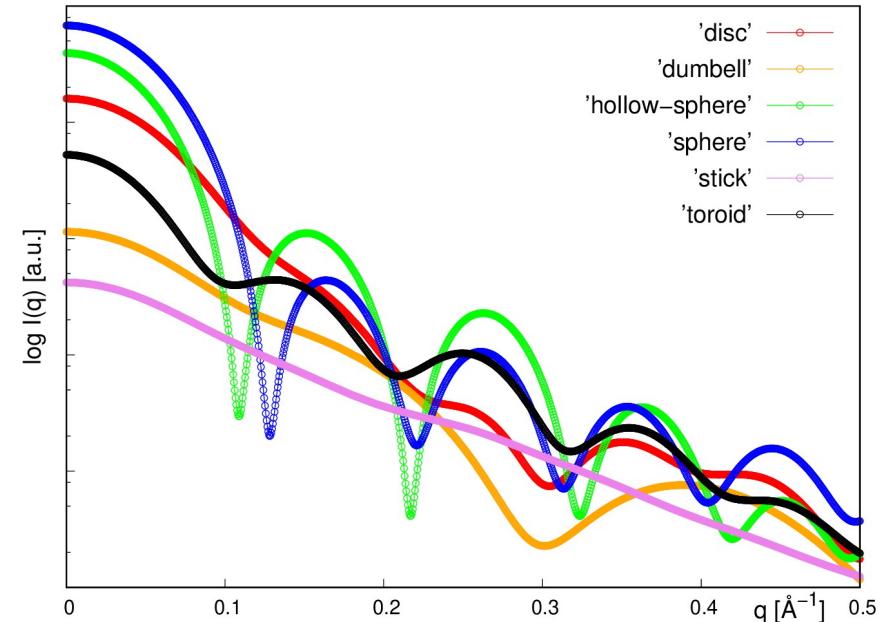
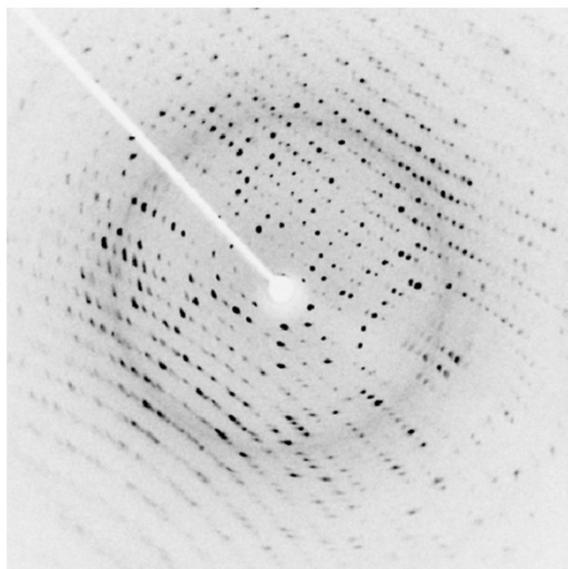
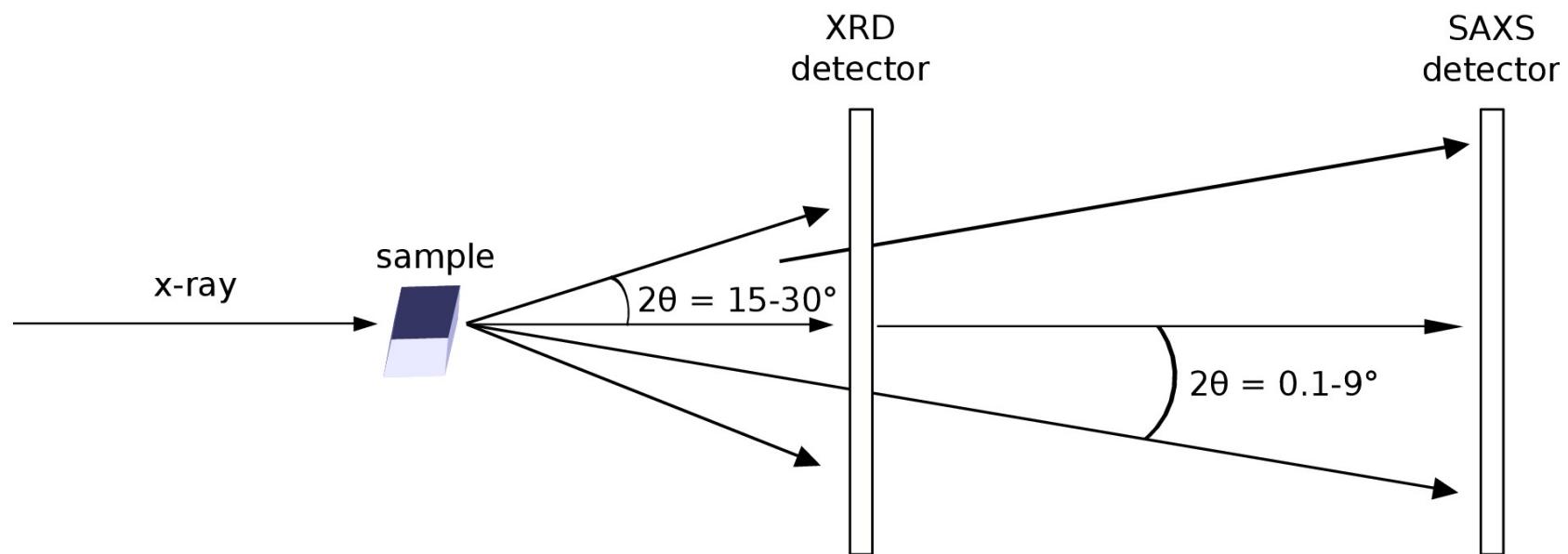


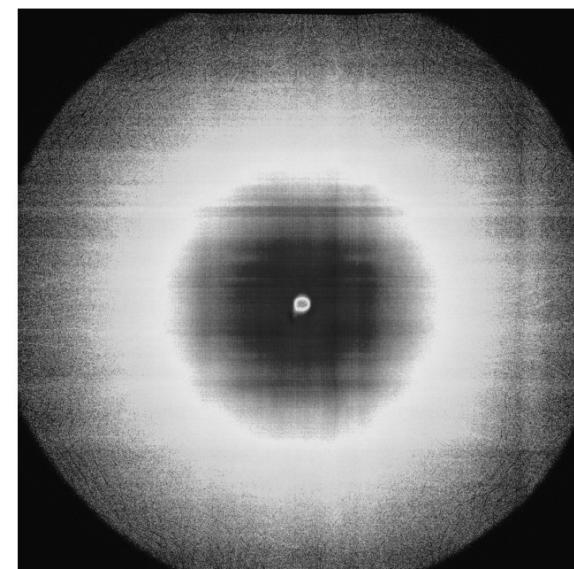
- size ( $R_g$ ,  $D_{max}$ , volume)
- folding state, compactness
- overall shape  $\sim 30 \text{ \AA}$  resolution
  
- oligomeric state
- quaternary structure
- flexible systems



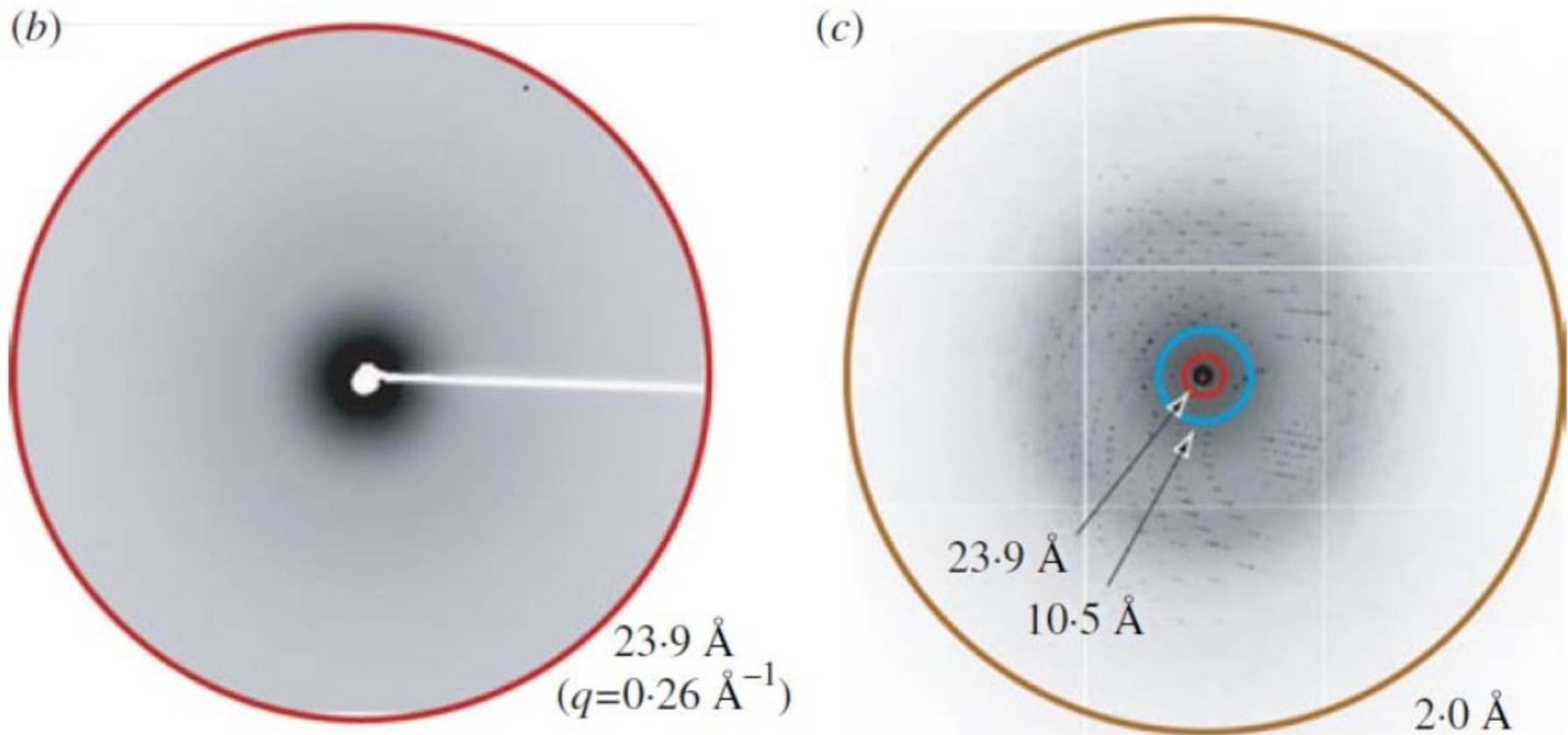
# Experimental setup XRD vs SAXS



XRD detector image  
diffractioin spots

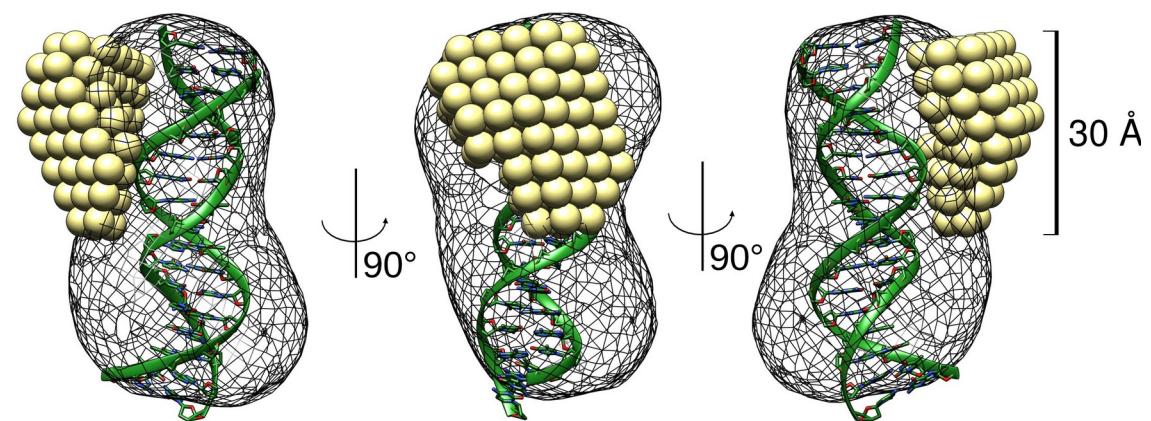
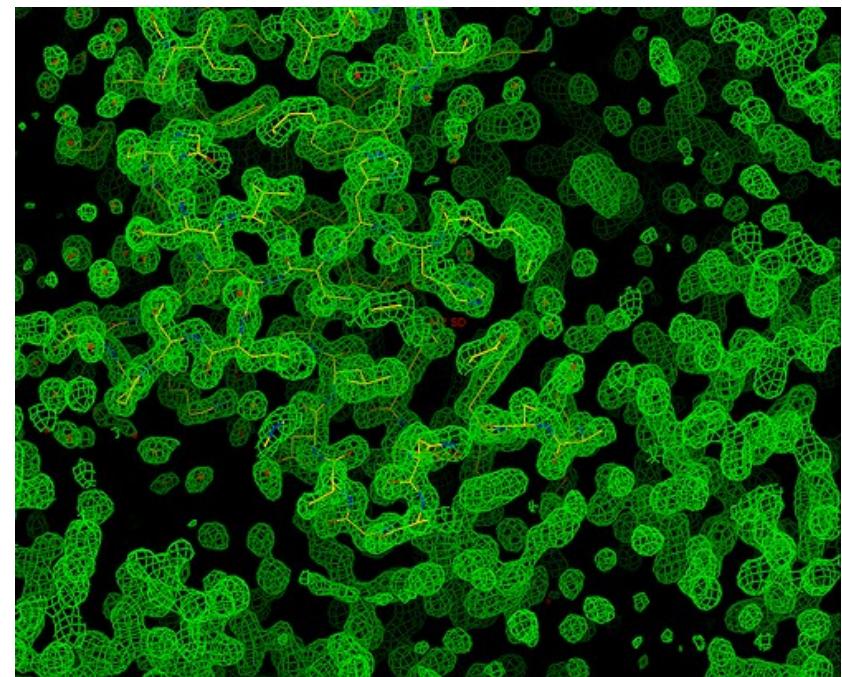


SAXS detector image  
x-ray scattering



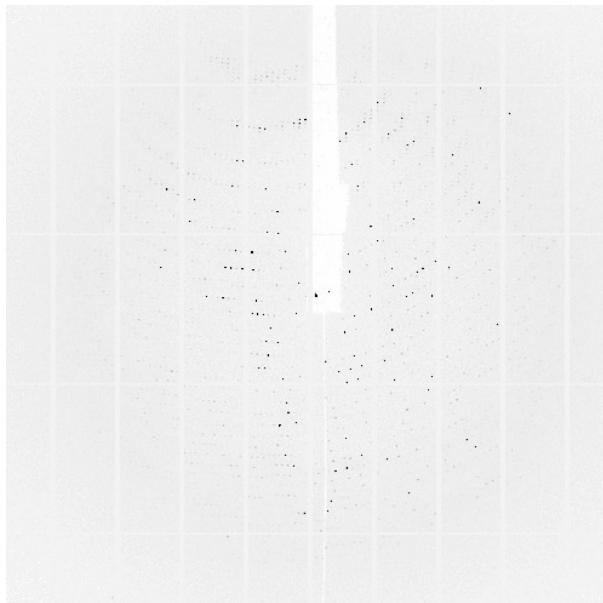
Putnam et al. (2007) *Quart. Rev. Biophys.* **40**(3), 191-285.

# Output structure XRD vs SAXS

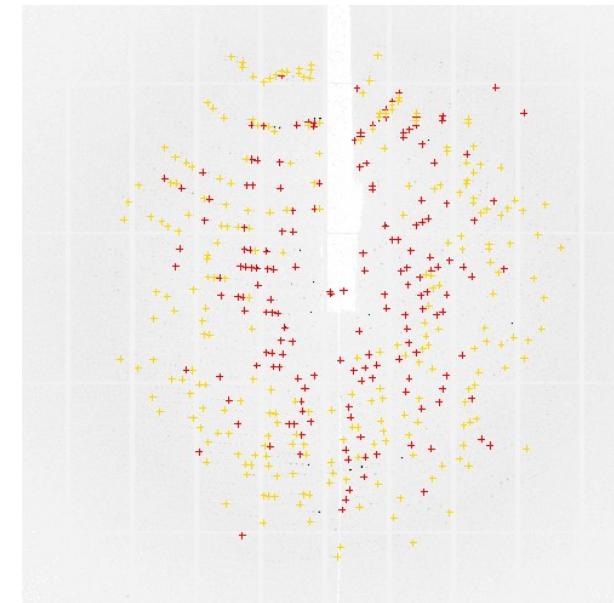


# XRD – data reduction

Set of diffraction images



indexing



Reduced data – intensities with h, k, l indices

	h	k	l	Orig. H	Orig. K	Orig. L	M/SYM	BATCH	I	SIGI	FRACTIONCALC	XDET	YDET	ROT	LP	FLAG	
1	0	0	5	0	0	5	5	1	706	0,45	0,34	1.00	1248.00	1285.20	70.50	0.01	0
2	0	0	6	0	0	6	6	1	710	11780.00	315.60	1.00	1260.90	1281.00	70.93	0.01	0
3	0	0	7	0	0	7	7	1	714	-0,17	0,52	1.00	1273.90	1276.90	71.36	0.01	0
4	0	0	13	0	0	13	13	1	740	0,54	0,98	1.00	1351.80	1252.00	73.93	0.02	0
5	0	0	14	0	0	14	14	1	744	1,26	1,08	1.00	1364.80	1247.90	74.35	0.02	0
6	0	0	15	0	0	15	15	1	748	3,08	1,15	1.00	1377.80	1243.80	74.78	0.02	0
7	0	0	16	0	0	16	16	1	753	-0,63	1,18	1.00	1390.90	1239.70	75.21	0.02	0
8	0	0	17	0	0	17	17	1	757	-0,12	1,25	1.00	1403.90	1235.60	75.64	0.03	0
9	0	0	18	0	0	18	18	1	761	157.10	5,16	1.00	1417.00	1231.50	76.07	0.03	0
10	0	0	19	0	0	19	19	1	766	1,74	1,48	1.00	1430.10	1227.50	76.51	0.03	0
11	0	0	20	0	0	20	20	1	770	3,40	1,58	1.00	1443.20	1223.40	76.94	0.03	0
12	0	0	21	0	0	21	21	1	774	2,61	1,65	1.00	1456.30	1219.30	77.37	0.03	0
13	0	0	21	0	0	-21	2	2	595	-0,20	1,56	0,79	910.30	1393.30	59.46	0.03	0
14	0	0	22	0	0	22	22	1	779	13,00	2,00	1.00	1469.50	1215.30	77.80	0.03	0
15	0	0	22	0	0	-22	2	2	591	10,20	1,68	1.00	897.20	1397.44	59.03	0.03	0
16	0	0	23	0	0	-23	23	2	587	6,14	1,68	1.00	884.00	1401.50	58.60	0.04	0
17	0	0	24	0	0	24	24	1	787	17610.00	472.70	1.00	1495.80	1207.20	78.67	0.04	0
18	0	0	24	0	0	-24	2	2	582	17080.00	458.50	1.00	870.90	1405.60	58.17	0.04	0
19	0	0	25	0	0	25	25	1	792	1,56	1,96	1.00	1509.10	1203.20	79.10	0.04	0
20	0	0	25	0	0	-25	2	2	578	-2,49	1,81	1.00	857.70	1409.60	57.73	0.04	0
21	0	0	26	0	0	26	26	1	796	1,20	2,02	0,89	1522.30	1199.20	79.54	0.04	0
22	0	0	26	0	0	-26	2	2	574	-1,04	1,93	1.00	844.40	1413.70	57.30	0.04	0
23	0	0	27	0	0	-27	27	2	569	-0,63	1,98	1.00	831.20	1417.70	56.87	0.04	0
24	0	0	28	0	0	-28	28	2	565	2,69	2,06	1.00	817.90	1421.80	56.43	0.04	0
25	0	0	29	0	0	-29	29	2	561	-0,08	2,09	1.00	804.70	1425.80	56.00	0.04	0
26	0	0	30	0	0	-30	30	2	556	3668.00	99.62	1.00	791.30	1429.80	55.56	0.05	0
27	0	0	31	0	0	-31	31	2	552	-0,04	2,20	1.00	778.00	1433.80	55.13	0.05	0
28	0	0	32	0	0	-32	32	2	547	0,58	2,33	1.00	764.60	1437.70	54.69	0.05	0

indexing  
integration



Space group	P 61 2 2
Space group confidence	S (spacegroup is known)
Cell	177.54 177.54 203.64 90 90 120
Resolution low	48.33
Resolution high	2.832
Number of lattices	1
Number of reflections	398868
Number of datasets	1

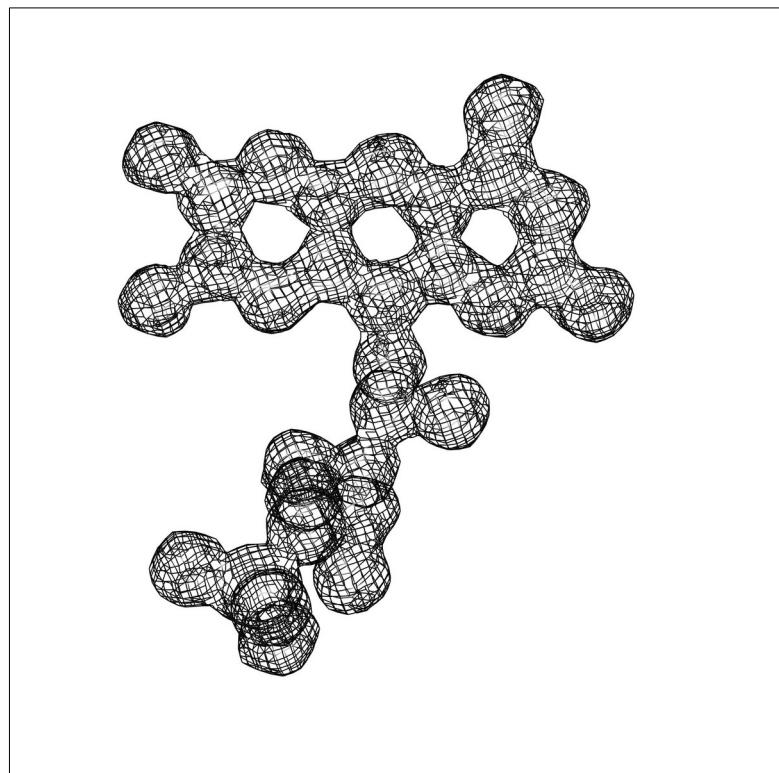
# XRD – outputs

xrd intensities ~ $10^5$  reflexions,  
reciprocal space, indices  $h, k, l$

	<b>h</b>	<b>k</b>	<b>l</b>	Orig. <b>H</b>	Orig. <b>K</b>	Orig. <b>L</b>	M/SYM	BATCH	I	SIGI	FRACTIONCALC	XDET	YDET	ROT	LP	FLAG	
1	0	0	5	0	0	5		1	706	0.45	0.34	1.00	1248.00	1285.20	70.50	0.01	0
2	0	0	6	0	0	6		1	710	11780.00	315.60	1.00	1260.90	1281.00	70.93	0.01	0
3	0	0	7	0	0	7		1	714	-0.17	0.52	1.00	1273.90	1276.90	71.36	0.01	0
4	0	0	13	0	0	13		1	740	0.54	0.98	1.00	1351.80	1252.00	73.93	0.02	0
5	0	0	14	0	0	14		1	744	1.26	1.08	1.00	1364.80	1247.90	74.35	0.02	0
6	0	0	15	0	0	15		1	748	3.08	1.15	1.00	1377.80	1243.80	74.78	0.02	0
7	0	0	16	0	0	16		1	753	-0.63	1.18	1.00	1390.90	1239.70	75.21	0.02	0
8	0	0	17	0	0	17		1	757	-0.12	1.25	1.00	1403.90	1235.60	75.64	0.03	0
9	0	0	18	0	0	18		1	761	157.10	5.16	1.00	1417.00	1231.50	76.07	0.03	0
10	0	0	19	0	0	19		1	766	1.74	1.48	1.00	1430.10	1227.50	76.51	0.03	0
11	0	0	20	0	0	20		1	770	3.40	1.58	1.00	1443.20	1223.40	76.94	0.03	0
12	0	0	21	0	0	21		1	774	2.61	1.65	1.00	1456.30	1219.30	77.37	0.03	0
13	0	0	21	0	0	-21		2	595	-0.20	1.56	0.79	910.30	1393.30	59.46	0.03	0
14	0	0	22	0	0	22		1	779	13.00	2.00	1.00	1469.50	1215.30	77.80	0.03	0
15	0	0	22	0	0	-22		2	591	10.20	1.68	1.00	897.20	1397.40	59.03	0.03	0
16	0	0	23	0	0	-23		2	587	6.14	1.68	1.00	884.00	1401.50	58.60	0.04	0
17	0	0	24	0	0	24		1	787	17610.00	472.70	1.00	1495.80	1207.20	78.67	0.04	0
18	0	0	24	0	0	-24		2	582	17080.00	458.50	1.00	870.90	1405.60	58.17	0.04	0
19	0	0	25	0	0	25		1	792	1.56	1.96	1.00	1509.10	1203.20	79.10	0.04	0
20	0	0	25	0	0	-25		2	578	-2.49	1.81	1.00	857.70	1409.60	57.73	0.04	0
21	0	0	26	0	0	26		1	796	1.20	2.02	0.89	1522.30	1199.20	79.54	0.04	0
22	0	0	26	0	0	-26		2	574	-1.04	1.93	1.00	844.40	1413.70	57.30	0.04	0
23	0	0	27	0	0	-27		2	569	-0.63	1.98	1.00	831.20	1417.70	56.87	0.04	0
24	0	0	28	0	0	-28		2	565	2.69	2.06	1.00	817.90	1421.80	56.43	0.04	0
25	0	0	29	0	0	-29		2	561	-0.08	2.09	1.00	804.70	1425.80	56.00	0.04	0
26	0	0	30	0	0	-30		2	556	3668.00	99.62	1.00	791.30	1429.80	55.56	0.05	0
27	0	0	31	0	0	-31		2	552	-0.04	2.20	1.00	778.00	1433.80	55.13	0.05	0
28	0	0	32	0	0	-32		2	547	0.58	2.33	1.00	764.60	1437.70	54.69	0.05	0
29	0	0	33	0	0	-33		2	543	0.79	2.38	1.00	751.20	1441.70	54.25	0.05	0
30	0	0	34	0	0	-34		2	539	2.15	2.44	1.00	737.80	1445.70	53.82	0.05	0
31	0	0	35	0	0	-35		2	534	1.52	2.54	1.00	724.30	1449.60	53.38	0.05	0
32	0	0	36	0	0	-36		2	530	531.30	16.14	1.00	710.90	1453.50	52.94	0.06	0
33	0	0	37	0	0	-37		2	526	0.98	2.68	1.00	697.30	1457.40	52.50	0.06	0
34	0	0	38	0	0	-38		2	521	-0.53	2.85	1.00	683.80	1461.30	52.05	0.06	0
35	0	0	39	0	0	-39		2	517	-4.92	3.00	0.95	670.20	1465.10	51.61	0.06	0
36	0	0	45	0	0	-45		2	490	2.37	3.73	1.00	587.80	1487.90	48.93	0.07	0

