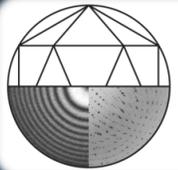


Structure and replication cycle of a virus infecting climate-modulating alga *Emiliana huxleyi*

Miroslav Homola



Plevka Lab
Structural virology

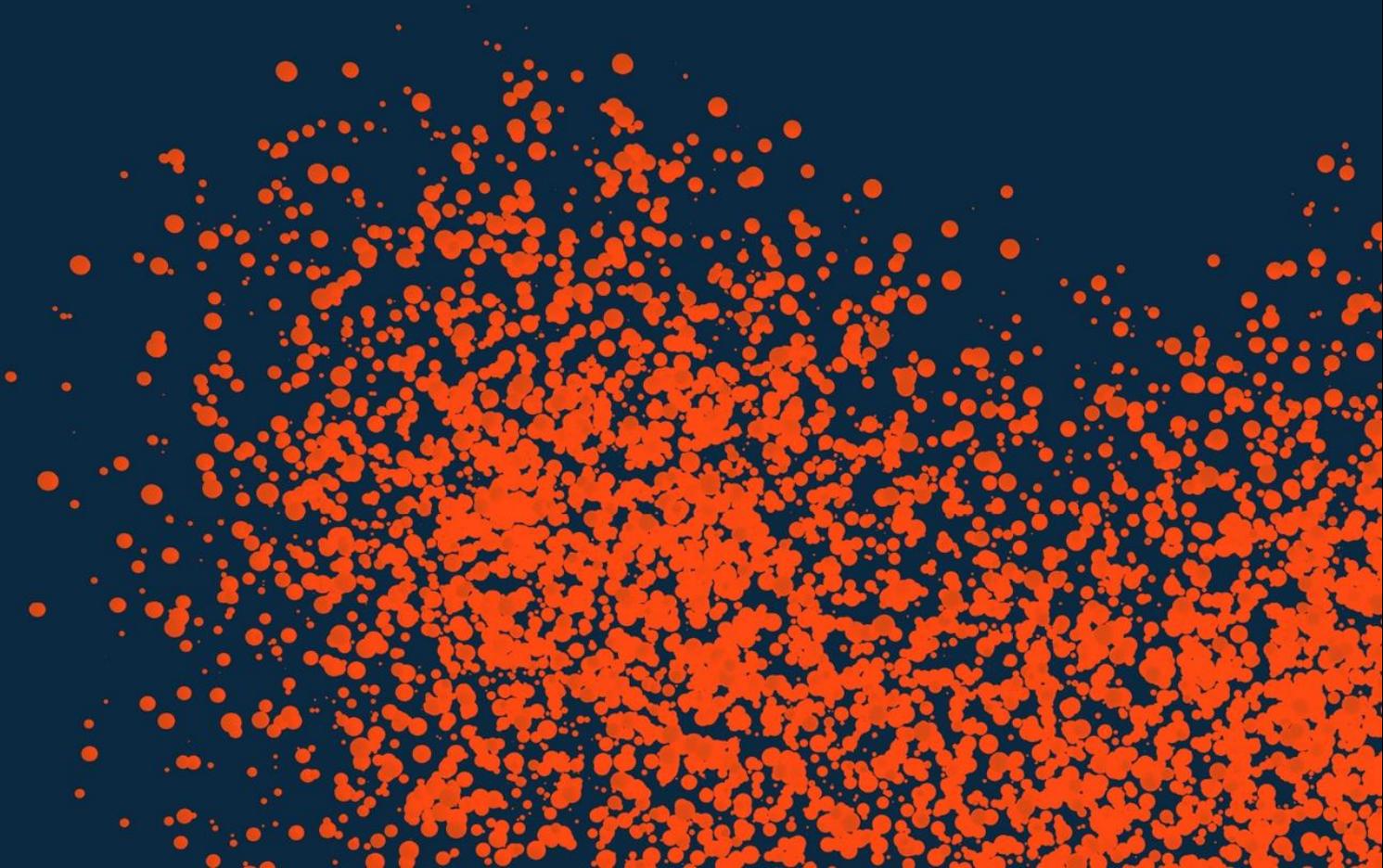


CEITEC
MASARYK UNIVERSITY

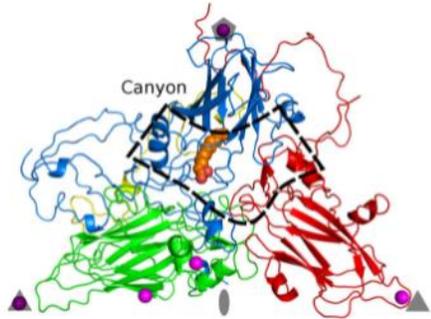
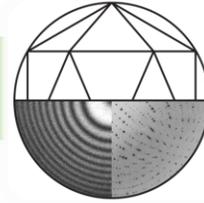


NIVB

National Institute
of Virology and Bacteriology



Structural virology lab

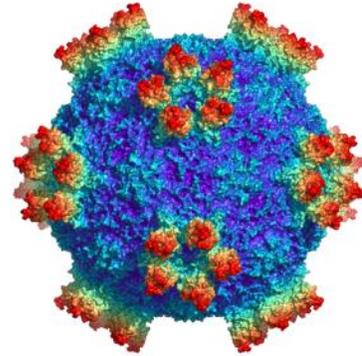


Human Viruses

We study human viruses from families *Flaviviridae*, *Togaviridae*, tick-borne encephalitis virus, *Herpesviridae*, Plevka viruses and *Picornaviridae*, including rhinoviruses and coronaviruses. We aim to structure and characterize replication cycles of these viruses and their interactions with the immune system.

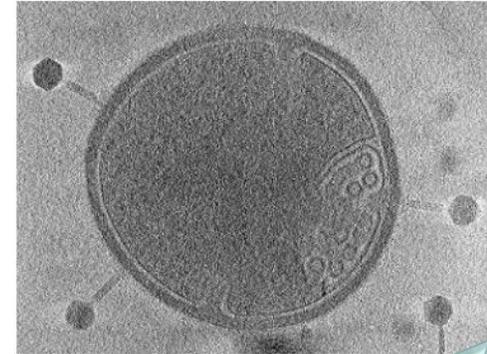


picoRNAviruses



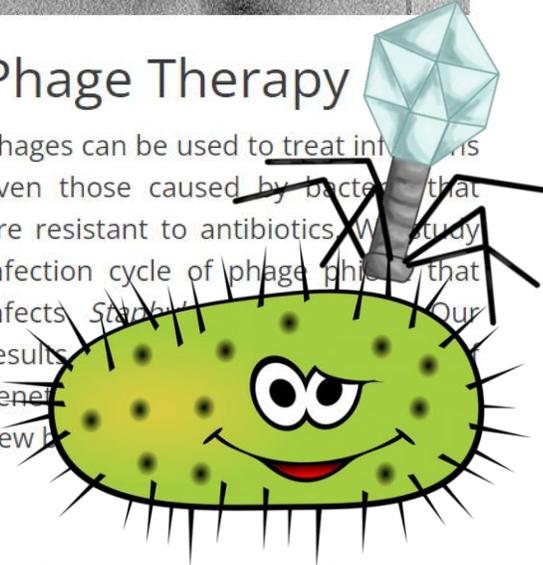
Honeybee Viruses

Honeybees (*Apis mellifera*) are vital for agricultural industry by providing pollination for many food crops. However, the beekeeper suffers from virus infections that cause colony losses. We study the structure of honeybee viruses and their replication cycles in order to develop antiviral compounds.



Phage Therapy

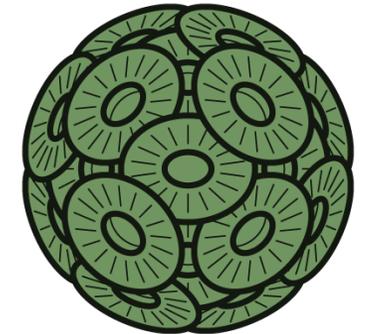
Phages can be used to treat infections, even those caused by bacteria that are resistant to antibiotics. We study the infection cycle of phage phi29 that infects *Staphylococcus aureus*. Our results show that phage phi29 generally does not cause new infections.



bacteriophages

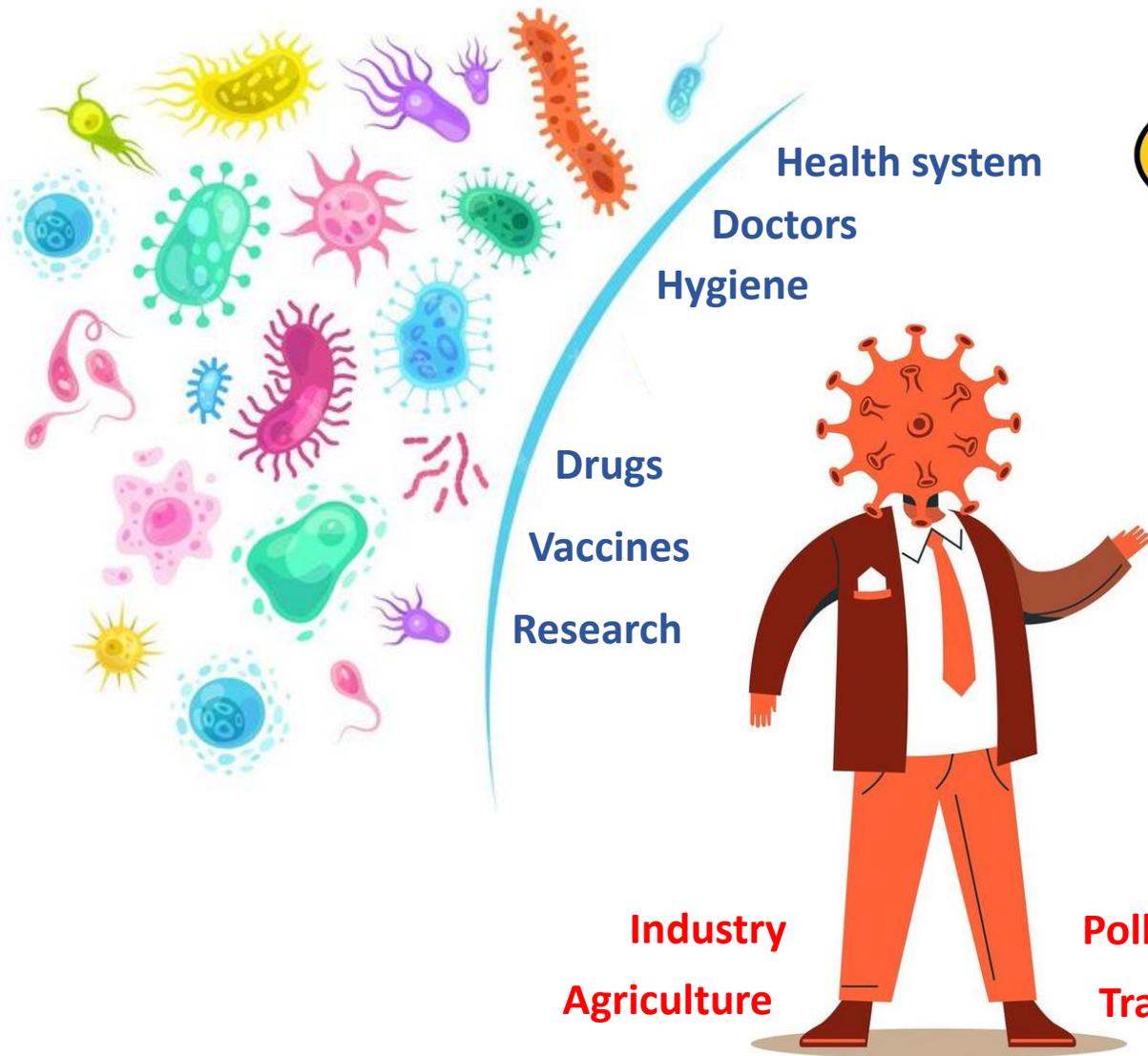


Marine algae viruses

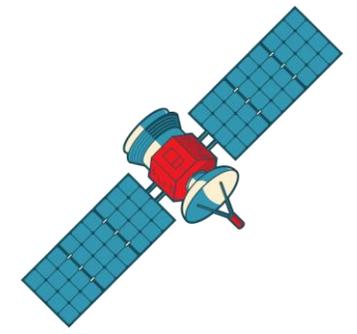
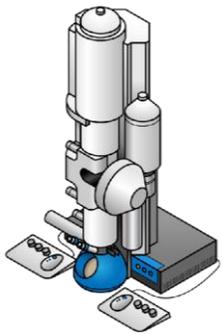
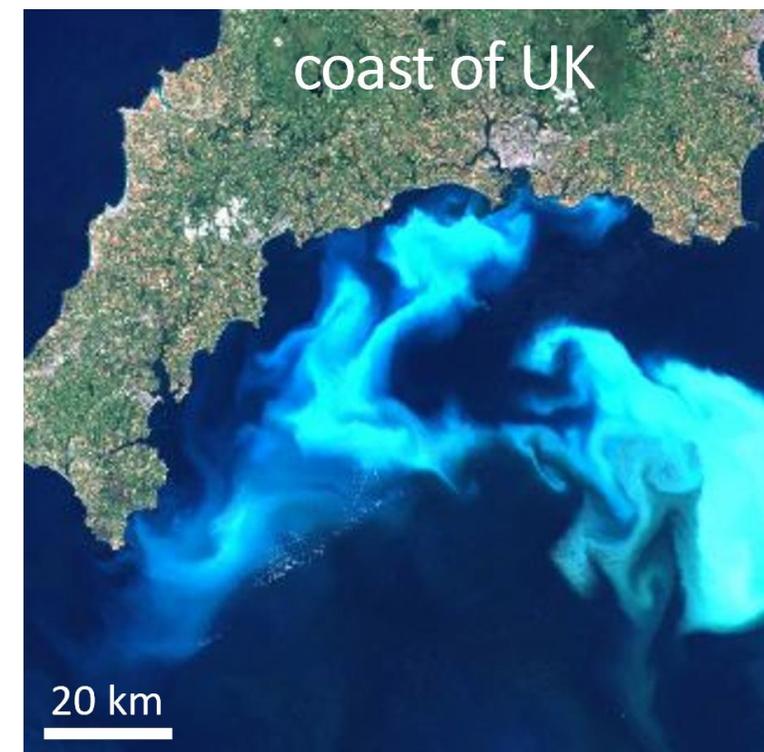
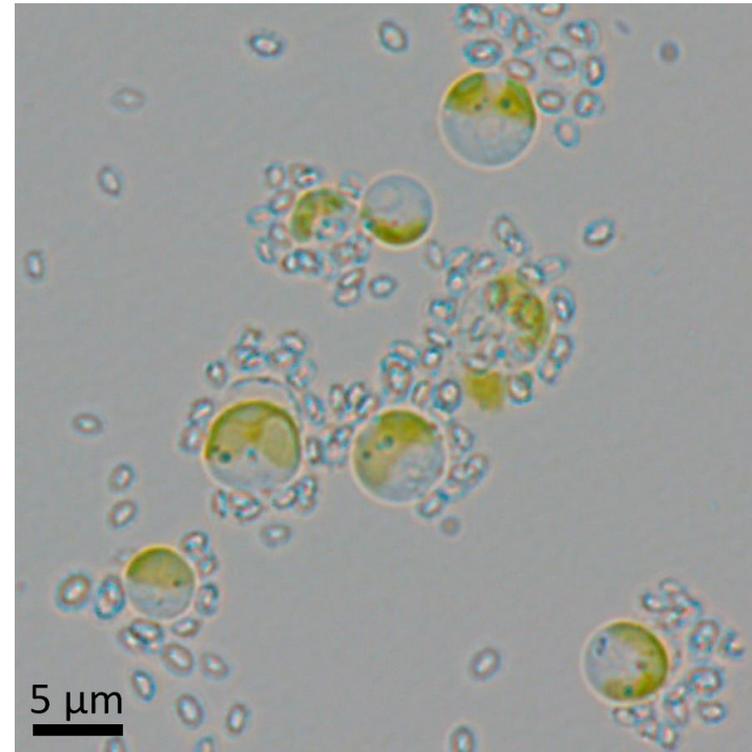
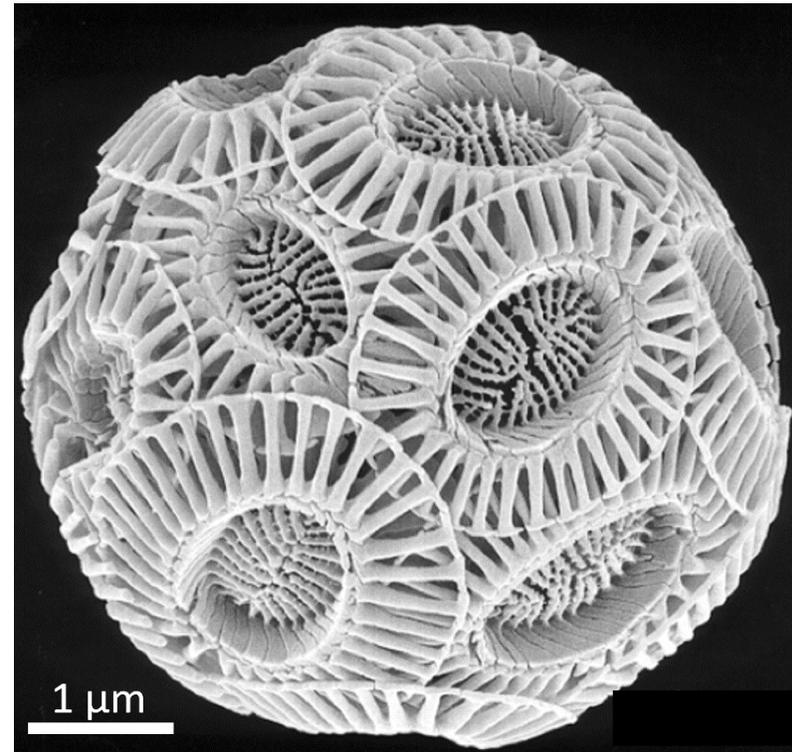


