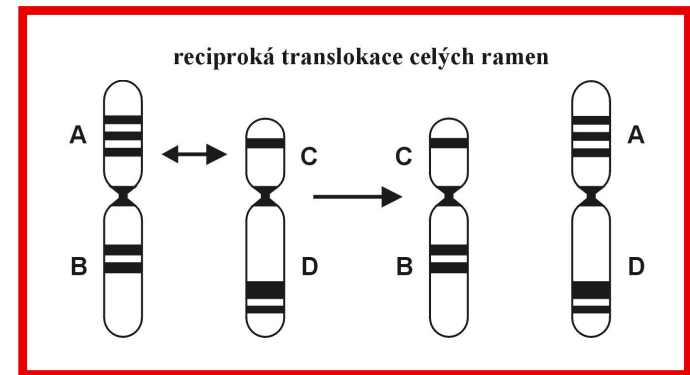
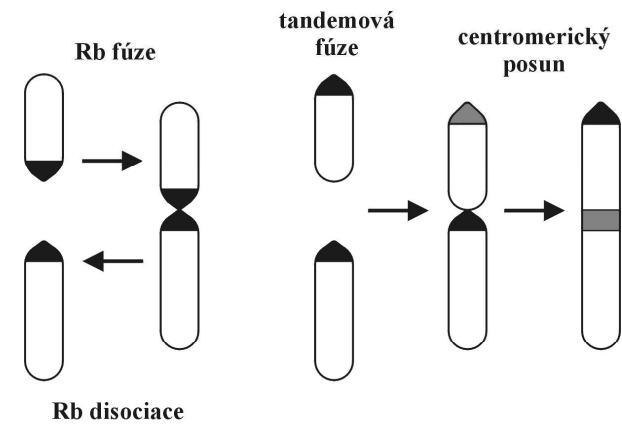
Fig. 13.30

translocations

fusions and dissociations

whole-arm reciprocal translocations (WART)

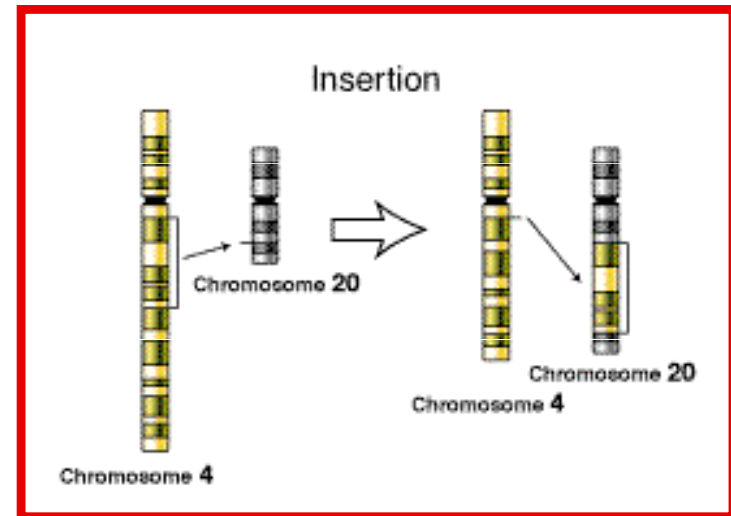
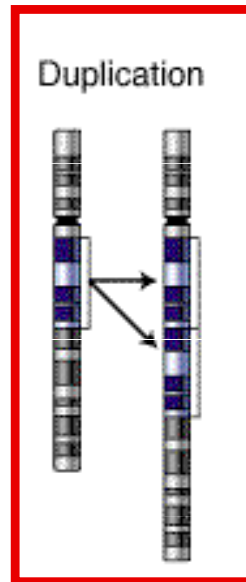
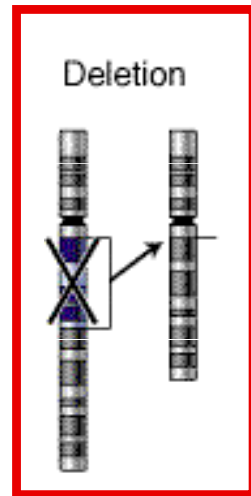
house mouse



deletions

duplications

insertions



Genome mutations

-somes (monosomies, trisomies)

mostly incompatible with life

monosomies: the only viable = X0 (Turner syndrom)

trisomies: imbalance in gene dosage (increased expression of the trisomic pair)

viable trisomies : XXY, XXX, XYY, Patau syndrom (chr. 13),
Edwards s. (chr. 18), Down s. (chr. 21)

-ploidies (polyploidy)

especially plants

in animals less frequent (invertebrates, fishes, amphibians)

during the vertebrate evolution 2 rounds of whole genome duplications
(2R-hypothesis)

polyploid individuals usually bigger (increased cell volume)

odd multiples of the genome → problems in meiosis ⇒ reproductive barrier
(not always – eg. triploid frogs)

autopolyploidy: combination of two identical genomes

fusion of cells

endoreplication

abortive cell cycle

allopolyploidy: combination of two different genomes

fusion of diploid gametes

polyspermy

Randomness and mutation rate (μ)

mutation effects random, position and rate nonrandom

transitions > transversions

mutation „hotspots“: CpG in animals (methylated C \rightarrow T); TpT in Procaryota
„SOS reactions“ in Bacteria, minisatellites (VNTR), microsatellites (STR)

mtDNA > nuclear DNA

sex chromosomes > autosomes

influence of proximity of the replication start, centromeres, telomeres, repetitive sequences, intensity of transcriptions

cold-blooded animals: > temperature \Rightarrow > μ

RNA viruses (HIV)

parasites

antigens, immunoglobulins

> μ of somatic mutations

males > females: humans 6x, rodents, fox: 2x ... more cell divisions in germ cells

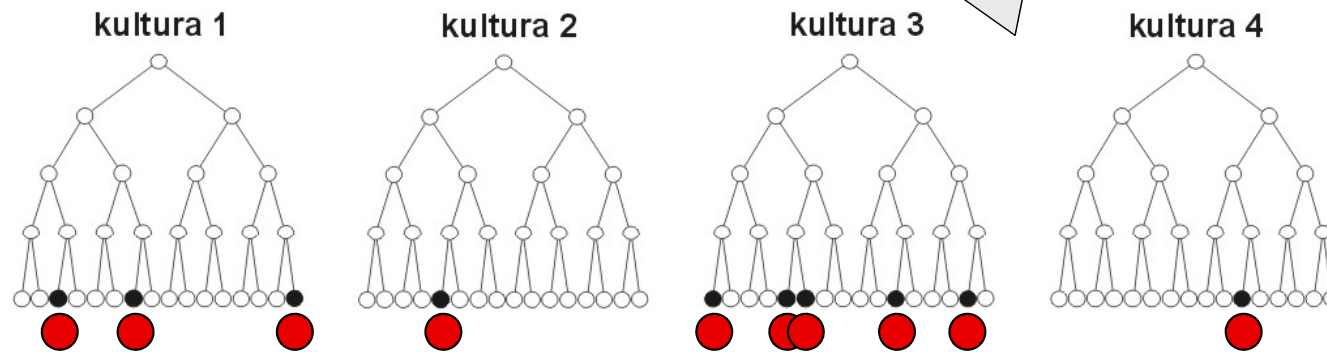
Adaptive (directed) mutations?