Phase problem → Fourier transform

Electron density map



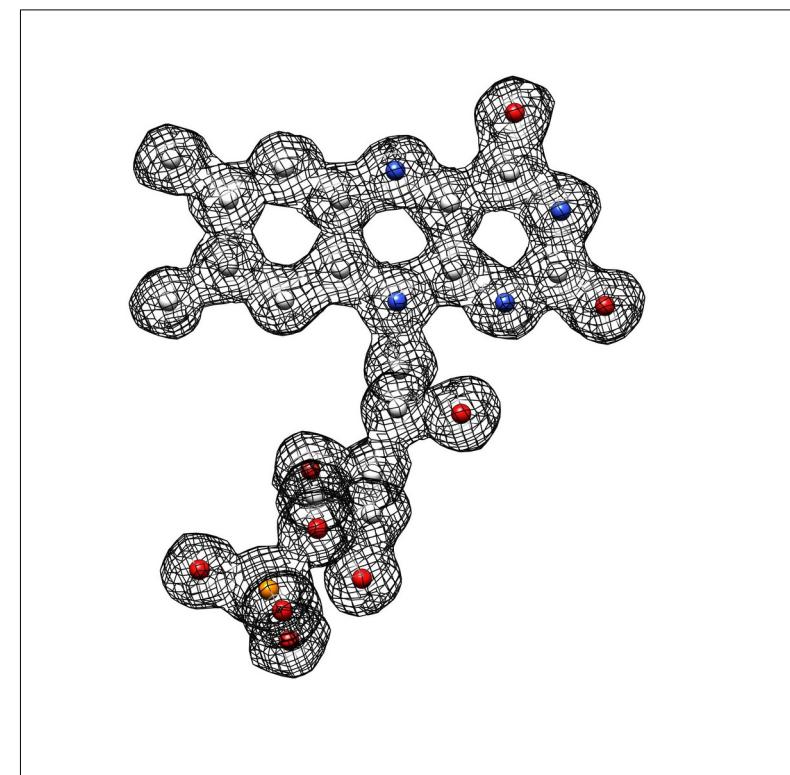
# XRD – outputs

xrd intensities ~ $10^5$  reflexions,  
reciprocal space, indices  $h, k, l$

	$h$	$k$	$l$	Orig. H	Orig. K	Orig. L	M/SYM	BATCH	I	SIGI	FRACTIONCALC	XDET	YDET	ROT	LP	FLAG	
1	0	0	5	0	0	5		1	706	0.45	0.34	1.00	1248.00	1285.20	70.50	0.01	0
2	0	0	6	0	0	6		1	710	11780.00	315.60	1.00	1260.90	1281.00	70.93	0.01	0
3	0	0	7	0	0	7		1	714	-0.17	0.52	1.00	1273.90	1276.90	71.36	0.01	0
4	0	0	13	0	0	13		1	740	0.54	0.98	1.00	1351.80	1252.00	73.93	0.02	0
5	0	0	14	0	0	14		1	744	1.26	1.08	1.00	1364.80	1247.90	74.35	0.02	0
6	0	0	15	0	0	15		1	748	3.08	1.15	1.00	1377.80	1243.80	74.78	0.02	0
7	0	0	16	0	0	16		1	753	-0.63	1.18	1.00	1390.90	1239.70	75.21	0.02	0
8	0	0	17	0	0	17		1	757	-0.12	1.25	1.00	1403.90	1235.60	75.64	0.03	0
9	0	0	18	0	0	18		1	761	157.10	5.16	1.00	1417.00	1231.50	76.07	0.03	0
10	0	0	19	0	0	19		1	766	1.74	1.48	1.00	1430.10	1227.50	76.51	0.03	0
11	0	0	20	0	0	20		1	770	3.40	1.58	1.00	1443.20	1223.40	76.94	0.03	0
12	0	0	21	0	0	21		1	774	2.61	1.65	1.00	1456.30	1219.30	77.37	0.03	0
13	0	0	21	0	0	-21		2	595	-0.20	1.56	0.79	910.30	1393.30	59.46	0.03	0
14	0	0	22	0	0	22		1	779	13.00	2.00	1.00	1469.50	1215.30	77.80	0.03	0
15	0	0	22	0	0	-22		2	591	10.20	1.68	1.00	897.20	1397.40	59.03	0.03	0
16	0	0	23	0	0	-23		2	587	6.14	1.68	1.00	884.00	1401.50	58.60	0.04	0
17	0	0	24	0	0	24		1	787	17610.00	472.70	1.00	1495.80	1207.20	78.67	0.04	0
18	0	0	24	0	0	-24		2	582	17080.00	458.50	1.00	870.90	1405.60	58.17	0.04	0
19	0	0	25	0	0	25		1	792	1.56	1.96	1.00	1509.10	1203.20	79.10	0.04	0
20	0	0	25	0	0	-25		2	578	-2.49	1.81	1.00	857.70	1409.60	57.73	0.04	0
21	0	0	26	0	0	26		1	796	1.20	2.02	0.89	1522.30	1199.20	79.54	0.04	0
22	0	0	26	0	0	-26		2	574	-1.04	1.93	1.00	844.40	1413.70	57.30	0.04	0
23	0	0	27	0	0	-27		2	569	-0.63	1.98	1.00	831.20	1417.70	56.87	0.04	0
24	0	0	28	0	0	-28		2	565	2.69	2.06	1.00	817.90	1421.80	56.43	0.04	0
25	0	0	29	0	0	-29		2	561	-0.08	2.09	1.00	804.70	1425.80	56.00	0.04	0
26	0	0	30	0	0	-30		2	556	3668.00	99.62	1.00	791.30	1429.80	55.56	0.05	0
27	0	0	31	0	0	-31		2	552	-0.04	2.20	1.00	778.00	1433.80	55.13	0.05	0
28	0	0	32	0	0	-32		2	547	0.58	2.33	1.00	764.60	1437.70	54.69	0.05	0
29	0	0	33	0	0	-33		2	543	0.79	2.38	1.00	751.20	1441.70	54.25	0.05	0
30	0	0	34	0	0	-34		2	539	2.15	2.44	1.00	737.80	1445.70	53.82	0.05	0
31	0	0	35	0	0	-35		2	534	1.52	2.54	1.00	724.30	1449.60	53.38	0.05	0
32	0	0	36	0	0	-36		2	530	531.30	16.14	1.00	710.90	1453.50	52.94	0.06	0
33	0	0	37	0	0	-37		2	526	0.98	2.68	1.00	697.30	1457.40	52.50	0.06	0
34	0	0	38	0	0	-38		2	521	-0.53	2.85	1.00	683.80	1461.30	52.05	0.06	0
35	0	0	39	0	0	-39		2	517	-4.92	3.00	0.95	670.20	1465.10	51.61	0.06	0
36	0	0	45	0	0	-45		2	490	2.37	3.73	1.00	587.80	1487.90	48.93	0.07	0

Phase problem → Fourier transform

Electron density map  
Atom coordinates

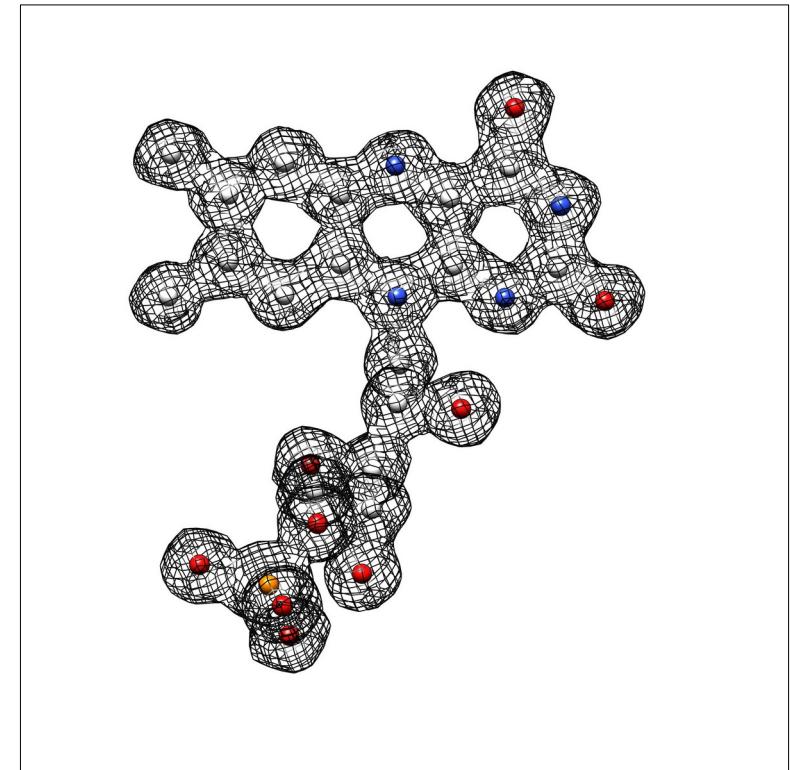


# XRD - output

Structure with high resolution

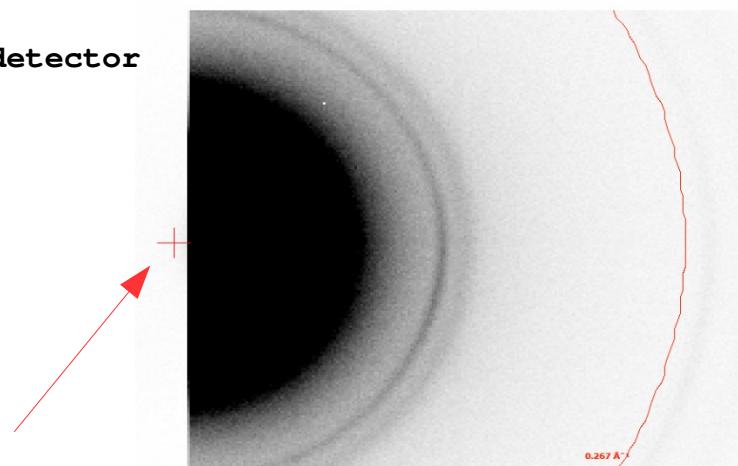
3D coordinates,  
PDB format, in real space x, y, z

HETATM	2968	C9A	FNR	B	201	18.093	-4.814	66.786	1.00	7.49	C
HETATM	2969	N10	FNR	B	201	18.926	-4.246	67.742	1.00	7.31	N
HETATM	2970	CAA	FNR	B	201	20.263	-4.072	67.478	1.00	7.09	C
HETATM	2971	N1	FNR	B	201	21.001	-3.377	68.290	1.00	7.03	N
HETATM	2972	C2	FNR	B	201	22.260	-3.219	68.029	1.00	7.67	C
HETATM	2973	O2	FNR	B	201	22.895	-2.495	68.786	1.00	8.20	O
HETATM	2974	N3	FNR	B	201	22.832	-3.652	66.907	1.00	7.06	N
HETATM	2975	C4	FNR	B	201	22.174	-4.394	65.997	1.00	6.83	C
HETATM	2976	O4	FNR	B	201	22.762	-4.851	64.973	1.00	6.36	O
HETATM	2977	C4A	FNR	B	201	20.769	-4.664	66.266	1.00	6.92	C
HETATM	2978	N5	FNR	B	201	19.988	-5.342	65.408	1.00	7.47	N
HETATM	2979	C5A	FNR	B	201	18.678	-5.408	65.569	1.00	7.59	C
HETATM	2980	C6	FNR	B	201	17.856	-5.955	64.611	1.00	7.52	C
HETATM	2981	C7	FNR	B	201	16.502	-5.982	64.805	1.00	7.90	C
HETATM	2982	C7M	FNR	B	201	15.729	-6.554	63.669	1.00	8.29	C
HETATM	2983	C8	FNR	B	201	15.929	-5.414	65.987	1.00	8.03	C
HETATM	2984	C8M	FNR	B	201	14.454	-5.501	66.238	1.00	8.78	C
HETATM	2985	C9	FNR	B	201	16.734	-4.854	66.964	1.00	7.60	C
HETATM	2986	C1'	FNR	B	201	18.395	-3.681	68.980	1.00	7.22	C
HETATM	2987	C2'	FNR	B	201	18.023	-2.264	68.917	1.00	7.07	C
HETATM	2988	O2'	FNR	B	201	19.202	-1.525	69.187	1.00	6.83	O
HETATM	2989	C3'	FNR	B	201	16.991	-1.922	69.974	1.00	7.10	C
HETATM	2990	O3'	FNR	B	201	15.809	-2.741	69.803	1.00	7.23	O
HETATM	2991	C4'	FNR	B	201	16.644	-0.457	69.874	1.00	7.17	C
HETATM	2992	O4'	FNR	B	201	15.924	-0.152	71.082	1.00	7.78	O
HETATM	2993	C5'	FNR	B	201	15.746	-0.178	68.686	1.00	6.50	C
HETATM	2994	O5'	FNR	B	201	15.556	1.235	68.495	1.00	6.21	O
HETATM	2995	P	FNR	B	201	14.156	1.902	68.939	1.00	5.78	P
HETATM	2996	O1P	FNR	B	201	14.298	3.344	68.517	1.00	5.61	O
HETATM	2997	O2P	FNR	B	201	13.020	1.211	68.280	1.00	5.79	O
HETATM	2998	O3P	FNR	B	201	14.088	1.723	70.433	1.00	6.03	O



# SAXS - isotropic scattering $\rightarrow$ radial averaging

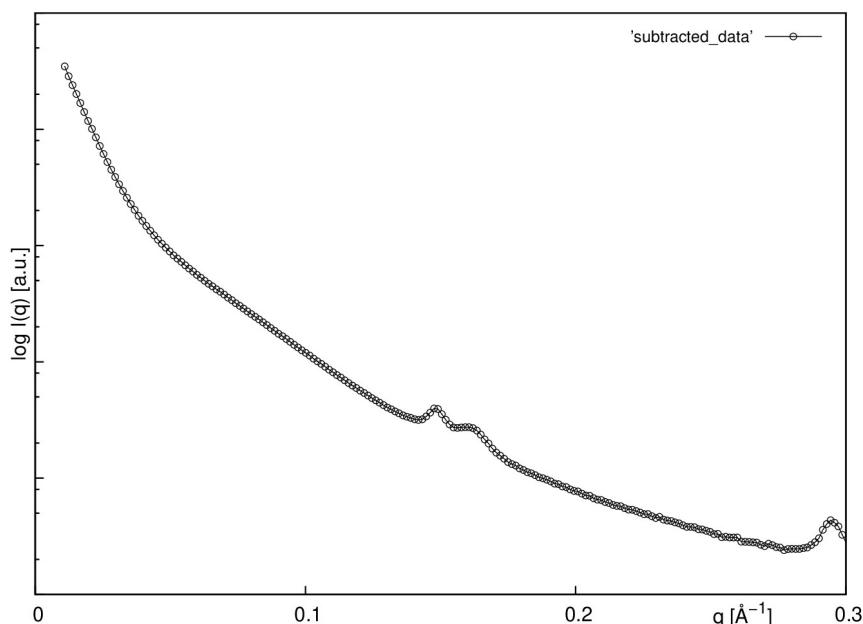
Image from detector



SAXS curve - txt file

S	I	SD
0.0166627	0.973409	0.0105135
0.0181009	0.949327	0.00825875
0.019539	0.926431	0.00710475
0.0209772	0.903951	0.00601258
0.0224154	0.874771	0.00508973
0.0238536	0.852858	0.00497113
0.0252918	0.814447	0.00488254
0.0267299	0.786807	0.00621834
0.0281681	0.764362	0.00503159
0.0296063	0.728444	0.00473319
0.0310445	0.715133	0.00407554
0.0324826	0.689458	0.00290202
0.0339208	0.673741	0.00323363
0.035359	0.643885	0.00306539
0.0367972	0.62527	0.00308635
0.0382354	0.607309	0.00260789
0.0396735	0.577026	0.00250923
0.0411117	0.559098	0.00284009
0.0425499	0.539921	0.00270961
0.0439881	0.517847	0.00237405
0.0454262	0.498033	0.00242969
0.0468644	0.479202	0.00216011
0.0483026	0.46049	0.00254124
0.0497408	0.442897	0.00241848
0.051179	0.430697	0.00202251
0.0526171	0.410743	0.00230559
0.0540553	0.396025	0.00210054
0.0554935	0.380624	0.00223789
0.0569317	0.362036	0.00205874
0.0583698	0.351014	0.002059
0.059808	0.336265	0.00196403
0.0612462	0.324684	0.00208857
0.0626844	0.311476	0.00170515
0.0641225	0.299744	0.00185129
0.0655607	0.290324	0.00174539
0.0669989	0.276878	0.00176257
0.0684371	0.267973	0.00167985
0.0698753	0.255922	0.00181061
0.0713134	0.245727	0.00178832

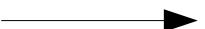
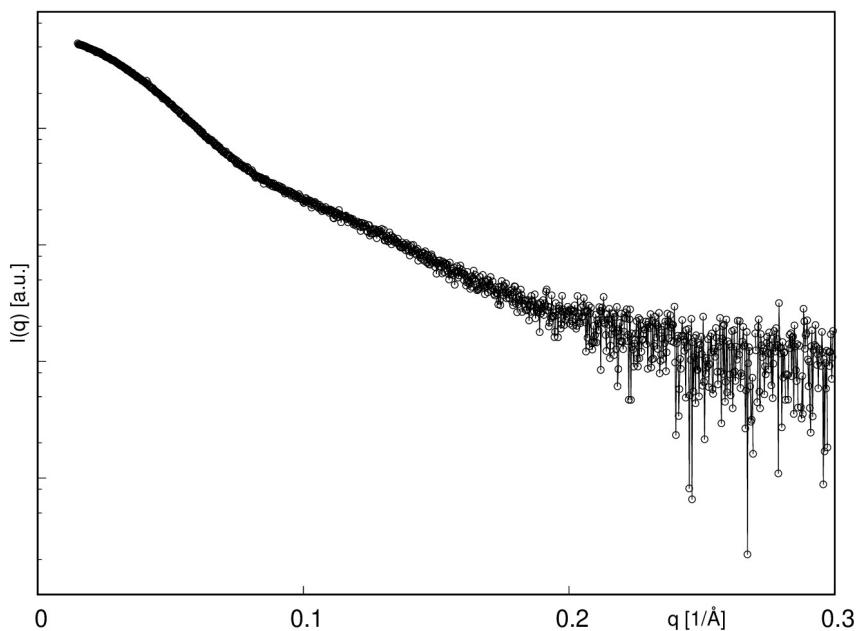
Radially averaged data - SAXS curve



# SAXS – output

Low resolution structure

SAXS data

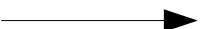
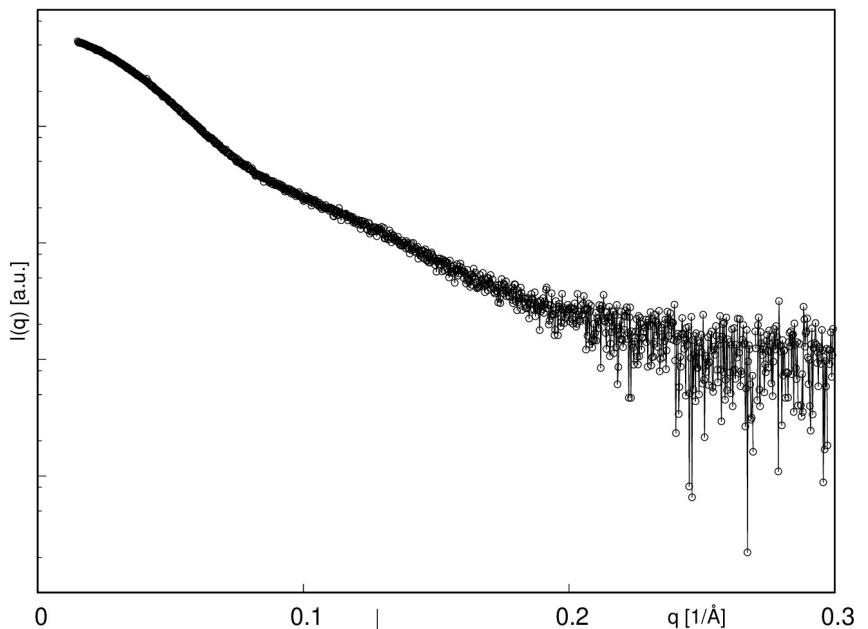


Data-collection parameters	
Instrument	BioSAXS-1000 (Rigaku)
Wavelength [ $\text{\AA}$ ]	1.5418
$q$ range [ $\text{\AA}^{-1}$ ]	0.0095 - 0.65
Exposure time [min]	60
Temperature [ $^{\circ}\text{C}$ ]	4
Concentration [ $\text{mg ml}^{-1}$ ]	5.0
Structural parameters	
$R_g$ [ $\text{\AA}$ ] (from Guinier)	18.163
$R_g$ [ $\text{\AA}$ ] (from $P(r)$ )	18.85
$D_{\max}$ [ $\text{\AA}$ ]	62.21
Porod volume estimate [ $\text{\AA}^3$ ]	15417
MW theoretical [kDa]	9.4
MW porod [kDa]	9.4
MW dammin [kDa]	10.5

# SAXS – output

Low resolution structure

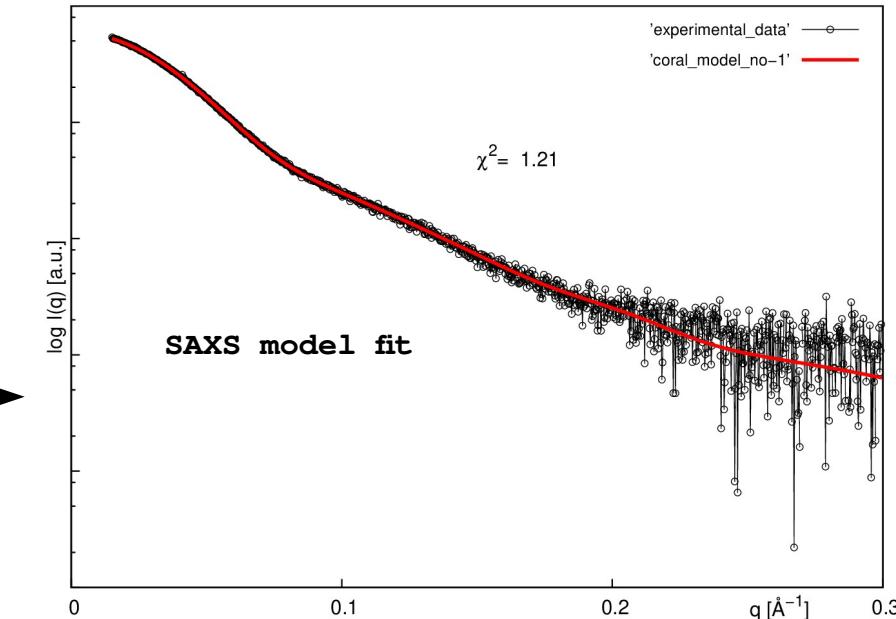
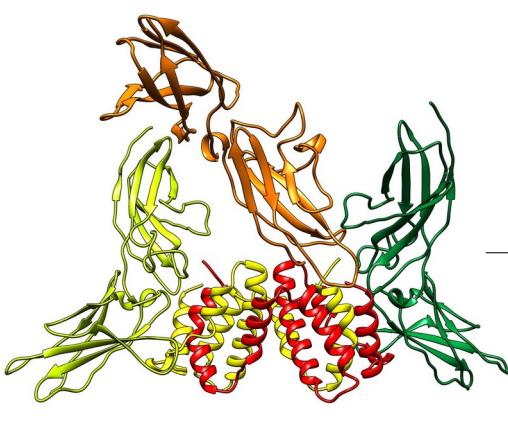
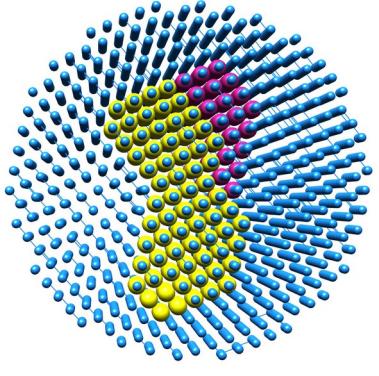
SAXS data



## SAXS table

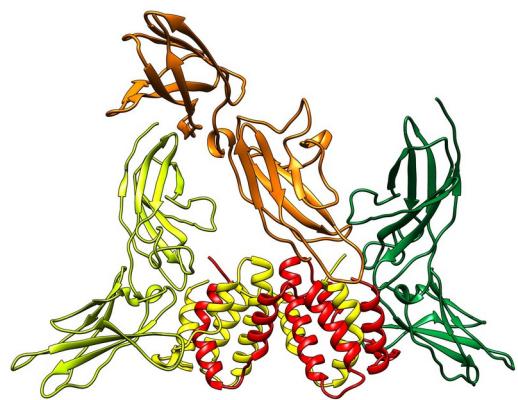
Data-collection parameters	
Instrument	BioSAXS-1000 (Rigaku)
Wavelength [ $\text{\AA}$ ]	1.5418
$q$ range [ $\text{\AA}^{-1}$ ]	0.0095 - 0.65
Exposure time [min]	60
Temperature [ $^{\circ}\text{C}$ ]	4
Concentration [ $\text{mg ml}^{-1}$ ]	5.0
Structural parameters	
$R_g$ [ $\text{\AA}$ ] (from Guinier)	18.163
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Porod volume estimate [ $\text{\AA}^3$ ]	15417
MW theoretical [kDa]	9.4
MW porod [kDa]	9.4
MW dammin [kDa]	10.5

reconstruction



# SAXS - output

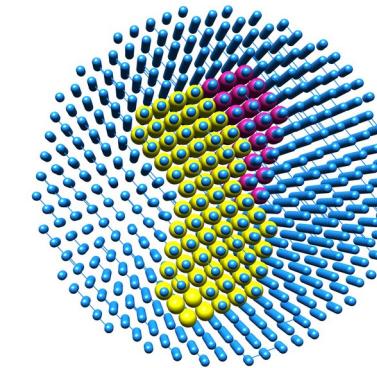
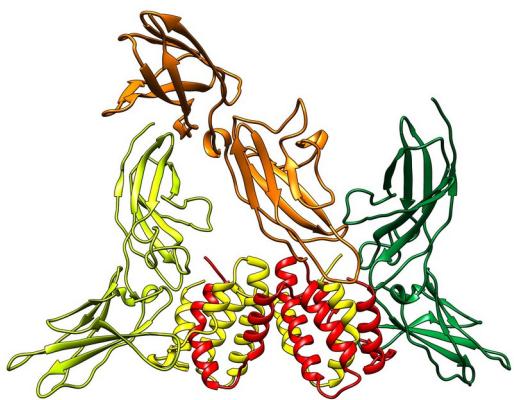
Low resolution structure



ATOM	1	N	ILE	A	4	-36.360	17.558	21.748	1.00	20.00
ATOM	2	CA	ILE	A	4	-36.096	16.392	22.585	1.00	20.00
ATOM	3	C	ILE	A	4	-35.687	16.837	23.988	1.00	20.00
ATOM	4	O	ILE	A	4	-34.781	17.649	24.159	1.00	20.00
ATOM	5	CB	ILE	A	4	-35.027	15.488	21.933	1.00	20.00
ATOM	6	CG1	ILE	A	4	-35.563	14.881	20.645	1.00	20.00
ATOM	7	CG2	ILE	A	4	-34.584	14.383	22.884	1.00	20.00
ATOM	8	CD1	ILE	A	4	-34.500	14.135	19.847	1.00	20.00
ATOM	9	N	GLY	A	5	-36.348	16.278	25.000	1.00	20.00
ATOM	10	CA	GLY	A	5	-36.166	16.769	26.356	1.00	20.00
ATOM	11	C	GLY	A	5	-34.790	16.441	26.907	1.00	20.00
ATOM	12	O	GLY	A	5	-34.256	15.350	26.703	1.00	20.00
ATOM	13	N	THR	A	6	-34.216	17.399	27.635	1.00	20.00
ATOM	14	CA	THR	A	6	-32.917	17.203	28.259	1.00	20.00
ATOM	15	C	THR	A	6	-33.002	16.952	29.758	1.00	20.00
ATOM	16	O	THR	A	6	-31.999	16.560	30.359	1.00	20.00
ATOM	17	CB	THR	A	6	-32.014	18.421	28.013	1.00	20.00
ATOM	18	CG2	THR	A	6	-31.912	18.722	26.539	1.00	20.00
ATOM	19	OG1	THR	A	6	-32.570	19.559	28.685	1.00	20.00
ATOM	20	N	GLY	A	7	-34.162	17.186	30.377	1.00	20.00
ATOM	21	CA	GLY	A	7	-34.280	17.005	31.806	1.00	20.00
ATOM	22	C	GLY	A	7	-34.519	15.562	32.194	1.00	20.00
ATOM	23	O	GLY	A	7	-34.870	14.716	31.368	1.00	20.00
ATOM	24	N	PHE	A	8	-34.320	15.295	33.486	1.00	20.00
ATOM	25	CA	PHE	A	8	-34.561	13.987	34.094	1.00	20.00
ATOM	26	C	PHE	A	8	-35.542	14.179	35.247	1.00	20.00
ATOM	27	O	PHE	A	8	-35.138	14.249	36.417	1.00	20.00