NCLDV's

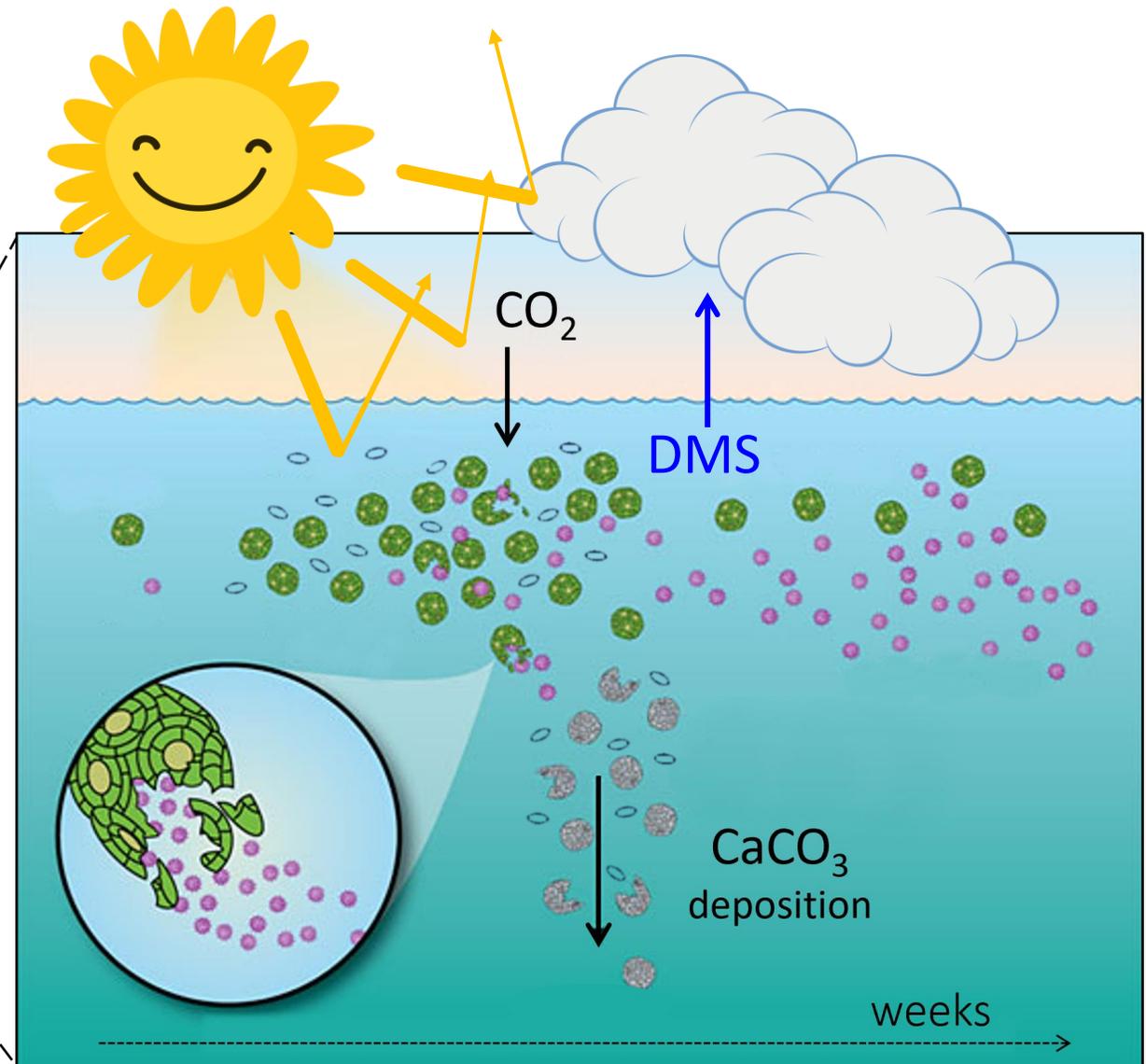
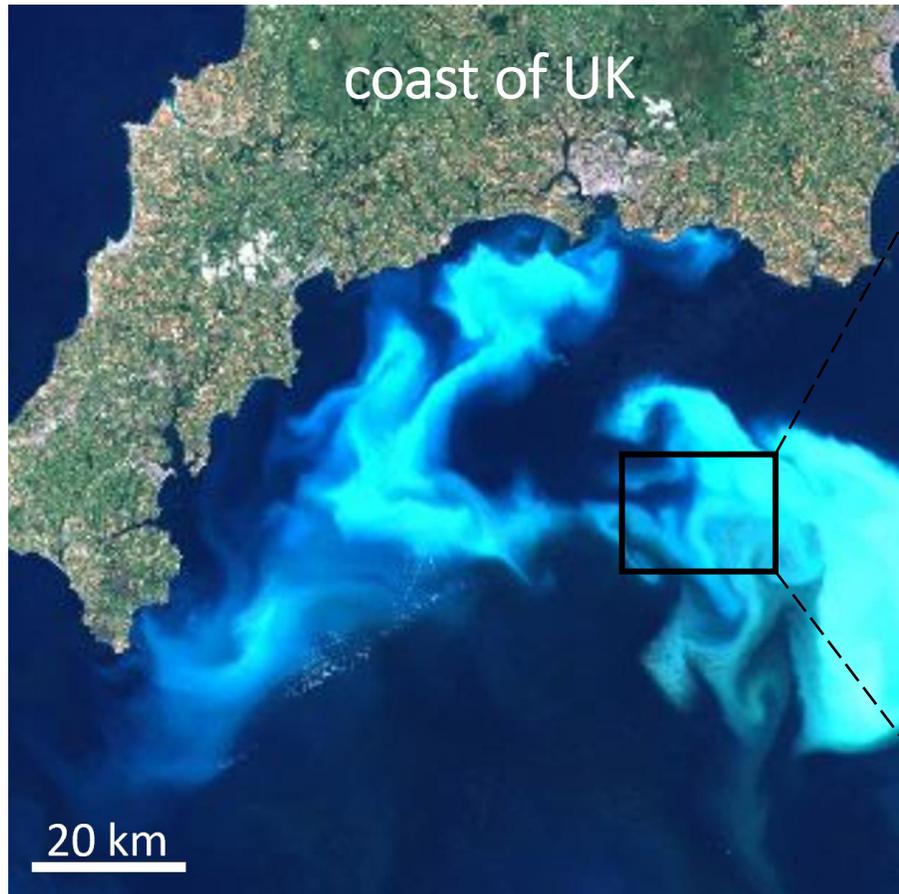
Human pathogen vs. human as a pathogen



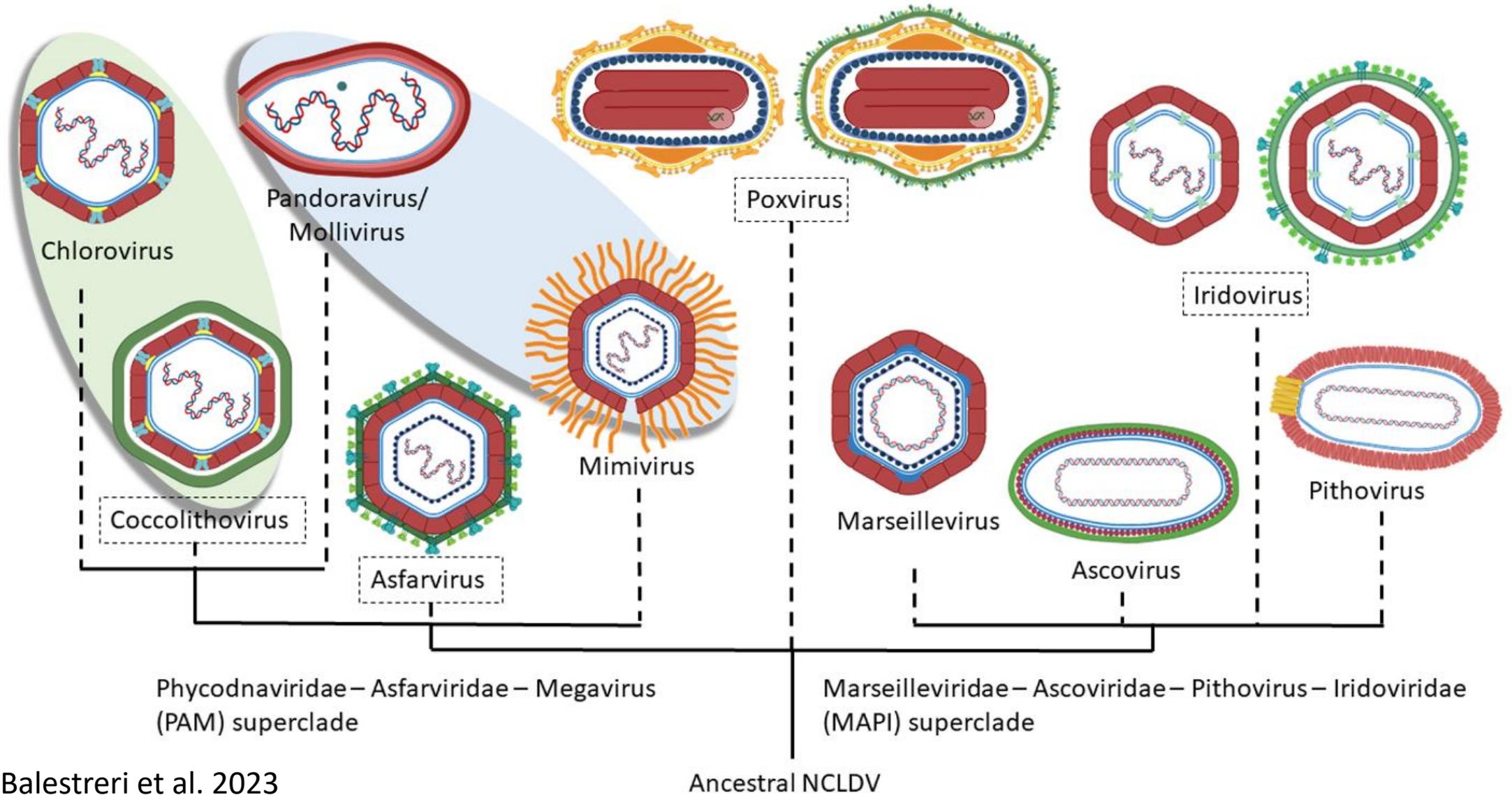
Marine alga *Emiliana huxleyi*



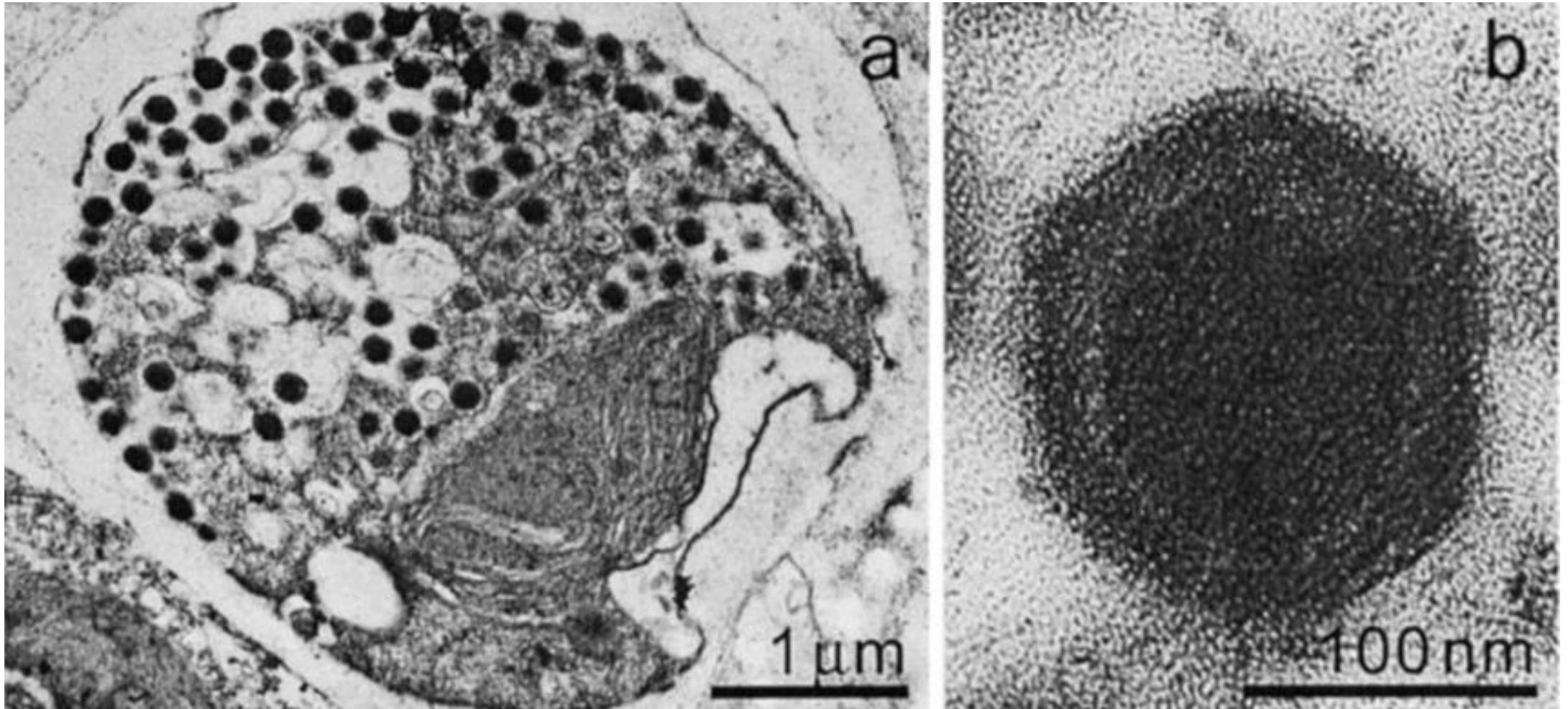
Blooms of the *Emiliana huxleyi* alga



Emiliana huxleyi virus 201 (EhV-201)