Max Delbrück, Salvador Luria (1943):
fluctuation test

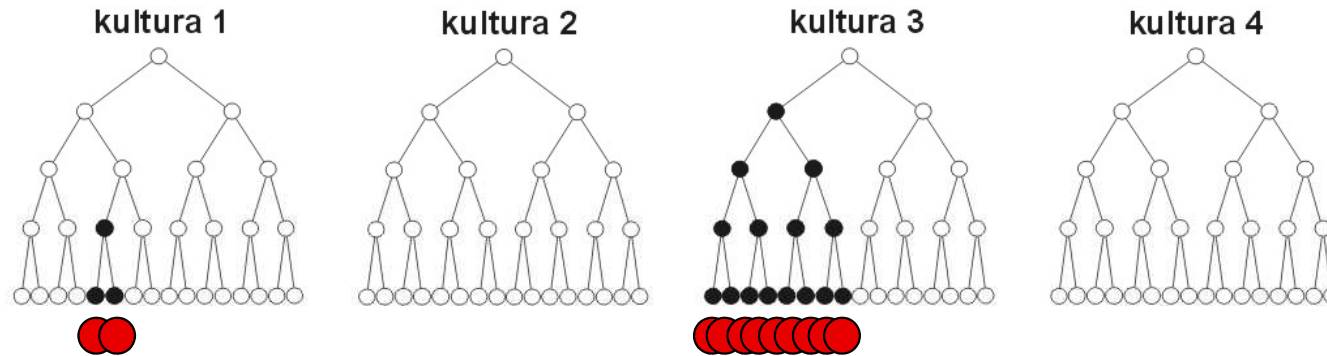


directed mutations

(a) mutace vyvolané prostředím



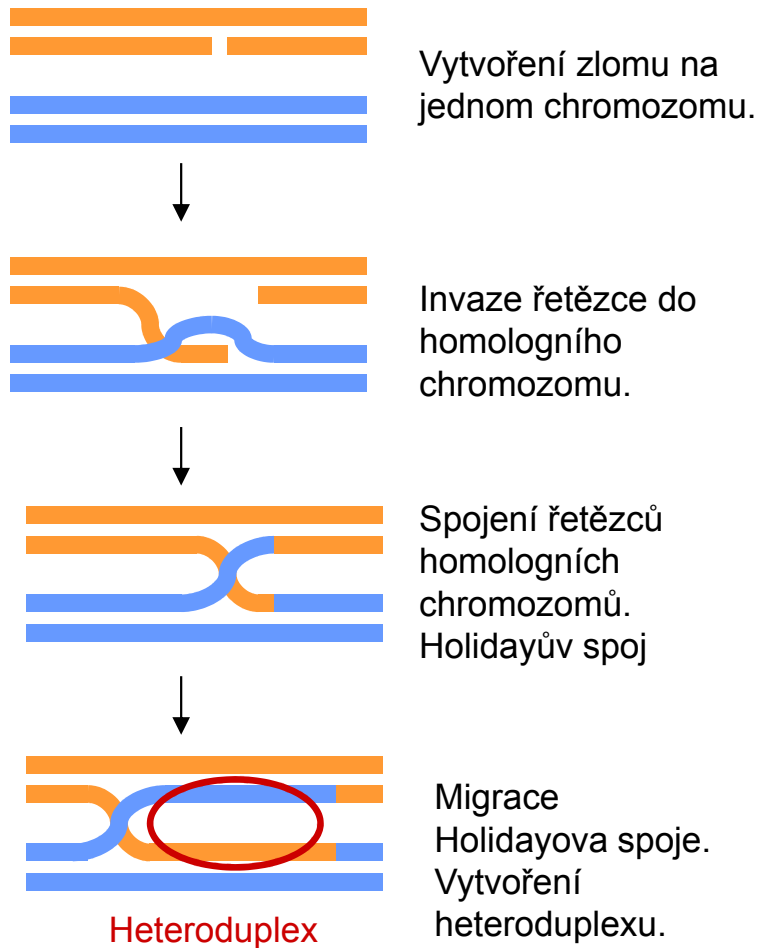
(b) náhodné mutace



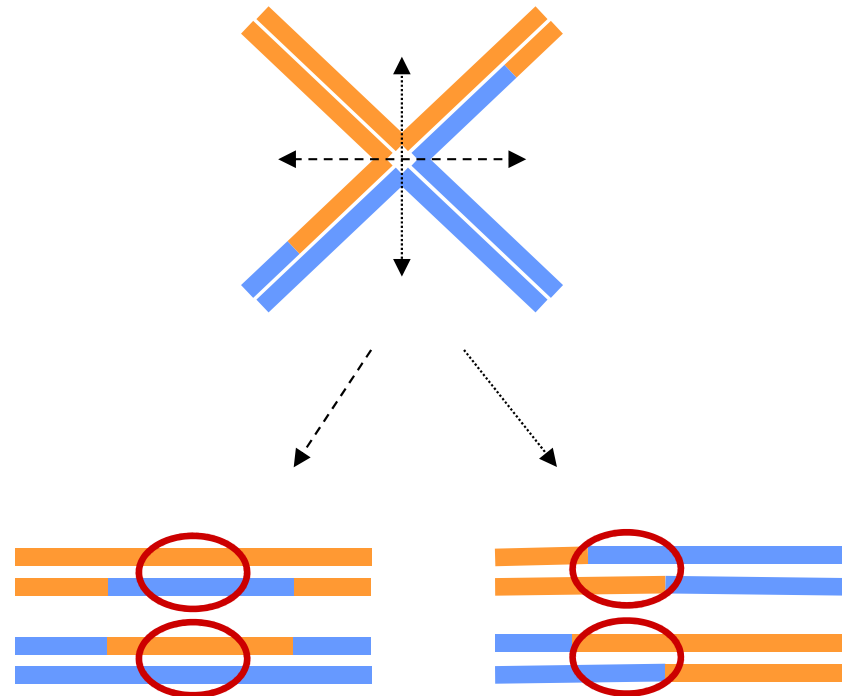
RECOMBINATIONS

mutations → new alleles

recombinations → new genotypes (exception = intragenomic recombination)



Štěpení Holidayova spoje



in many organisms crossing-over important for right meiosis
(at least 1 c-o per chromosome, otherwise aneuploidies)

women with > c-o → > children

children of older women → > recombinations

differences in various parts of chromosome (near centromeres and telomeres etc.,
differences among organisms)

small chromosomes > recombination frequencies

recombination „hotspots“:

humans ~25 000

absent in *Drosophila* and *Caenorhabditis elegans*

frequent appearance and disappearance

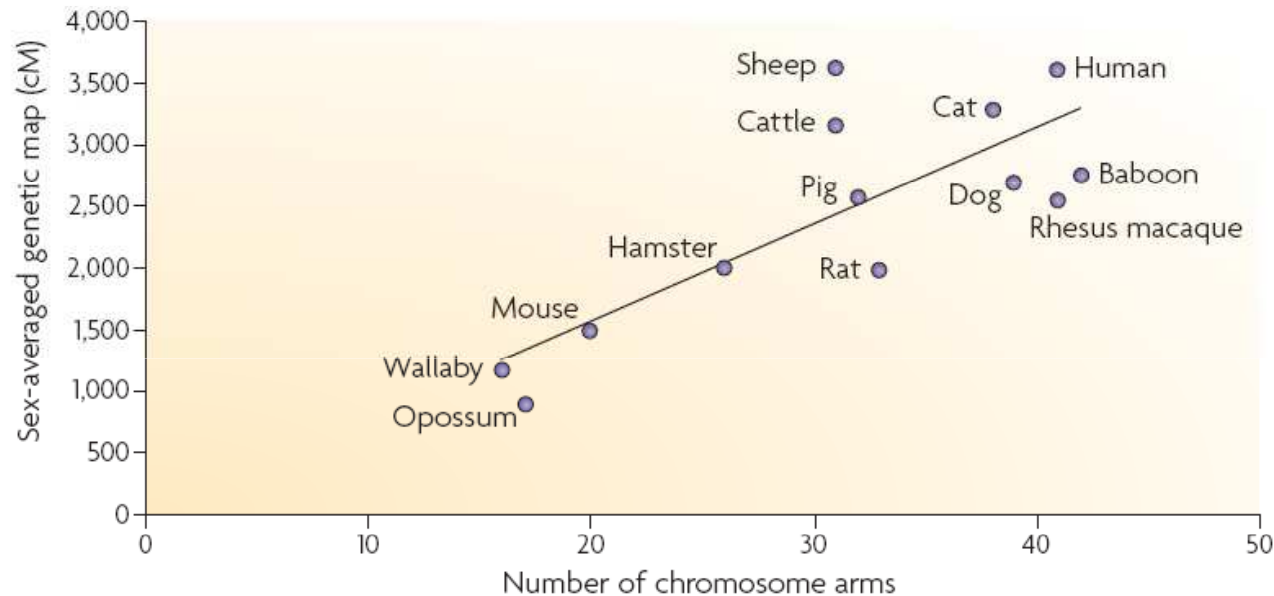
demise of 1 point often compensated by increased activity of a neighbour point

differences in recombination rate between sexes:

- **Haldane-Huxley rule**: if one sex doesn't recombine, it is the heterogametic sex
- if both sexes recombine, mostly in females > recombinations (man 1,7x, mouse 1,3x)

differences between species:

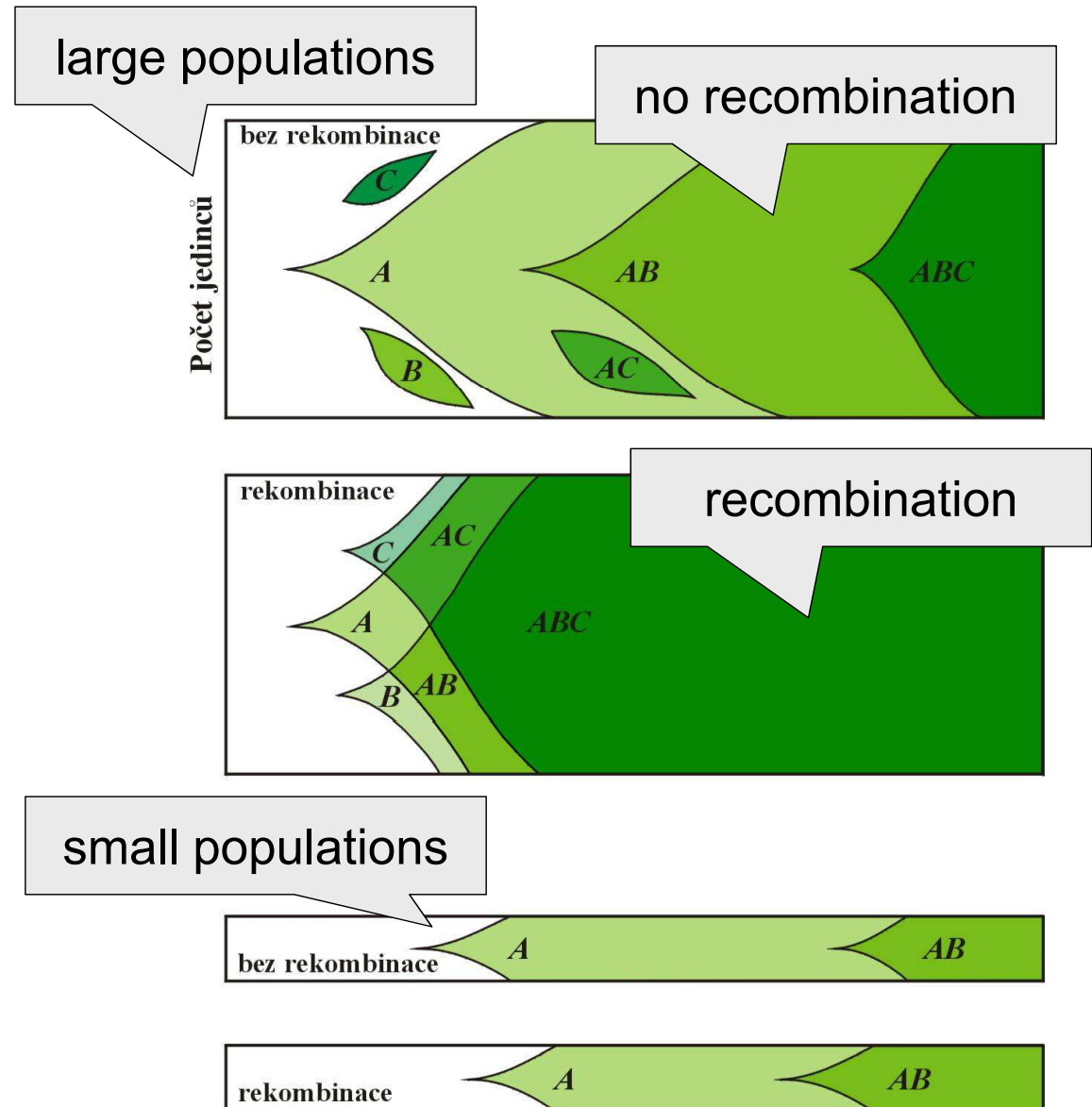
- species with more small chromosomes → more recombinations than species with less large chromosomes
- correlation with the number of arms: more recombinations in karyotypes with large numbers of chrom. arms (at least 1 c-o/arm to avoid aneuploidies?)



EVOLUTIONARY CONSEQUENCES OF RECOMBINATION:

Recombination and polymorphism:

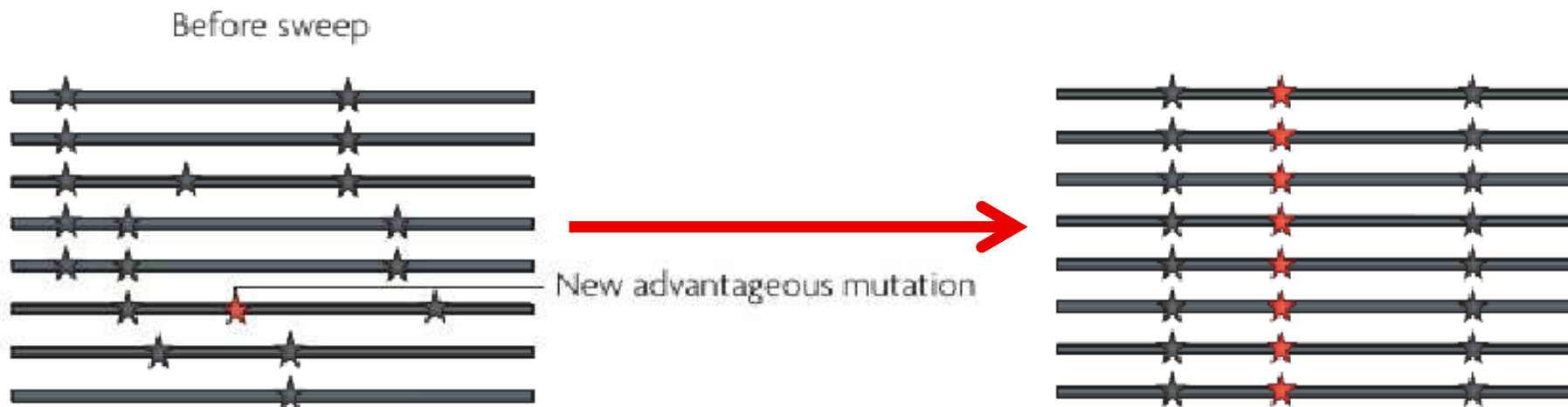
absence of recombination
⇒ linkage disequilibrium



EVOLUTIONARY CONSEQUENCES OF RECOMBINATION:

Recombination and polymorphism:

pozitivní selekce: *hitchhiking (draft)* → *selective sweep* (selekční smetení)

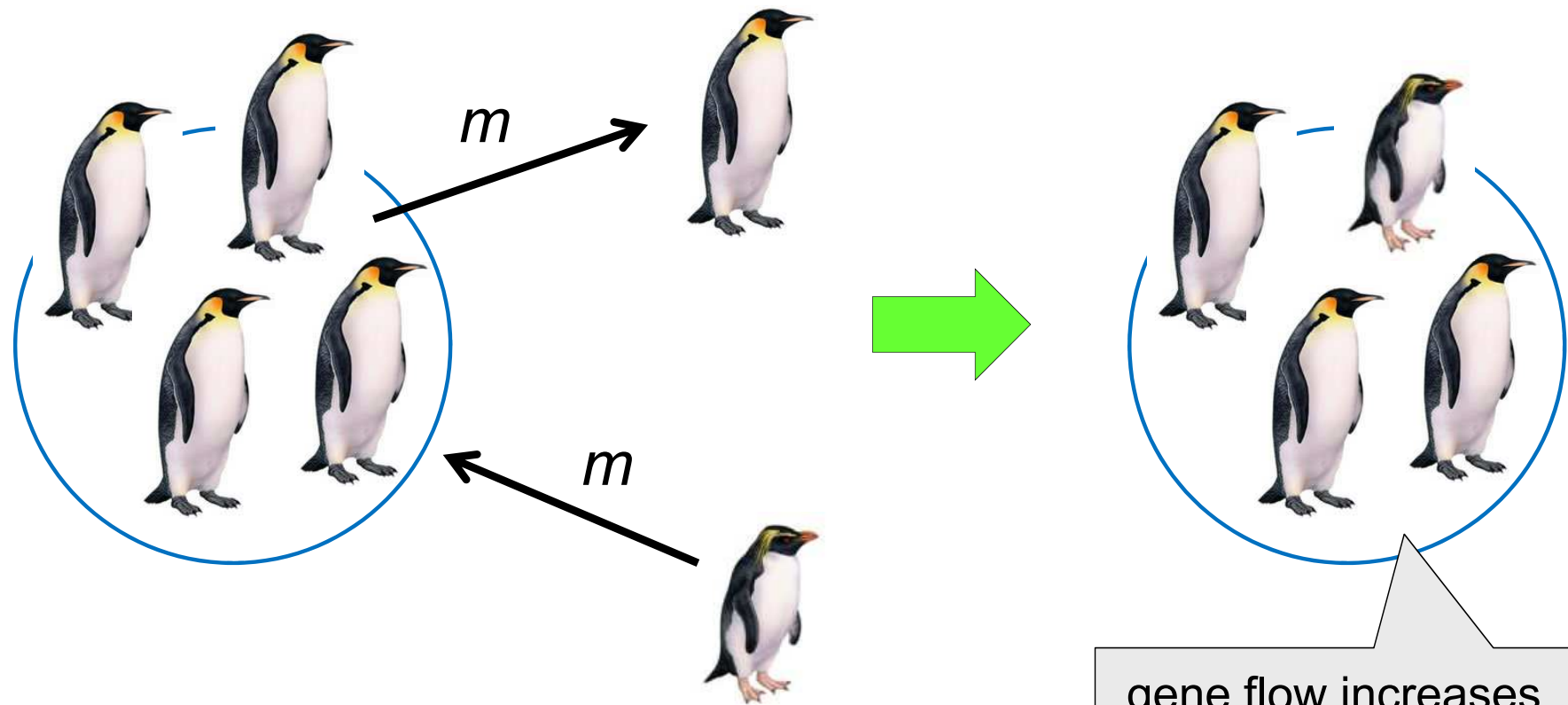


negativní selekce: *background selection*

→ **loss of polymorphism**

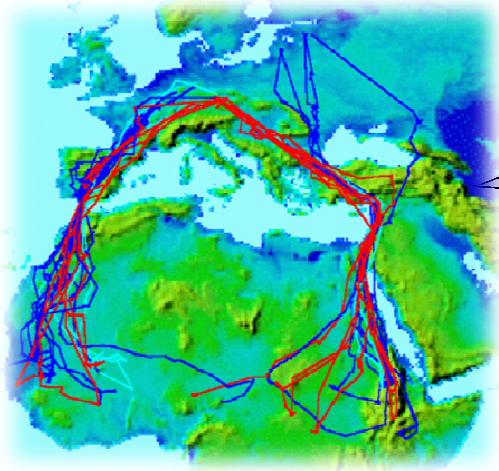
MIGRATION (GENE FLOW)