# SAXS - output

Low resolution structure

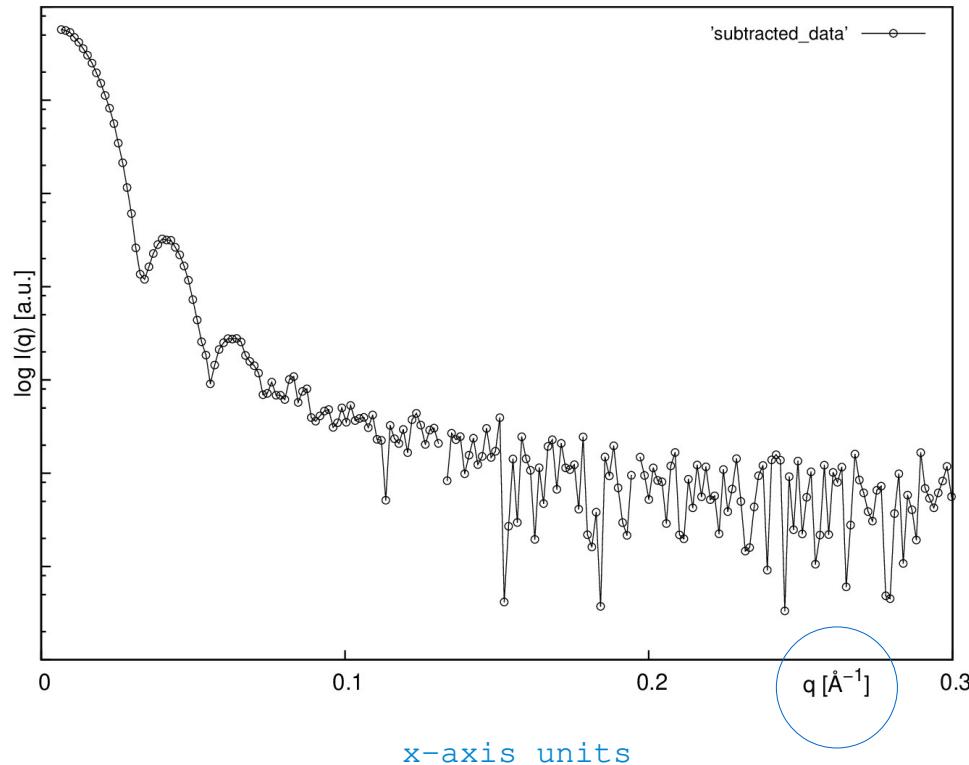


ATOM	1	N	ILE	A	4	-36.360	17.558	21.748	1.00	20.00
ATOM	2	CA	ILE	A	4	-36.096	16.392	22.585	1.00	20.00
ATOM	3	C	ILE	A	4	-35.687	16.837	23.988	1.00	20.00
ATOM	4	O	ILE	A	4	-34.781	17.649	24.159	1.00	20.00
ATOM	5	CB	ILE	A	4	-35.027	15.488	21.933	1.00	20.00
ATOM	6	CG1	ILE	A	4	-35.563	14.881	20.645	1.00	20.00
ATOM	7	CG2	ILE	A	4	-34.584	14.383	22.884	1.00	20.00
ATOM	8	CD1	ILE	A	4	-34.500	14.135	19.847	1.00	20.00
ATOM	9	N	GLY	A	5	-36.348	16.278	25.000	1.00	20.00
ATOM	10	CA	GLY	A	5	-36.166	16.769	26.356	1.00	20.00
ATOM	11	C	GLY	A	5	-34.790	16.441	26.907	1.00	20.00
ATOM	12	O	GLY	A	5	-34.256	15.350	26.703	1.00	20.00
ATOM	13	N	THR	A	6	-34.216	17.399	27.635	1.00	20.00
ATOM	14	CA	THR	A	6	-32.917	17.203	28.259	1.00	20.00
ATOM	15	C	THR	A	6	-33.002	16.952	29.758	1.00	20.00
ATOM	16	O	THR	A	6	-31.999	16.560	30.359	1.00	20.00
ATOM	17	CB	THR	A	6	-32.014	18.421	28.013	1.00	20.00
ATOM	18	CG2	THR	A	6	-31.912	18.722	26.539	1.00	20.00
ATOM	19	OG1	THR	A	6	-32.570	19.559	28.685	1.00	20.00
ATOM	20	N	GLY	A	7	-34.162	17.186	30.377	1.00	20.00
ATOM	21	CA	GLY	A	7	-34.280	17.005	31.806	1.00	20.00
ATOM	22	C	GLY	A	7	-34.519	15.562	32.194	1.00	20.00
ATOM	23	O	GLY	A	7	-34.870	14.716	31.368	1.00	20.00
ATOM	24	N	PHE	A	8	-34.320	15.295	33.486	1.00	20.00
ATOM	25	CA	PHE	A	8	-34.561	13.987	34.094	1.00	20.00
ATOM	26	C	PHE	A	8	-35.542	14.179	35.247	1.00	20.00
ATOM	27	O	PHE	A	8	-35.138	14.249	36.417	1.00	20.00

ATOM	1	CA	ASP	1	0.778	-0.275	-0.227	1.00	20.00
ATOM	2	CA	ASP	1	-7.972	1.475	12.148	1.00	20.00
ATOM	3	CA	ASP	1	9.528	-5.525	-12.601	1.00	20.00
ATOM	4	CA	ASP	1	13.028	-5.525	-12.601	1.00	20.00
ATOM	5	CA	ASP	1	-0.972	15.475	7.198	1.00	20.00
ATOM	6	CA	ASP	1	11.278	6.725	14.623	1.00	20.00
ATOM	7	CA	ASP	1	9.528	8.475	12.148	1.00	20.00
ATOM	8	CA	ASP	1	-14.972	1.475	-2.701	1.00	20.00
ATOM	9	CA	ASP	1	-6.222	-0.275	14.623	1.00	20.00
ATOM	10	CA	ASP	2	11.278	-3.775	-15.076	1.00	20.00
ATOM	11	CA	ASP	2	2.528	-2.025	2.248	1.00	20.00
ATOM	12	CA	ASP	2	2.528	15.475	7.198	1.00	20.00
ATOM	13	CA	ASP	2	-6.222	3.225	14.623	1.00	20.00
ATOM	14	CA	ASP	2	11.278	13.725	-5.176	1.00	20.00
ATOM	15	CA	ASP	2	0.778	17.225	4.723	1.00	20.00
ATOM	16	CA	ASP	2	-14.972	4.975	-2.701	1.00	20.00
ATOM	17	CA	ASP	2	-16.721	3.225	-0.227	1.00	20.00
ATOM	18	CA	ASP	2	7.778	-3.775	-15.076	1.00	20.00
ATOM	19	CA	ASP	2	16.529	4.975	-2.701	1.00	20.00
ATOM	20	CA	ASP	3	0.778	13.725	4.723	1.00	20.00
ATOM	21	CA	ASP	3	-7.972	1.475	17.098	1.00	20.00
ATOM	22	CA	ASP	3	7.778	6.725	14.623	1.00	20.00
ATOM	23	CA	ASP	3	-2.722	13.725	4.723	1.00	20.00
ATOM	24	CA	ASP	3	11.278	-7.275	-15.076	1.00	20.00
ATOM	25	CA	ASP	3	9.528	-9.025	-12.601	1.00	20.00
ATOM	26	CA	ASP	3	-13.222	3.225	-0.227	1.00	20.00

# SAXS curve - SAXS data

averaged data - SAXS profil



scattering vector, momentum transfer:  $s = q \text{ [nm}^{-1}, \text{\AA}^{-1}\text{]}$

$$k = \frac{2\pi}{\lambda}$$

A geometric diagram illustrating the scattering geometry. A horizontal arrow labeled  $k = \frac{2\pi}{\lambda}$  represents the wave vector. A diagonal vector labeled  $q = \frac{4\pi \sin(\theta)}{\lambda}$  represents the scattering vector. The angle between the horizontal axis and the scattering vector is labeled  $2\theta$ .

## SAXS – application

**Sample: solution of proteins, nucleic acids, and complexes**

1. size
2. shape
3. homogenous vs. aggregated sample
4. folded vs. flexible protein
5. oligomeric state (mixtures)
6. interaction, quaternary structure
7. modeling of flexible systems

## **SAXS – advantages**

1. sample preparation
2. in solution – close to „natural conditions“
3. usefull before and after molecular structure is known
4. time consumption
5. not limited by size or flexibility (almost)

## **SAXS – disadvantages**

1. low resolution
2. non unambiguous interpretation

# SAXS - resolution

## 1. Bragg's

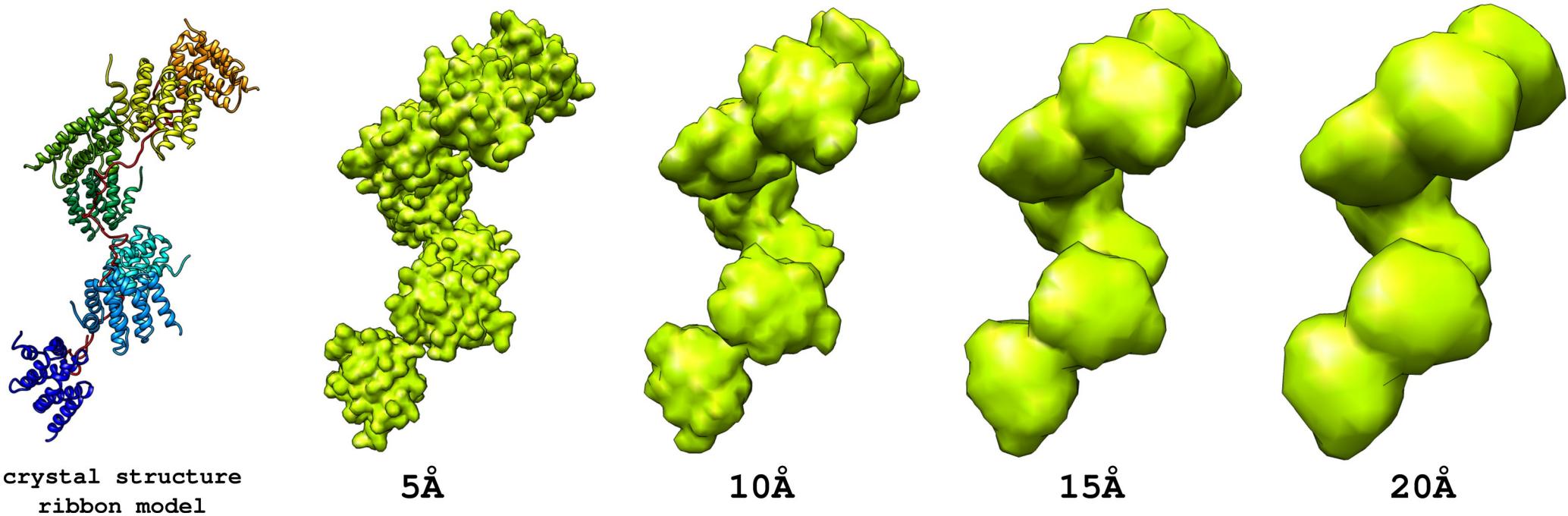
$$q_{\min} = 0.006 \text{ \AA}^{-1} \sim d = 1000 \text{ \AA}$$

$$q_{\max} = 0.6 \text{ \AA}^{-1} \sim d = 10 \text{ \AA}$$

$$d = \frac{2\pi}{q}$$

## 2. Effective

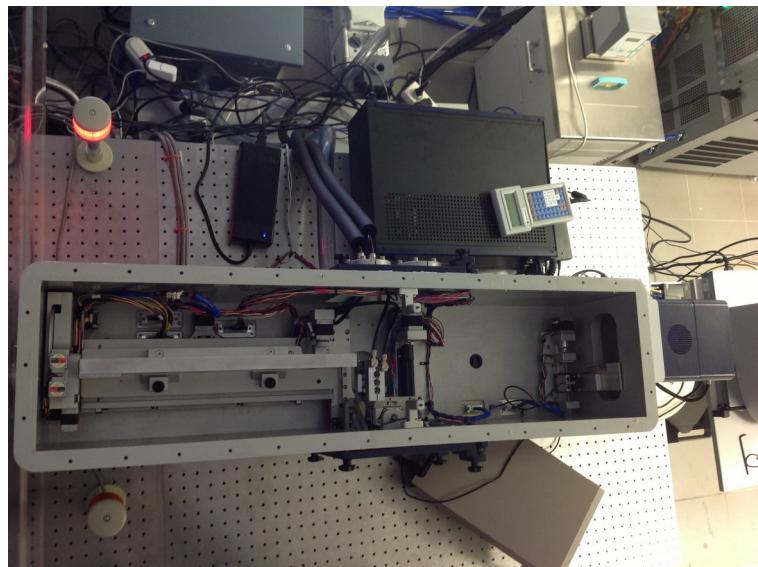
„low resolution structure“ 20–30 Å



# SAXS - experiment - home source



- rotating anode
- point focus
- colimation: 2D Kratky
- Pilatus 100K
- temperature range -30 to +65°C



# SAXS - experiment - home source



Data-collection parameters	
Instrument	BioSAXS-1000 (Rigaku)
Wavelength [Å]	1.5418
q range [ $\text{\AA}^{-1}$ ]	0.0095 - 0.65
Exposure time [min]	60
Temperature [°C]	4
Concentration [mg ml $^{-1}$ ]	5.0
Structural parameters	
Rg [Å] (from Guinier)	
Rg [Å] (from P(r))	
Dmax [Å]	
Porod volume estimate [ $\text{\AA}^3$ ]	
MW theoretical [kDa]	9.4
MW porod [kDa]	
MW dammin [kDa]	

## SAXS – experiment – sample

### **Sample quality:**

- pure as possible
- monodisperse
- free of aggregation

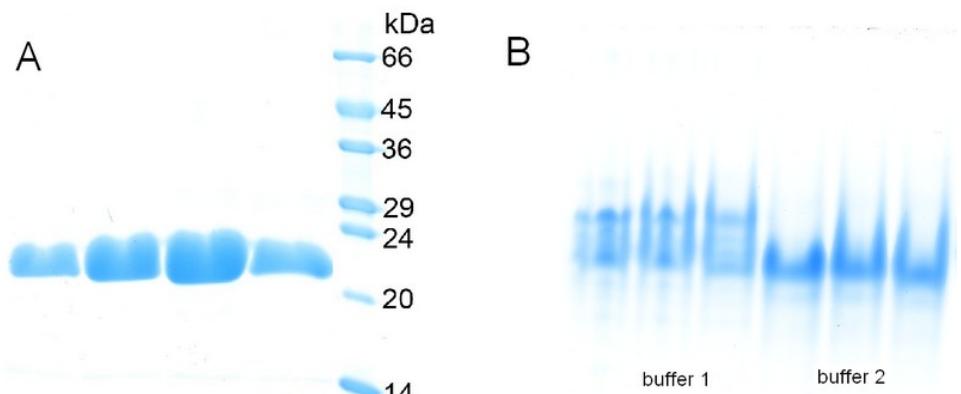
# SAXS – experiment – sample

## Sample quality:

- pure as possible
- monodisperse
- free of aggregation

## Quality control:

- native gels
- DLS



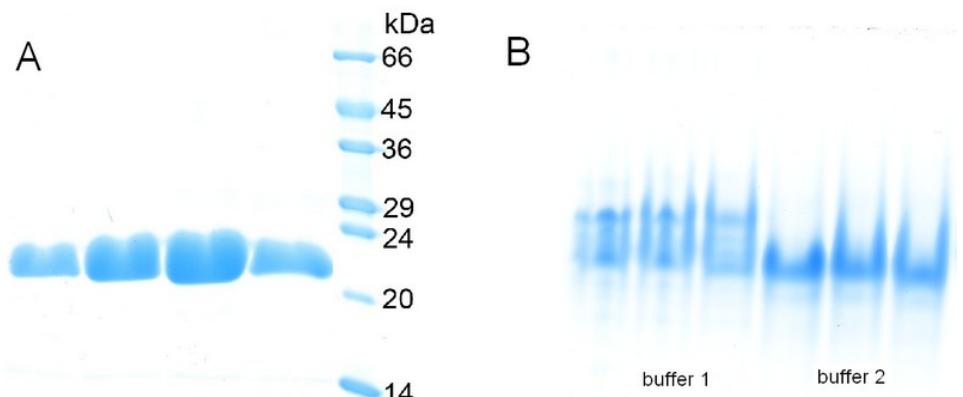
# SAXS – experiment – sample

## Sample quality:

- pure as possible
- monodisperse
- free of aggregation

## Quality control:

- native gels
- DLS



## Sample volume and concentration:

- ~25 microL
- concentration series 10; 5; 2.5 mg/ml

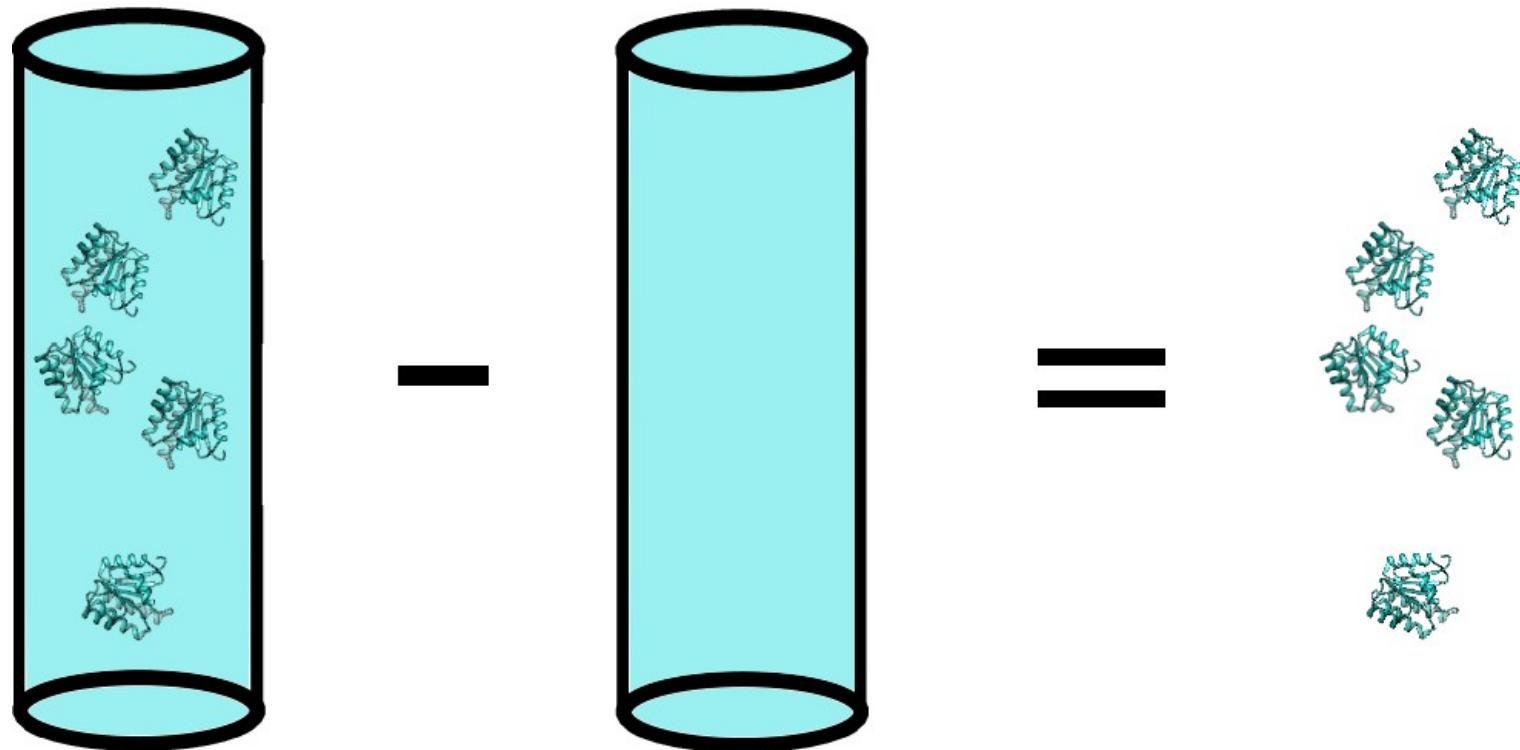
**Forward scattering intensity is proportional to size (number of electrons) :**

15 kDa protein needs 2.0 mg/ml

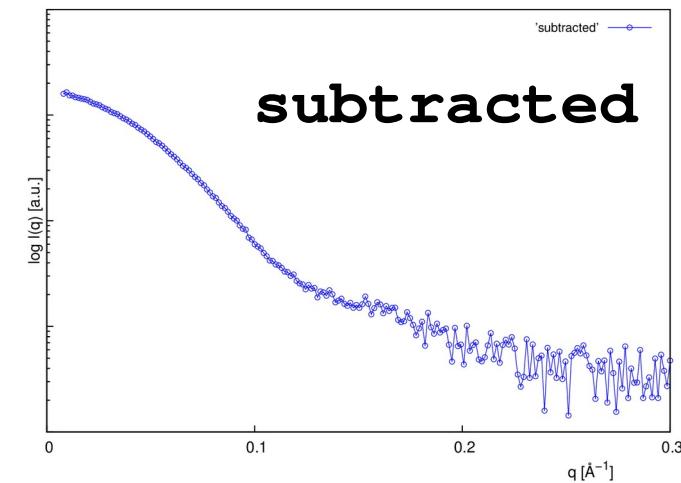
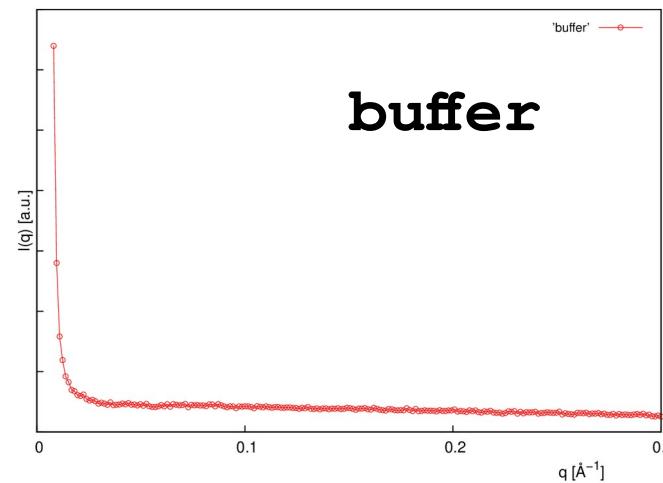
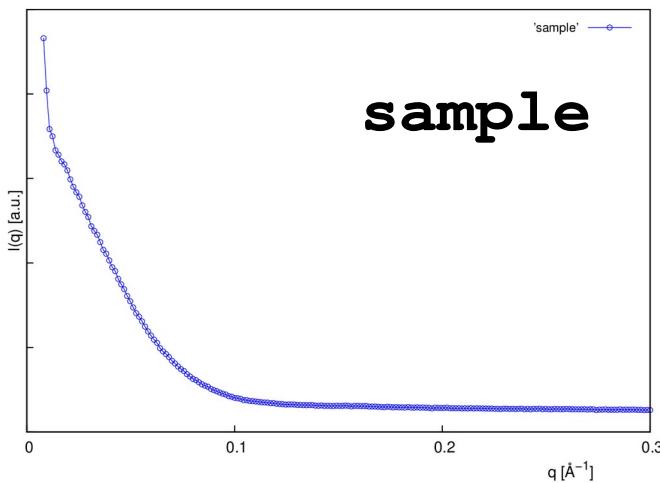
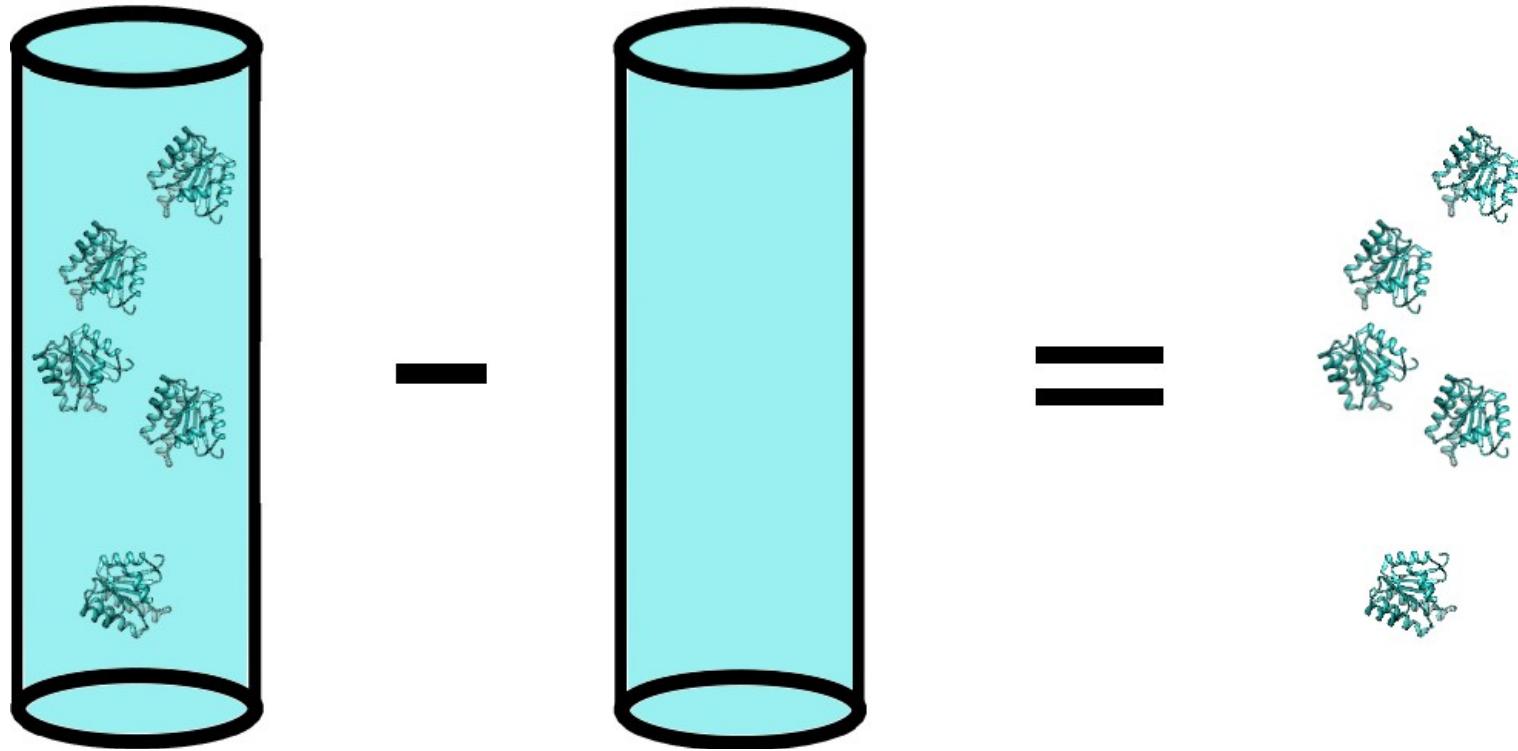
300 kDa protein ..... 0.1 mg/ml

=> Aggregates ruins meaningful data interpretation

# SAXS – subtraction



# SAXS – subtraction

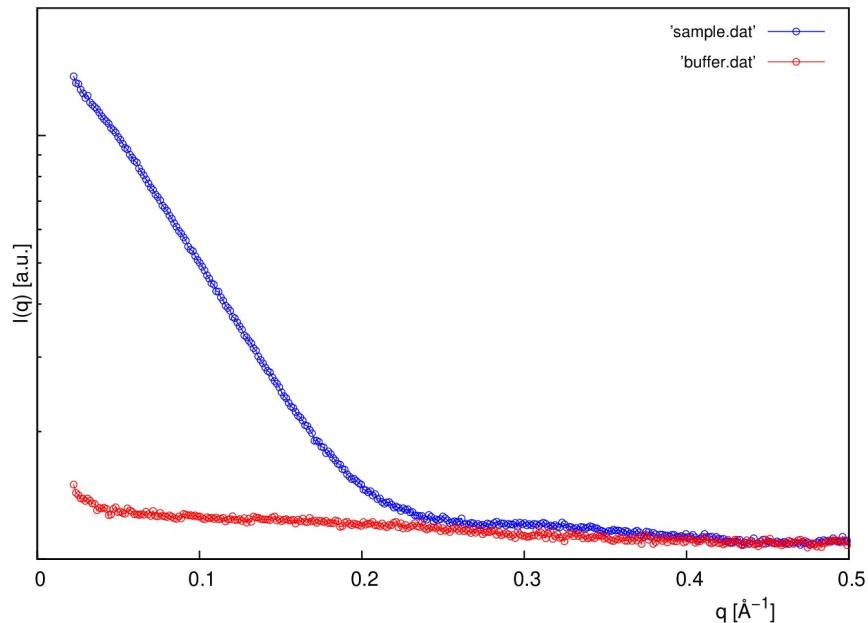


# SAXS – buffer

## Buffer requirements:

- identical to last step of purification (dialysis)
- salt < 1 M
- glycerol < 5% v/v
- detergents < CMC

**Matching buffer** = no oversubtraction or undersubtraction

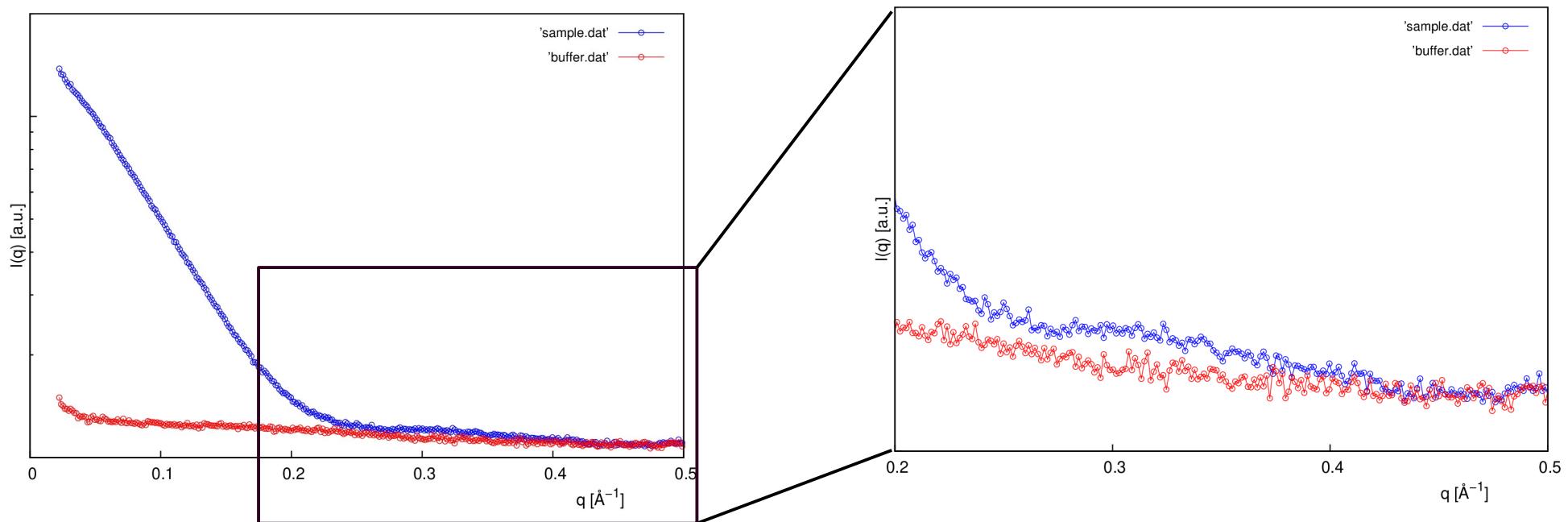


# SAXS – buffer

## Buffer requirements:

- identical to last step of purification (dialysis)
- salt < 1 M
- glycerol < 5% v/v
- detergents < CMC