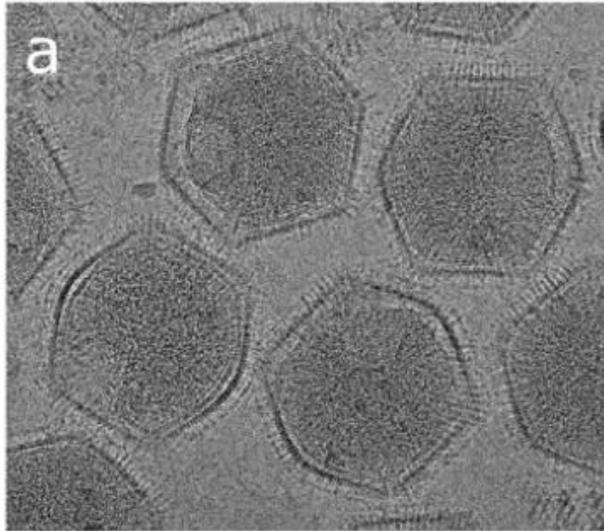


Emiliana huxleyi virus 201 (EhV-201)

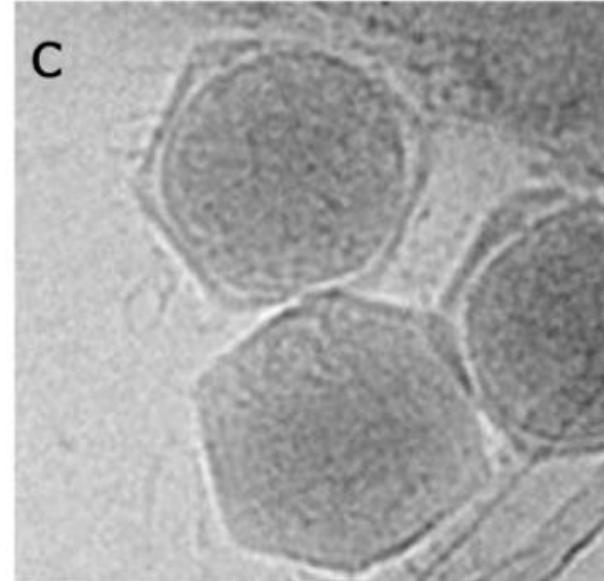
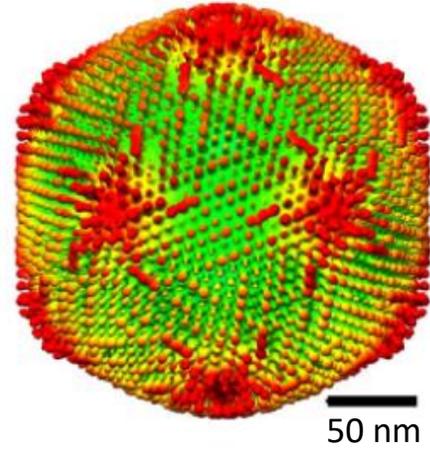


Castberg et al. 2001

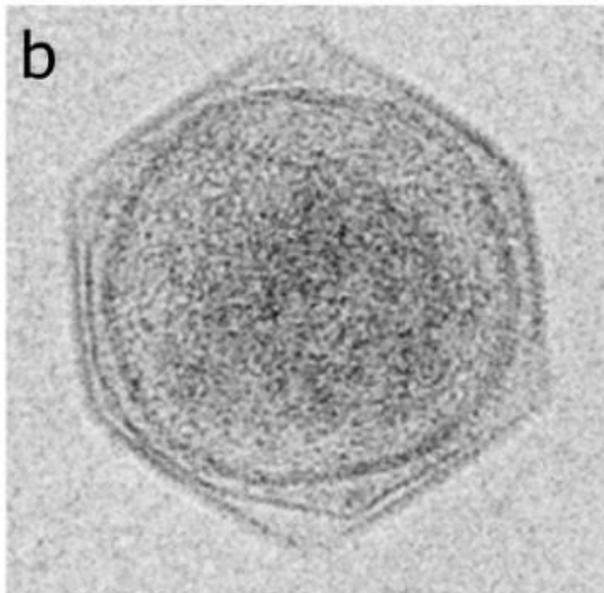
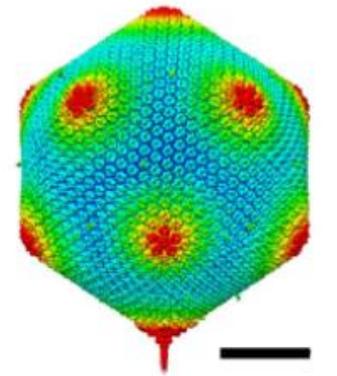
Related viruses



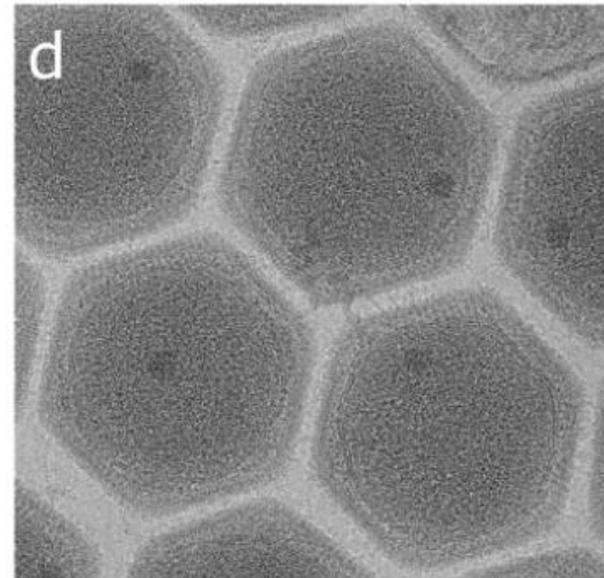
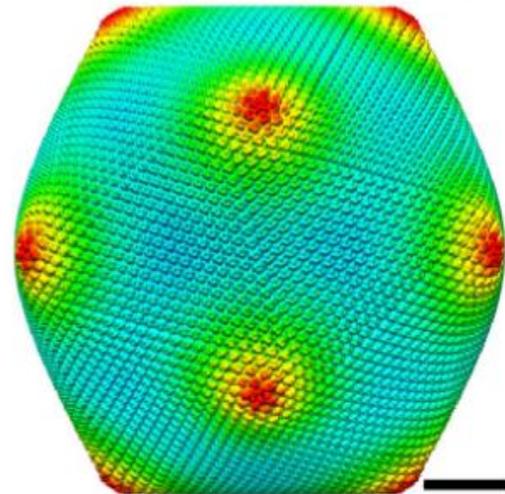
Medusavirus T=277



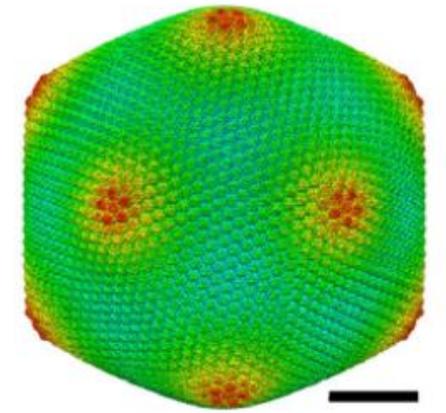
PBCV-1 T=169



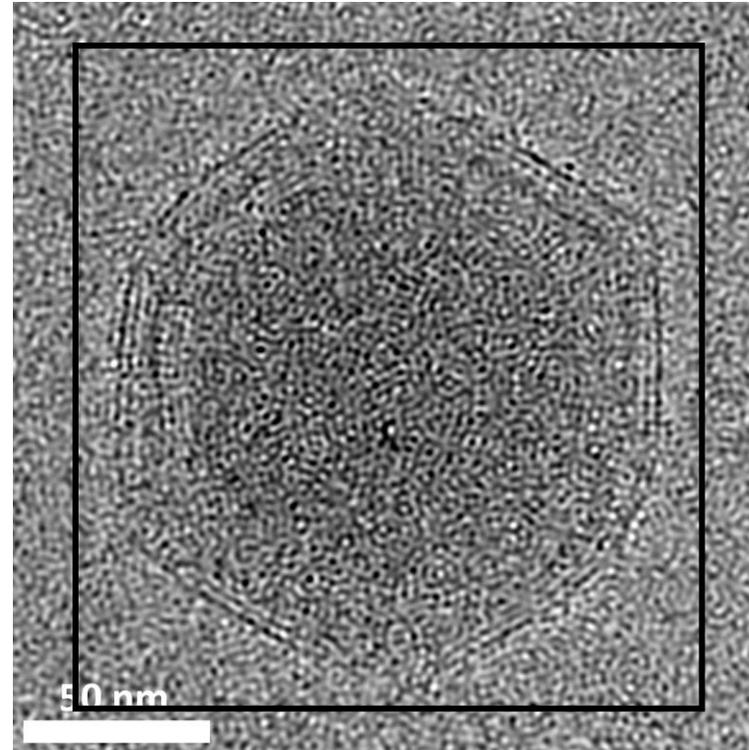
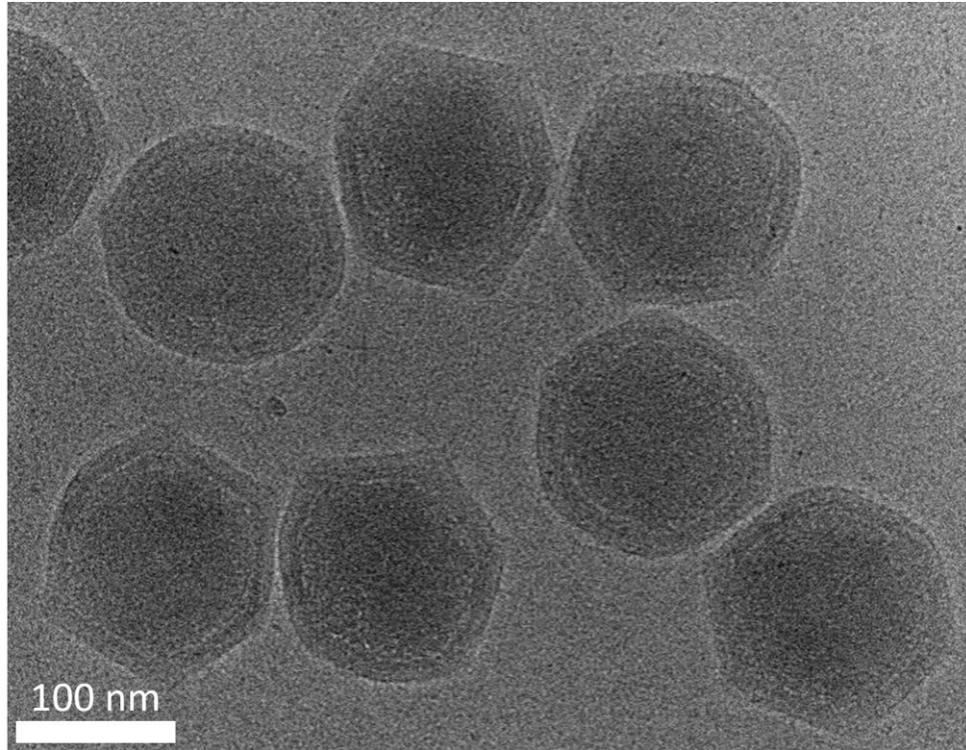
CroV T=499



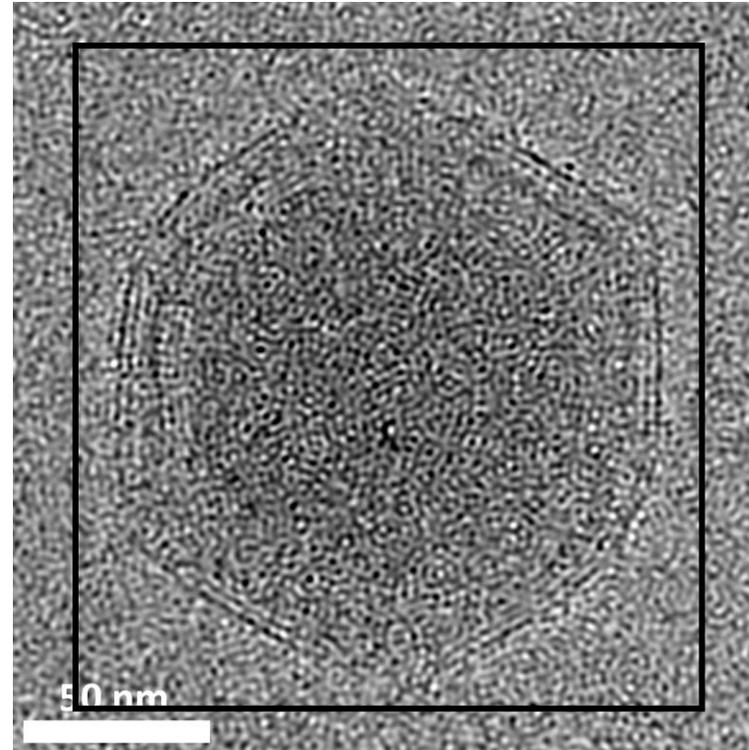
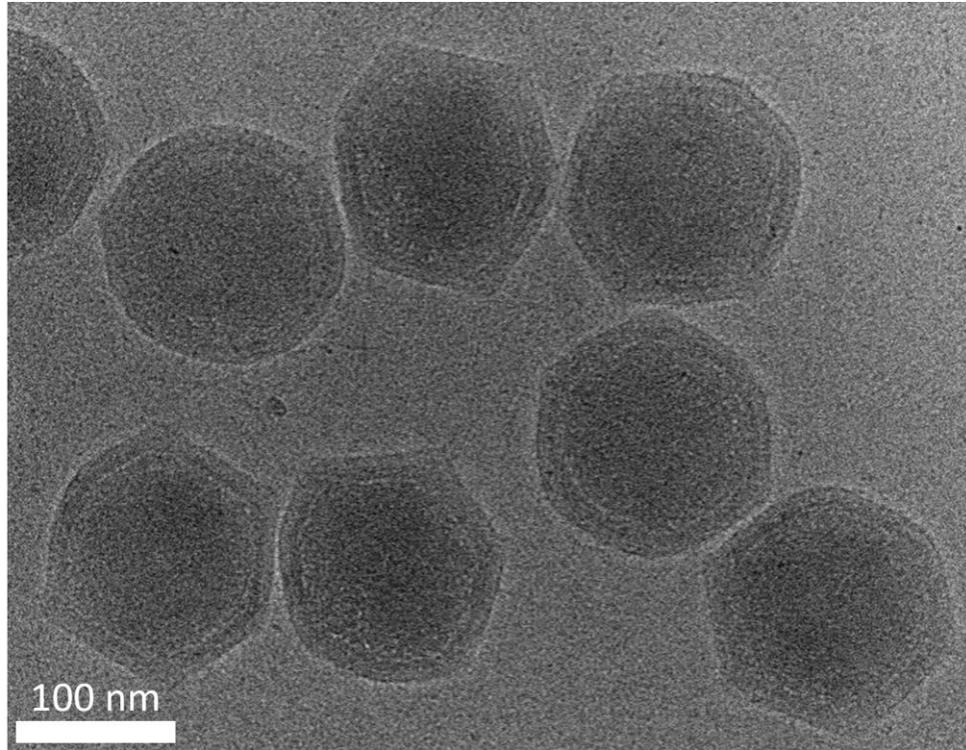
Melbournevirus T=309