Migration rate, m = proportion of gene copies appearing in the population by immigration from other populations in the given generation



gene flow increases variation in the deme

MIGRATION (GENE FLOW)



long-distance
migration but no
gene flow



gene flow but no
migration

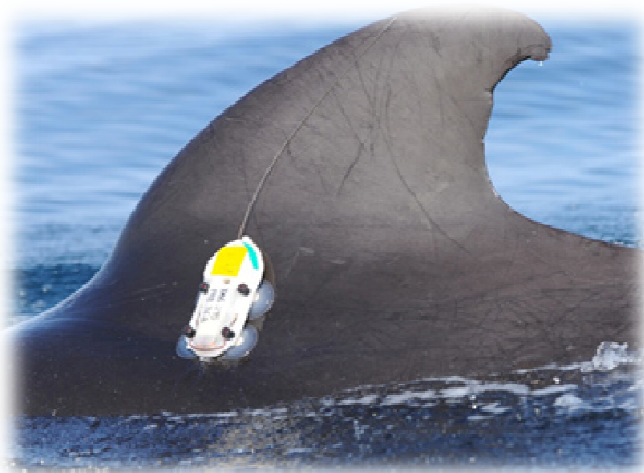


METHODS OF GENE FLOW ESTIMATION:

1. direct

capture-mark-recapture (CMR)

finger clipping, special dyes, tattooing, tags, rings, collars, genetic marking

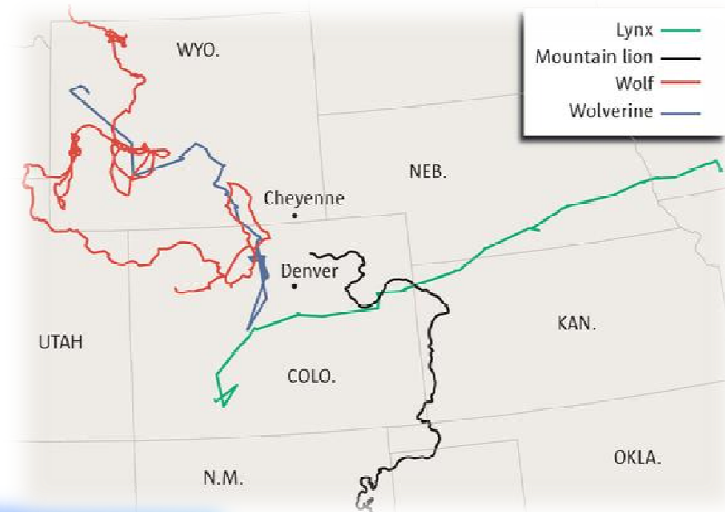


1. Direct methods

remote tracking – telemetry

transmitters, antennas; GPS systems

... more expensive, time consuming



Risk of underestimation of gene flow!!

2. Indirect methods

molecular markers

gene flow models

maximum likelihood and Bayesian programs

dispersal: distance between parents and offspring

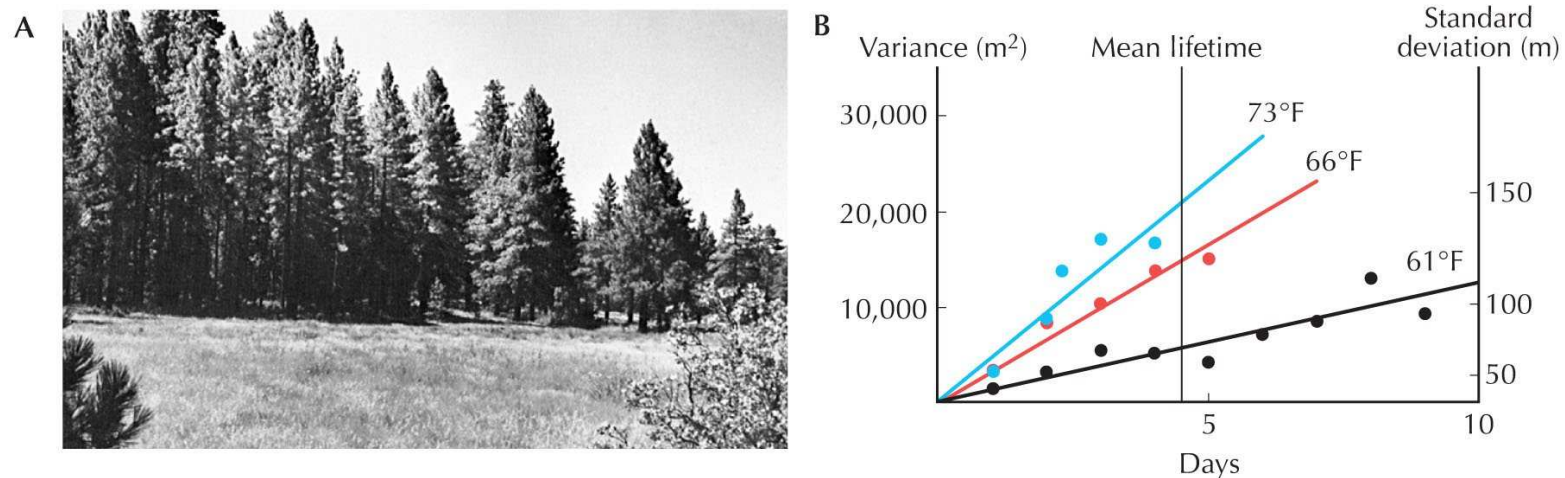
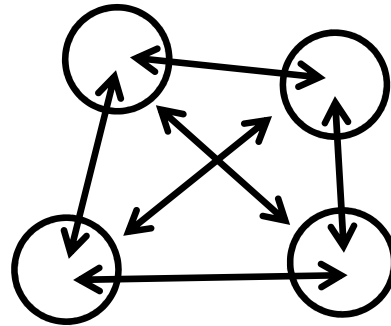


FIGURE 16.4. Dobzhansky and Wright (1943) measured the rate of dispersal of *Drosophila pseudoobscura* by releasing marked flies at sites in the Sierra Nevada, California (A). Over the following days, flies were caught in a series of traps. The graphs (B) show how the variance of the distribution of marked flies increased over time. The three sets of points show results from experiments at different times during the summer: Rates of movement increase strongly with temperature. The rate of diffusion of genes is estimated by assuming a mean lifetime of 4.5 days (vertical line).

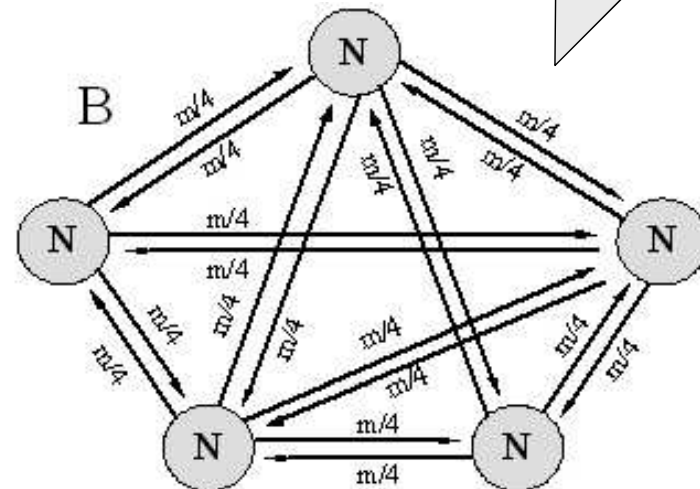
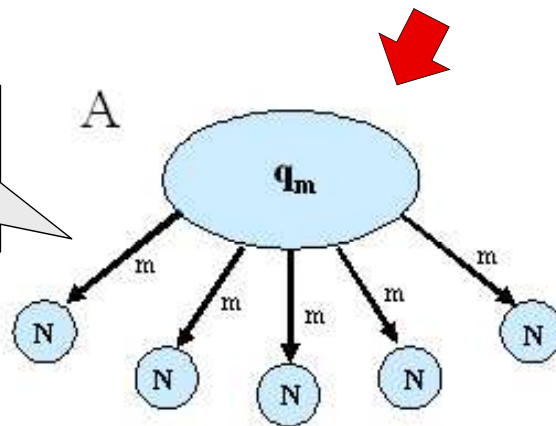
A) Island model



S. Wright (F-statistics): $F_{ST} = 1/(4Nm + 1) \Rightarrow Nm = (1/F_{ST} - 1)/4$
 ... Nm = number of migrants per generation