**Matching buffer** = no oversubtraction or undersubtraction

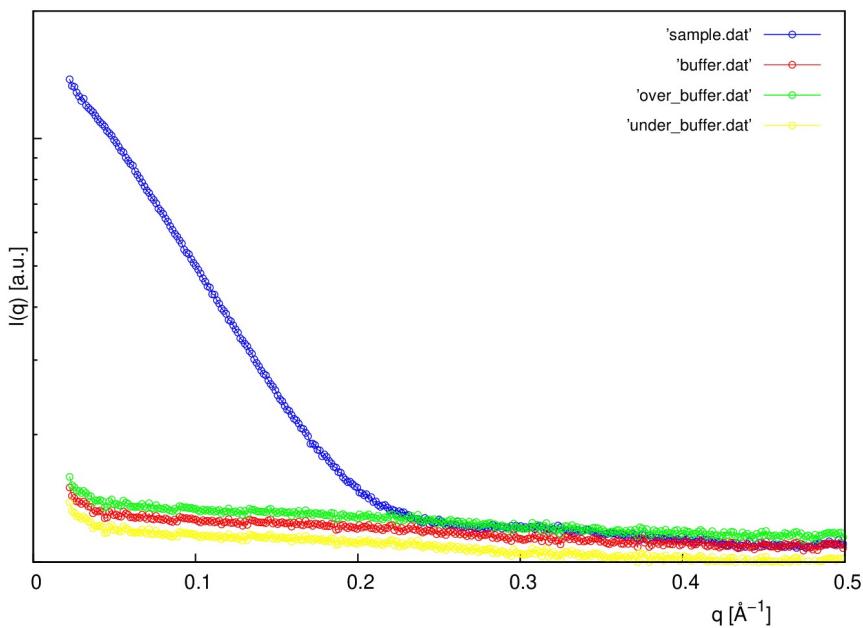


# SAXS – buffer

## Buffer requirements:

- identical to last step of purification (dialysis)
- salt < 1 M
- glycerol < 5% v/v
- detergents < CMC

**Matching buffer** = no over- or under-subtraction

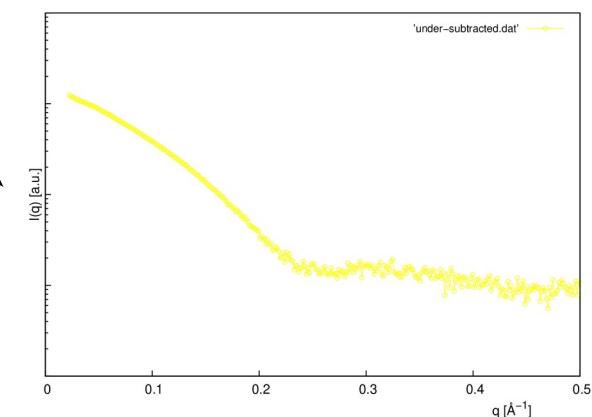
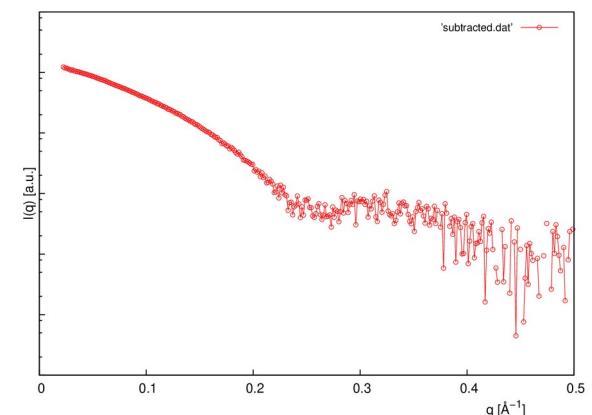
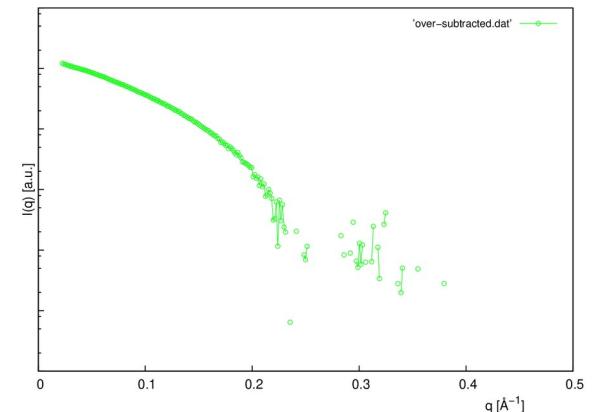
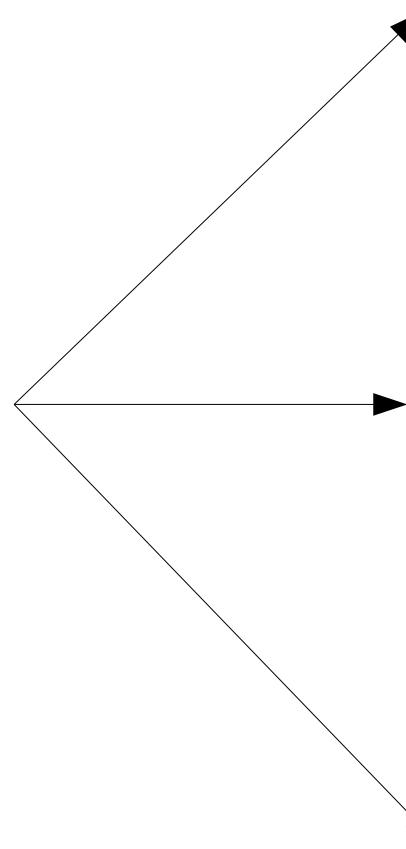
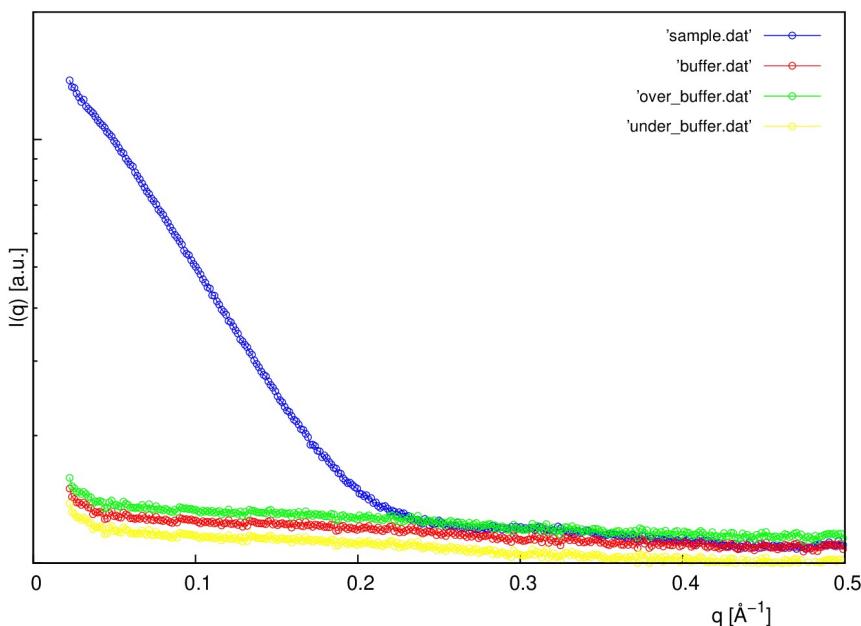


# SAXS – buffer

## Buffer requirements:

- identical to last step of purification (dialysis)
- salt < 1 M
- glycerol < 5% v/v
- detergents < CMC

**Matching buffer** = no over- or under-subtraction

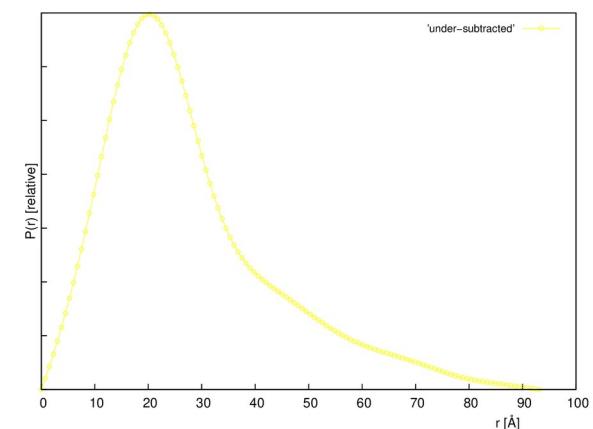
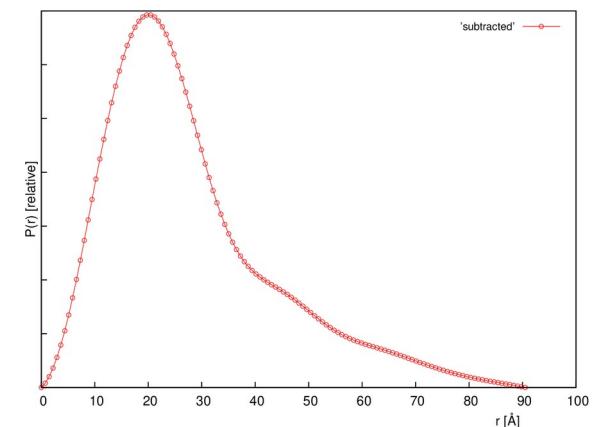
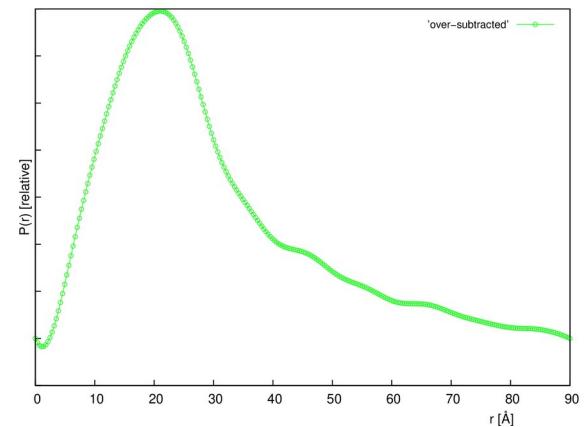
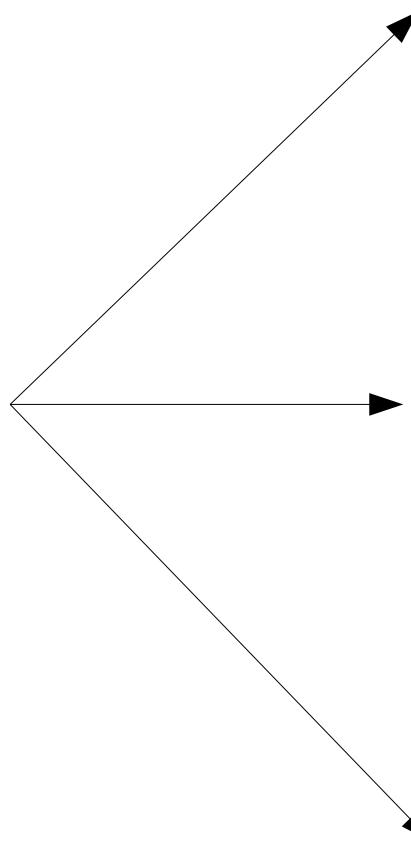
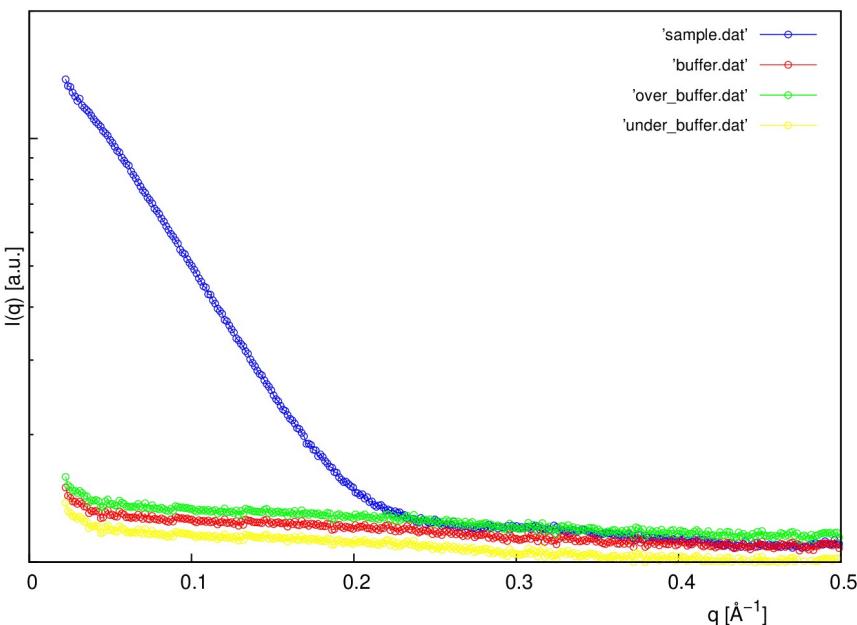


# SAXS – buffer

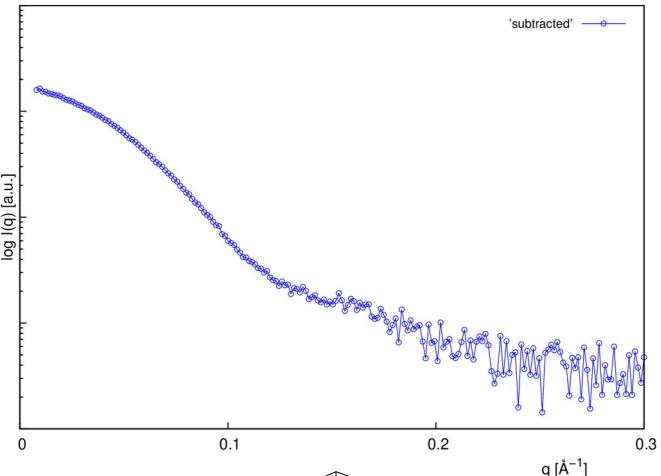
## Buffer requirements:

- identical to last step of purification (dialysis)
- salt < 1 M
- glycerol < 5% v/v
- detergents < CMC

**Matching buffer** = no over- or under-subtraction



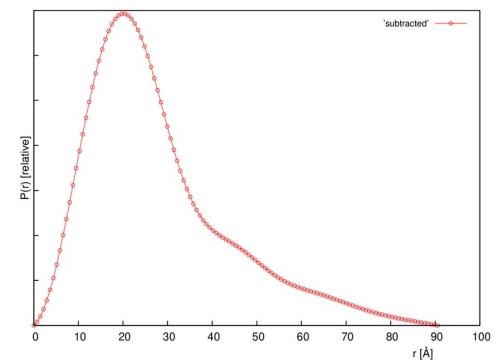
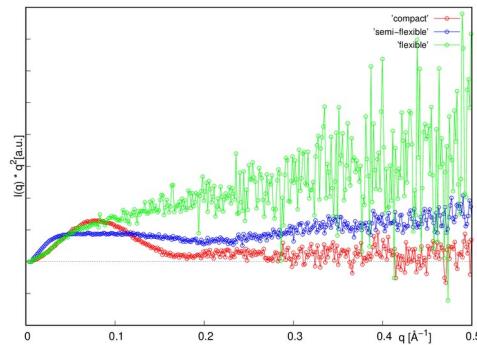
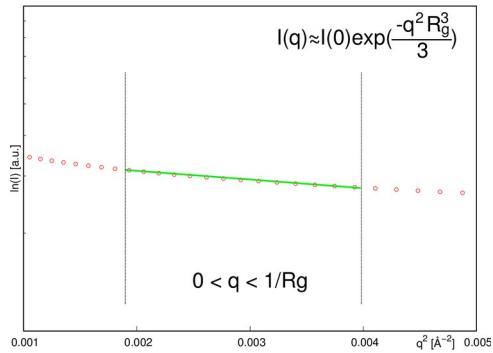
# Subtracted SAXS data



Guinier plot

Kratky plot

$P(r)$  Pair distance distribution function



# Guinier plot

- analysis of very low angle
- slope of decay proportional to size ( $R_g$ )
- $R_g$  = radius of gyration
- aggregation/repulsion detection

$$K = \frac{R_g^2}{3}$$

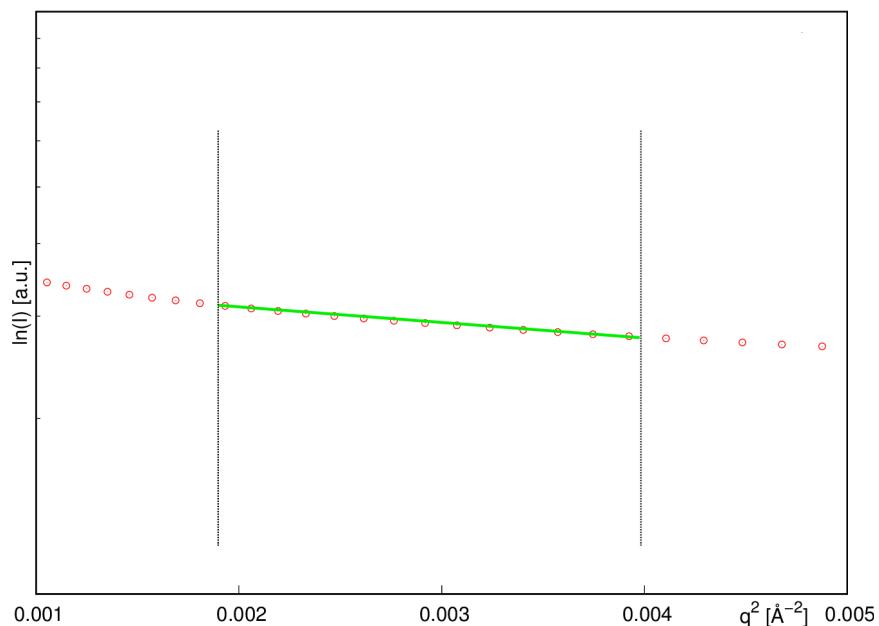
$$R_g = \sqrt{\frac{1}{n} \sum_{i=1}^n (r_i - r_{cm})^2}$$

$$\ln[I(q)] \approx \ln[I(0)] \exp\left(\frac{-R_g^2 q^2}{3}\right)$$

Valid in:

$$0 < R_g q < 1.2$$

$$0 < R_g q < \frac{1}{R_g}$$



# Guinier plot

- analysis of very low angle
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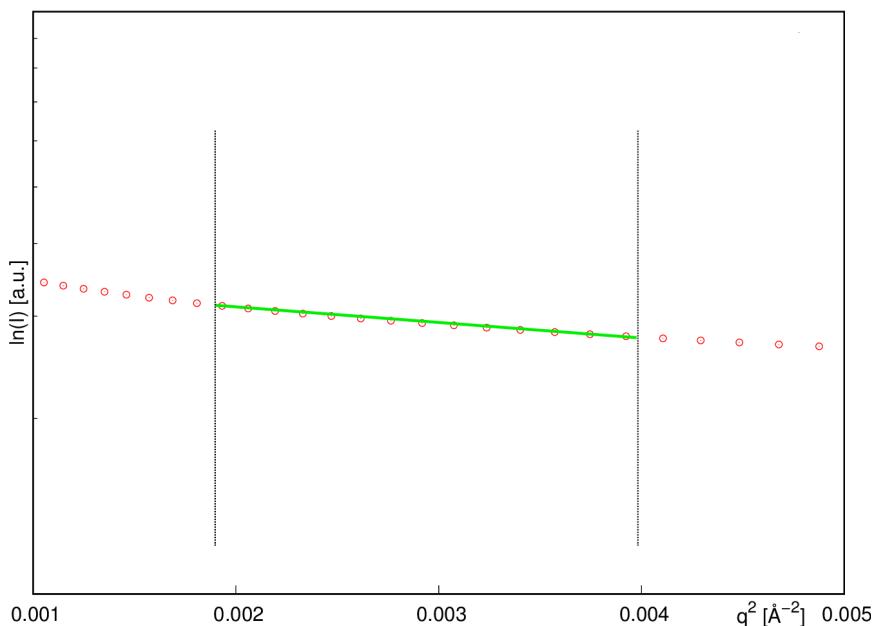
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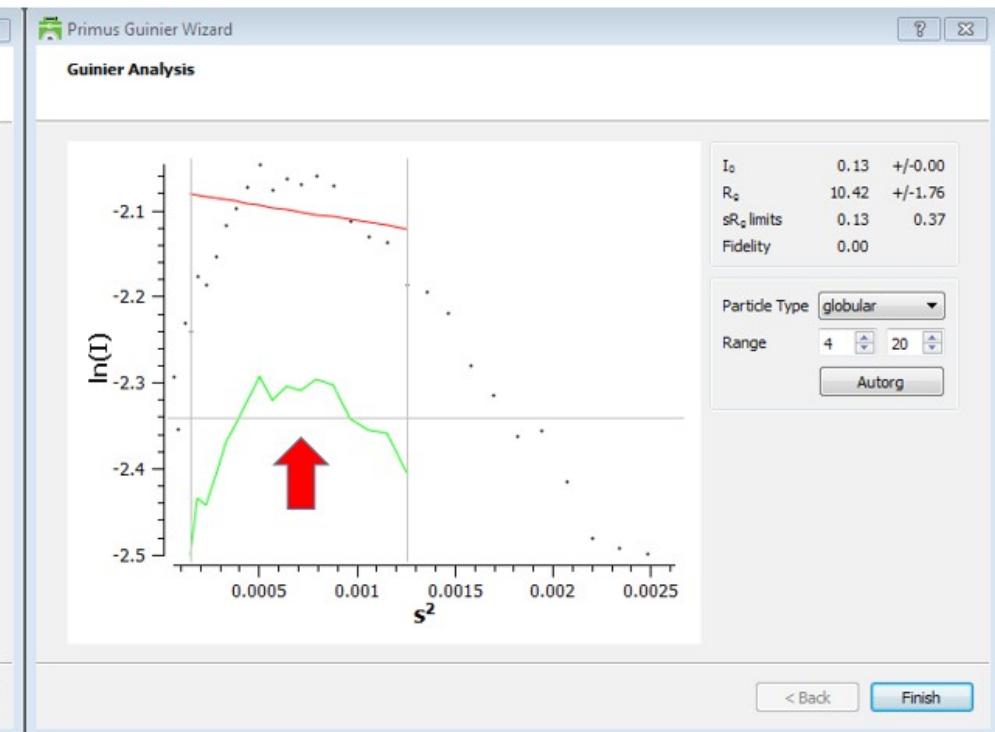
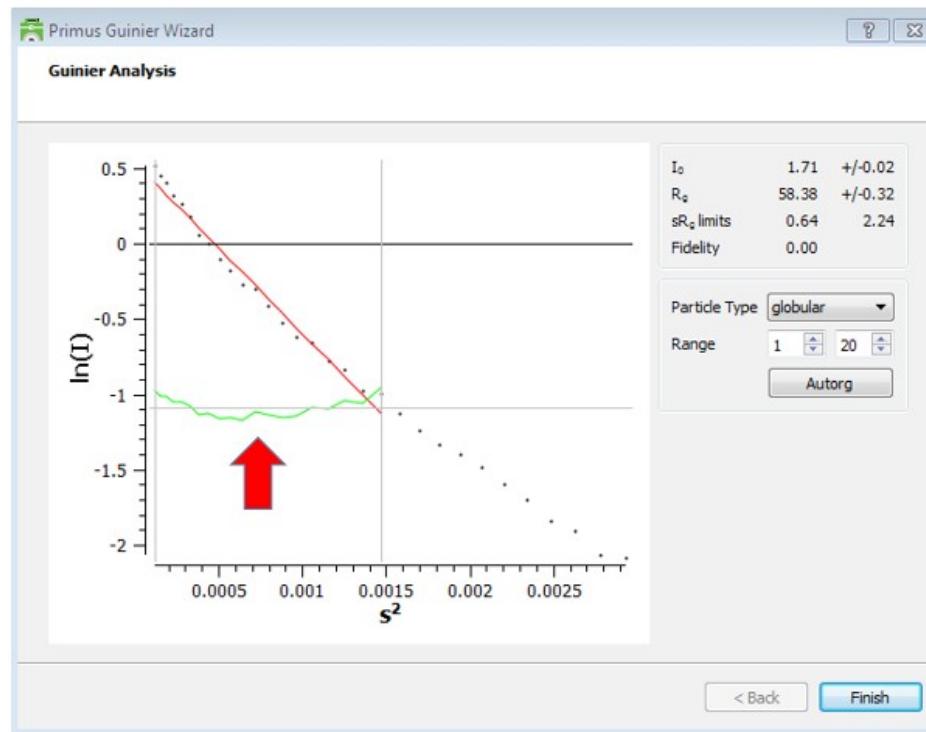


Data-collection parameters	
Instrument	BioSAXS-1000 (Rigaku)
Wavelength [ $\text{\AA}$ ]	1.5418
$q$ range [ $\text{\AA}^{-1}$ ]	0.0095 - 0.65
Exposure time [min]	60
Temperature [ $^{\circ}\text{C}$ ]	4
Concentration [mg ml $^{-1}$ ]	5.0
Structural parameters	
$R_g$ [ $\text{\AA}$ ] (from Guinier)	18.163
$R_g$ [ $\text{\AA}$ ] (from $P(r)$ )	
$D_{max}$ [ $\text{\AA}$ ]	
Porod volume estimate [ $\text{\AA}^3$ ]	
MW theoretical [kDa]	9.4
MW porod [kDa]	
MW dammin [kDa]	

# Guinier plot

- analysis of very low angle
- slope of decay proportional to size ( $R_g$ )
- $R_g$  = radius of gyration
- aggregation/repulsion detection

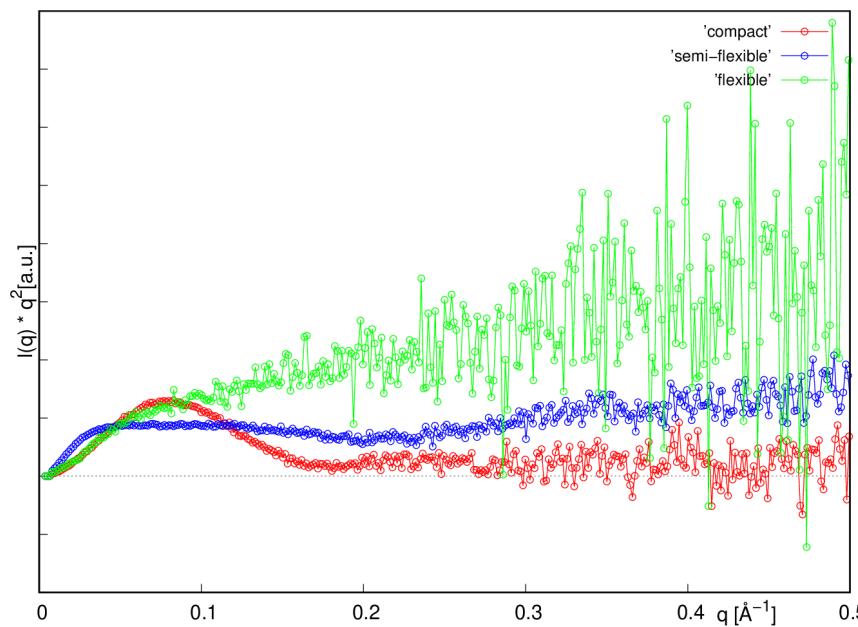
$$K = \frac{R_g^2}{3}$$



# Kratky plot

Chain compactness

- slope of decay of intensity in „Kratky“ region
- compact well folded chains have clear minima
- dependent on data quality (subtraction)