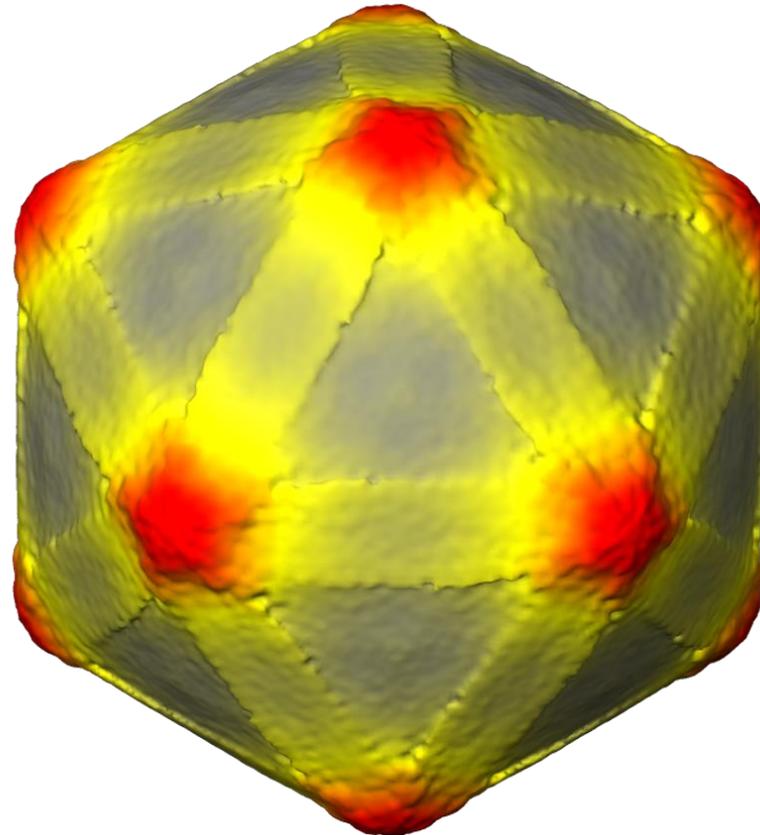
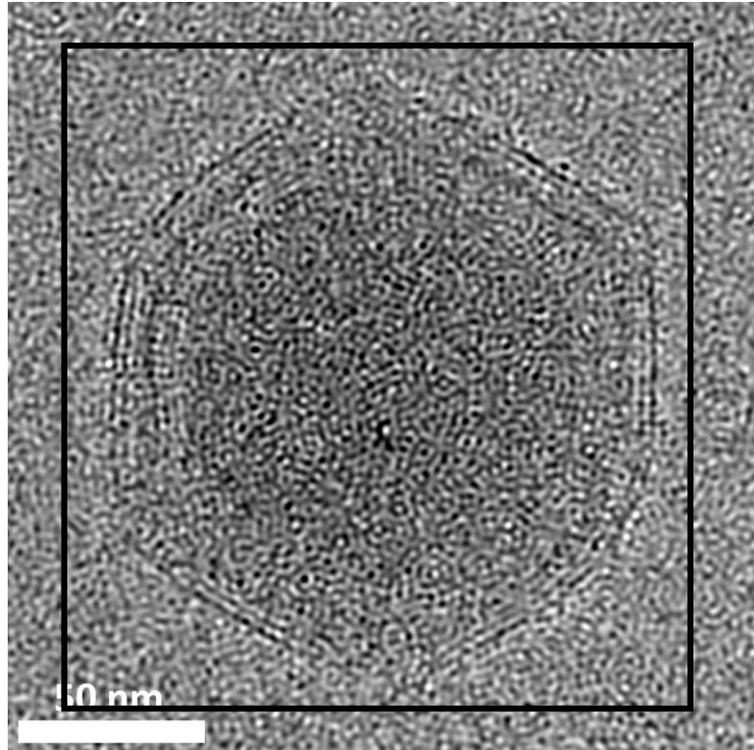
Single-particle analysis of EhV-201 cryo-EM data



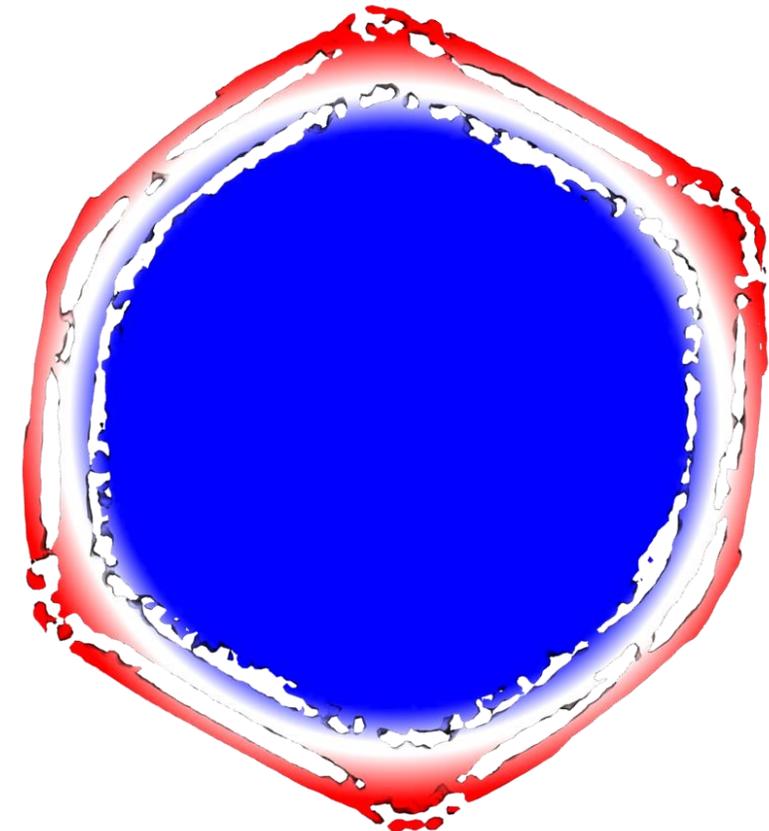
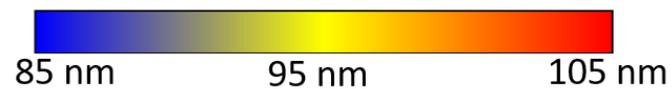
Single-particle analysis of EhV-201 cryo-EM data



Single-particle analysis of EhV-201 cryo-EM data

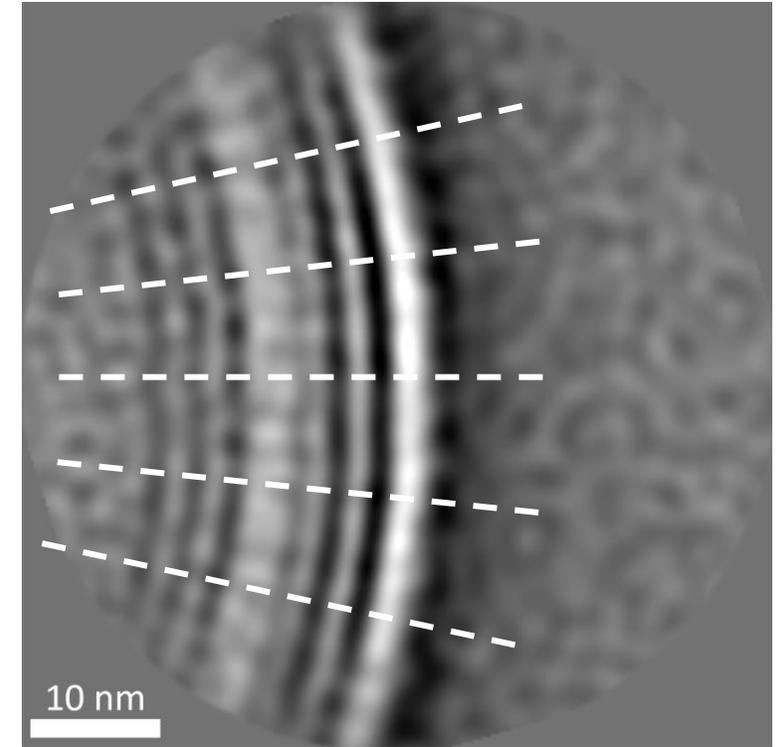
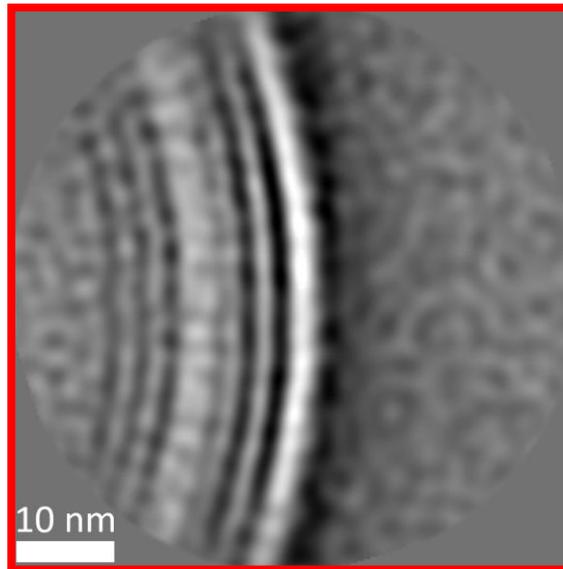
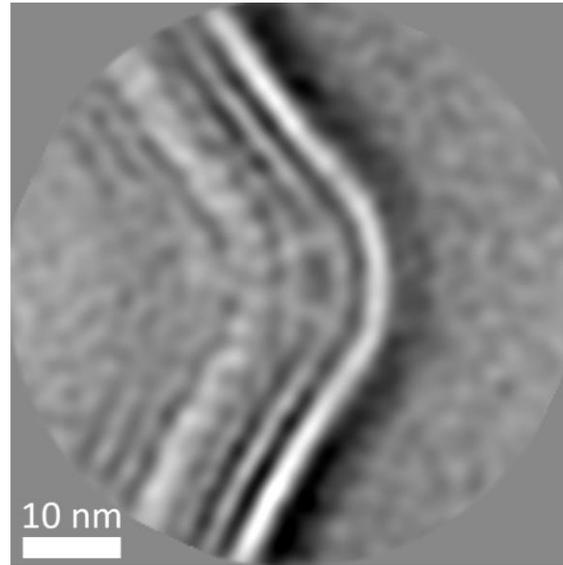
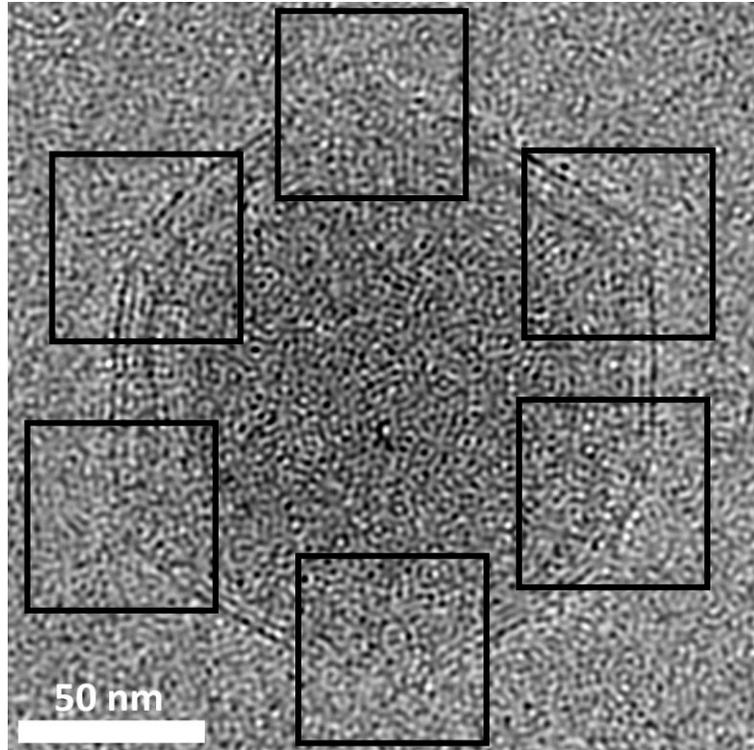


distance from particle center

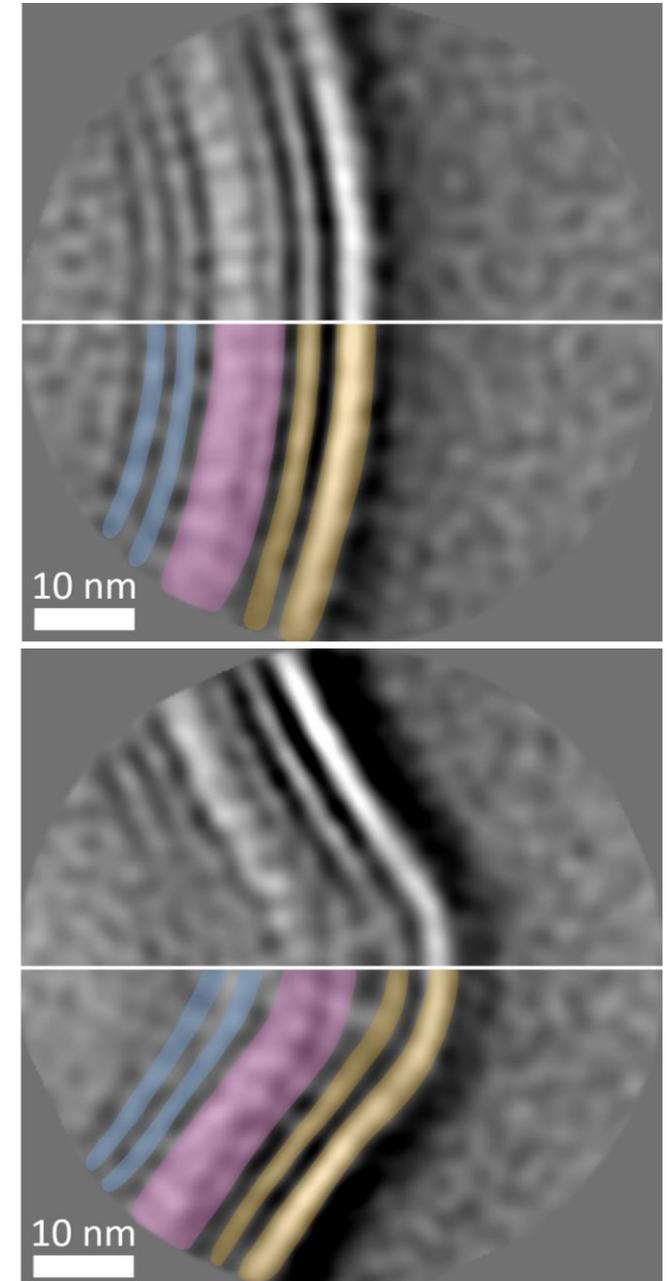
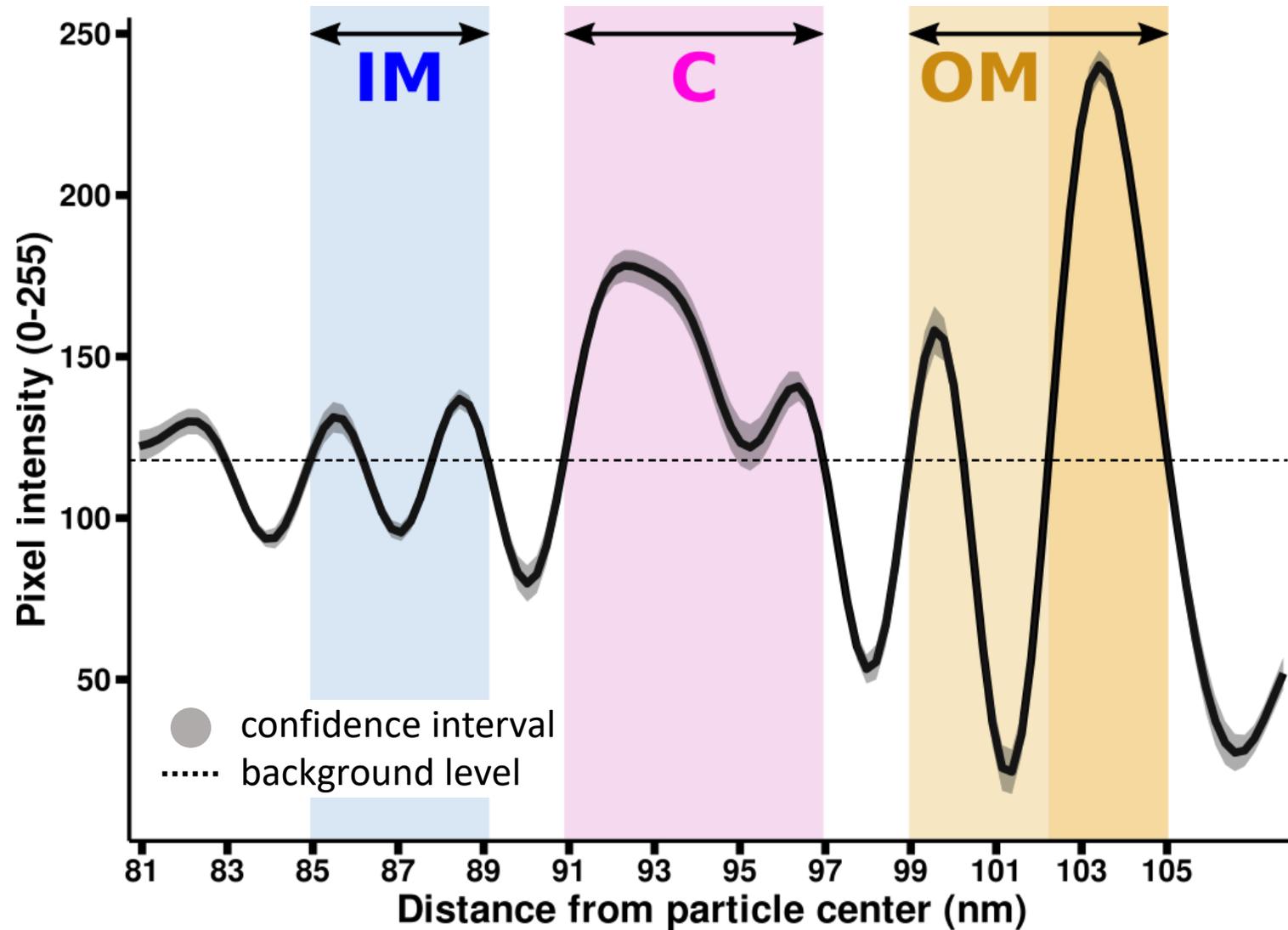