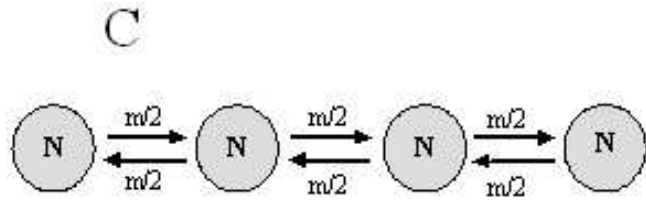
Island model can be also asymmetric:

continent-island model

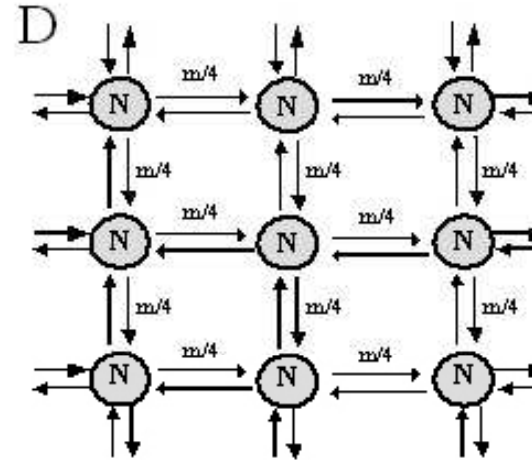


island model

B) Isolation by distance models discontinuous = stepping stone model



1D stepping-stone model



2D stepping-stone model



B) Isolation by distance models continuous

Linanthus parryae (Polemoniaceae), Mojave Desert (California)
T. Dobzhansky, Sewall Wright



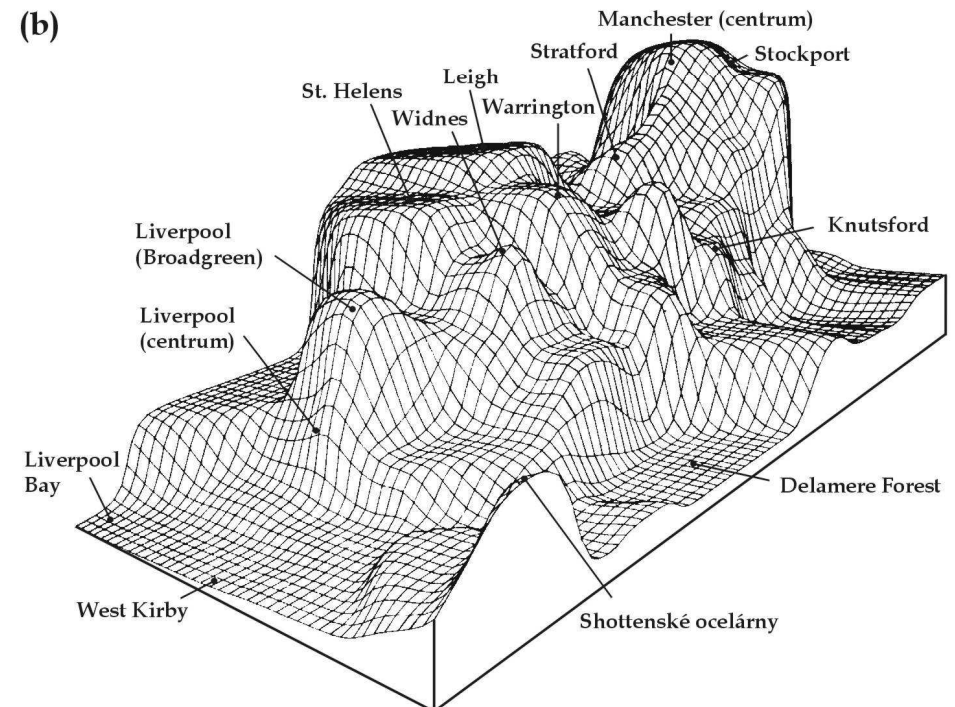
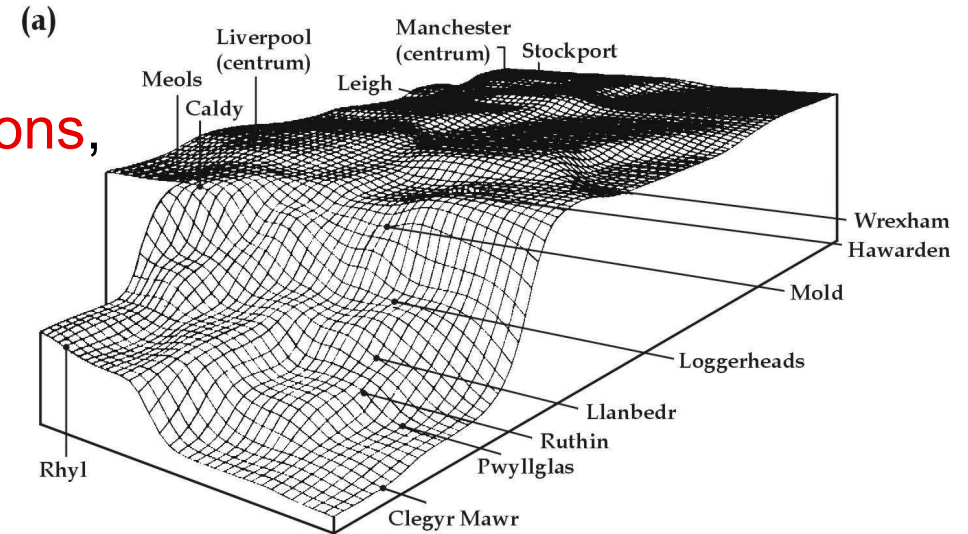
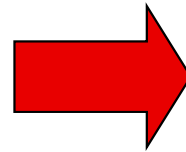
L. parryae

Gene flow consequences:

genetic homogenization of subpopulations,
preventing their genetic divergence

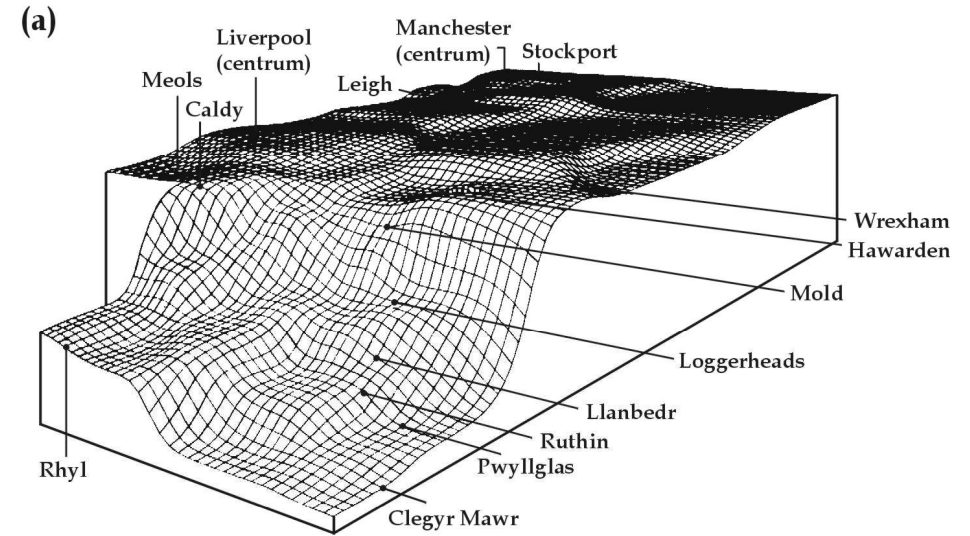
in many species migration severely
reduced