# Pair distance distribution function P (r)

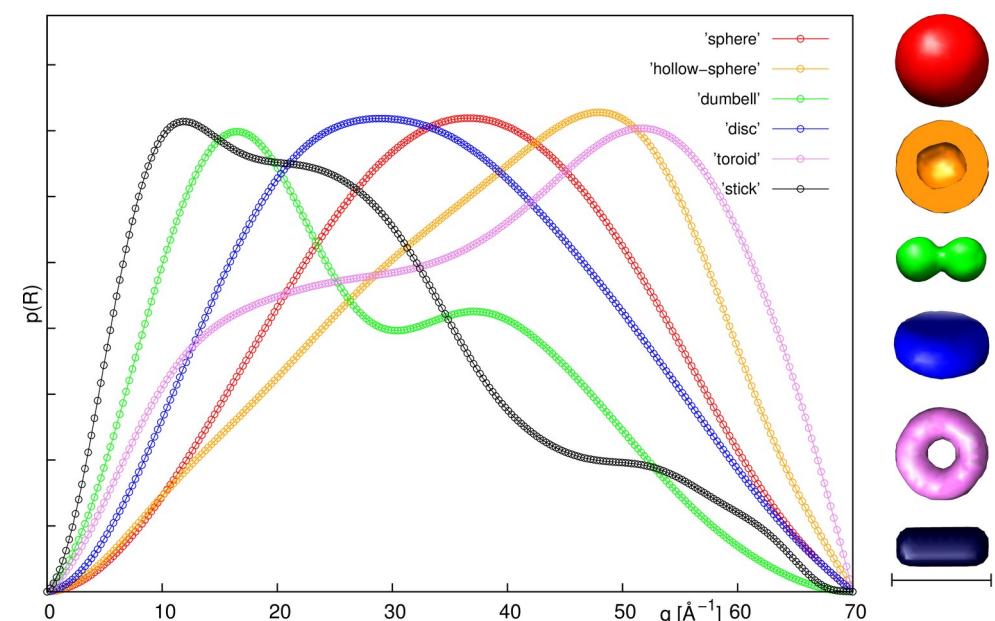
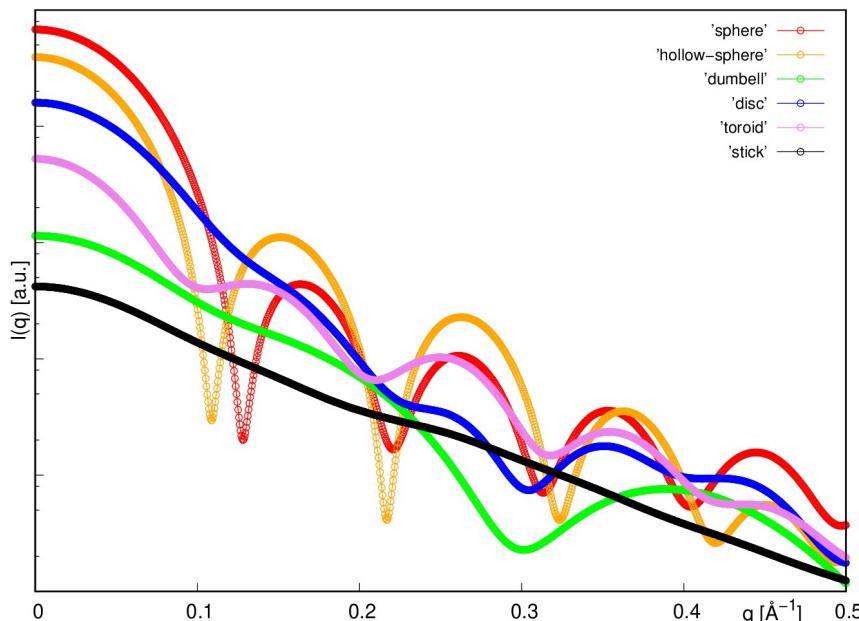
- indirect Fourier transform of scattered intensity
- GNOM - Svergun's perceptual criteria (smoothness, systematic deviation, ... )
- distances of all pairs of points (electrons) of the particle (weighted)

$$p(r) = \gamma(r) \pi r^2$$

$$I(q) = 4\pi \int_0^{D_{max}} p(r) \frac{\sin(qr)}{qr} dr$$



$$p(r) = \frac{r^2}{2\pi^2} \int_0^{\infty} q^2 I(q) \frac{\sin(qr)}{qr} dq$$



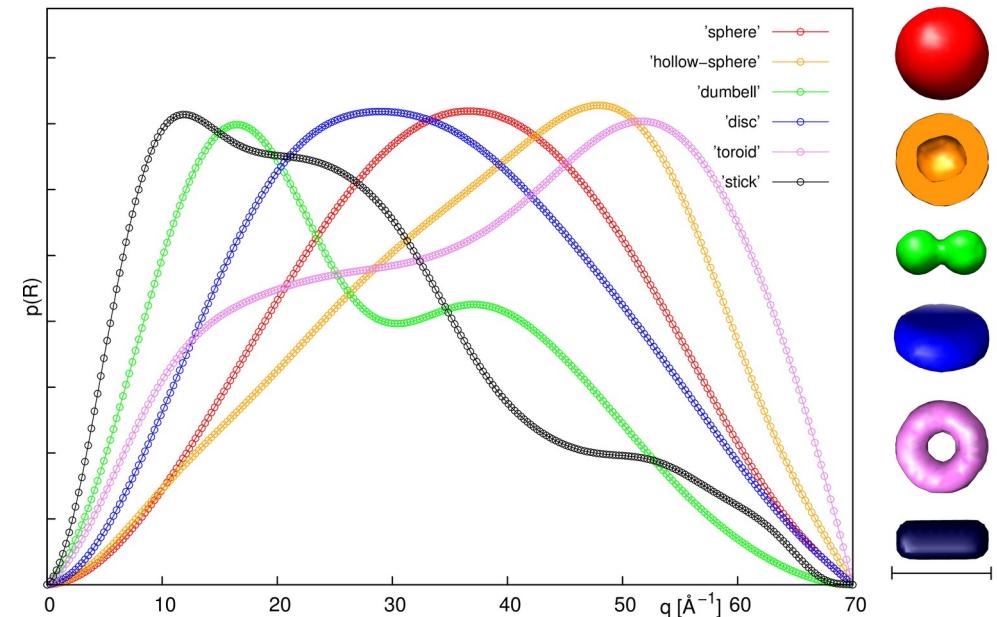
# Pair distance distribution function P (r)

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Data-collection parameters	
Instrument	BioSAXS-1000 (Rigaku)
Wavelength [Å]	1.5418
q range [Å <sup>-1</sup> ]	0.0095 - 0.65
Exposure time [min]	60
Temperature [°C]	4
Concentration [mg ml <sup>-1</sup> ]	5.0
Structural parameters	
Rg [Å] (from Guinier)	18.163
Rg [Å] (from P(r))	18.85
Dmax [Å]	62.21
Porod volume estimate [Å <sup>3</sup> ]	
MW theoretical [kDa]	9.4
MW porod [kDa]	
MW dammin [kDa]	

$$p(r) = \frac{r^2}{2\pi^2} \int_0^\infty q^2 I(q) \frac{\sin(qr)}{qr} dq$$



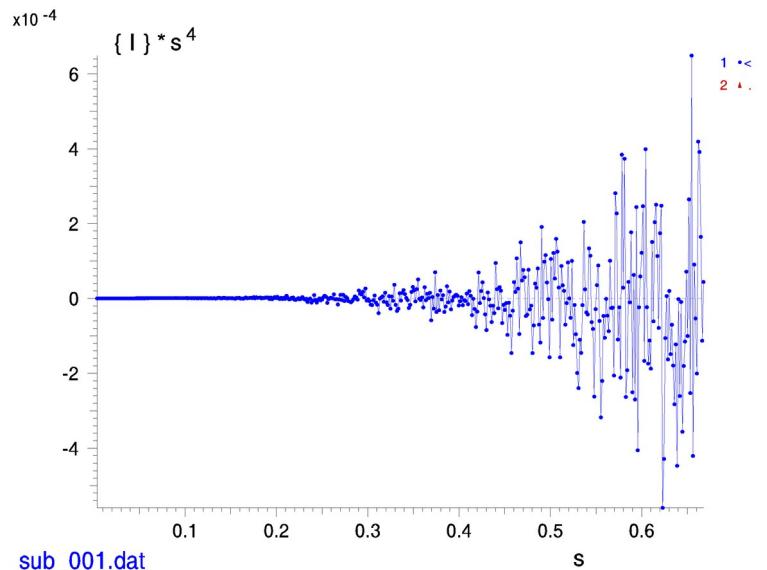
# Porod volume

- applicable for folded macromolecules
- Porod's law - intensity decay  $I(q) = q^{-4}$
- Porod's invariant

$$Q = \int_0^{\infty} q^2 [I(q) - K] dq$$

$$V_p = \frac{2\pi^2 I(0)}{Q}$$

<b>Data-collection parameters</b>	
Instrument	BioSAXS-1000 (Rigaku)
Wavelength [ $\text{\AA}$ ]	1.5418
q range [ $\text{\AA}^{-1}$ ]	0.0095 - 0.65
Exposure time [min]	60
Temperature [ $^{\circ}\text{C}$ ]	4
Concentration [mg ml $^{-1}$ ]	5.0
<b>Structural parameters</b>	
Rg [ $\text{\AA}$ ] (from Guinier)	18.163
Rg [ $\text{\AA}$ ] (from P(r))	18.85
Dmax [ $\text{\AA}$ ]	62.21
Porod volume estimate [ $\text{\AA}^3$ ]	15417
MW theoretical [kDa]	9.4
MW porod [kDa]	
MW dammin [kDa]	



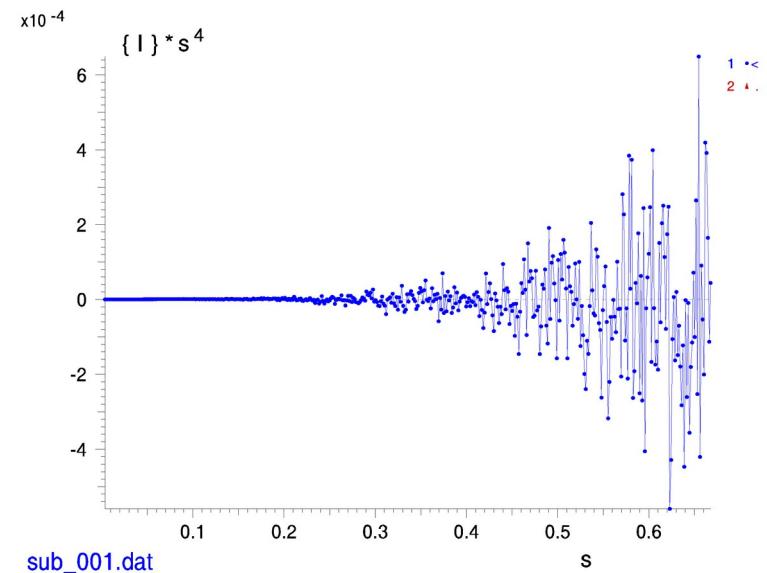
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Structural parameters	
Rg [\AA] (from Guinier)	18.163
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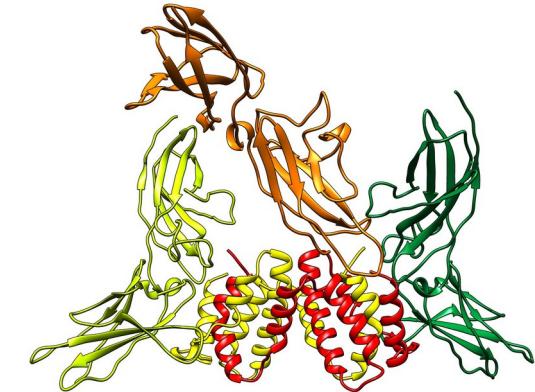


MW magic number for proteins = 0.625

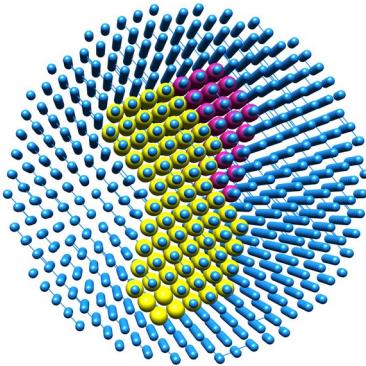
$$V_p [\text{\AA}^3] * 0.625 = MW [\text{Da}]$$

MW dammin [kDa]

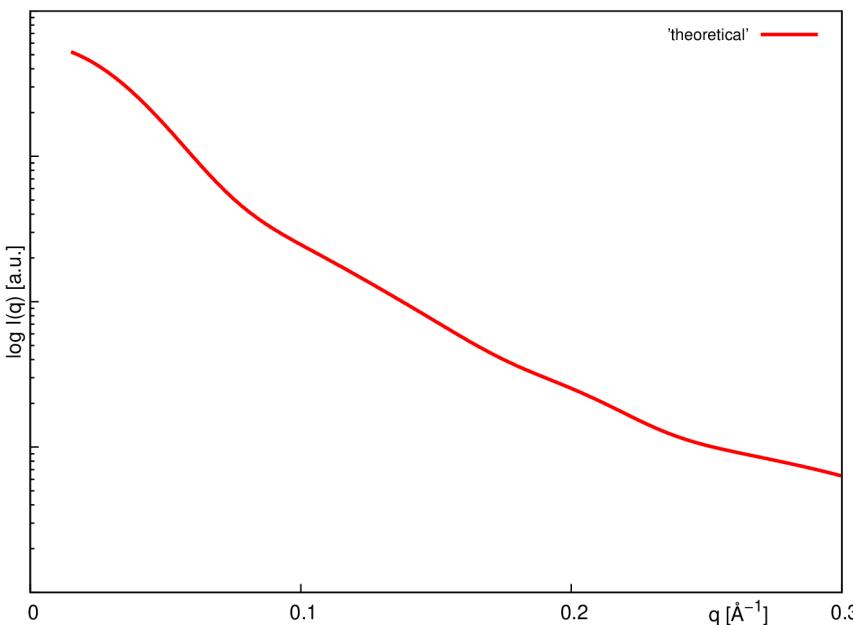
# SAXS – theoretical scattering Debye's formula



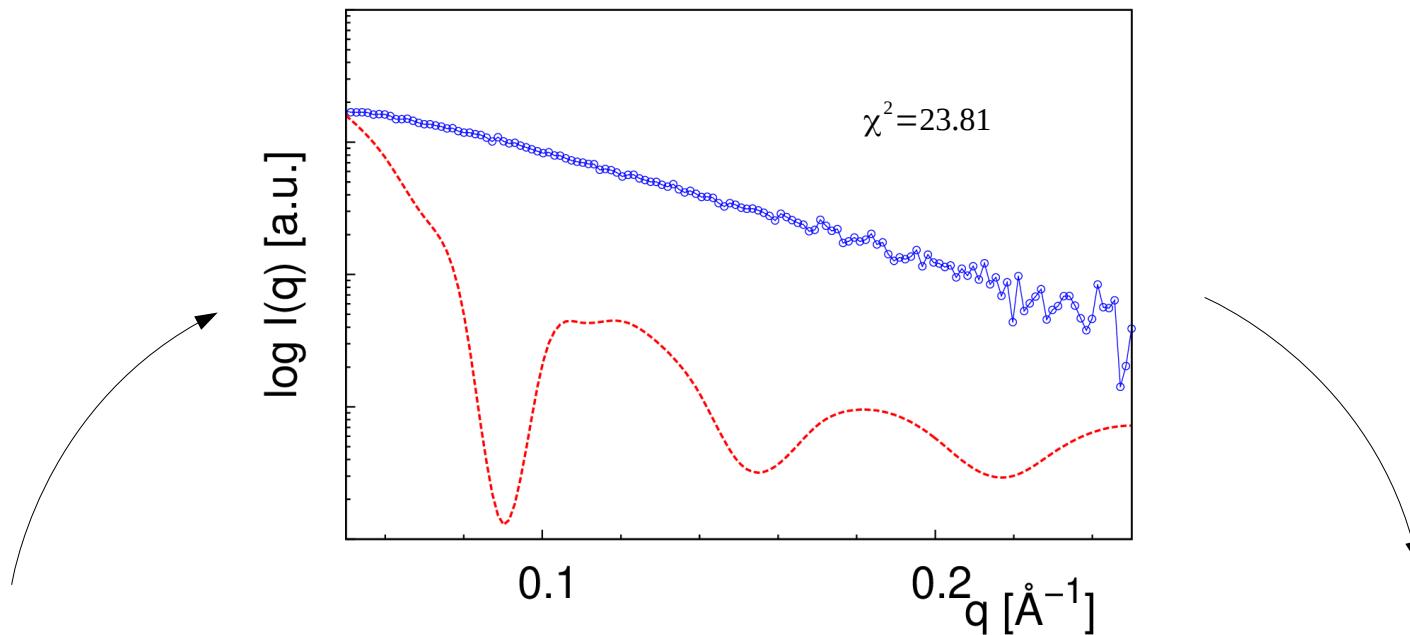
PDB coordinates



$$I(q) = \sum_{i=1}^{N_{\text{sph}}} F_i^2(q) + 2 \sum_{i=1}^{N_{\text{sph}}-1} \sum_{j=i+1}^{N_{\text{sph}}} F_i(q) F_j(q) \frac{\sin(qr_{ij})}{qr_{ij}}$$



# SAXS – modeling

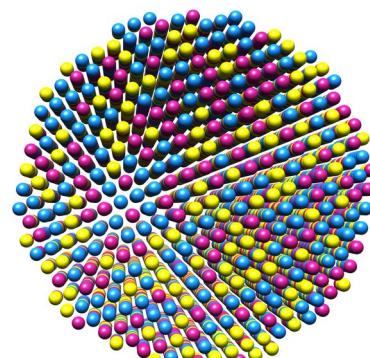


$$I(q) = \sum_{i=1}^{N_{sph}} F_i^2(q) + 2 \sum_{i=1}^{N_{sph}-1} \sum_{j=i+1}^{N_{sph}} F_i(q) F_j(q) \frac{\sin(qr_{ij})}{qr_{ij}}$$

Debye's formula

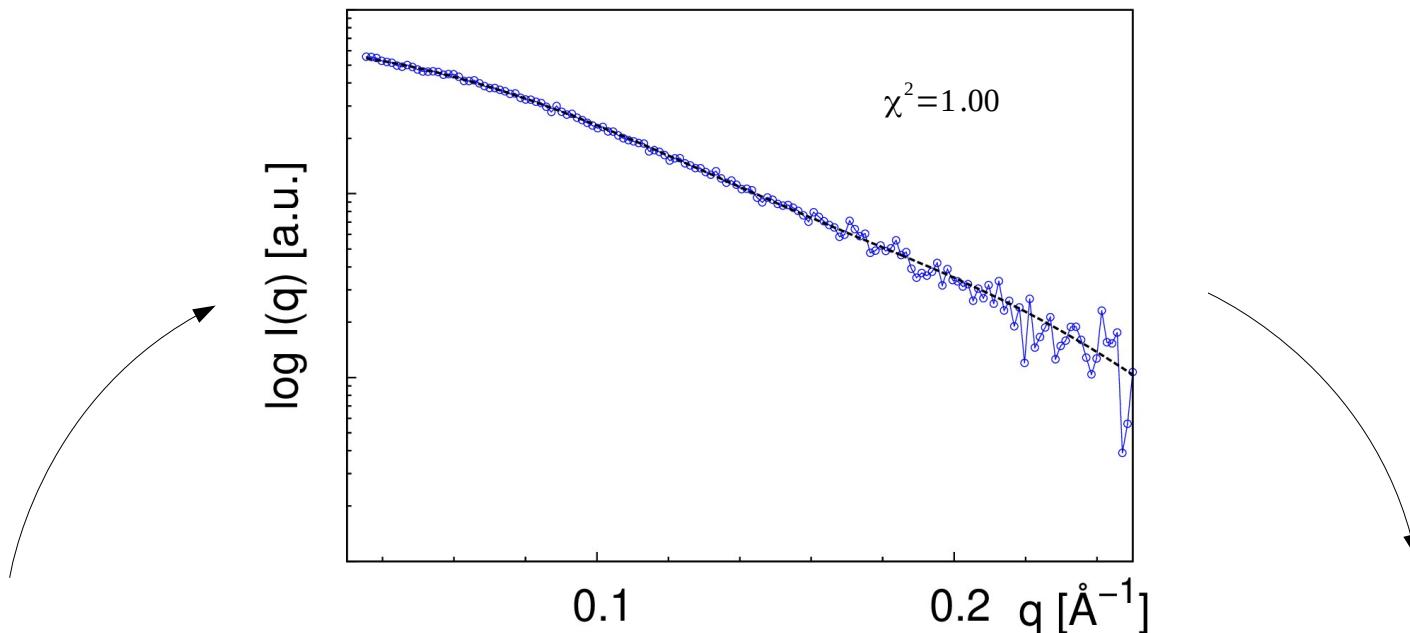
$$\chi^2 = \frac{1}{N_p} \sum_{k=1}^{N_p} \left[ \frac{I_{\text{exp}}(q_k) - c \cdot I_{\text{calc}}(q_k)}{\sigma_k} \right]^2$$

Quality of fit



**initial model**  
(ab initio, rigid body)

# SAXS – modeling

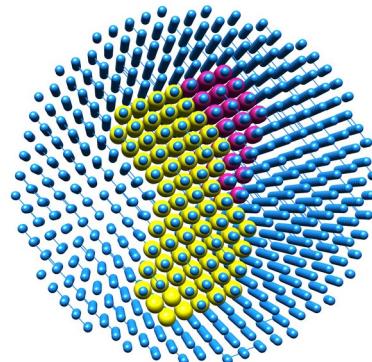


$$I(q) = \sum_{i=1}^{N_{sp}} F_i^2(q) + 2 \sum_{i=1}^{N_{sp}-1} \sum_{j=i+1}^{N_{sp}} F_i(q) F_j(q) \frac{\sin(qr_{ij})}{qr_{ij}}$$

Debye's formula

$$\chi^2 = \frac{1}{N_p} \sum_{k=1}^{N_p} \left[ \frac{I_{\text{exp}}(q_k) - c \cdot I_{\text{calc}}(q_k)}{\sigma_k} \right]^2$$

Quality of fit



**final model**  
(ab initio, rigid body)

## SAXS software

- ATSAS (Svergun group)
- SCATTER (Rambo group)
- <http://smallangle.org/content/Software#Model-Fitting>

# ATSAS software



Secure | <https://www.embl-hamburg.de/biosaxs/software.html> | EUROPÄISCHES LABORATORIUM FÜR MOLEKULARBIOLOGIE [DE] | <https://www.saxler.org/forum/>

**EMBL** Biological Small Angle Scattering 

**ATSAS software**

Home > ATSAS software

**Data analysis software ATSAS 2.8.0**  
A program suite for small-angle scattering data analysis from biological macromolecules

**Data processing**  
[PRIMUS](#) - manipulations with experimental 1D SAS data  
[GNOM](#) - indirect transform program that evaluates the particle distance distribution function p(r)  
[Data manipulation and analysis tools](#) - AUTORG, ALMERGE, DATGNOM, DATPOROD etc.

**Ab initio methods**  
[DAMMIN](#) - ab initio shape determination using a dummy atom model  
[DAMMIE](#) - rapid shape determination  
[GASBOR](#) - reconstruction of a protein structure by a chain-like ensemble of dummy residues  
[MONSA](#) - shape determination using a multiphase dummy atom model

**Rigid body modelling**  
[SASREF](#) - modelling of multisubunit complexes  
[BUNCH](#) - modelling of multidomain proteins against multiple data sets  
[CORAL](#) - modelling of multidomain protein complexes against multiple data sets  
[MASSHA](#) - interactive modelling of atomic structures and shape analysis  
[GLOBSYMM](#) - rigid body modelling of symmetric oligomers

**Mixtures and flexible systems**  
[OLIGOMER](#) - volume fractions of mixtures with known scattering intensities from the components  
[MIXTURE](#) - modelling of multicomponent systems  
[EOM](#) - Ensemble Optimization Method for flexible proteins  
[SREFLEX](#) - flexible refinement of high-resolution models combining SAXS and NMA

**PDB oriented tools**  
[CRYSL](#) - X-ray scattering patterns from known hi-res structures  
[CRYSON](#) - neutron scattering patterns from known hi-res structures  
[SUPCOMB](#) - superimposes one 3D structure onto another  
[DAMAVER](#) - align ab initio models, select the most typical one

**Manuals**

If you use ATSAS please cite:  
Petoukhov, M.V., Franke, D., Shkumatov, A.V., Tria, G., Kikhney, A.G., Gajda, M., Gorba, C., Mertens, H.D.T., Konarev, P.V. and Svergun, D.I. (2012) [New developments in the ATSAS program package for small-angle scattering data analysis](#) *J. Appl. Cryst.* 45, 342-350 © International Union of Crystallography DOI

**SAXS**  
Small Angle X-ray Scattering Initiative for Europe :: Forum

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- [SAS Experiment Design](#)  
Plan your small angle X-ray or neutron scattering measurements
- [Literature](#)  
Books and reviews on small angle scattering. Discussions and criticism on recent SAXS papers.
- [Feedback](#)  
Suggestions for new topics to discuss, questions on the use of the Forum

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- [Ab Initio Shape Determination](#)  
Ab initio modeling: DAMMIF, DAMMIN, GASBOR, MONSA
- [Rigid Body Modelling](#)  
Interactive modelling (MASSHA, SASPy) and global minimization programs (SASREF, BUNCH, CORAL, GLOBSYMM)
- [Mixtures and Flexible Systems](#)  
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**Other Software**

- [SAXS & SANS Software](#)  
Small angle scattering software (except ATSAS)

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# ATSAS software



Secure | <https://www.embl-hamburg.de/biosaxs/software.html>

**EMBL** Biological Small Angle Scattering 

Home > ATSAS software

**Group members**

**ATSAS software**  
Download

BUNCH  
CORAL  
CRY SOL  
CRYSON  
DAMAVER  
DAMMIF  
DAMMIN  
DATtools  
EOM  
GASBOR  
GLOBSYMM  
GNOM  
MASSHA  
MIXTURE  
MONSA  
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**If you use ATSAS please cite:**  
Petoukhov, M.V., Franke, D., Shkumatov, A.V., Tria, G., Kikhney, A.G., Gajda, M., Gorba, C., Mertens, H.D.T., Konarev, P.V. and Svergun, D.I. (2012) *New developments in the ATSAS program package for small-angle scattering data analysis*. *J. Appl. Cryst.* 45, 342-350 © International Union of Crystallography DOI

Last modified: October 8, 2015

JM FÜR MOLEKULARBIOLOGIE [DE] | <https://www.saxler.org/forum/>

**SAXS**  
Small Angle X-ray Scattering Initiative for Europe :: Forum

Forum

Manipulation tools, add-ons (Raman spectrometer, HPLC), gas phase scattering etc.

Measurements

Discussions and criticism on recent SAXS papers.