● membrane ○ capsid ● genome

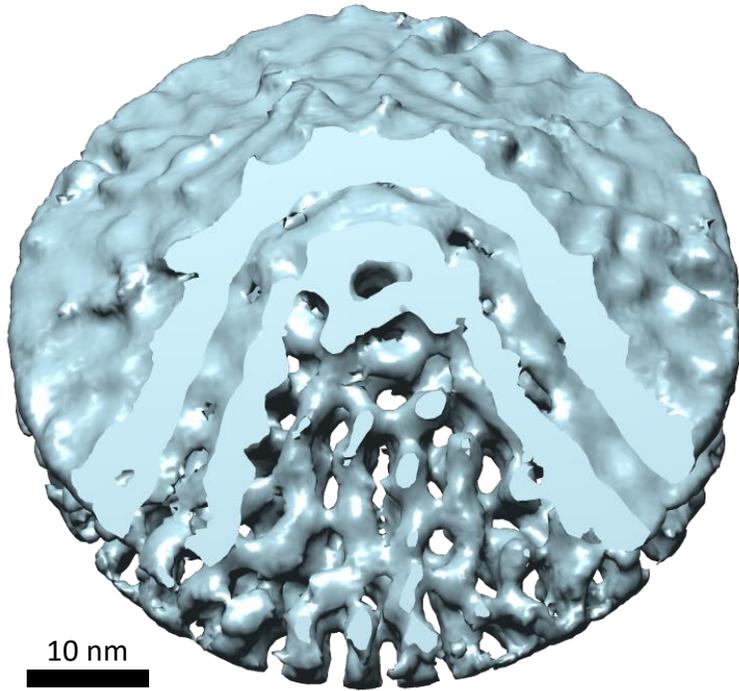
Localized single-particle analysis of EhV-201 cryo-EM data



Analysis of EhV-201 virion surface layers

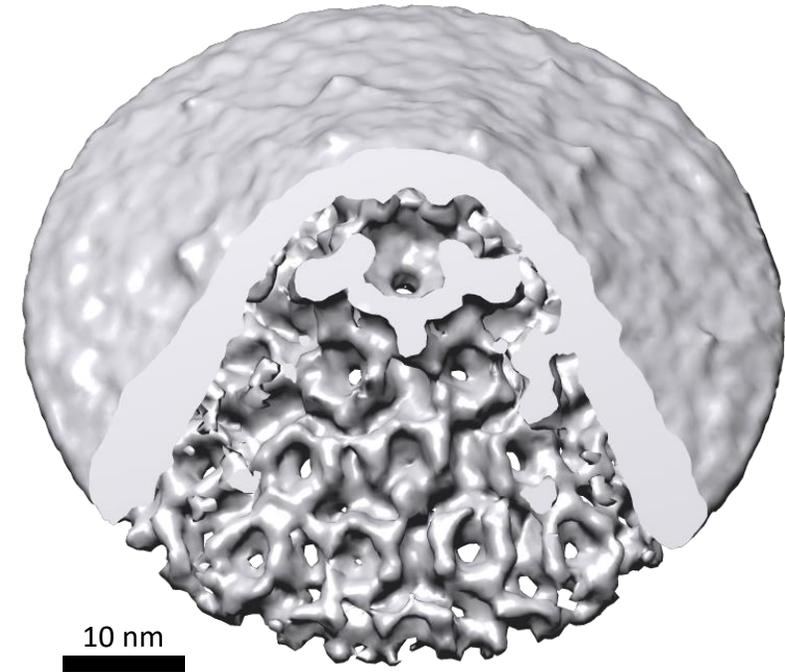
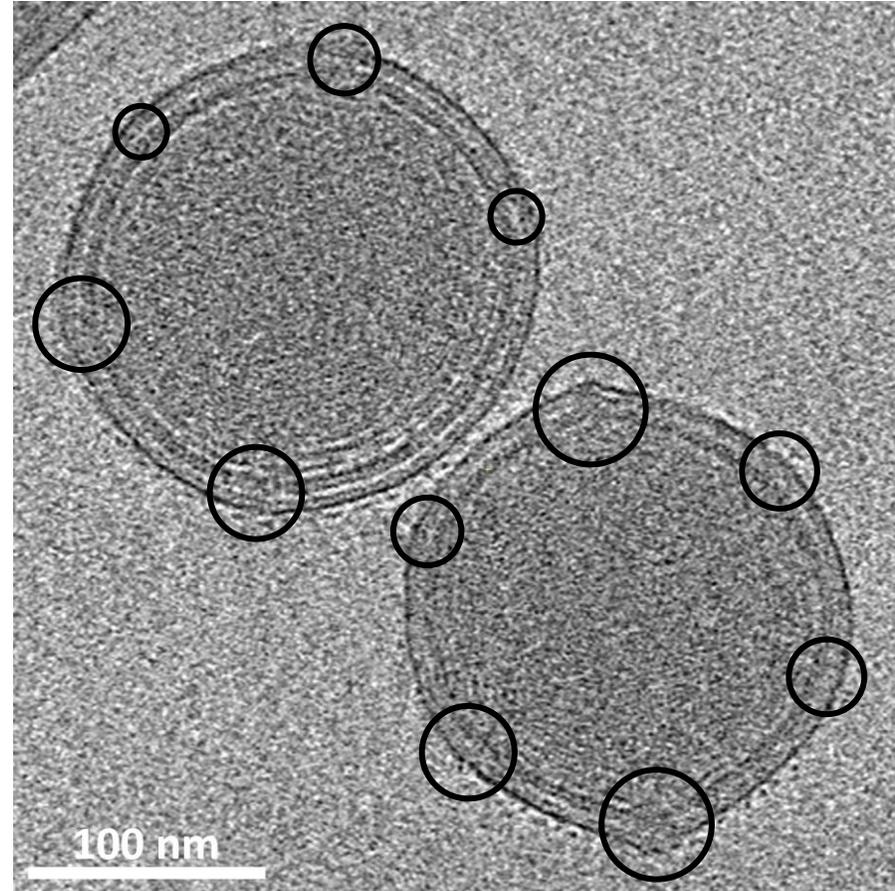


Sub-tomogram averaging of EhV-201



Single particle analysis

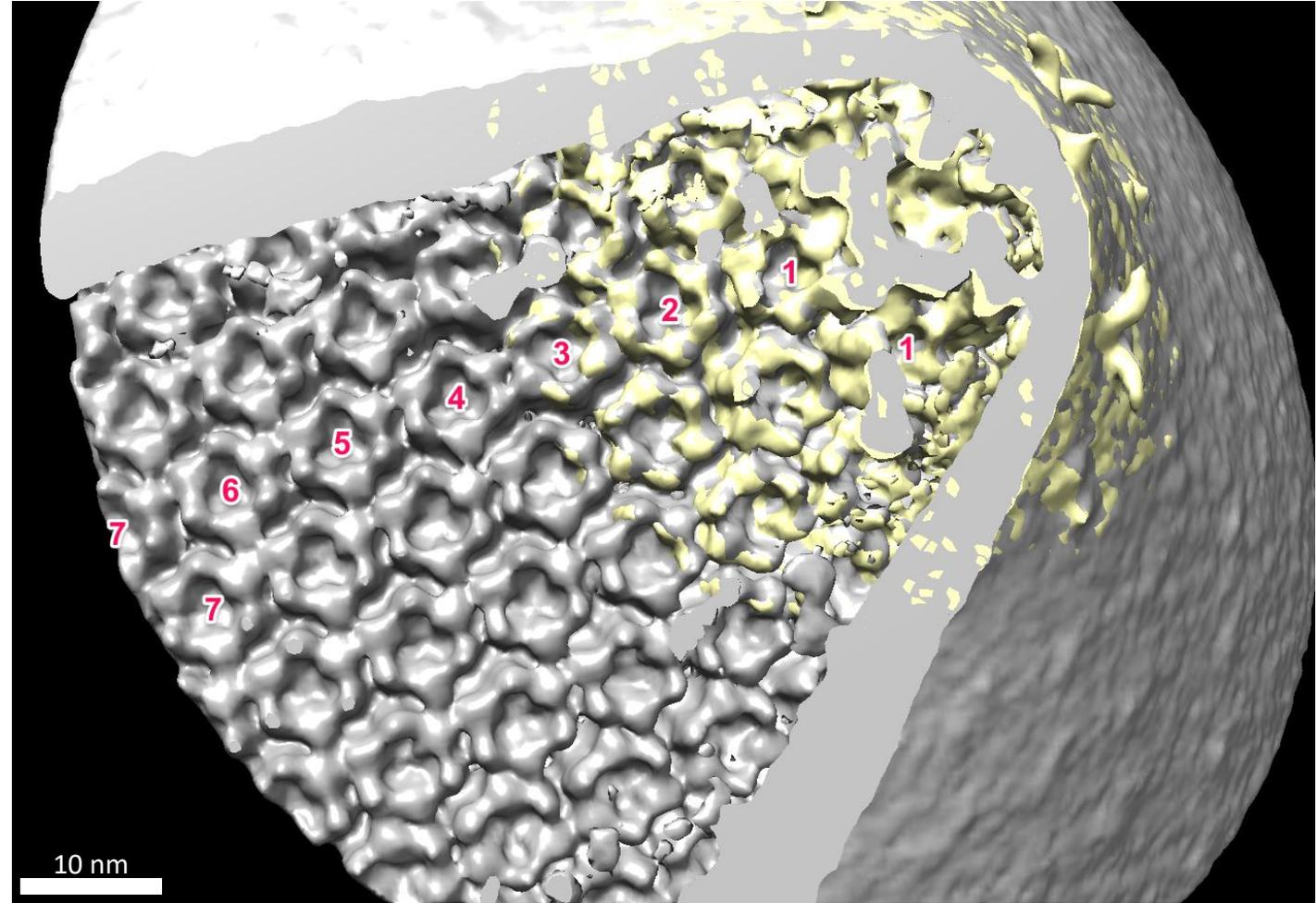
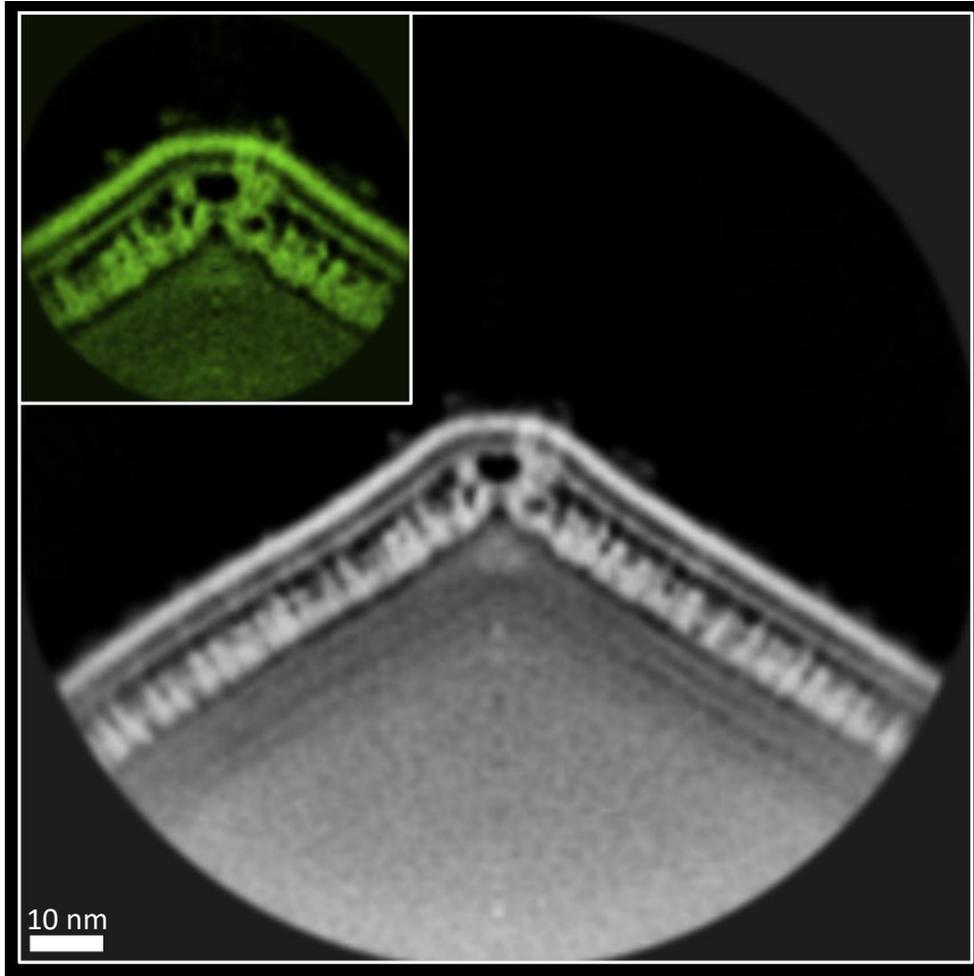
2.27 apix
24 Å res.
3 000 particles



