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RESEARCH

Weak coupling of neurons enables very high-frequency and ultra-fast oscillations through the interplay of synchronized phase shifts

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ABSTRACT

Recently, in the past decade, high-frequency oscillations (HFOs), very high-frequency oscillations (VHFOs), and ultra-fast oscillations (UFOs) were reported in epileptic patients with drug-resistant epilepsy. However, to this day, the physiological origin of these events has yet to be understood. Our study establishes a mathematical framework based on bifurcation theory for investigating the occurrence of VHFOs and UFOs in depth EEG signals of patients with focal epilepsy, focusing on the potential role of reduced connection strength between neurons in an epileptic focus. We demonstrate that synchronization of a weakly coupled network can generate very and ultra high-frequency signals detectable by nearby microelectrodes. In particular, we show that a bistability region enables the persistence of phase-shift synchronized clusters of neurons. This phenomenon is observed for different hippocampal neuron models, including Morris-Lecar, Destexhe-Paré, and an interneuron model. The mechanism seems to be robust for small coupling, and it also persists with random noise affecting the external current. Our findings suggest that weakened neuronal connections could contribute to the production of oscillations with frequencies above 1000 Hz, which could advance our understanding of epilepsy pathology and potentially improve treatment strategies. However, further exploration of various coupling types and complex network models is needed.

AUTHOR SUMMARY

We have built a mathematical framework to examine how a reduced neuronal coupling within an epileptic focus could lead to very high-frequency (VHFOs) and ultra-fast oscillations (UFOs) in depth EEG signals. By analyzing weakly coupled neurons, we found a bistability synchronization region where in-phase and anti-phase synchrony persist. These dynamics can be detected as very high-frequency EEG signals. The principle of weak coupling aligns with the disturbances in neuronal connections often observed in epilepsy; moreover, VHFOs are important markers of epileptogenicity. Our findings point to the potential significance of weakened neuronal connections in producing VHFOs and UFOs related to focal epilepsy. This could enhance our understanding of brain disorders. We emphasize the need for further investigations of weakly coupled neurons.

INTRODUCTION

It is known that hippocampal pyramidal neurons can switch from integrators to resonators, that is, from class I excitability to class II (Prescott, Ratté, De Koninck, & Seinowski, 2008). This dual operating mode allows for individual spikes, tonic spiking, and bursting, as well as different firing rates (Ermentrout & Terman, 2010; Izhikevich, 2006). Specifically, there is a maximum firing frequency above which the potential on the neuron membrane cannot oscillate faster; oscillations occur at a certain natural frequency for a given external stimulus. Although different types of neurons have different rates of action potential firing, these are physiologically limited (Gabbiani & Cox, 2017; Purves et al., 2019). The f-I curve (frequency given a fixed input I) typically saturates to a finite level not much above 350 Hz, even for the fastest spiking neurons; see Wang et al. (2016). Using mathematical conductance-based models, we can explain the onset and offset of the frequency-input curve. For the offset, one typically encounters a Hopf bifurcation or a limit point of cycles (Izhikevich, 2006; Kuznetsov, 2023). Biophysically plausible parameter settings then show we cannot expect higher frequencies. The dependence of the individual neuron model dynamics on the external current and other parameters has been studied extensively (Cessac & Samuelides, 2007; Duan & Lu, 2006; Shilnikov, 2012; Tsumoto, Kitajima, Yoshinaga, Aihara, & Kawakami, 2006; Xing, Song, Wang, Yang, & Chen, 2022). This biophysical basis of the mechanism is also confirmed by comparison with a real neuronal signal (Prescott, De Koninck, & Sejnowski, 2008).

Nevertheless, hippocampal electroencephalographic signals (EEG) show that highfrequency oscillations (HFOs) are reported in the seizure onset zone and are connected with epileptic seizure generation in humans with focal epilepsy (Cimbalnik et al., 2018, 2020; Jacobs et al., 2008, 2012; Jiruska et al., 2017; Pail et al., 2020; Řehulka et al., 2019; Staba & Bragin, 2011; Worrell & Gotman, 2011). Moreover, the extent of HFOs correlates with seizure frequency and disease severity (Zijlmans, Jacobs, Zelmann, Dubeau, & Gotman, 2009). HFOs are most commonly distinguished by frequency bands as interictal high gamma (65-100 Hz), ripples (100–250 Hz), and fast ripples (250–600 Hz). Recently, very high-frequency oscillations (VHFOs, 600-2000 Hz; Brázdil et al., 2017; Travnicek et al., 2021; Vasickova et al., 2023), very fast ripples (VFRs, 600–1000 Hz), and ultra-fast ripples (UFRs, 1000– 2000 Hz), and most recently ultra-fast oscillations (UFOs, over 2000 Hz) were reported in depth EEG recordings of patients with epilepsy (Brazdil et al., 2023). Since VHFOs were more spatially restricted in the brain than HFOs with lower frequencies, they have been suggested as novel biomarkers of the epileptogenic zone. Moreover, VFRs present intriguing synchronization phenomena, demonstrating strong phase-locking with slow oscillations (Hao et al., 2021). This highlights the crucial role synchronization mechanisms may play in the pathological progression and underlying propagation of epileptiform discharges within the hippocampal network during status epilepticus in temporal lobe epilepsy. However, the neurons themselves are unable to produce such high oscillation frequencies. This brings us to the question of whether multiple coupled neurons are capable of producing a synchronized signal of such high frequency.