The use of the Forum

The scattering analysis program package

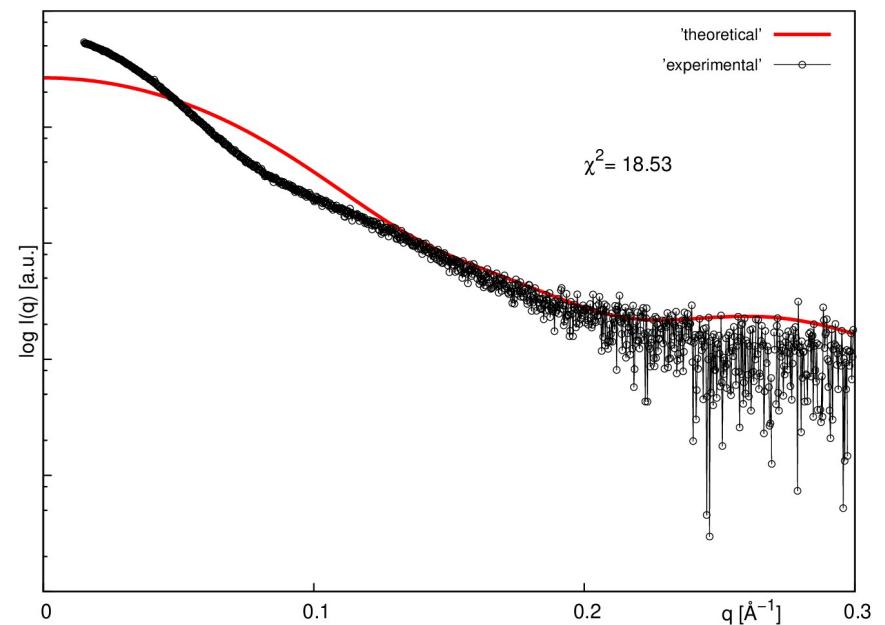
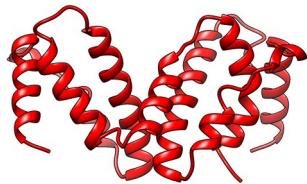
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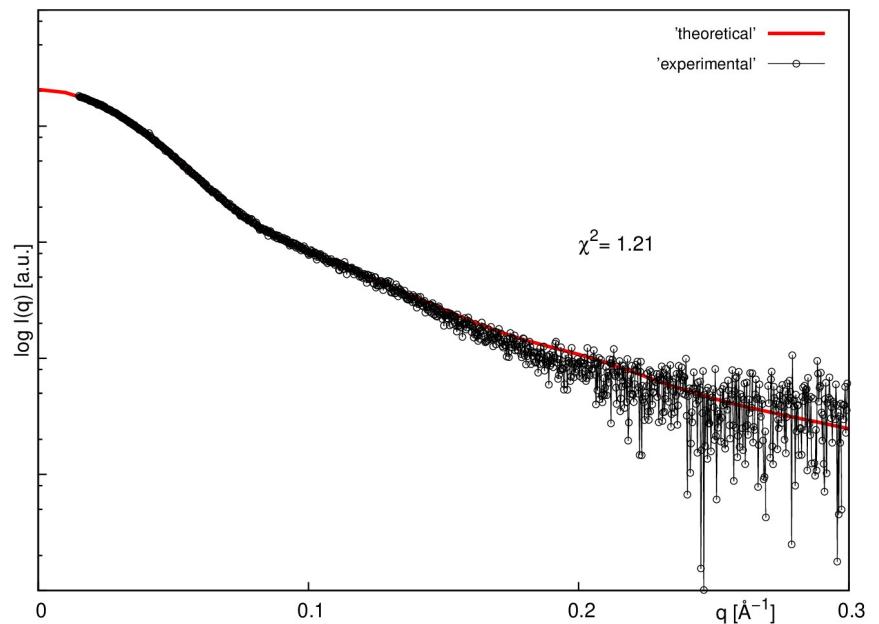
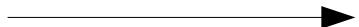
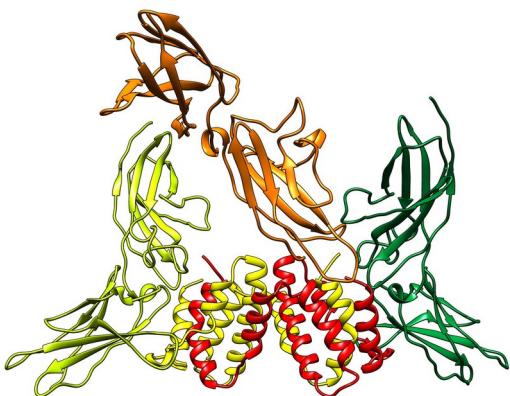
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- evaluation of solution scattering of known atomic structure
- fitting to experimental data
- validation tool

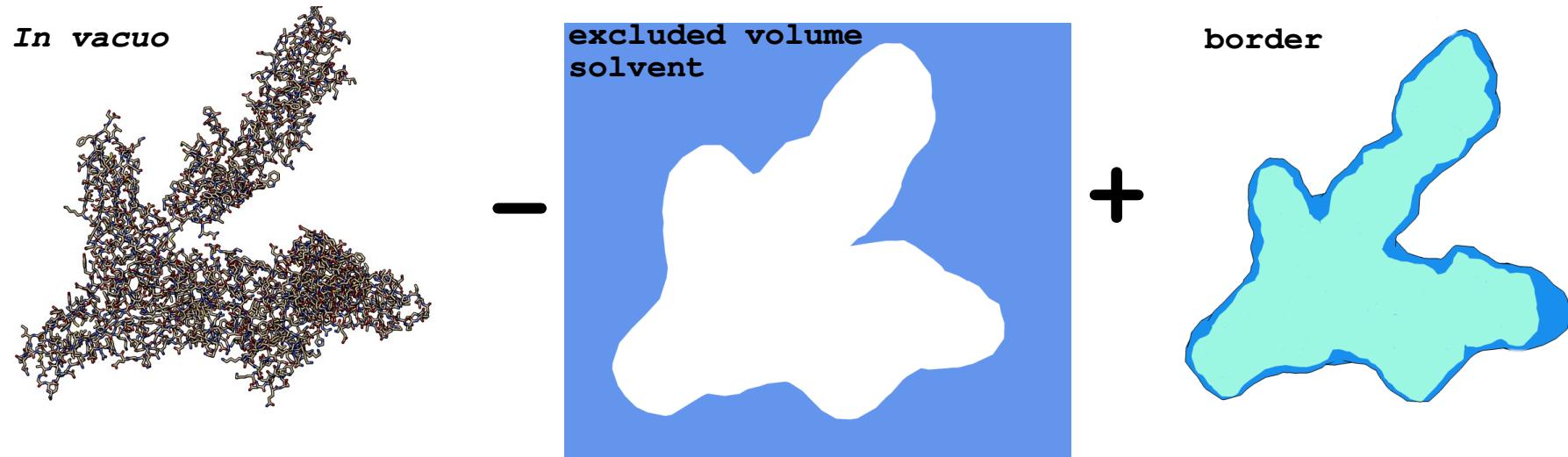


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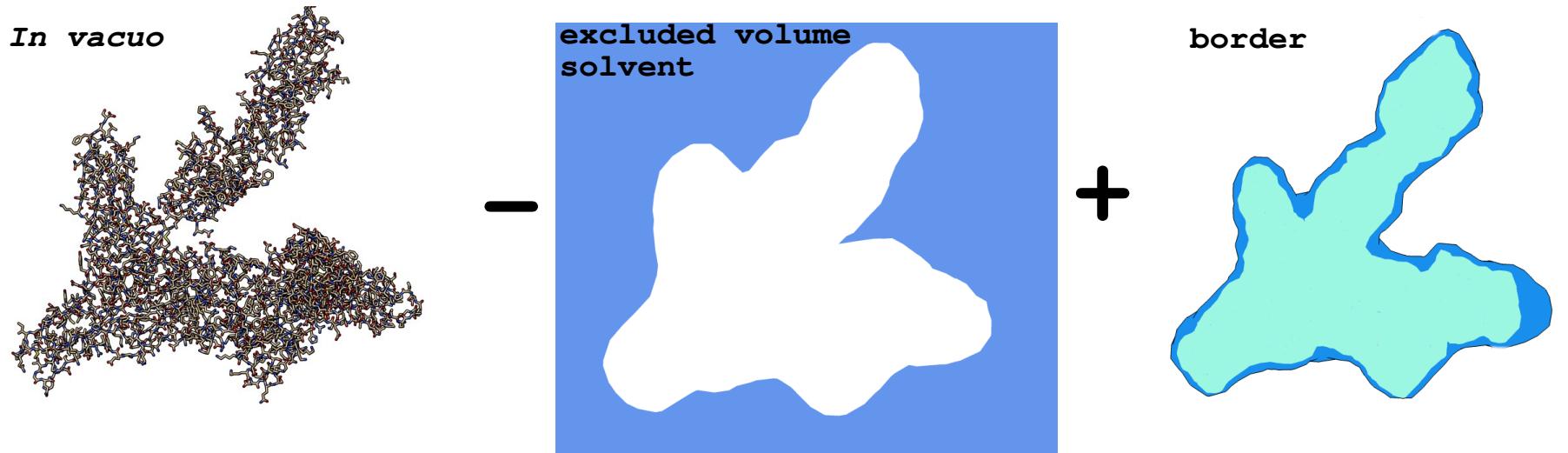


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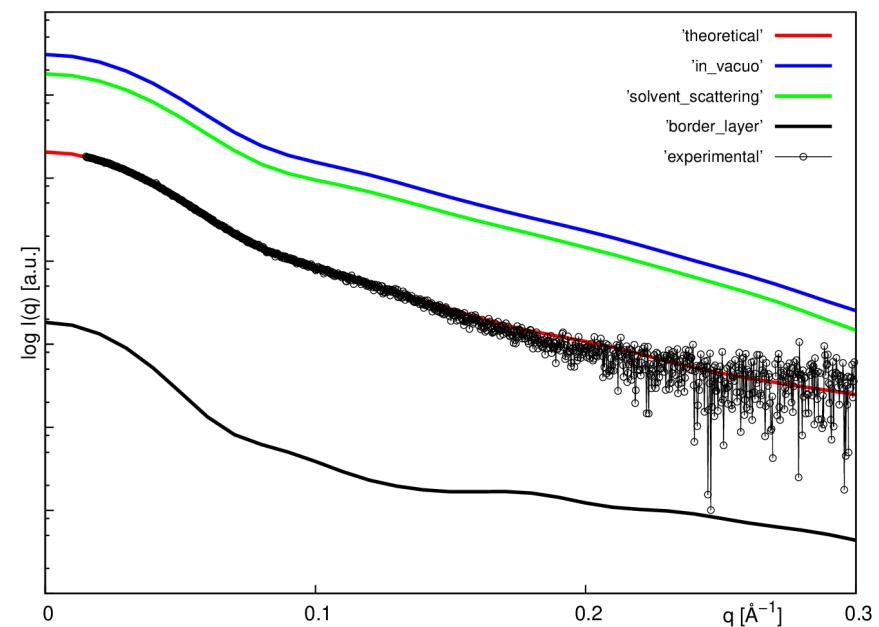
$$I(q) = \langle |A_a(q) - \rho_s A_s(q) + \delta \rho_b A_b(q)|^2 \rangle_{\Omega}$$

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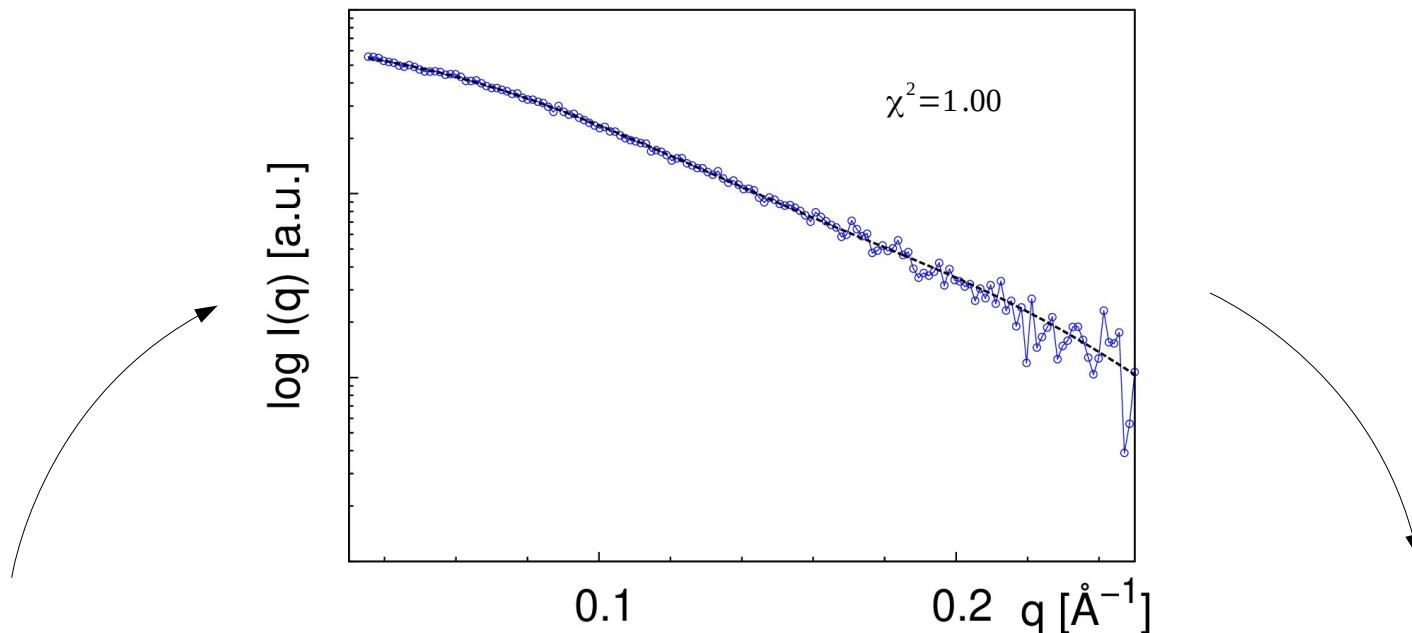


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$$\chi^2 = \frac{1}{N_p} \sum_{k=1}^{N_p} \left[ \frac{I_{\text{exp}}(q_k) - c \cdot I_{\text{calc}}(q_k, r_0, \delta \rho)}{\sigma_k} \right]^2$$



# SAXS – modeling

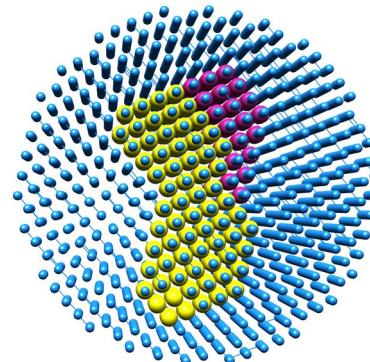


$$I(q) = \sum_{i=1}^{N_{sp}} F_i^2(q) + 2 \sum_{i=1}^{N_{sp}-1} \sum_{j=i+1}^{N_{sp}} F_i(q) F_j(q) \frac{\sin(qr_{ij})}{qr_{ij}}$$

Debye's formula

$$\chi^2 = \frac{1}{N_p} \sum_{k=1}^{N_p} \left[ \frac{I_{\text{exp}}(q_k) - c \cdot I_{\text{calc}}(q_k)}{\sigma_k} \right]^2$$

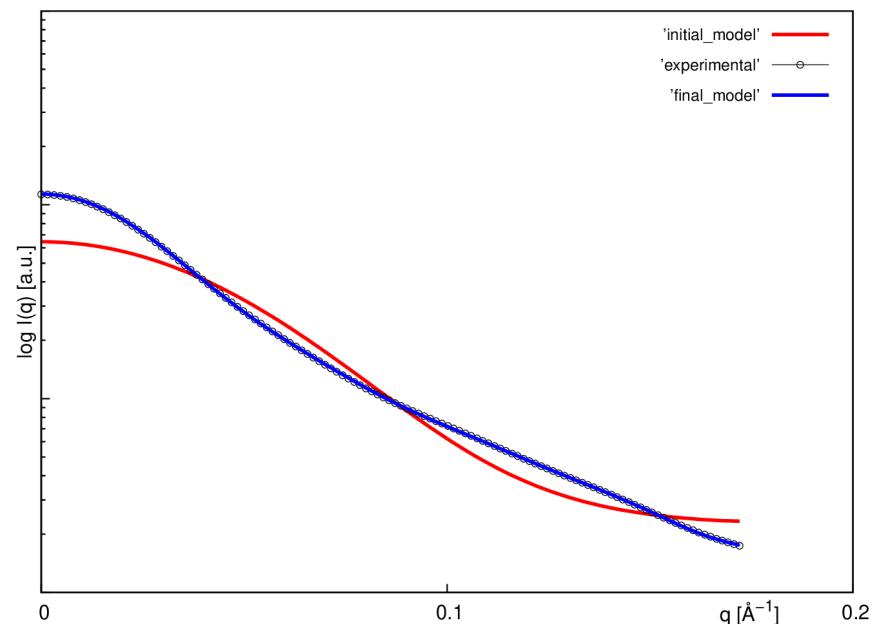
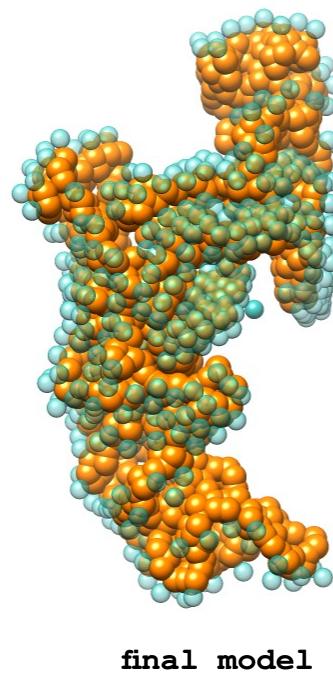
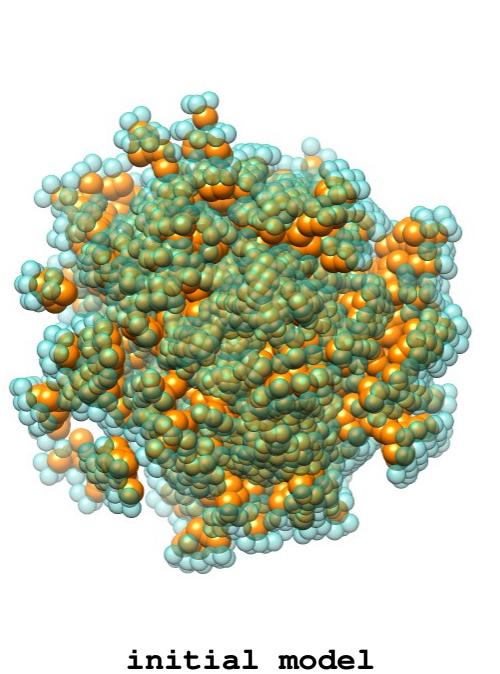
Quality of fit



**final model**  
(ab initio, rigid body)

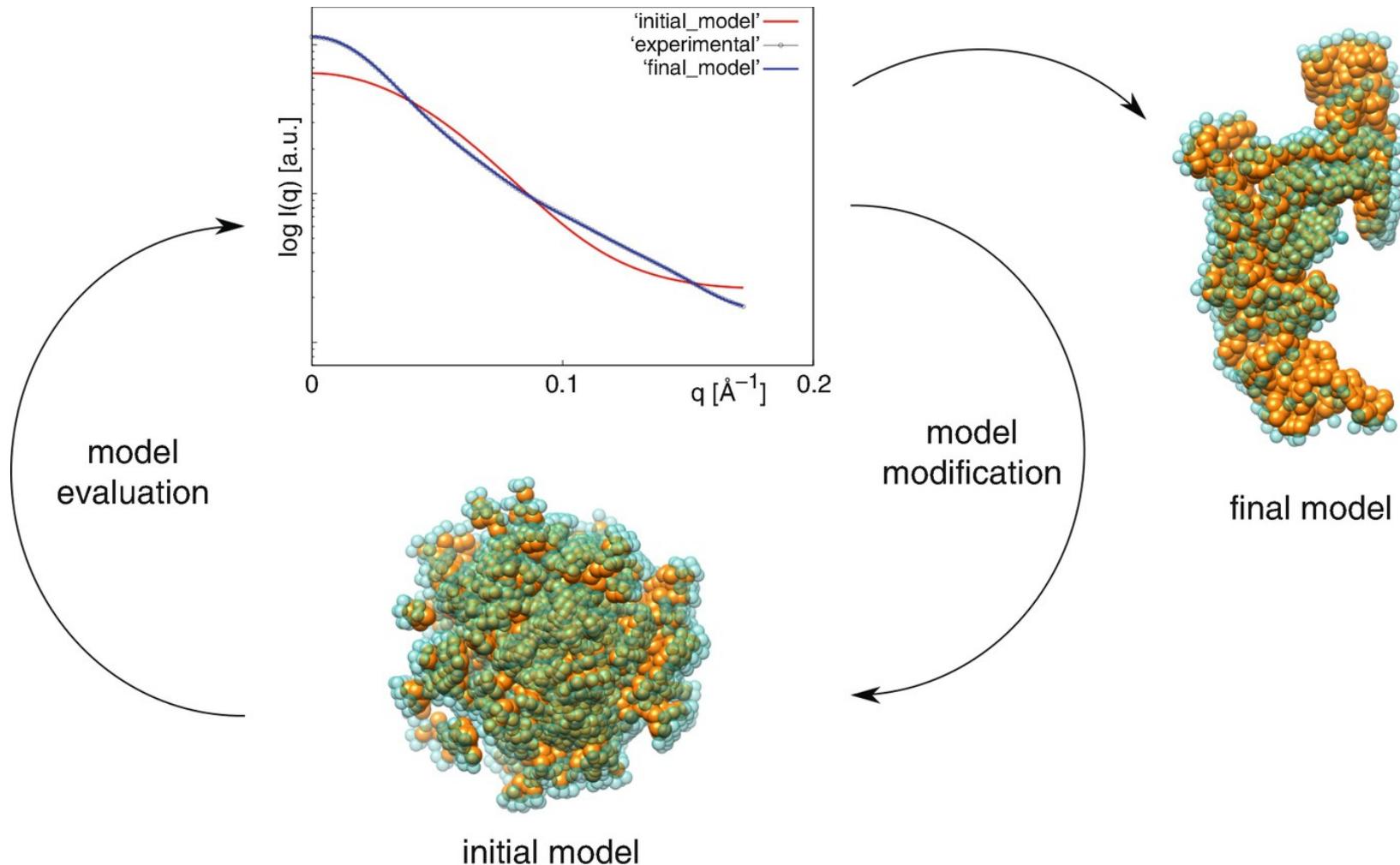
# *Ab initio* modeling dummy residues – GASBOR

- low resolution shape reconstruction
- dummy residues: spheres with electron density as average AA residue
- distance constrain 3.8 Å as approx. Cα-Cα position
- fixed number of dummy residues (protein sequence)



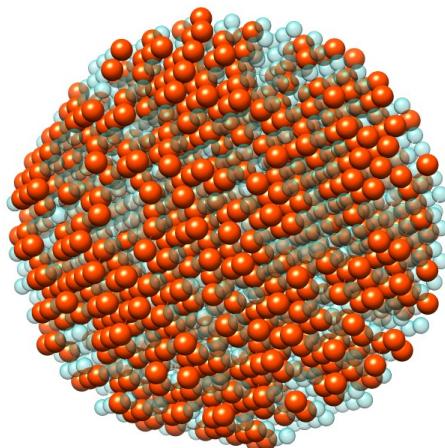
- simulated annealing method to minimize goal function:  $f(x) = \chi^2 + \sum \alpha P(x)$
- penalties: bond length, histogram distances, peripheral, discontiguity

# *Ab initio* modeling dummy residues - GASBOR



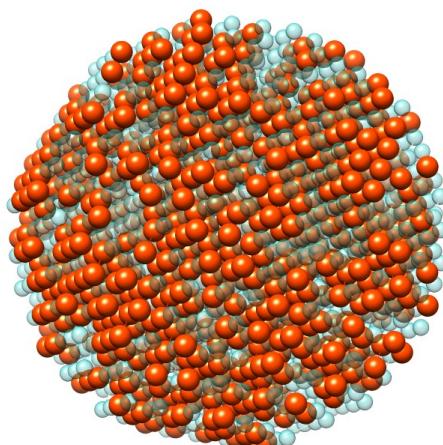
## *Ab initio* modeling dummy atoms – DAMMIN

- low resolution shape reconstruction
- dummy atoms – densely packed spheres (beads)
- search volume  $\sim D_{\max}$ , radius of beads  $\ll D_{\max}$
- two phases: solute (protein) and solvent (water)

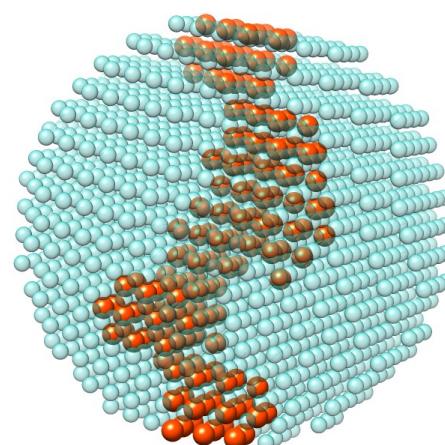


# *Ab initio* modeling dummy atoms – DAMMIN

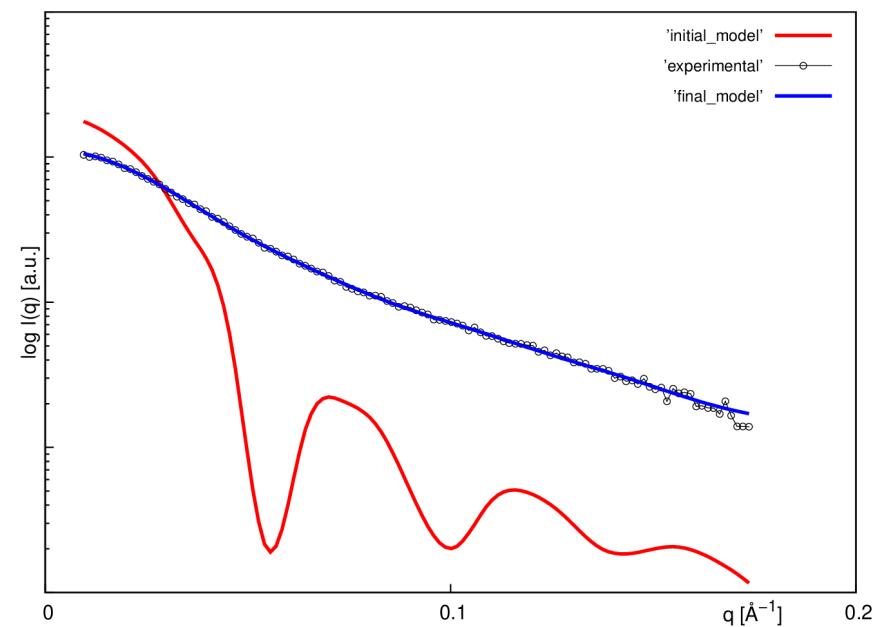
- low resolution shape reconstruction
- dummy atoms – densely packed spheres (beads)
- search volume  $\sim D_{\text{max}}$ , radius of beads  $\ll D_{\text{max}}$
- two phases: solute (protein) and solvent (water)



initial model

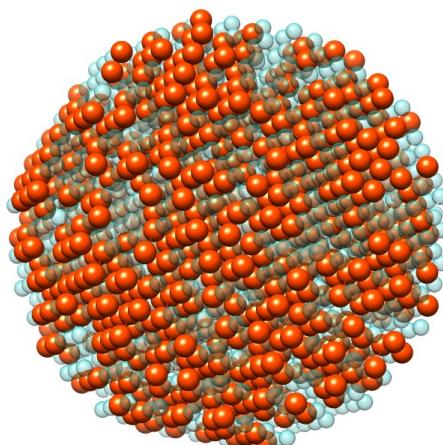


final model

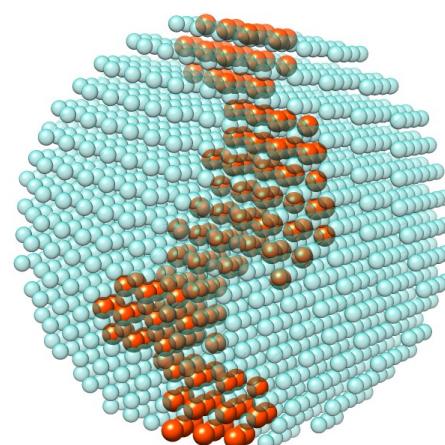


# *Ab initio* modeling dummy atoms – DAMMIN

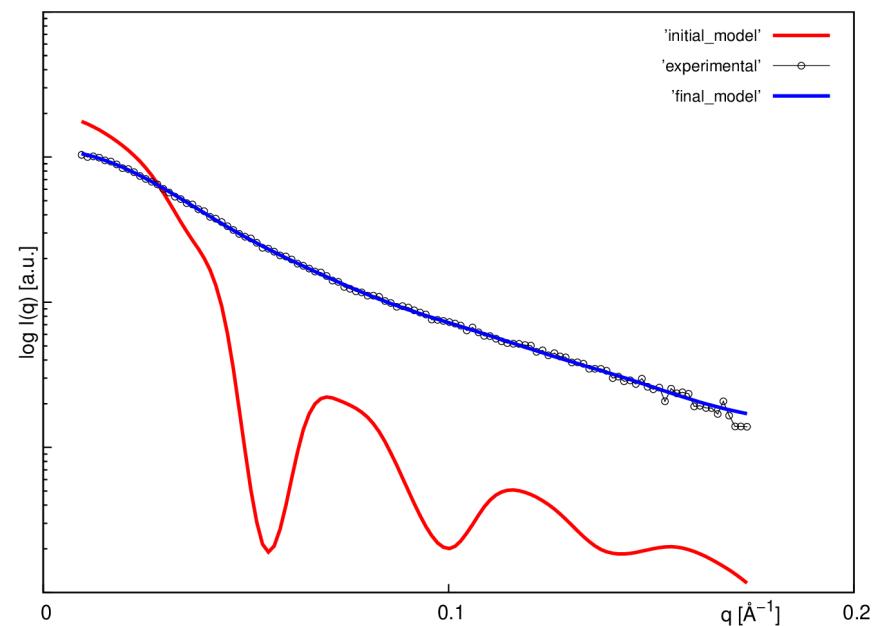
- low resolution shape reconstruction
- dummy atoms – densely packed spheres (beads)
- search volume  $\sim D_{\text{max}}$ , radius of beads  $\ll D_{\text{max}}$
- two phases: solute (protein) and solvent (water)



initial model



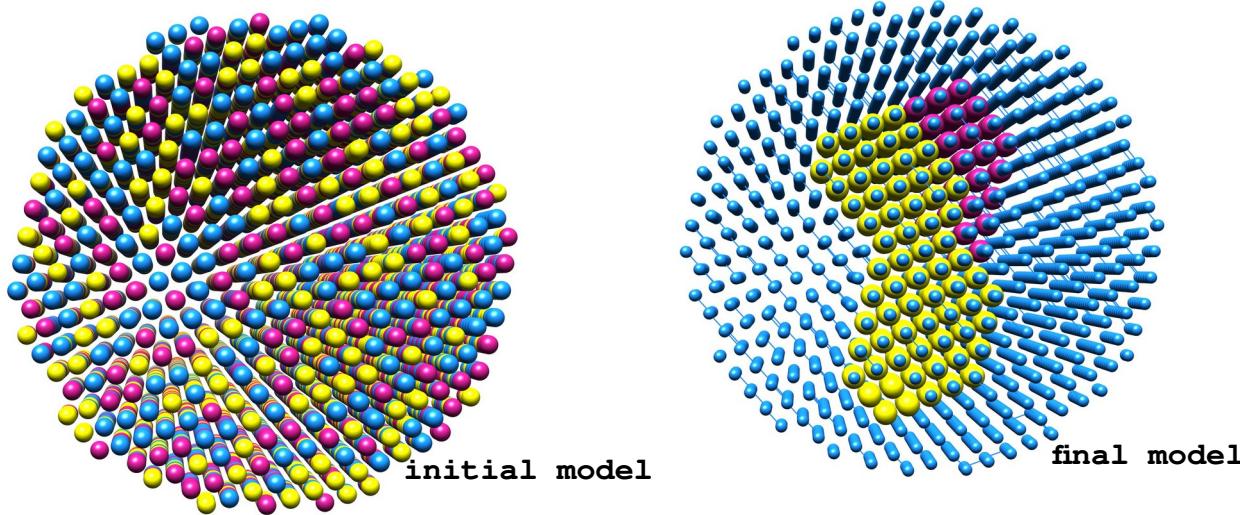
final model



- simulated annealing method to minimize goal function:  $f(x)=\chi^2+\sum \alpha P(x)$
- penalties: looseness, disconnectivity, peripheral