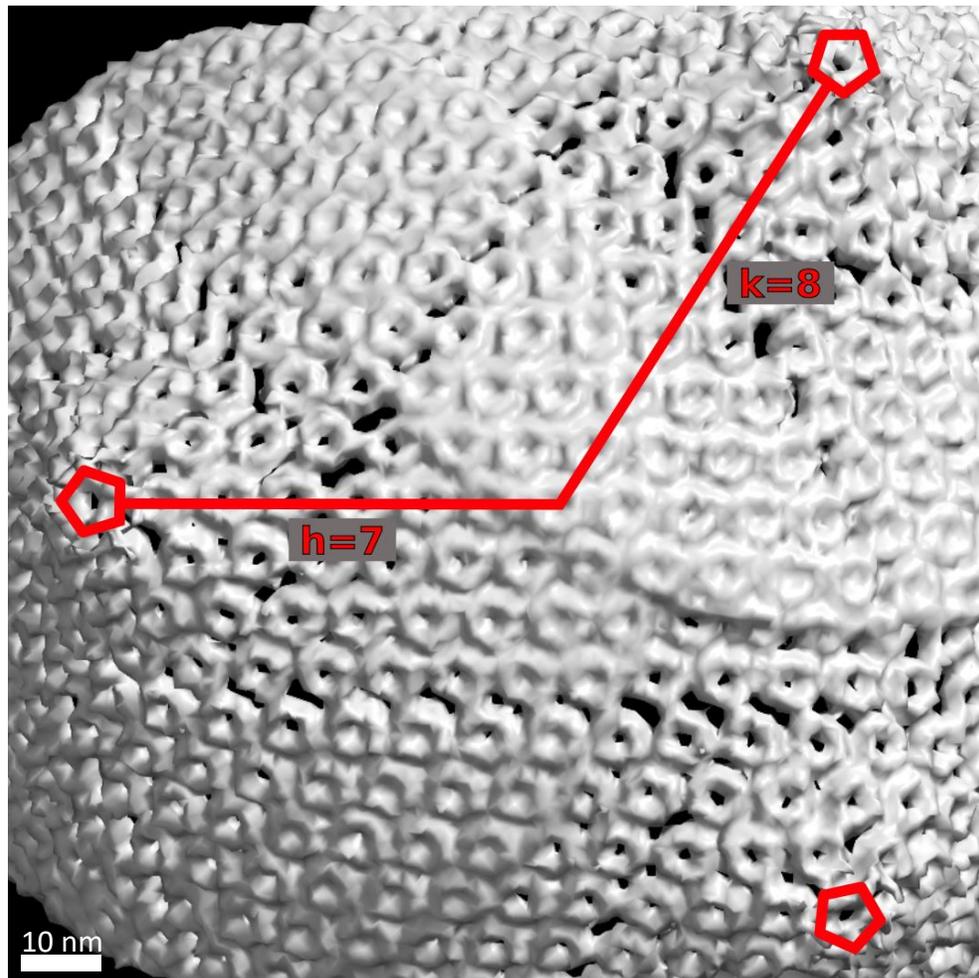
Sub-tomogram averaging

5.36 apix
22 Å res.
800 particles

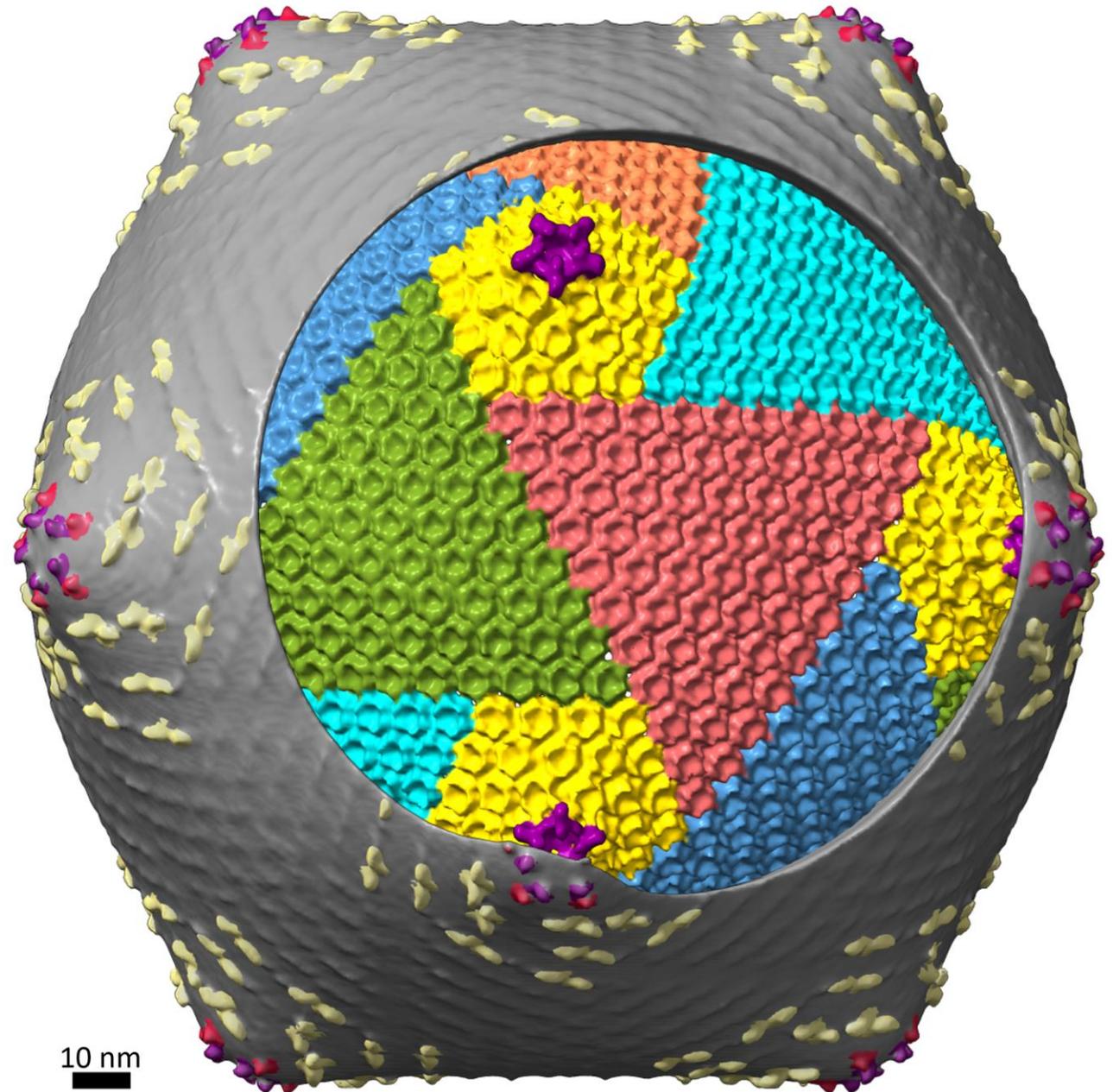
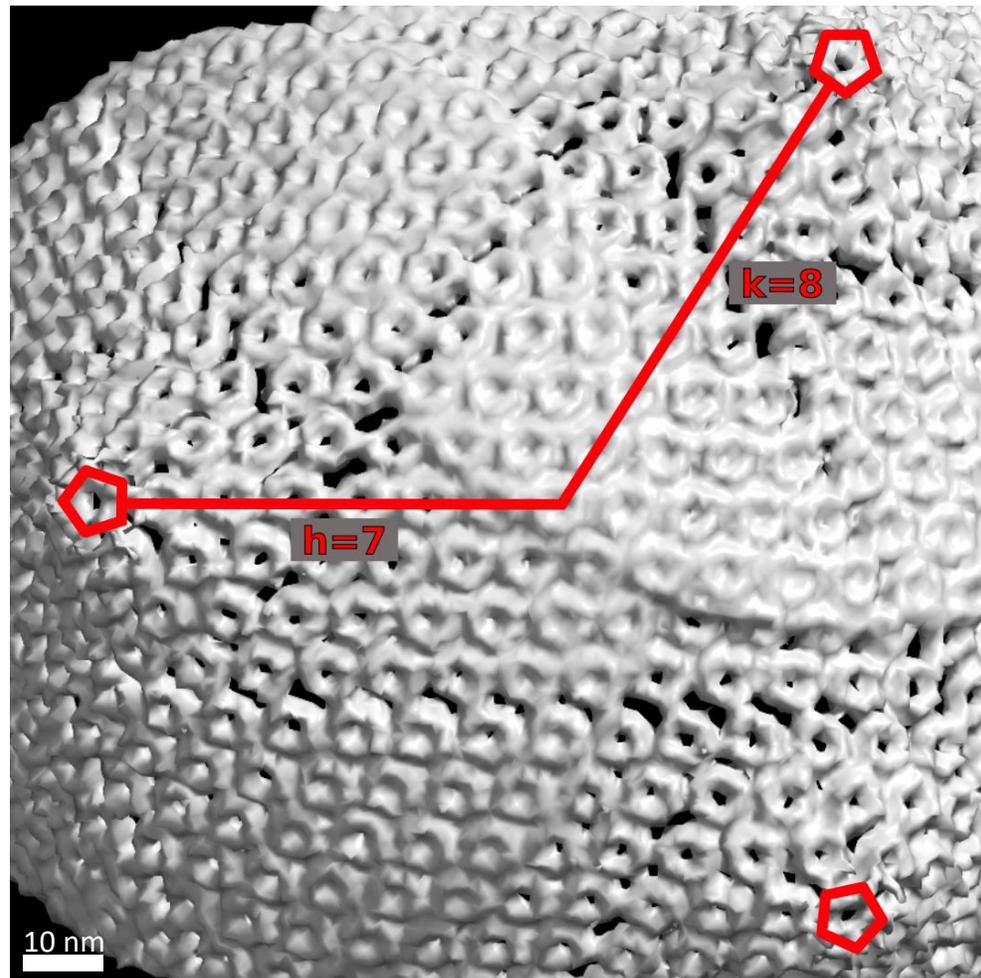
Bigger piece of capsid around virion vertex



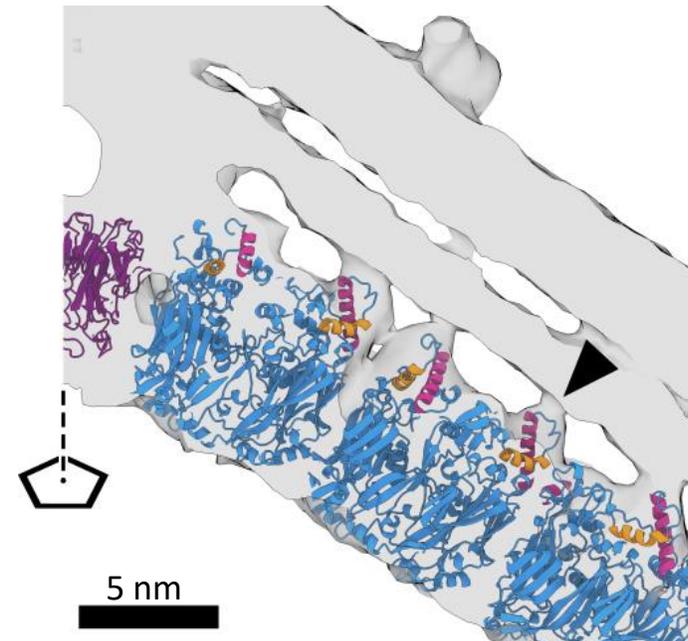
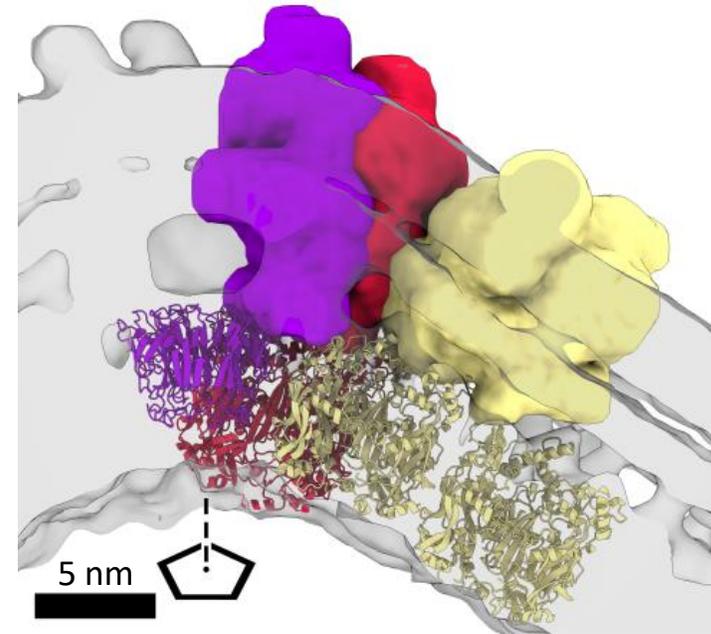
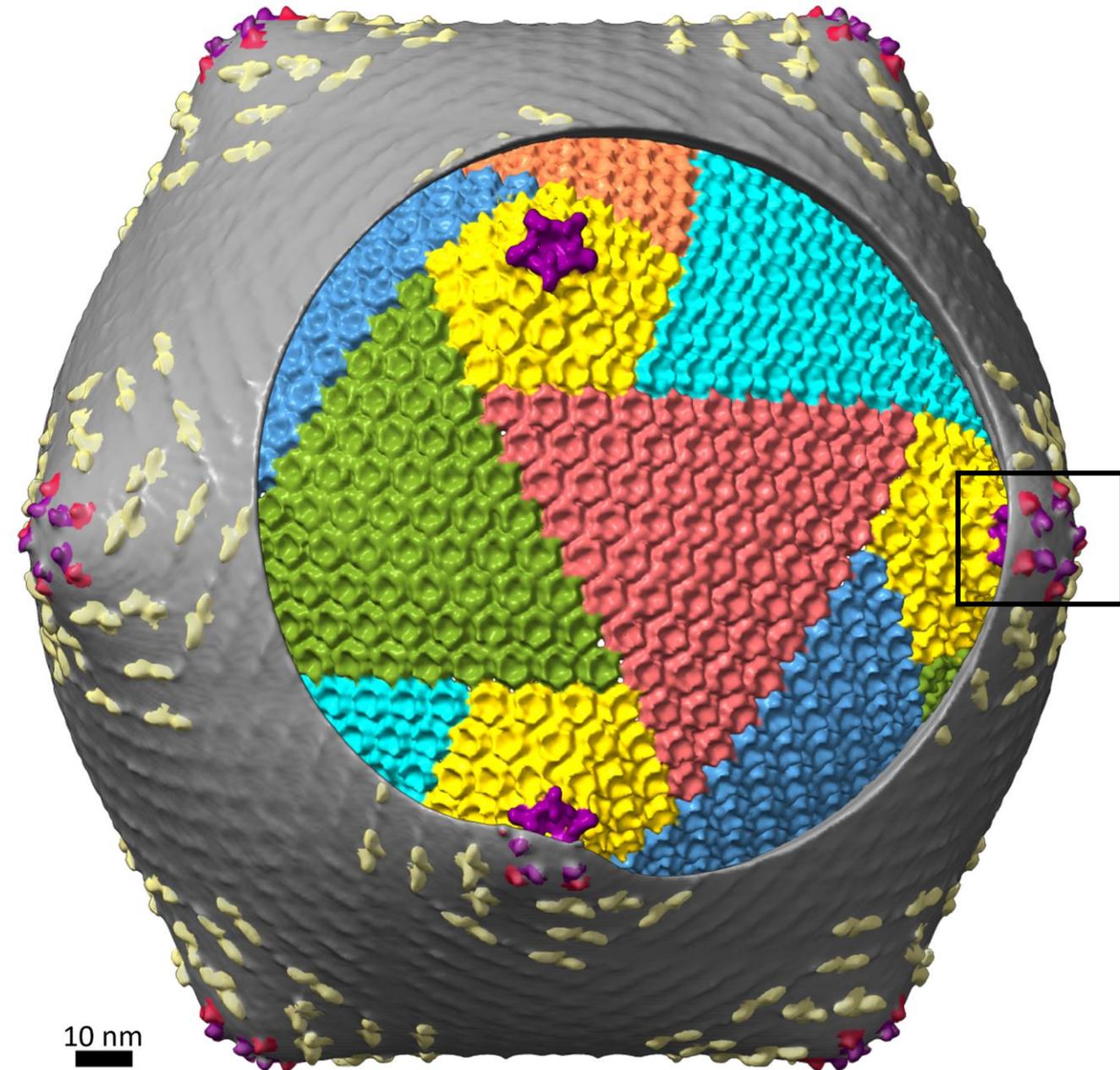
T number and composite map of EhV-201 virion