Eg.: melanic forms of moths
in England

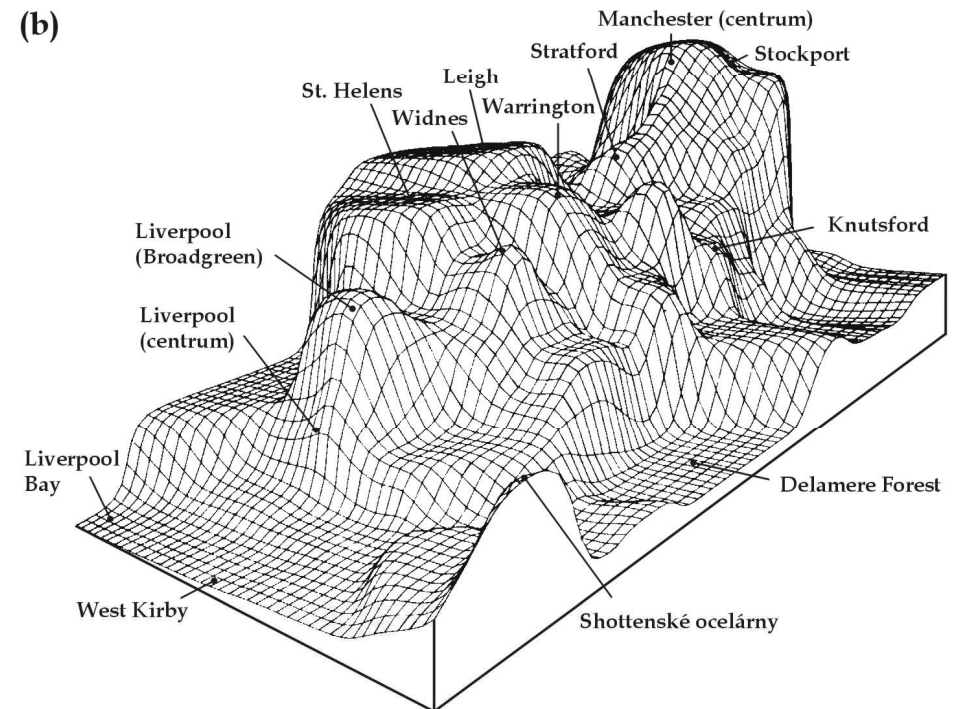




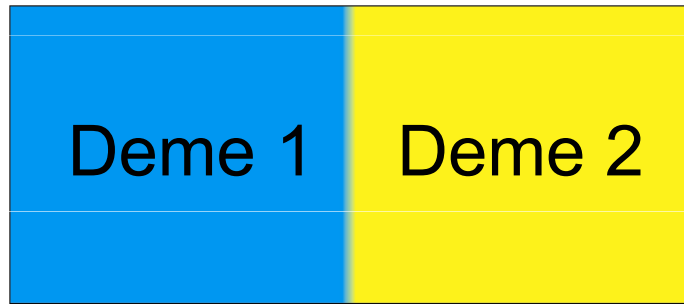
peppered moth (*Biston betularia*)



scalloped hazel
(*Odontoptera [Gonodontis] bidentata*)

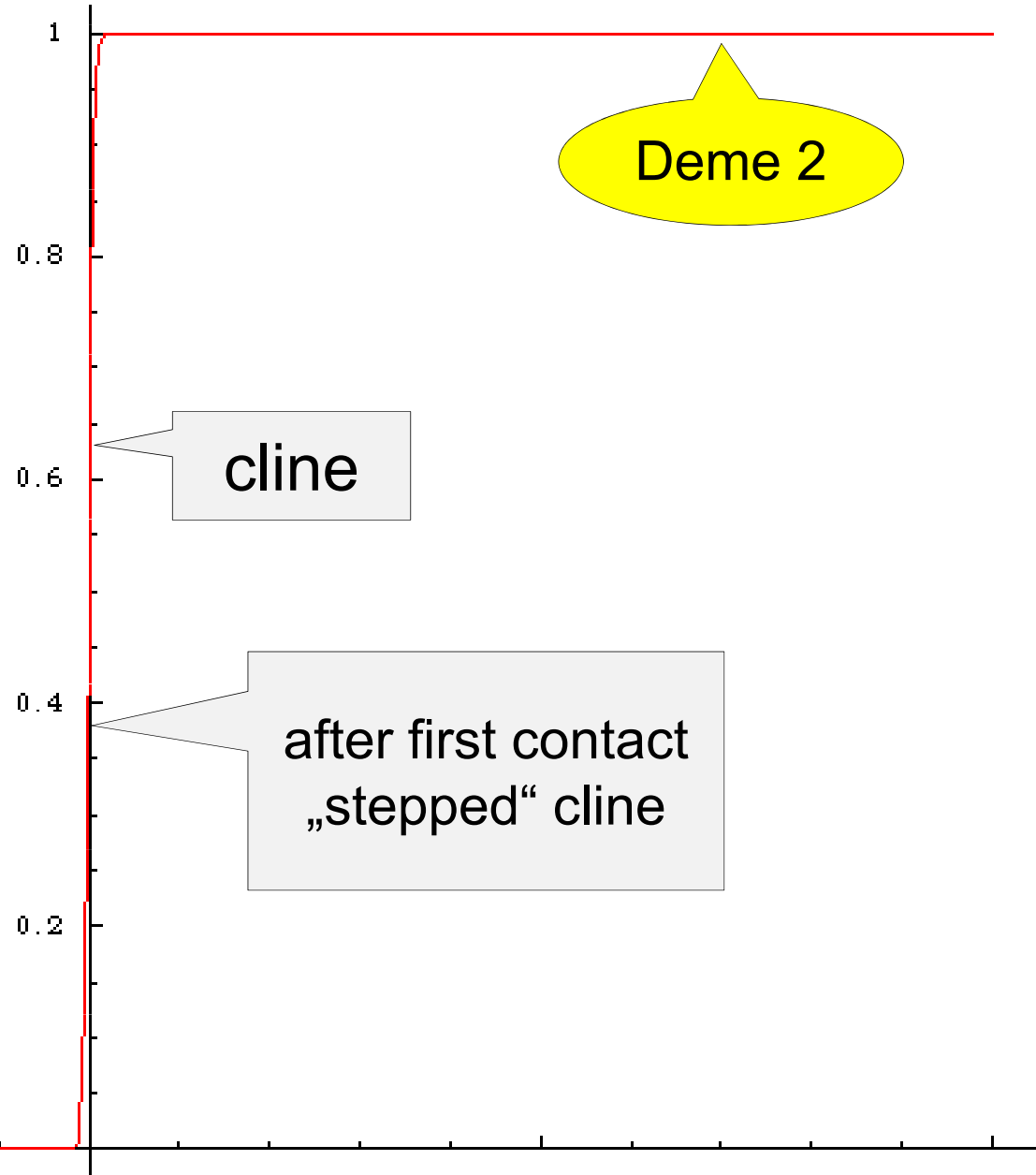


Diffusion of neutral alleles due to gene flow between demes



-100 -50 50 100

Geographic scale

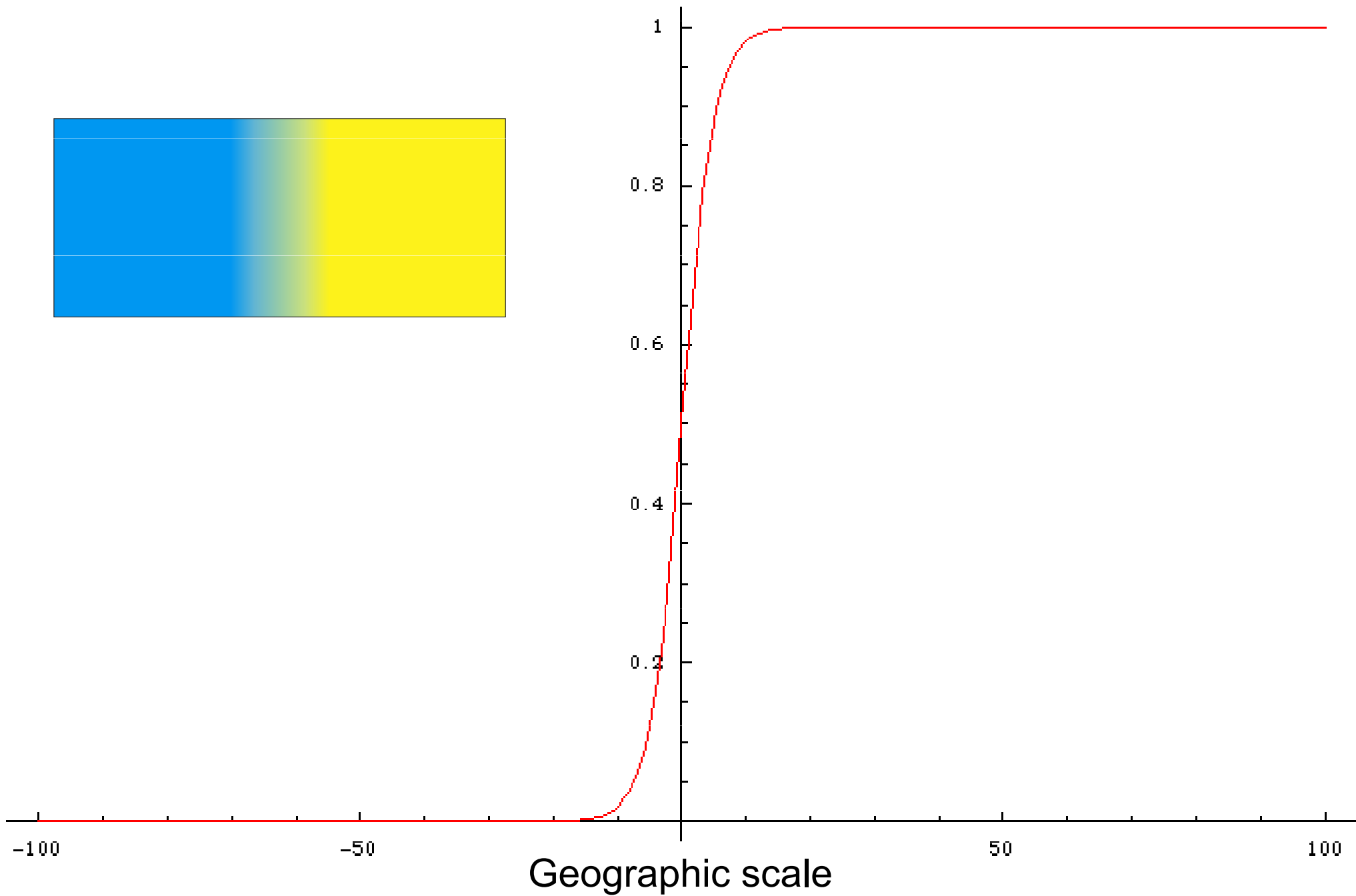
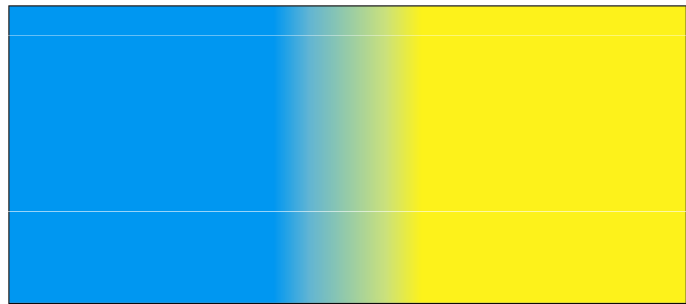