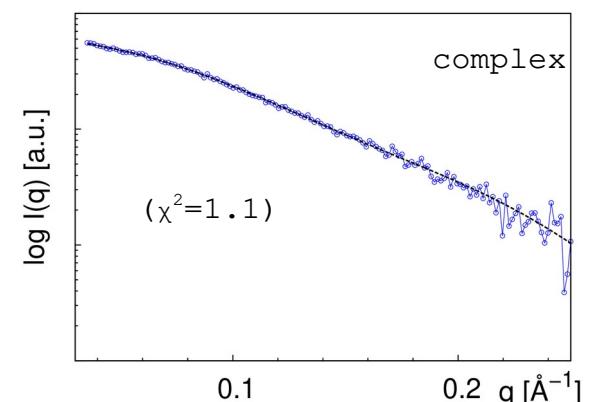
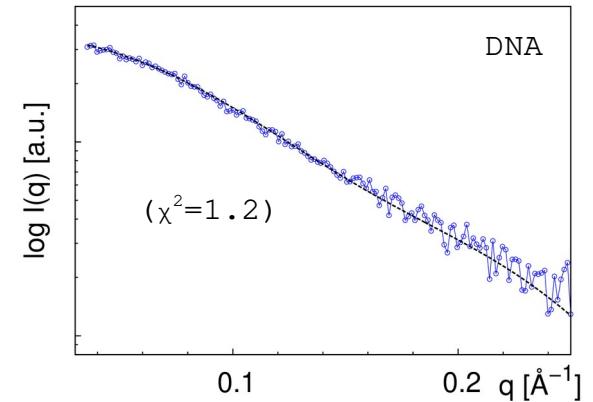
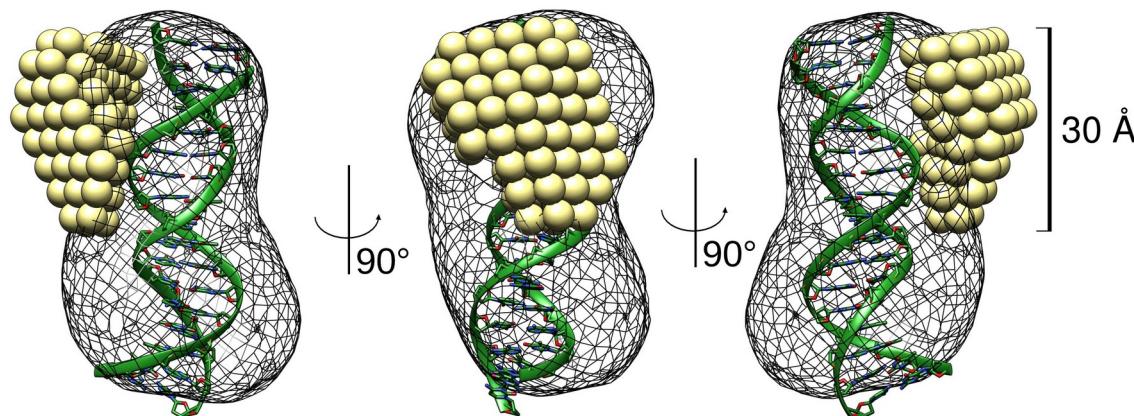
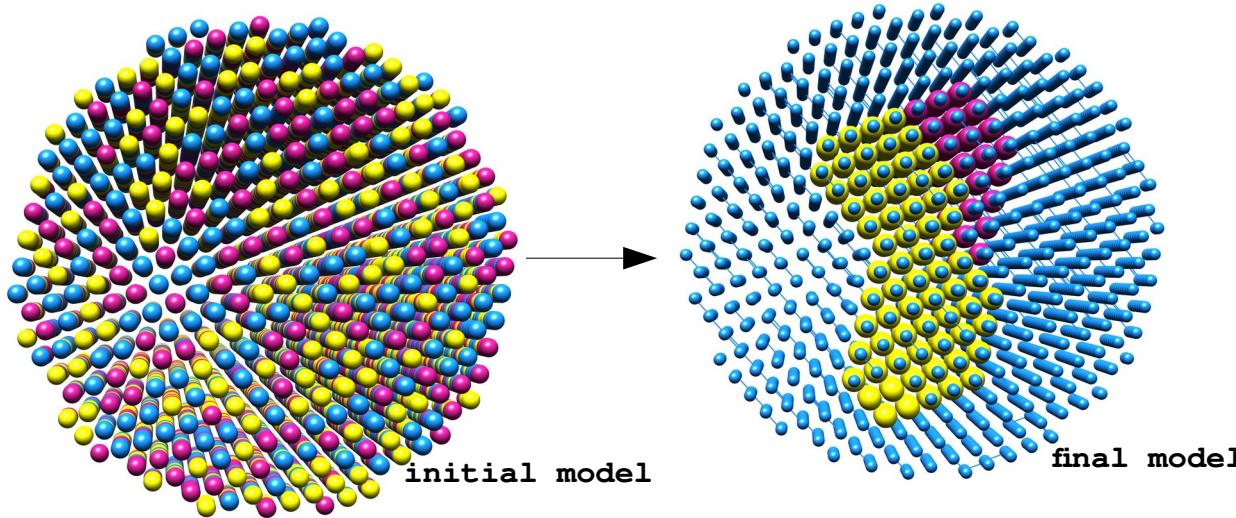
# *Ab initio* multi-phase modeling dummy atoms – MONSA

- low resolution shape reconstruction
- dummy atoms – densely packed spheres (beads)
- search volume  $\sim D_{\text{max}}$ , radius of beads  $\ll D_{\text{max}}$
- up to 4 phases: protein-protein complexes, protein::nucleic acid complexes



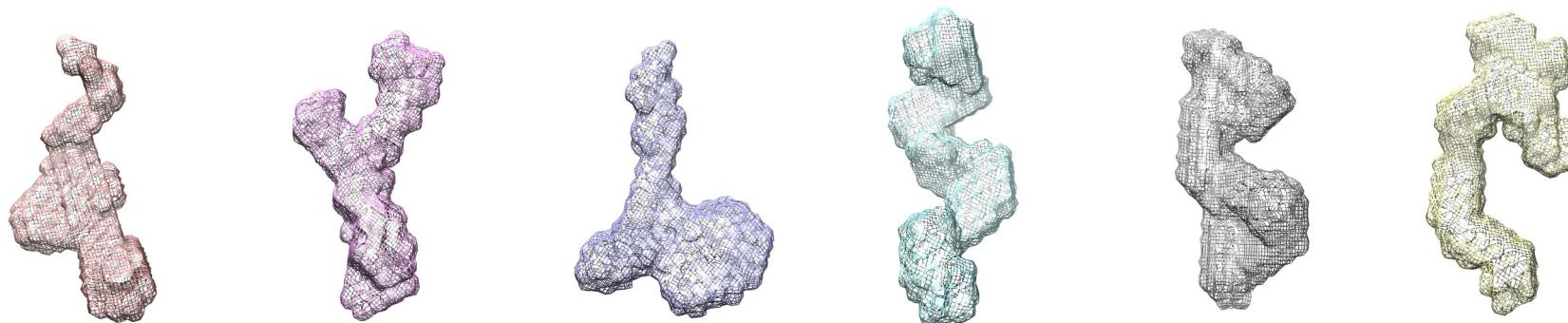
# *Ab initio* multi-phase modeling dummy atoms – MONSA

- low resolution shape reconstruction
- dummy atoms – densely packed spheres (beads)
- search volume  $\sim D_{\text{max}}$ , radius of beads  $\ll D_{\text{max}}$
- up to 4 phases: protein-protein complexes, protein::nucleic acid complexes

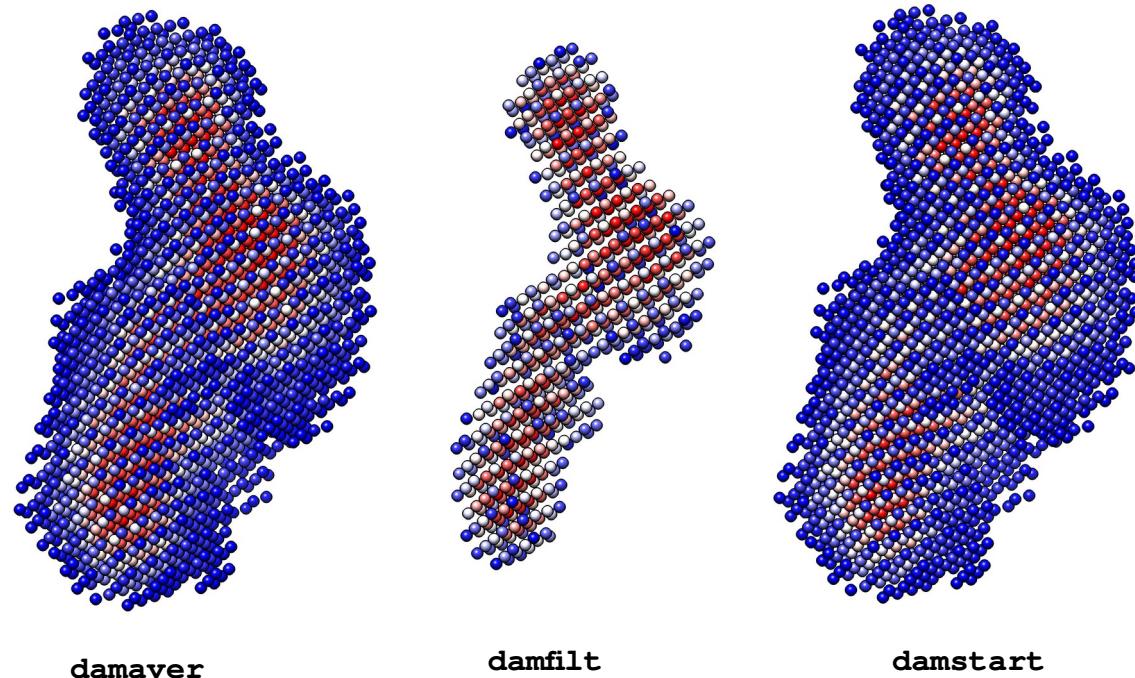


# *Ab initio* modeling refinement

- multiple runs of ab initio reconstruction give different results
- refinement: alignment, averaging, clustering, most typical model
- dummy atoms/res. with highest 'occupancy' used as starting structure



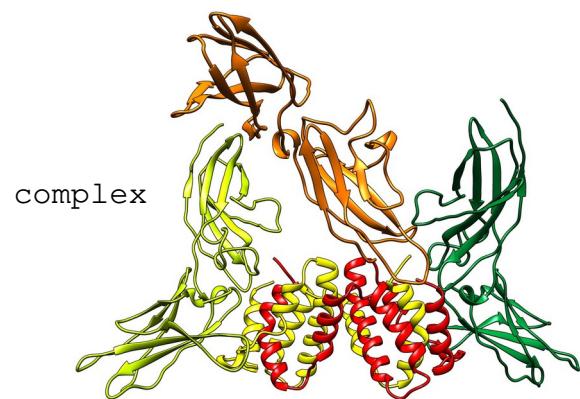
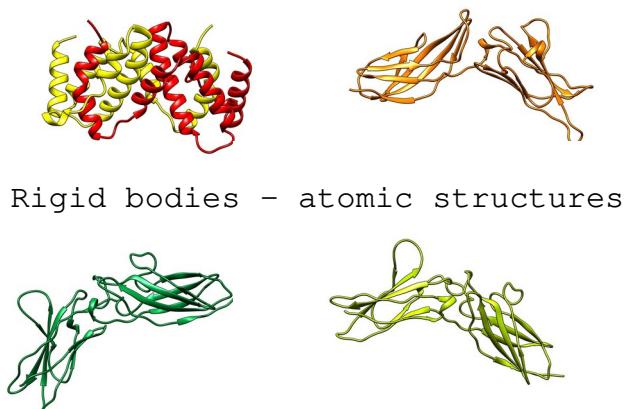
Recommendation	NSD	File
Include	0.735	dammif3.pdb
Include	0.739	dammif1.pdb
Include	0.739	dammif8.pdb
Include	0.740	dammif4.pdb
Include	0.750	dammif9.pdb
Include	0.752	dammif7.pdb
Include	0.754	dammif10.pdb
Include	0.756	dammif6.pdb
Include	0.756	dammif2.pdb
Discard	0.851	dammif5.pdb



# Rigid body modeling

## CORAL

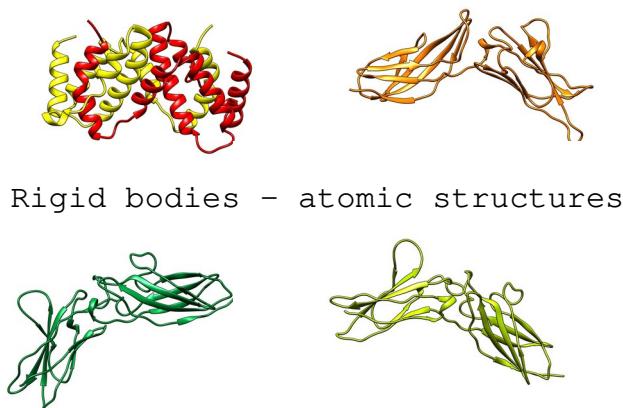
- Complexes with Random Loops
- translation and rotation search
- library of random loops composed of dummy residues (5-100AA)
- contact constrains, if known
- penalties: clashes, shift, contacts



# Rigid body modeling

## CORAL

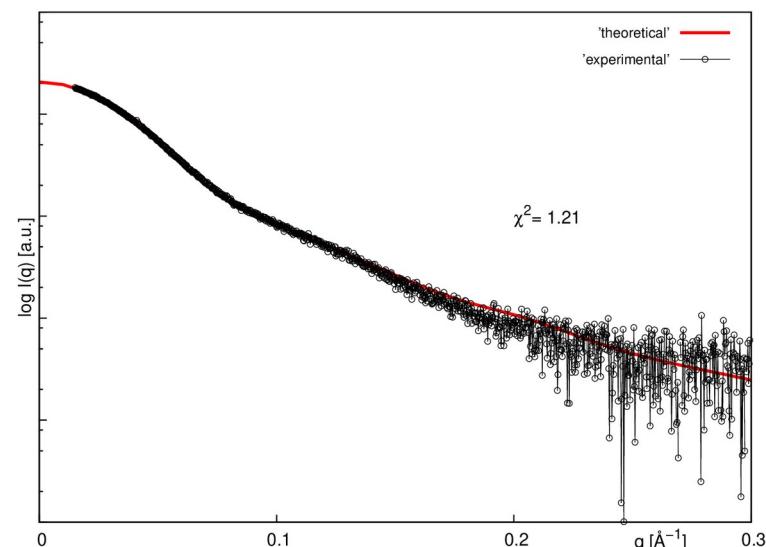
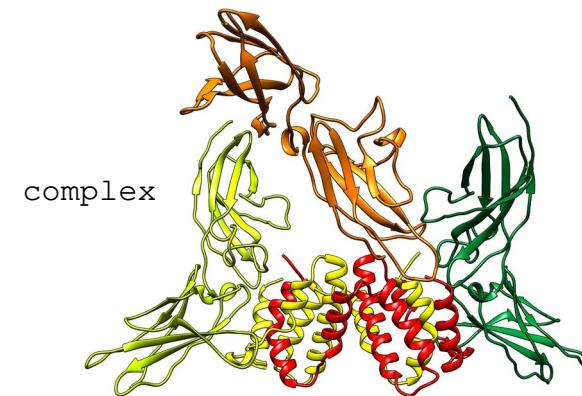
- Complexes with Random Loops
- translation and rotation search
- library of random loops composed of dummy residues (5-100AA)
- contact constrains, if known
- penalties: clashes, shift, contacts



**Simulated annealing**

Aim: find vector of M variables minimizing a function  $f(x)$

1. start from random configuration  $x$  at high temperature  $T$
2. make a small random modification of configuration  $x \rightarrow x'$  and compute the difference  $\Delta = f(x') - f(x)$
3. if  $\Delta > 0$ , accept it with probability  $e^{-\Delta/T}$
4. make another step from previous configuration  
if previous step was rejected  
or from new configuration, if was accepted
5. repeat 2-4 many times, then decrease the temperature
6. continue the cooling until no improvement is observed

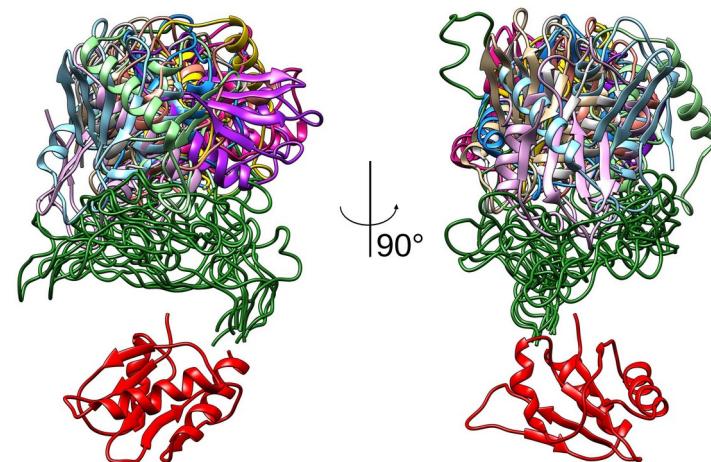
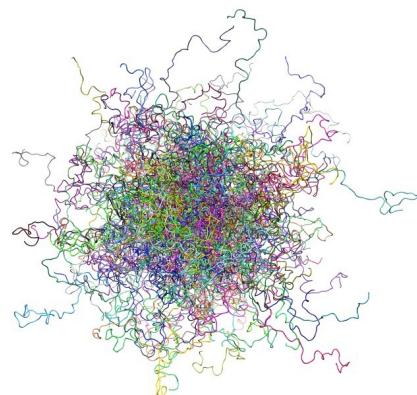
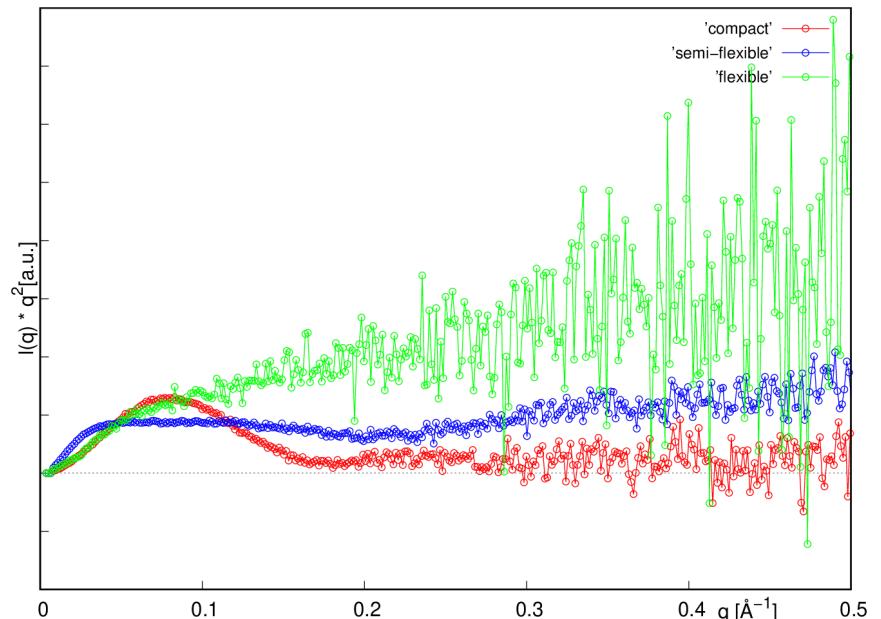


# Flexible systems

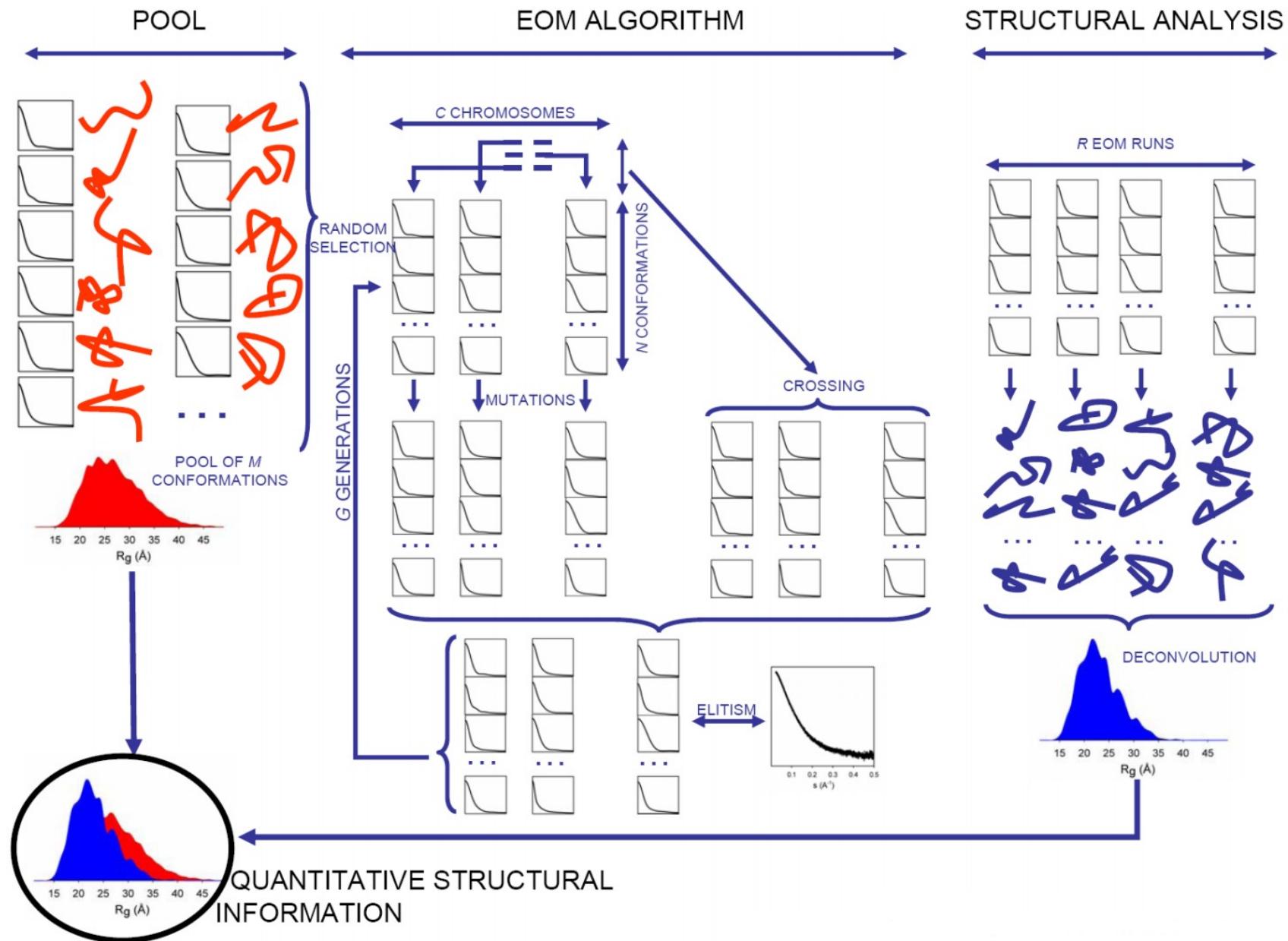
Conformational polydispersity  
Fully or partially unstructured chains

- conformational polydispersity
- fully/partially unstructured chains
- ensemble modeling (selection)

- multidomain proteins with flexible linkers, IDPs

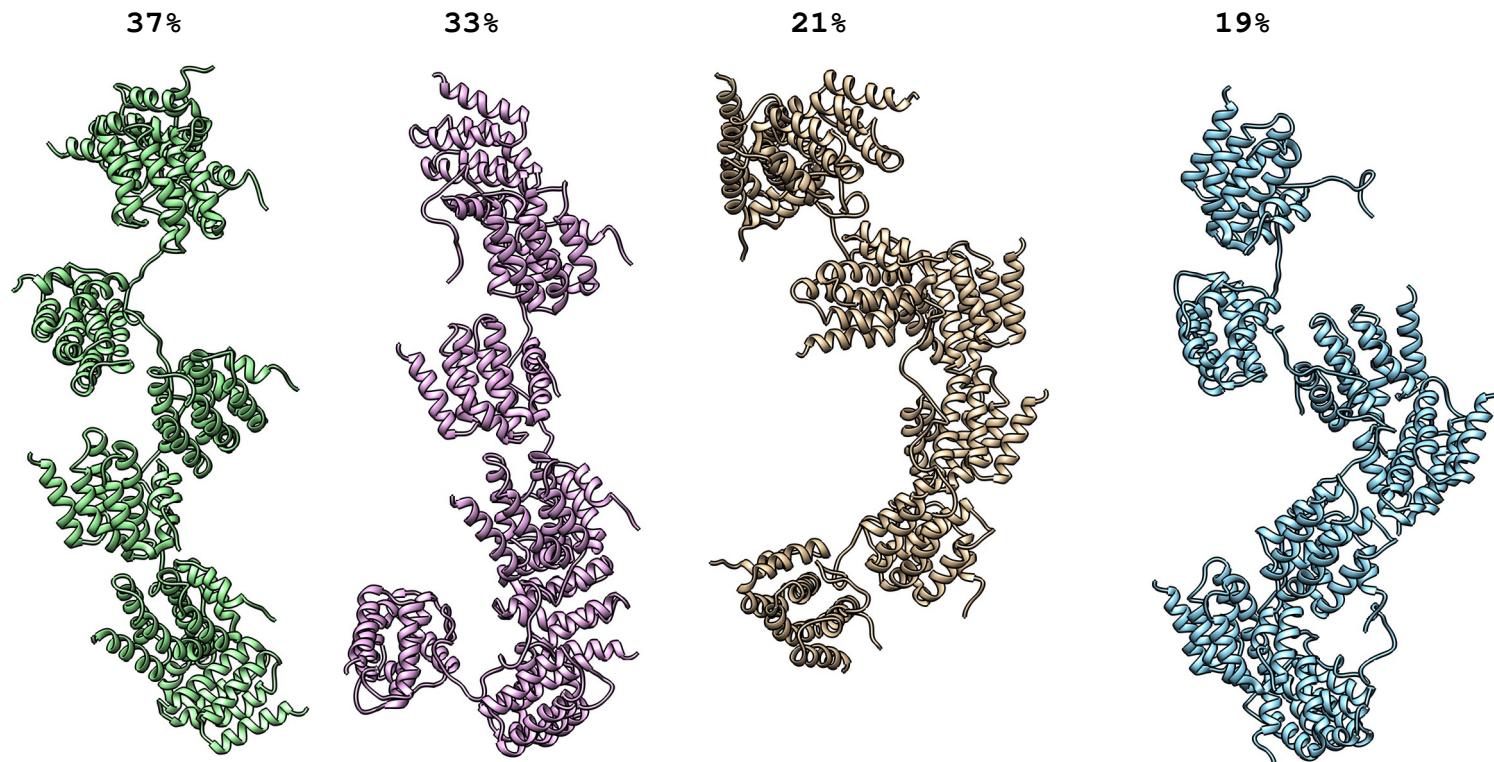
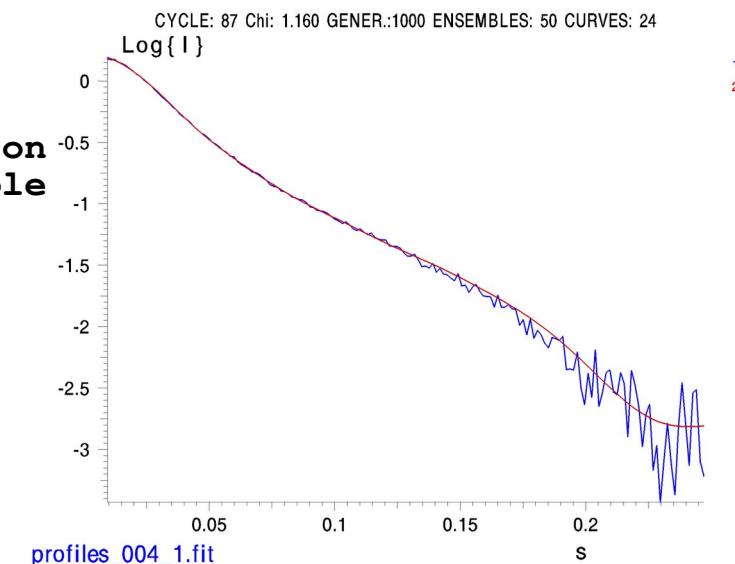