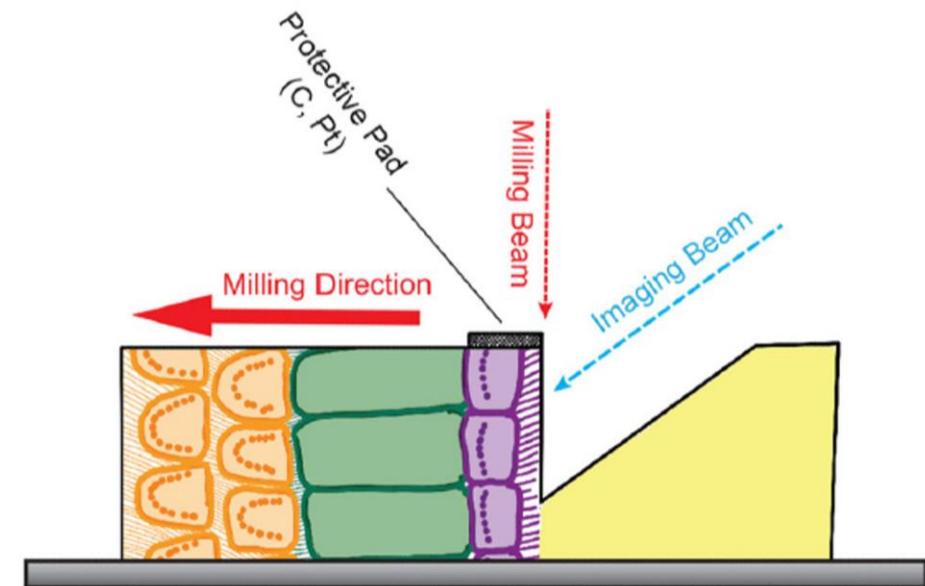
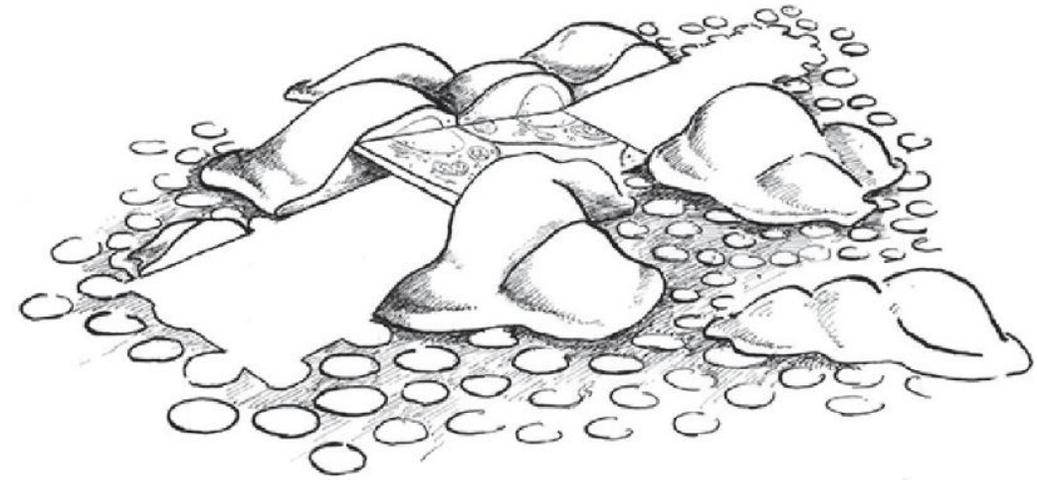
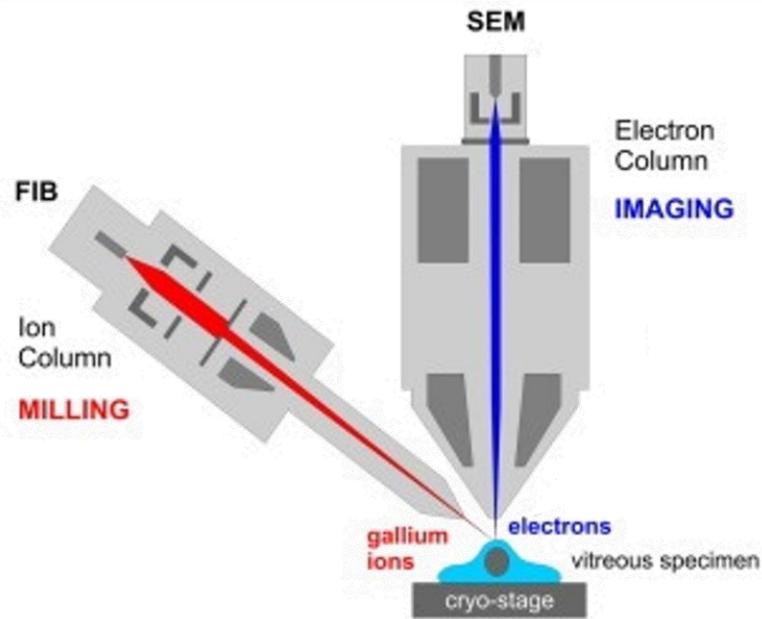
T number and composite map of EhV-201 virion



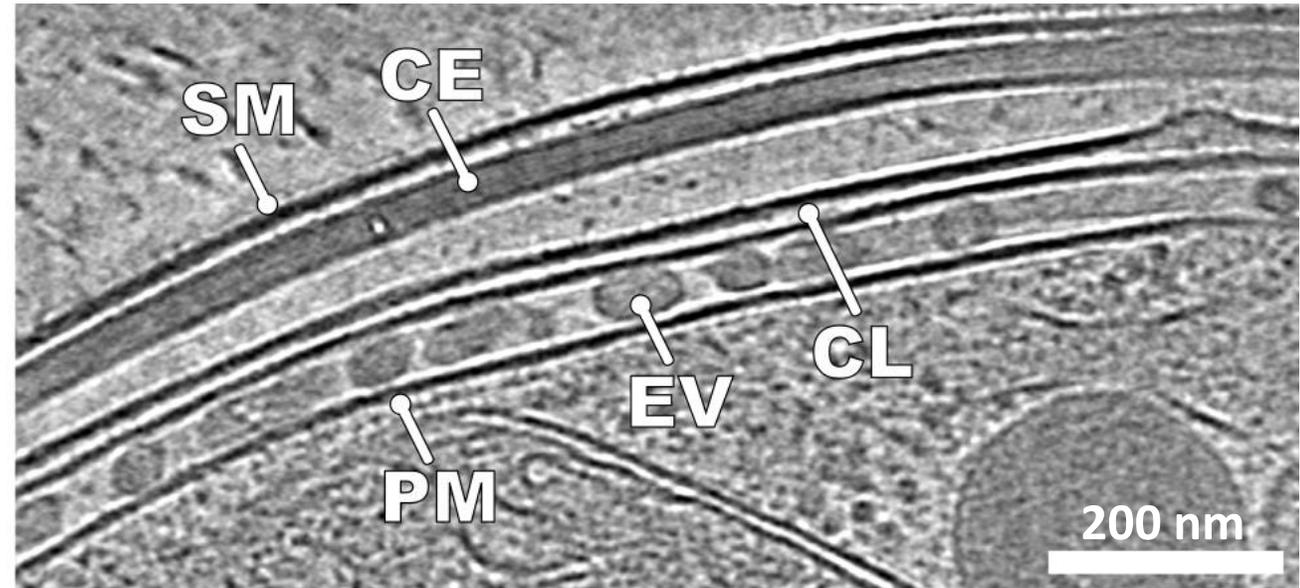
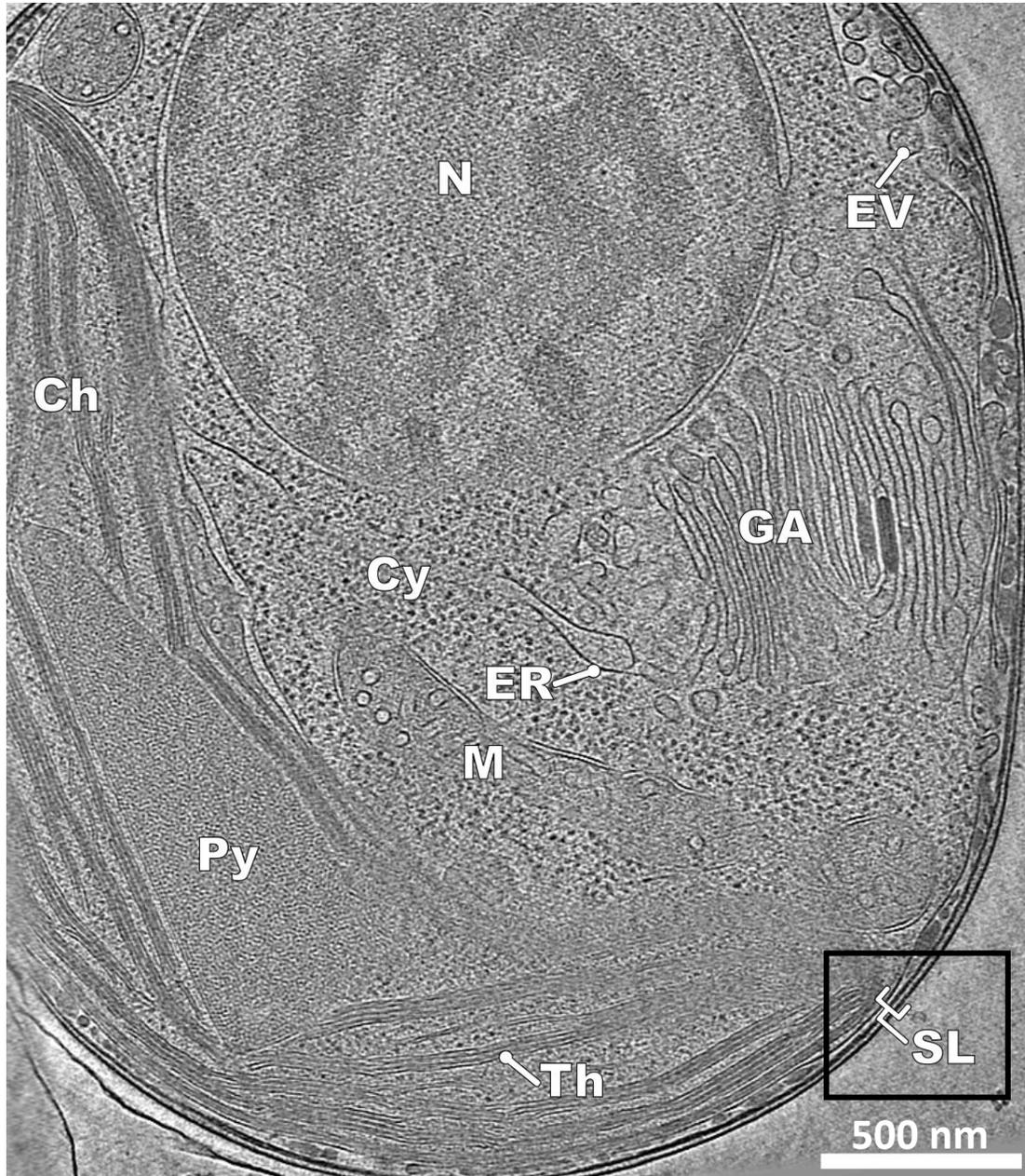
Composite map of EhV-201 virion



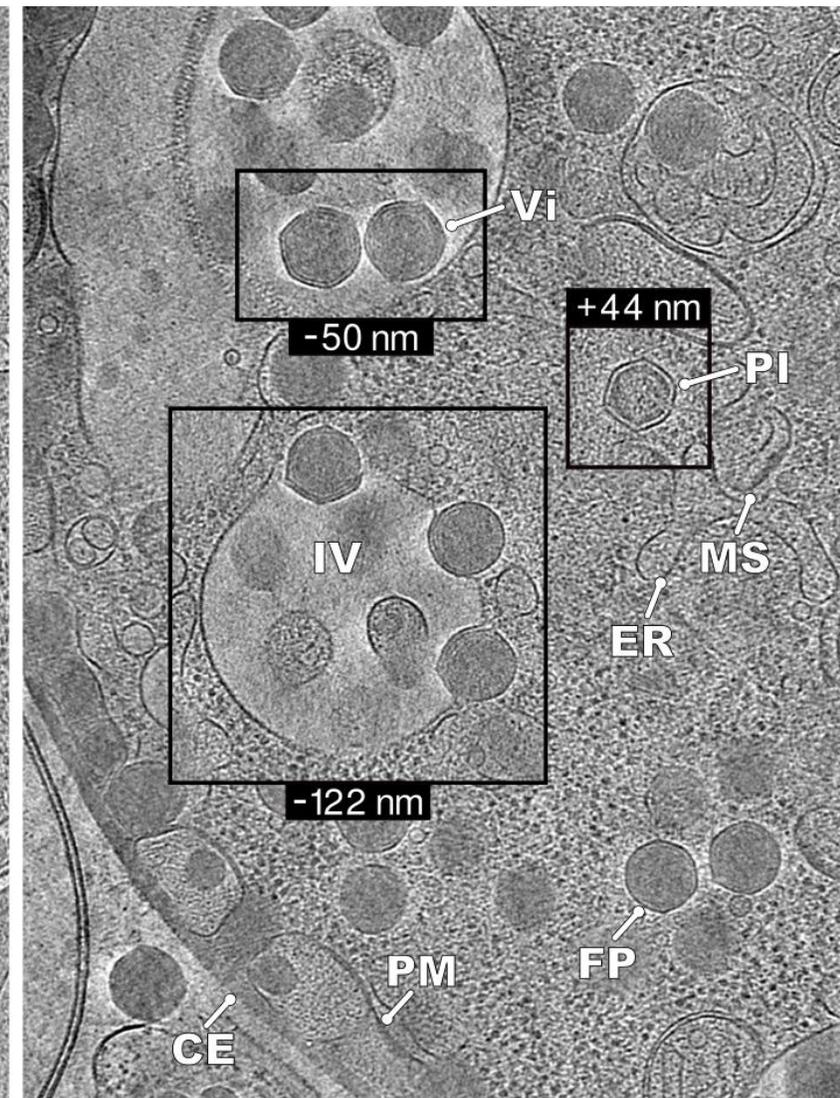
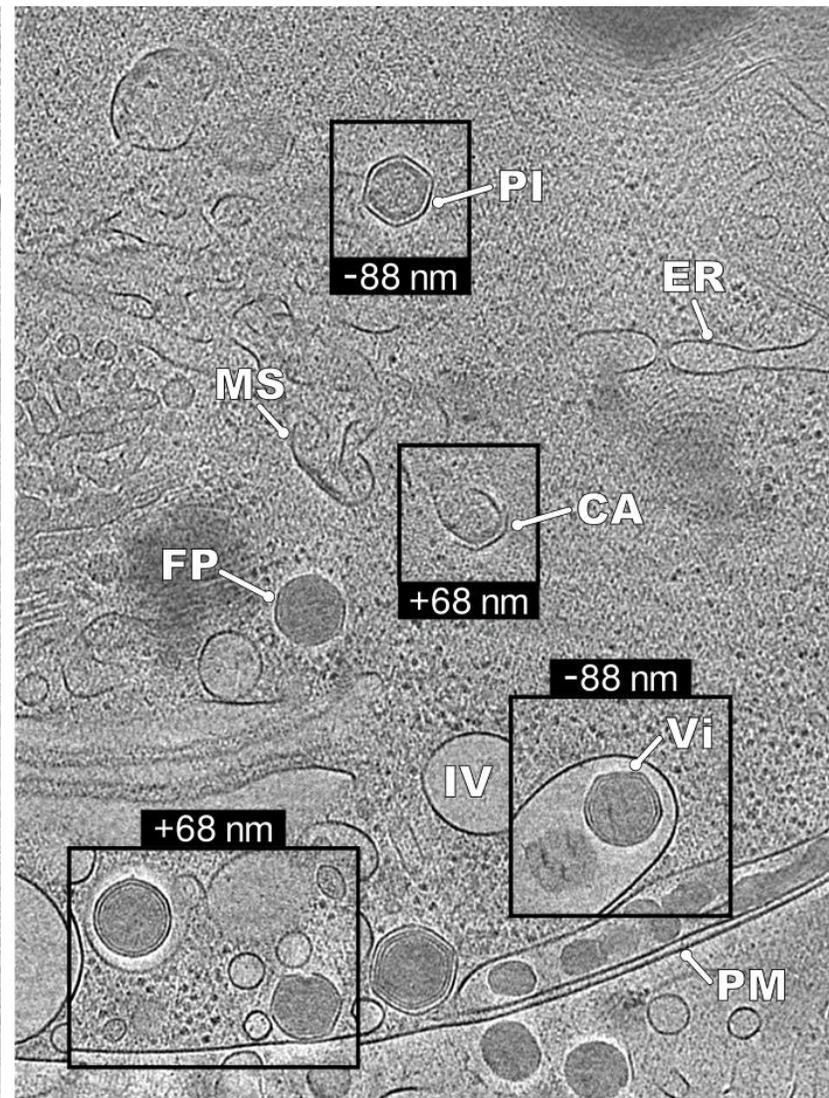
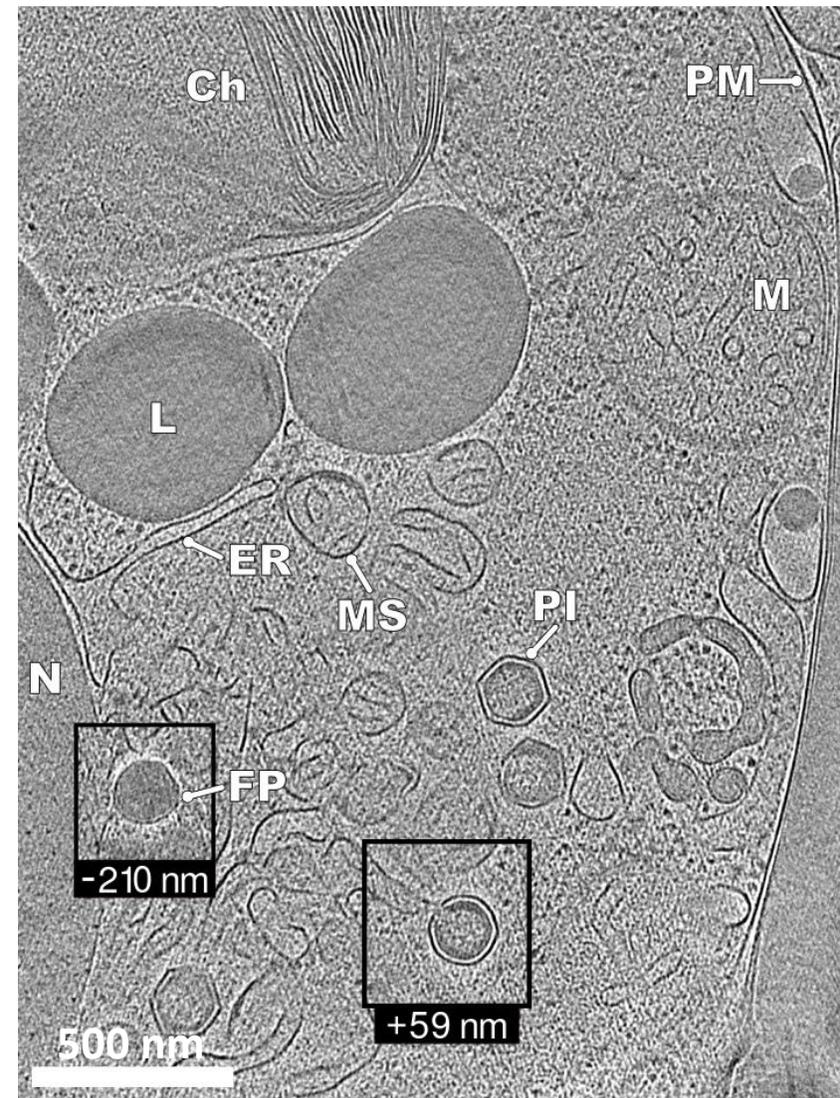
In situ EhV-201 life cycle - sample preparation