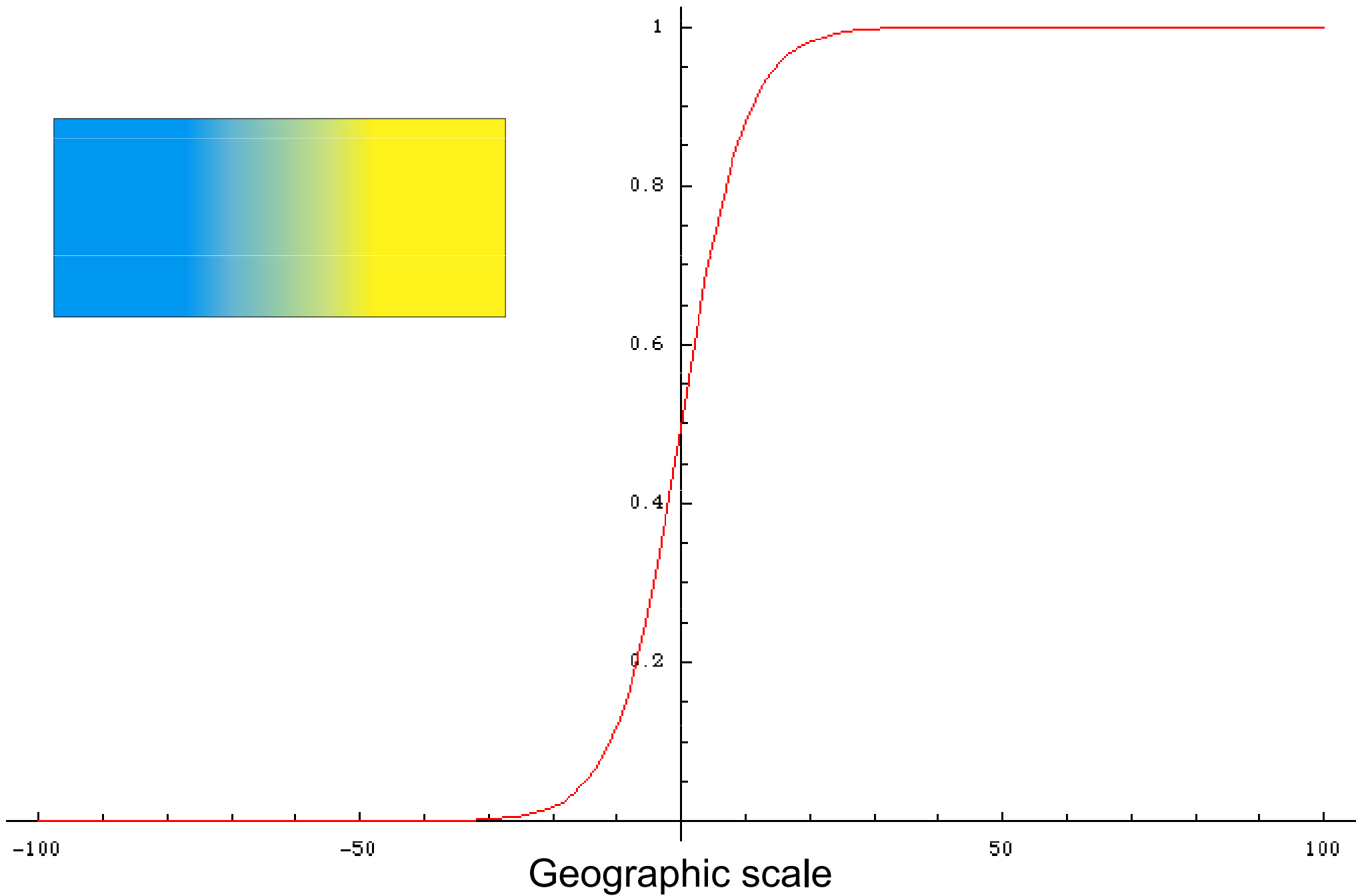
Deme 1

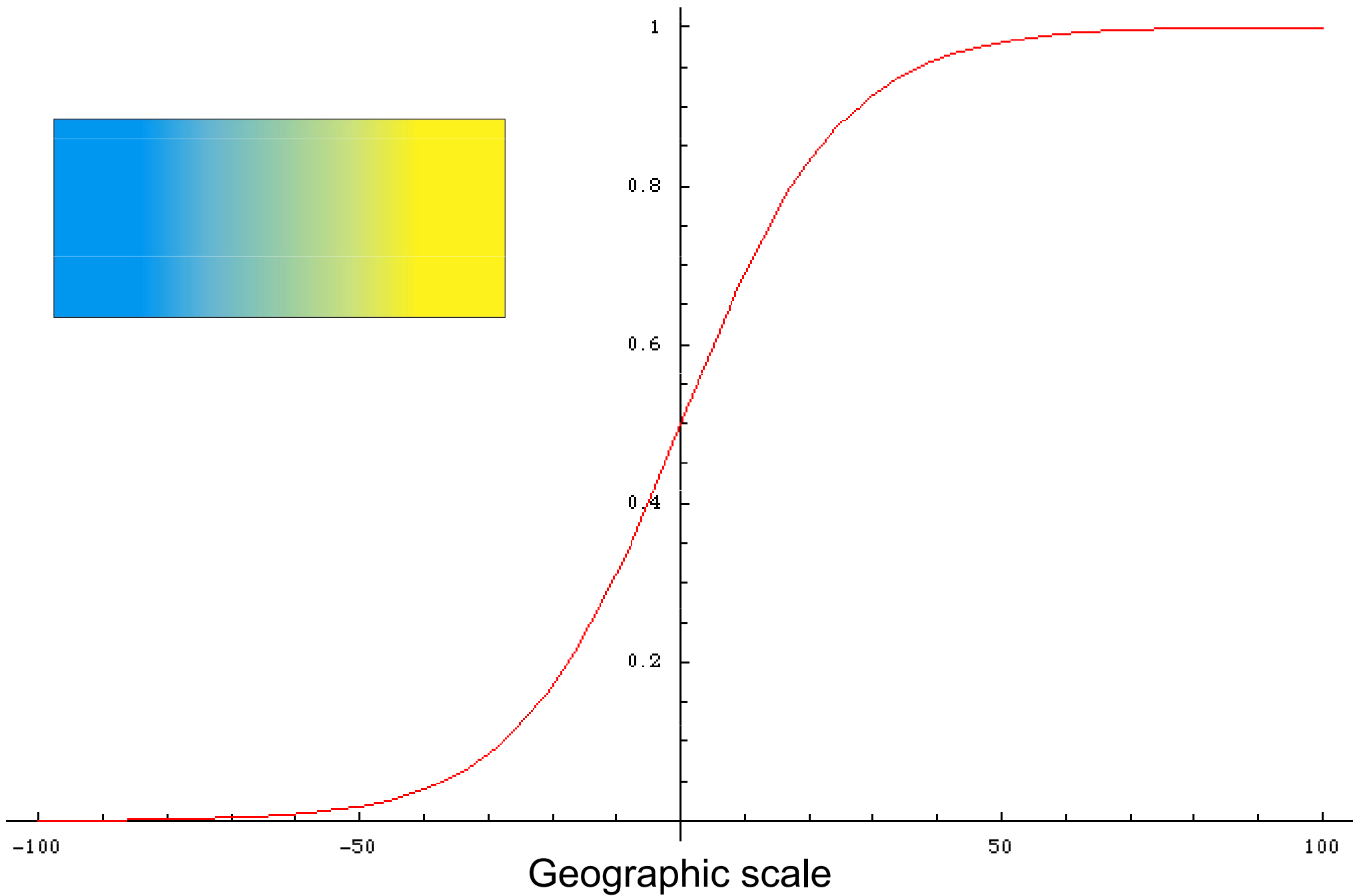
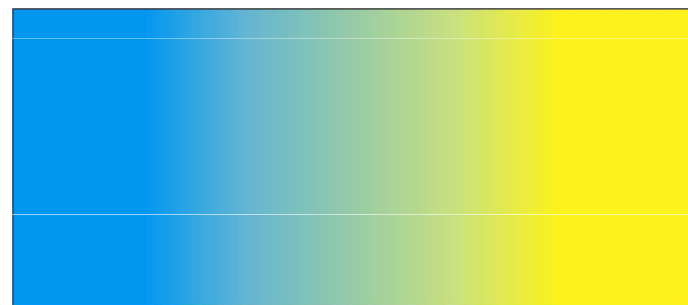
Deme 2