# Ensemble modeling EOM

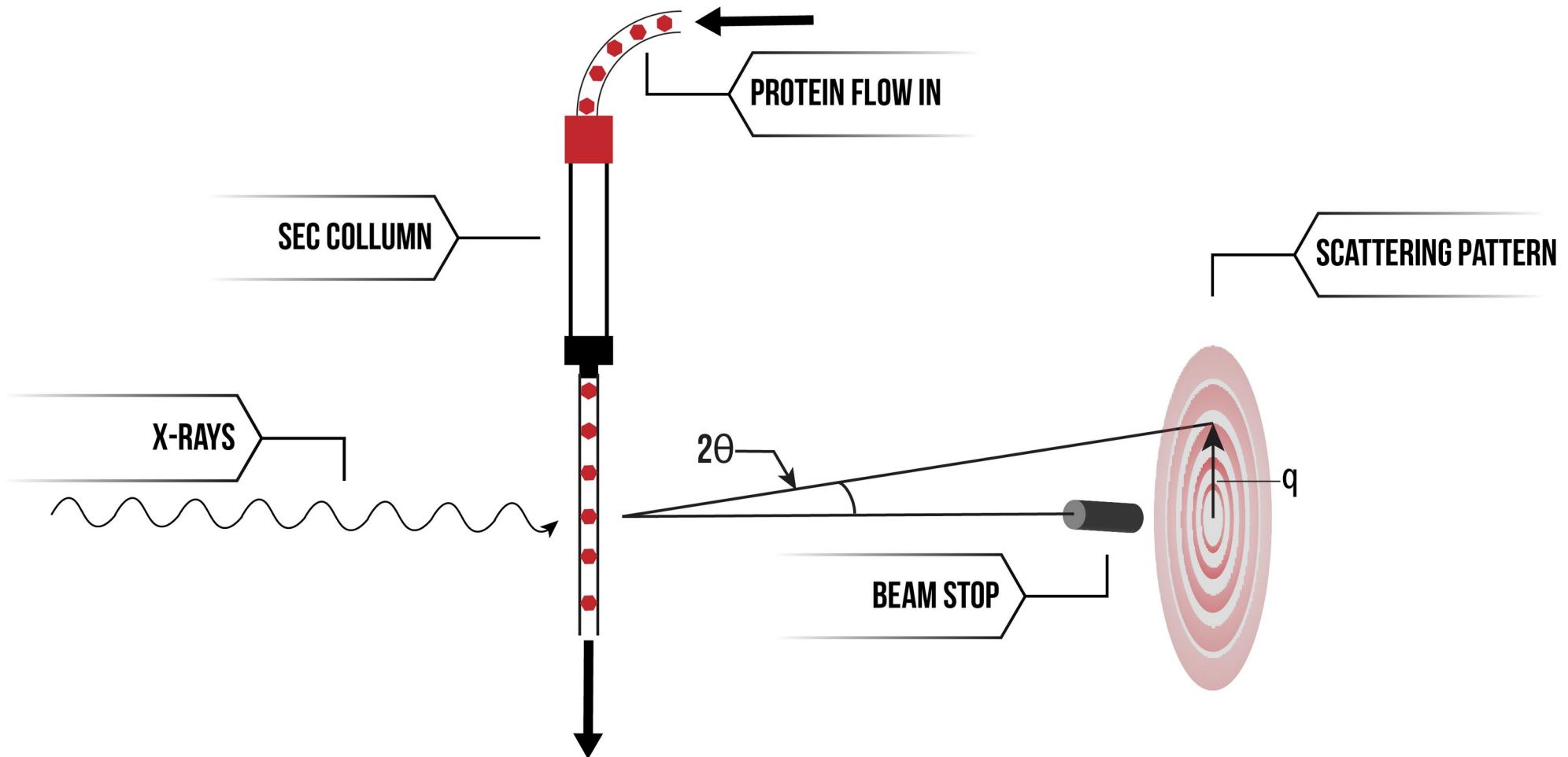


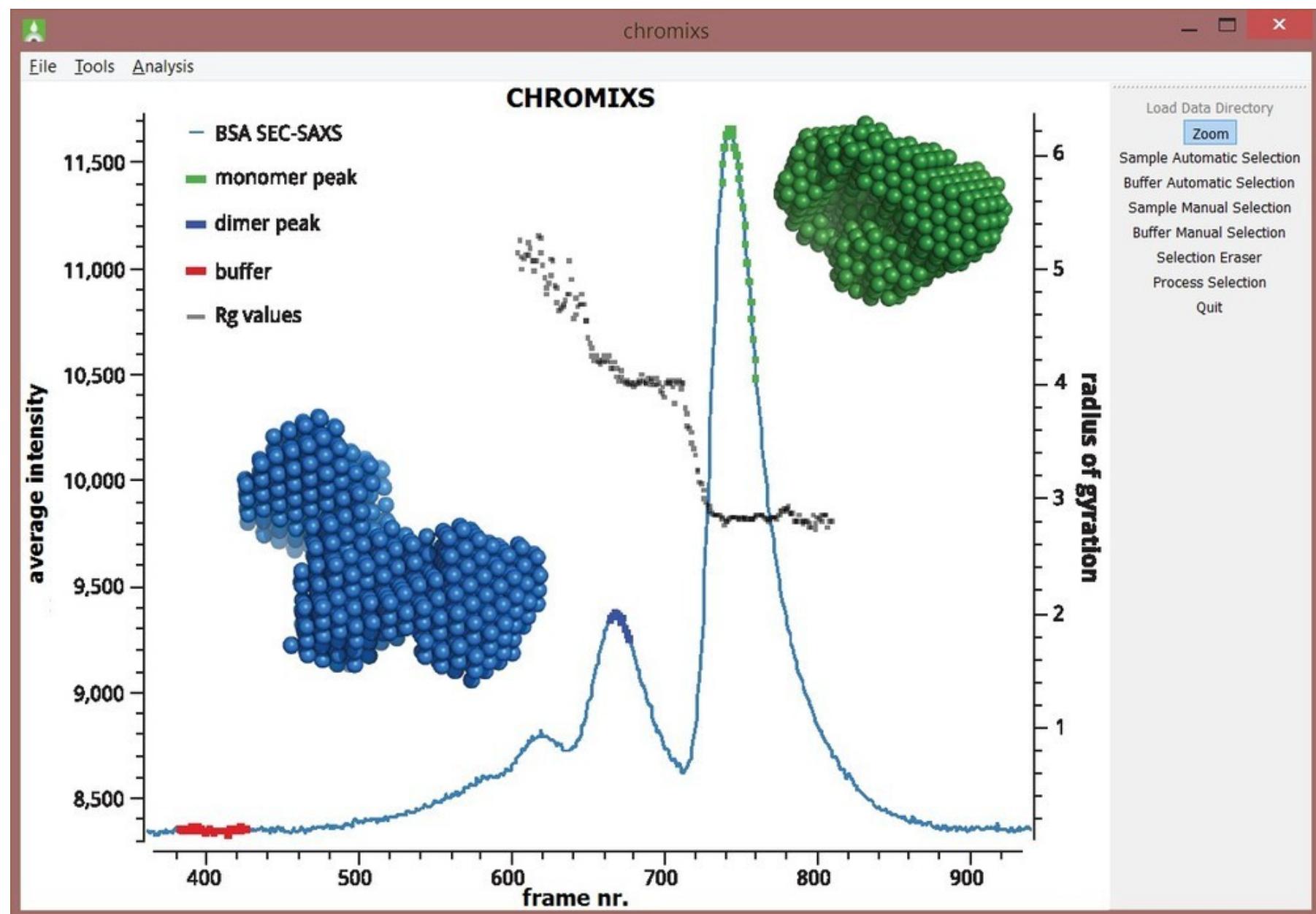
# Ensemble modeling EOM

- ensemble representation of possible flexibility
- model conformations not necessarily exists in solution
- $R_g$  and  $D_{max}$  distribution of pool and selected ensemble



# SEC-SAXS





**Group members****ATSAS software**

Download

BUNCH

CORAL

CRY SOL

CRYSON

DAMAVER

DAMMIF

DAMMIN

DATtools

EOM

GASBOR

GLOBSYMM

GNOM

MASSHA

MIXTURE

MONSA

OLIGOMER

PEAK

PRIMUS

SASFLOW

SASREF

SREFLEX

SUPCOMB

Manuals

**Web services****Facilities****Courses****Contact us**

## Data analysis software ATSAS 2.8.0

### A program suite for small-angle scattering data analysis from biological macromolecules

#### Data processing

[PRIMUS](#) - manipulations with experimental 1D SAS data[GNOM](#) - indirect transform program that evaluates the particle distance distribution function  $p(r)$ [Data manipulation and analysis tools](#) - AUTORG, ALMERGE, DATGNOM, DATPOROD etc.

#### Ab initio methods

[DAMMIN](#) - *ab initio* shape determination using a dummy atom model[DAMMIF](#) - rapid shape determination[GASBOR](#) - reconstruction of a protein structure by a chain-like ensemble of dummy residues[MONSA](#) - shape determination using a multiphase dummy atom model

#### Rigid body modelling

[SASREF](#) - modelling of multisubunit complexes[BUNCH](#) - modelling of multidomain proteins against multiple data sets[CORAL](#) - modelling of multidomain protein complexes against multiple data sets[MASSHA](#) - interactive modelling of atomic structures and shape analysis[GLOBSYMM](#) - rigid body modelling of symmetric oligomers

#### Mixtures and flexible systems

[OLIGOMER](#) - volume fractions of mixtures with known scattering intensities from the components[MIXTURE](#) - modelling of multicomponent systems[EOM](#) - Ensemble Optimization Method for flexible proteins[SREFLEX](#) - flexible refinement of high-resolution models combining SAXS and NMA

#### PDB oriented tools

[CRY SOL](#) - X-ray scattering patterns from known hi-res structures[CRYSON](#) - neutron scattering patterns from known hi-res structures[SUPCOMB](#) - superimposes one 3D structure onto another[DAMAVER](#) - align *ab initio* models, select the most typical one

#### Manuals

#### If you use ATSAS please cite:

Petoukhov, M.V., Franke, D., Shkumatov, A.V., Tria, G., Kikhney, A.G., Gajda, M., Gorba, C., Mertens, H.D.T., Konarev, P.V. and Svergun, D.I. (2012) [New developments in the ATSAS program package for small-angle scattering data analysis](#)

J. Appl. Cryst. 45, 242–252

## Curated repository for small angle scattering data and models

Small angle scattering (SAS) of X-ray and neutrons provides structural information on biological macromolecules in solution at a resolution of 1-2 nm.

SASBDB is a fully searchable curated repository of freely accessible and downloadable experimental data, which are deposited together with the relevant experimental conditions, sample details, derived models and their fits to the data.

**SASBDB currently contains:**

**1262** experimental data sets

**1892** models

428 experimental data sets on hold

565 models on hold

### Recent depositions:

#### SASDGB6 – Resistance to inhibitors of cholinesterase 8 homolog A (Ric8A) miniGi complex



Sample: Resistance to inhibitors of cholinesterase 8 homolog A monomer, 56 kDa *Bos taurus* protein

MiniGi monomer, 25 kDa synthetic construct protein

$R_g$  Guinier 3.2 nm

$D_{max}$  10.7 nm

Buffer: 20 mM Tris, 150 mM KCl, 5 % glycerol, 1 mM TCEP, pH: 8

Experiment: SAXS data collected at BioCAT 18ID, Advanced Photon Source (APS), Argonne National Laboratory on 2018 Oct 27

Large-scale conformational rearrangement of the  $\alpha 5$ -helix of  $G\alpha$  subunits in complex with the guanine nucleotide exchange factor Ric8A. *J Biol Chem* (2019)  
Srivastava D, Artemyev NO

#### SASDG76 – Inhibitor of apoptosis



Flexible Tethering of ASPP Protein

#### SASDGJ4 – Human alpha-amino



Structural Analysis of Pathogenic

#### SASDFB6 – The periplasmal



Protease-associated import sys

#### SASDDR3 – Yeast tRNA Nm34



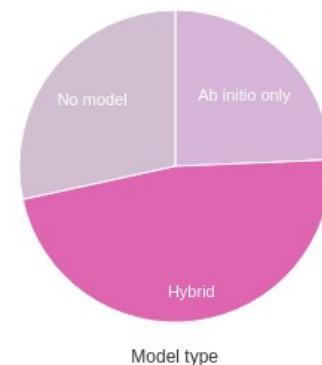
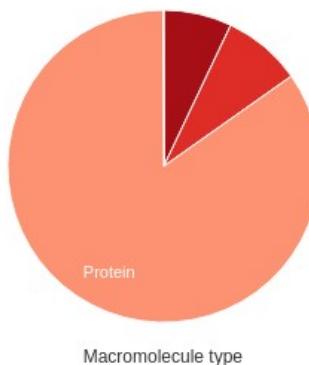
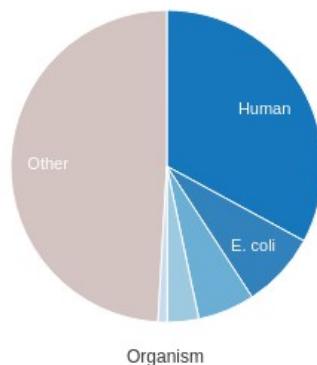
Structure of tRNA methyltransfer

#### SASDFL5 – Plasmodium falci



Structural studies of the Hsp70/H

### Browse the contents according to:



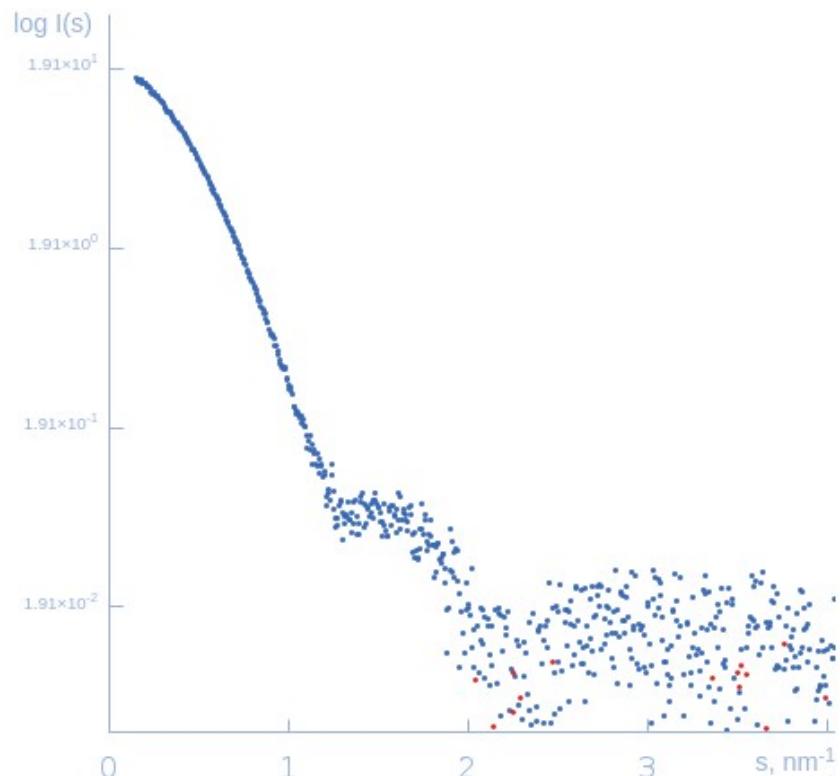
Combination of SAXS and Protein Painting Discloses the Three-Dimensional Organization of the Bacterial Cysteine Synthase Complex, a Potential Target for Enhancers of Antibiotic Action

Rosa B, Marchetti M, Paredi G, Amenitsch H, Franko N, Benoni R, Giabbai B, De Marino M, Mozzarelli A, Ronda L, Storici P, Campanini B, Bettati S, *International Journal of Molecular Sciences* 20(20):5219 (2019) DOI

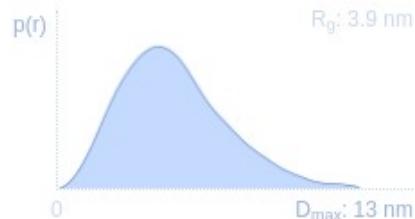
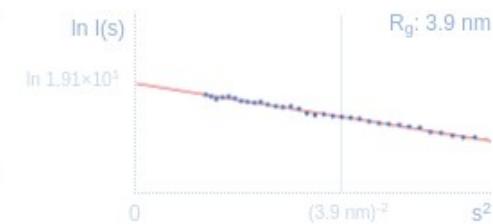
[Download files ▾](#)

## SASDGW6 – Serine acetyltransferase

### Serine acetyltransferase

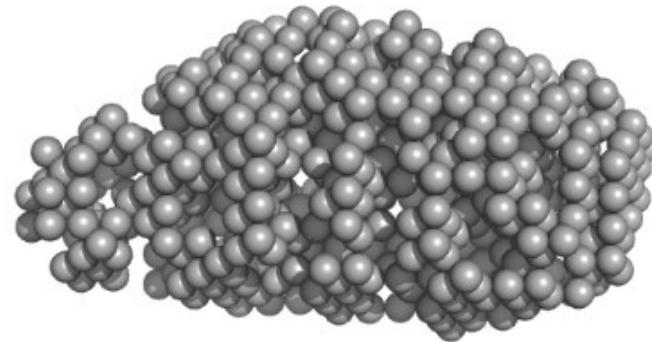
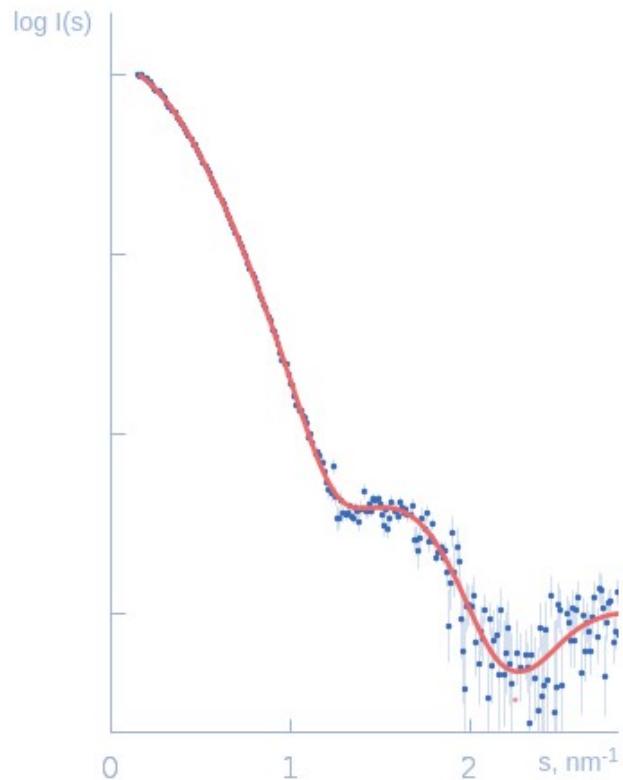


$MW^{\text{experimental}}$	189 kDa
$MW^{\text{expected}}$	177 kDa
$V^{\text{Porod}}$	280 $\text{nm}^3$



## Fits and models

---



Synchrotron SAXS data from solutions of serine acetyltransferase in 20 mM sodium phosphate, 85 mM NaCl, 2 mM EDTA, 10 mM 2-MCE, pH 7.5 were collected on the Austrian SAXS beamline 5.2L beam line at the ELETTRA - Sincrotrone Trieste storage ring (Trieste, Italy) using a Dectris / Pilatus3 1M detector at a wavelength of  $\lambda = 0.154 \text{ nm}$  ( $I(s)$  vs  $s$ , where  $s = 4\pi s \sin\theta/\lambda$ , and  $2\theta$  is the scattering angle). Solute concentrations ranging between 0.5 and 1.5 mg/ml were measured at 20°C. 12 successive 10 second frames were collected. The data were normalized to the intensity of the transmitted beam and radially averaged; the scattering of the solvent-blank was subtracted. The low angle data collected at lower concentration were merged with the highest concentration high angle data to yield the final composite scattering curve.

**Serine acetyltransferase (CysE)**

Mol. type Protein

Organism *Escherichia coli* K-12

Olig. state Hexamer

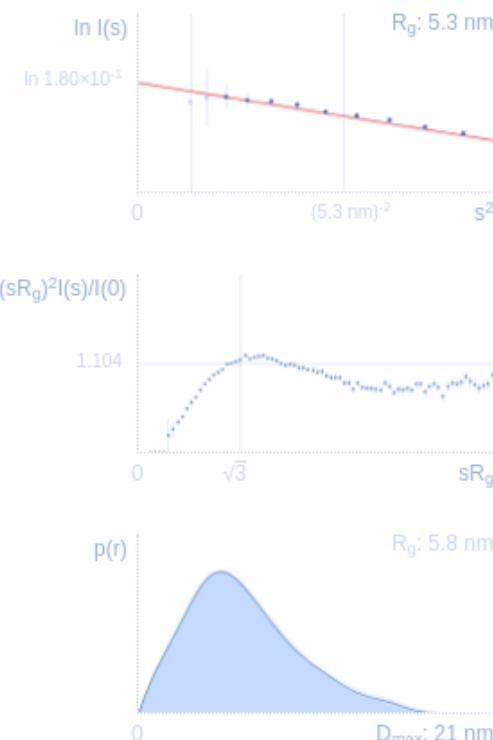
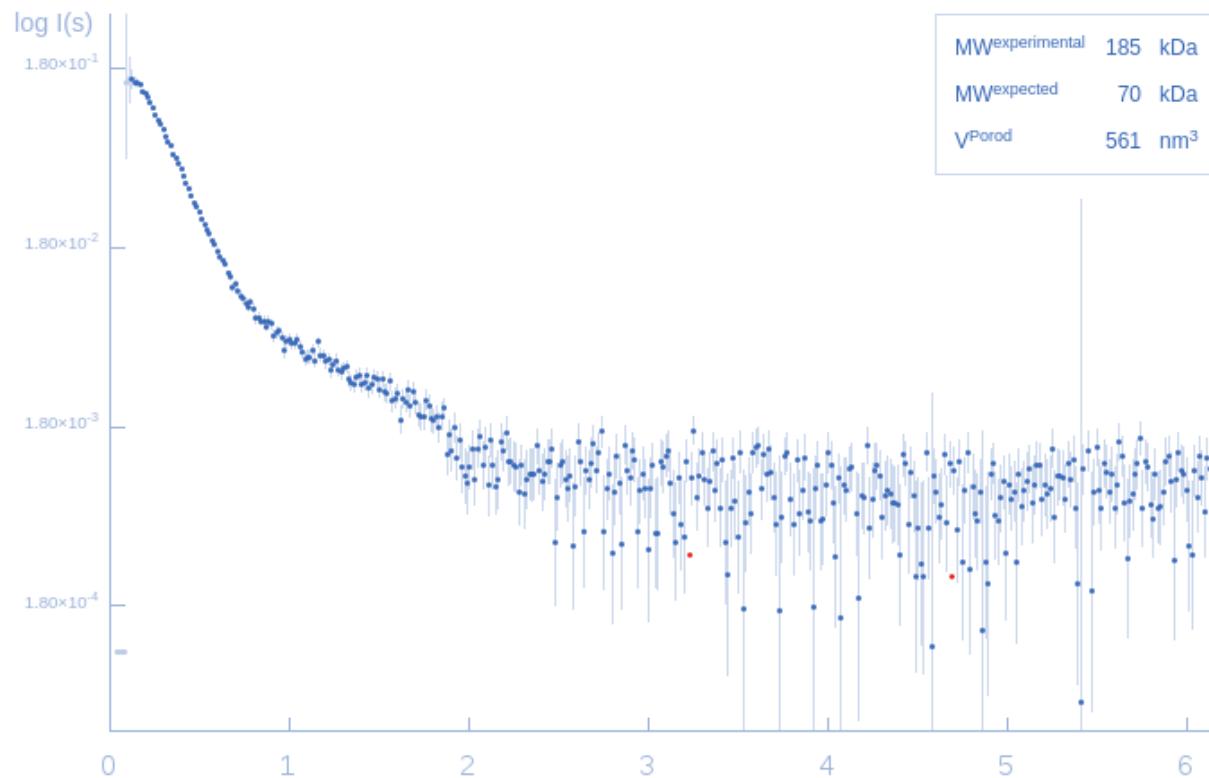
Mon. MW 29.4 kDa

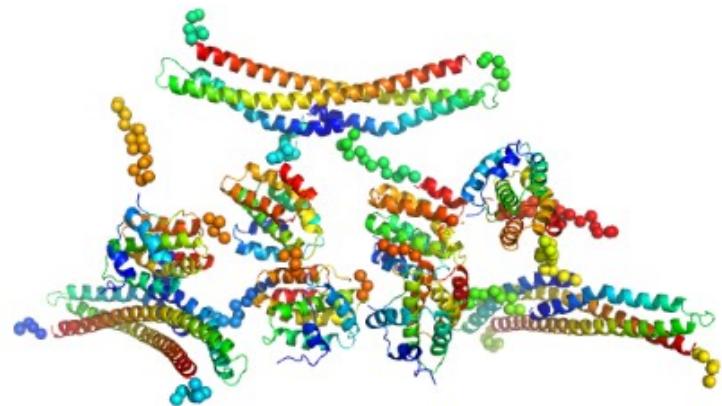
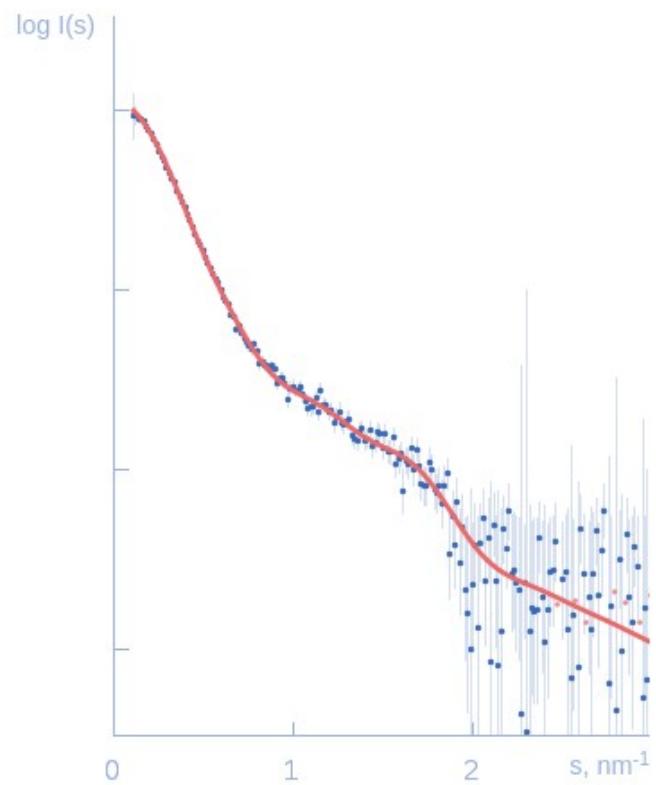
UniProt [P0A9D4 \(2-273\)](#)

Sequence [FASTA](#)

## SASDCZ2 – Yeast CTDsome: transcription termination factor Rtt103p/C-terminal domain of DNA-directed RNA polymerase II subunit RPB1 (Rtt103 1-246-phospho-mimetic complex)

Regulator of Ty1 transposition protein 103  
DNA-directed RNA polymerase II subunit RPB1





SAXS data from solutions of Rtt103 1-246-phospho-mimetic complex in 25 mM KH<sub>2</sub>PO<sub>4</sub>, 300 mM KCl, 10 mM β-mercaptoethanol, pH 6.5 were collected using a BioSAXS-1000 instrument at CEITEC (Brno, Czech Republic) equipped with a Pilatus 100K detector at a sample-detector distance of 0.5 m and at a wavelength of  $\lambda = 0.154$  nm ( $I(s)$  vs  $s$ , where  $s = 4\pi s \sin \theta / \lambda$  and  $2\theta$  is the scattering angle). Five successive 3600 second frames were collected at a sample temperature of 4°C. The data were normalized to the intensity of the transmitted beam and radially averaged and the corresponding scattering from the solvent-blank was subtracted to produce the scattering profile displayed in this entry.

**Regulator of Ty1 transposition protein 103**

Mol. type Protein

Organism *Saccharomyces cerevisiae* (strain ATCC 204508 / S288c)

Olig. state Dimer

Mon. MW 28.6 kDa

# **bioSAXS – summary**

**With good quality SAXS data:**

**1. model independent:**

integral structural parameters: Rg, Dmax, Vporod  
agregation, folding/flexibility  
*overall shape ab initio*

**2. model dependent:**

validation of molecular structures (XRD, NMR, molecular dynamics, docking)  
oligomeric state, mixtures  
quaternary structures  
missing parts in XRD structures  
study of flexible systems