Tomography of healthy alga cell



Tomography of EhV-201 viral factories



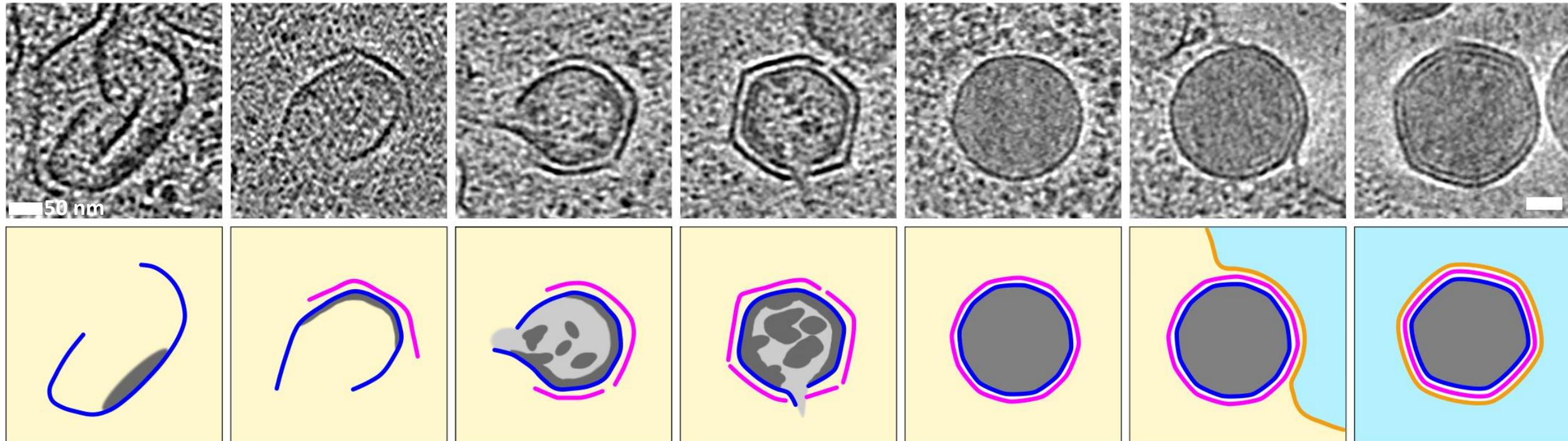
E. huxleyi infected cell

Late stage of infection with EhV-201

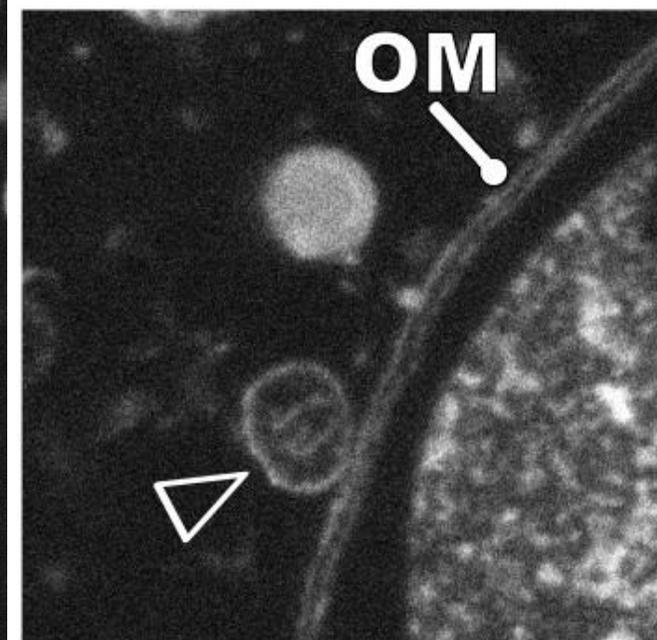
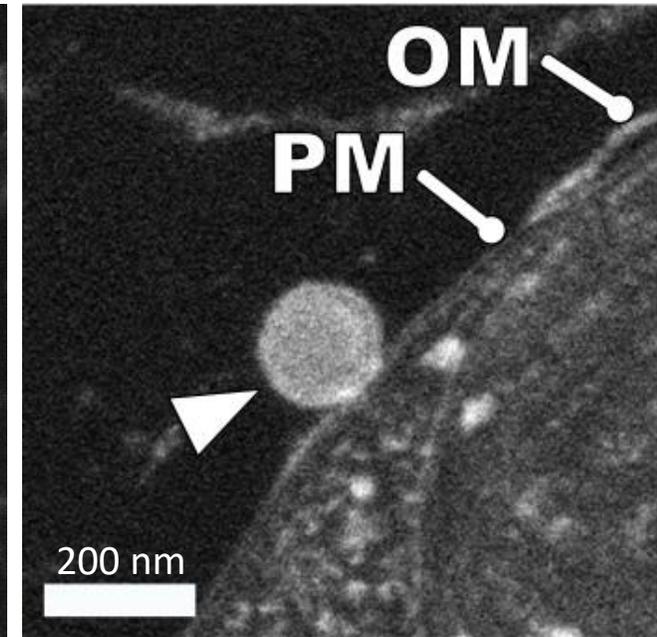
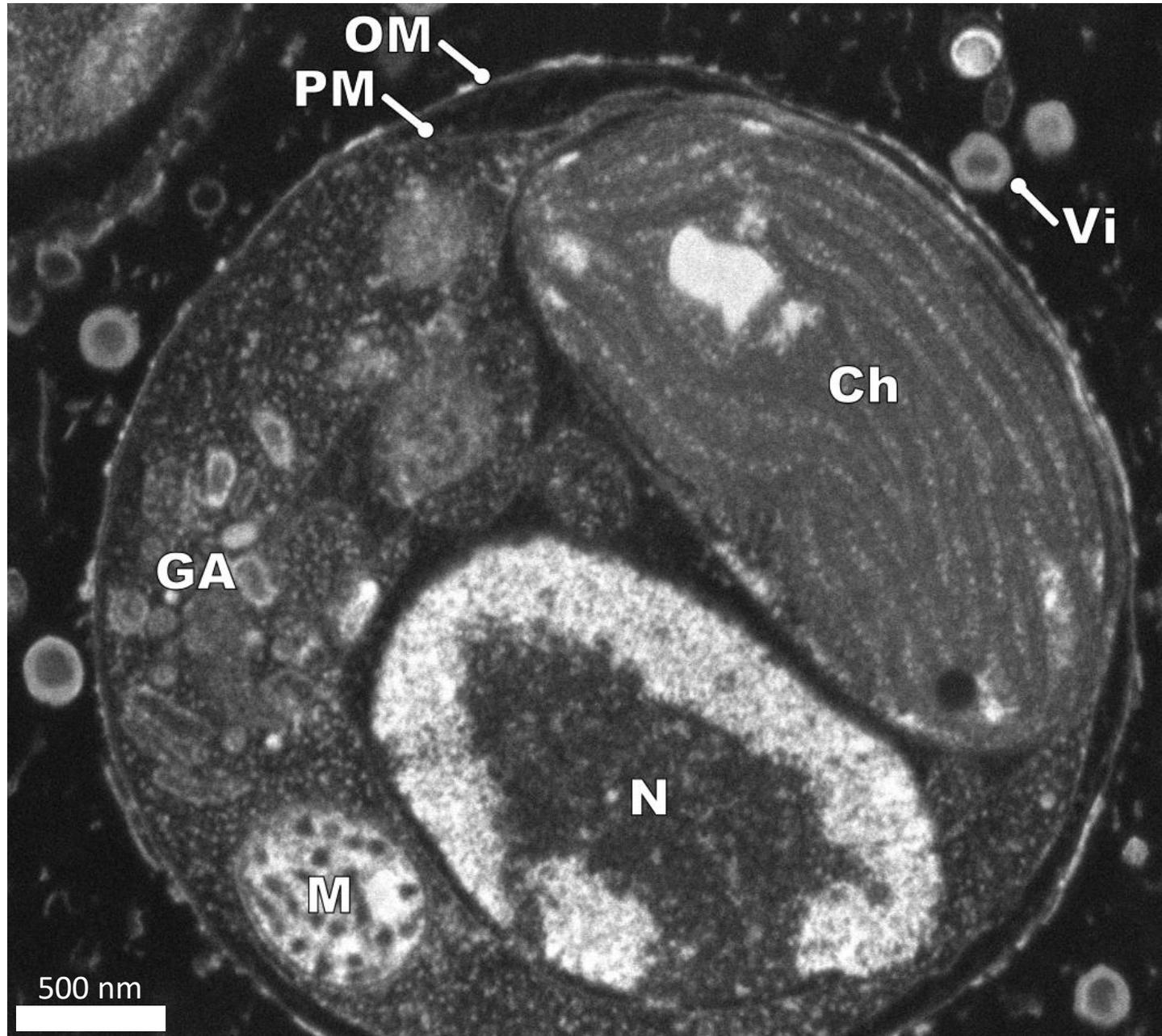
Cryo-lamella sample

Segmentation of a tomographic volume

EhV-201 virion assembly stages



EhV-201 attachment and genome delivery



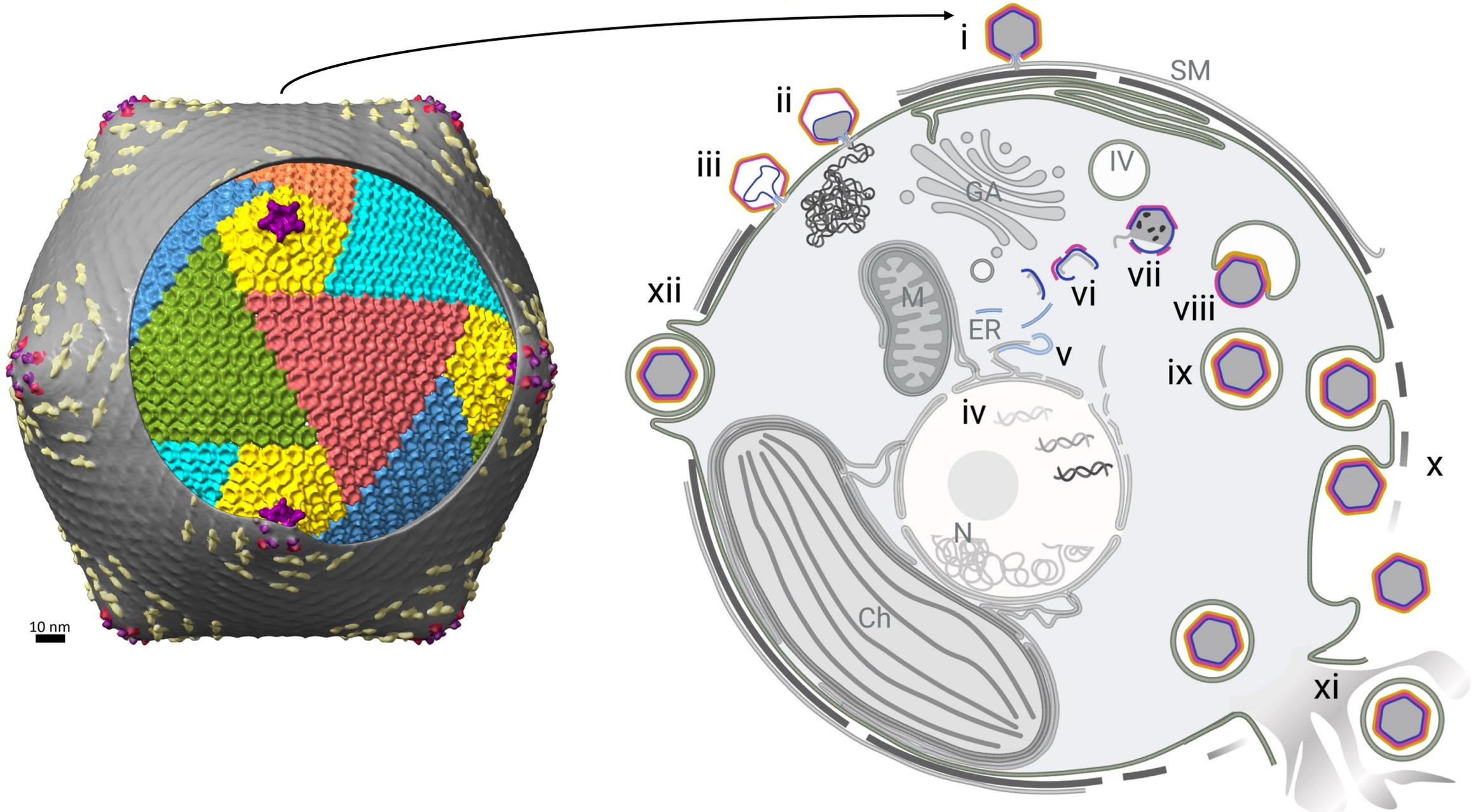
E. huxleyi infected cell

Early stage of infection with EhV-201 (30 min)

High-pressure frozen/freeze-substituted/resin-embedded sample

Serial block-face imaging

Conclusions



Acknowledgement



Pavel Plevka



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Jiří Nováček



Willie Wilson



Declan Schröder



Marten Chaillet



Friedrich Förster