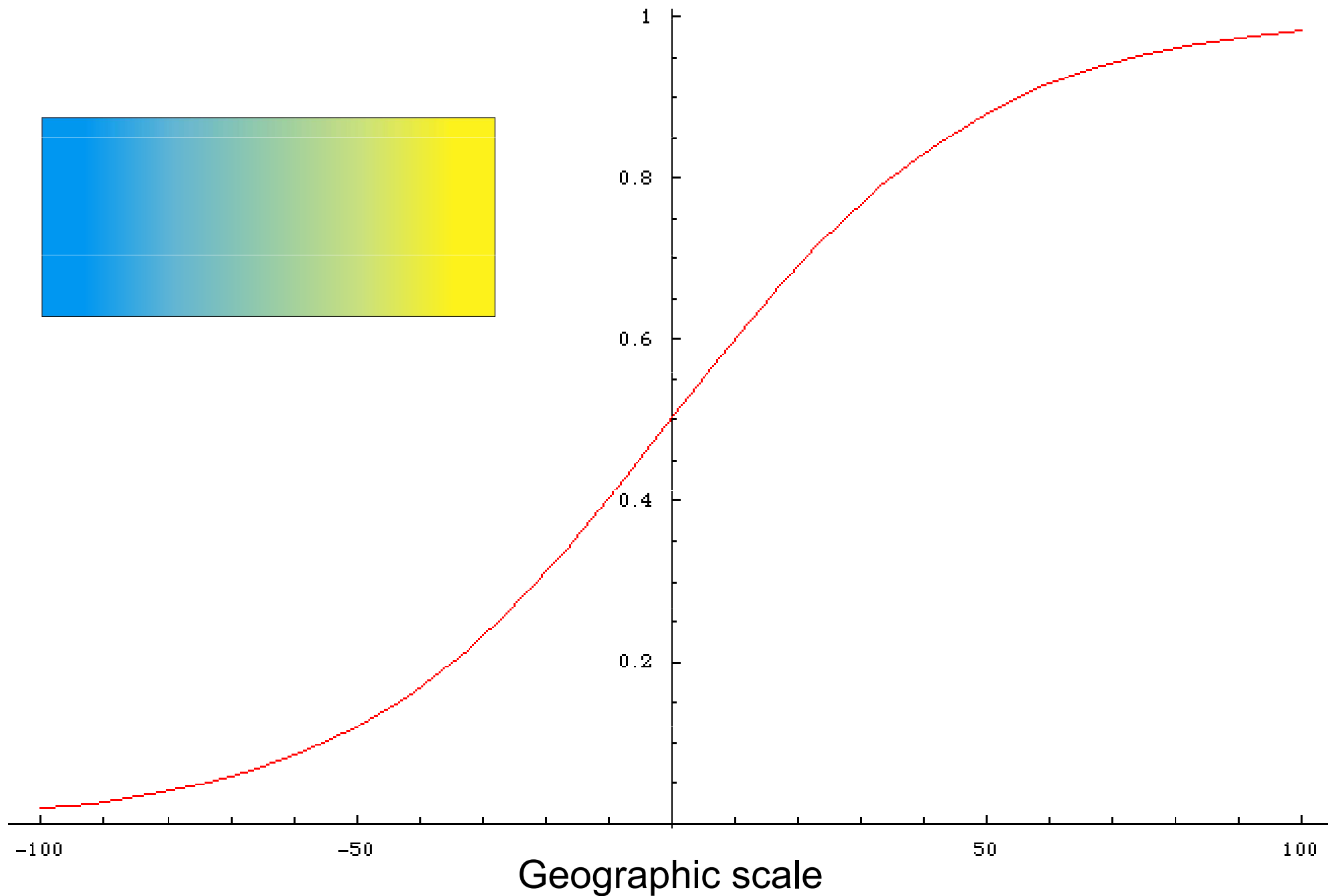
cline

after first contact
„stepped“ cline

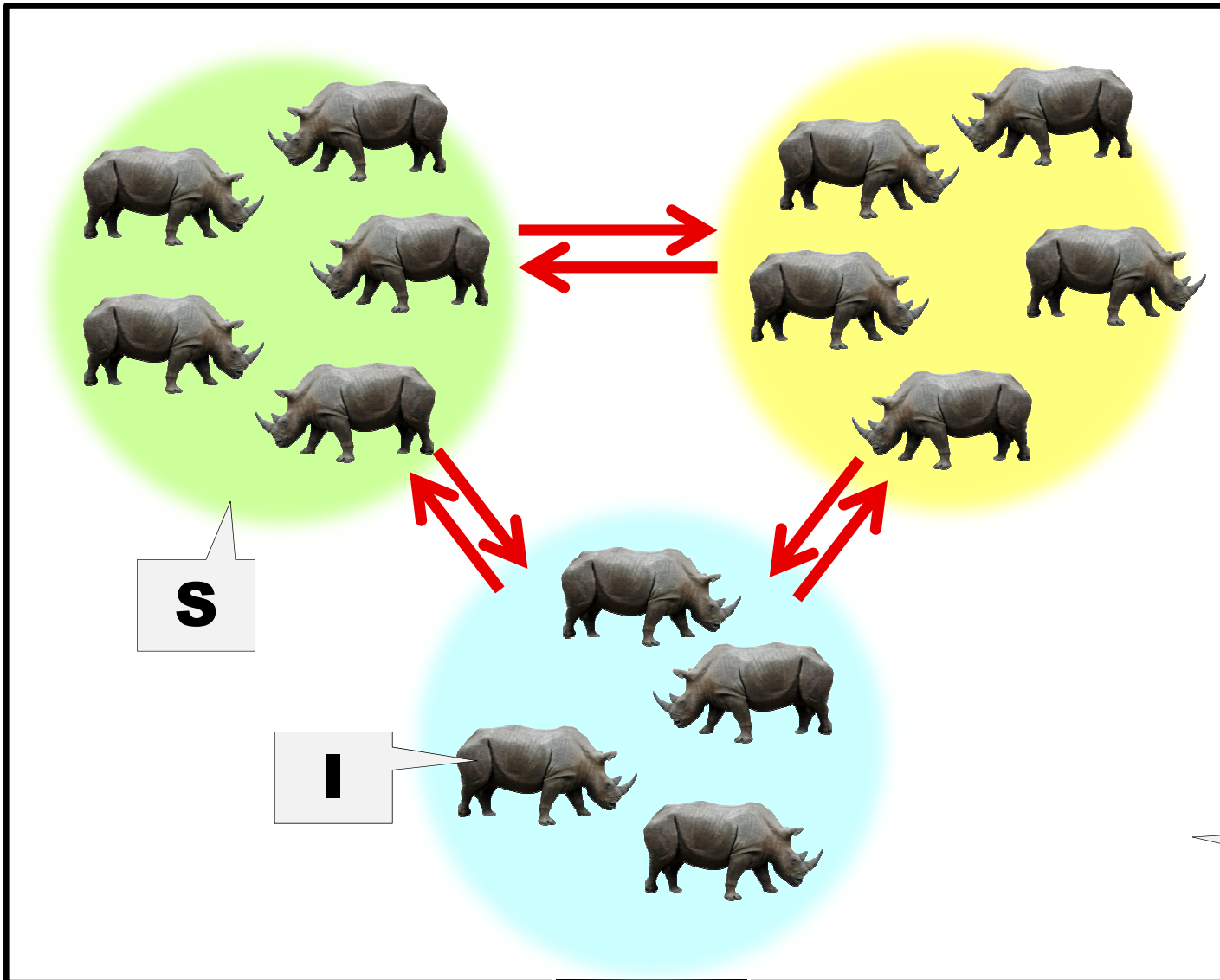








Sewall Wright - F-statistics:



F_{IS} (= inbreeding coefficient)

→ reduction of HZ in a subpopulation due to inbreeding

$$F_{IS} = (H_S - H_I)/H_S \quad -1 \leq F_{IS} \leq +1$$



F_{ST} (= fixation index) → reduction of HZ due to population substructuring

$$F_{ST} = (H_T - H_S)/H_T \quad 0 \leq F_{ST} \leq +1$$

F_{IT} → reduction of HZ both due to population substructuring and inbreeding

$$F_{IT} = (H_T - H_I)/H_T \quad (1 - F_{IS})(1 - F_{ST}) = 1 - F_{IT}$$