Hopf bifurcation (H): A change in a dynamical system that is related to cycle birth from equilibrium.

Limit point of cycles (LPC): A point in a dynamical system representing the fold of a sequence of periodic orbits or cycles as a parameter changes.

Phase-locking:

A state in a dynamical system where oscillators synchronize their frequencies, maintaining a constant relative phase difference over time. It is known that synchronization occurs in oscillators for sufficiently strong coupling (Boccaletti, Pisarchik, Del Genio, & Amann, 2018; Pikovsky, Rosenblum, & Kurths, 2001; Strogatz, 2018; Wiggins, 2003) and even that it can occur through external noise (Lang, Lu, & Kurths, 2010; Nagai & Kori, 2010; Zhou & Kurths, 2002). At the same time, the dynamics of coupled oscillators may be complex, chaotic, or consist of chimera states (Abrams & Strogatz, 2004; Aihara, Takabe, & Toyoda, 1990; Calim, Hövel, Ozer, & Uzuntarla, 2018; Hoppensteadt & Izhikevich, 1997; Majhi, Bera, Ghosh, & Perc, 2019). Therefore, it is important to determine how coupled neuron models behave dynamically for physiological parameters and whether and, if so, how they can be synchronized with higher than natural frequencies, especially within VHFO and UFO frequency bands.

Within this paper, we consider near-identical neurons weakly connected through gap junctions. In this setup, we show that local field potentials can exhibit VHFOs and UFOs through phase synchronization. Moreover, we claim that it is not an exceptional phenomenon but quite the opposite in the case of very weak neuronal couplings. It arises for various connectivity types and neuron models. The mechanism is based on theoretical works on the symmetry of dynamical networks that can also be spatiotemporal and can lead to a rigid phase-shift synchrony (Golubitsky, Messi, & Spardy, 2016; Golubitsky, Romano, & Wang, 2012; Golubitsky & Stewart, 2006, 2016; Golubitsky, Stewart, & Török, 2005). We show that the occurrence of these higher frequencies is possible both in the case of different values of model parameters and in the case of random noise affecting the magnitude of the external current input.

In the Results section, we first discuss the Morris-Lecar neuron model, which we examine thoroughly. This includes a discussion of its fundamental dynamics, followed by an exploration of Morris-Lecar neuronal networks. We then analyze the behavior of two coupled Morris-Lecar neuron models, explain the phase shift, and reveal the concept of anti-phase collective synchrony in a model of a neuronal network. We show that we can achieve transient anti-phase synchrony by stimulating a subnetwork of neurons due to the existence of the bistability region. This concept can lead to an apparently higher frequency in the EEG signal. We want to demonstrate that the observed phenomena of phase-shift synchrony and bistability are generic for weak coupling and still hold for more physiological models of neurons. The analysis of the Morris-Lecar model is straightforward as the single-neuron model is twodimensional. The hippocampal neuronal network, however, is considerably more complex than the simple networks composed of Morris-Lecar neuron models, which exhibit a specific course of spike dynamics. We show that the bistability and anti-phase synchronization appear robustly in network models with more complex neuronal dynamics. One model is an interneuron network that exhibits VHFOs and UFOs with anti-phase locking, and the other is a Destexhe-Paré neuron network that shows VHFOs transiently. Similarly to the Morris-Lecar model, investigating the dynamics of two coupled interneurons, we find regions with synchronization and bistability and present simulations of the collective anti-phase behavior. We discuss transient ultra-fast ripples and ultra-fast oscillations in a two-level neuronal network. Lastly, simulations of the collective anti-phase behavior are presented for the Destexhe-Paré neuron model. In the Discussion and Conclusions section, we detail our primary objective to investigate whether weakened connections between neurons in an epileptic focus could result in high, very high-frequency, and ultra-fast oscillations; we examine how our findings support this theory and how they provide a mathematical framework for interpreting EEG signals from patients with focal epilepsy, while also highlighting the need for future research on the effects of different coupling types, strengths, and parameters on the dynamics of coupled neurons.

Local field potentials: Electrophysiological signals reflecting the summed electrical activity of a group of neurons in a local area.

Rigid:

In the context of dynamical systems, refers to behaviors that do not vary or change under a set of conditions.

Phase-shift synchrony:

A synchronous state where oscillating systems align their cycles but with a constant time delay or phase difference. Saddle-node on an invariant curve

system where a stable limit cycle is

A state in a dynamical system where

two equilibrium points, a stable and

created or destroyed alongside an

A critical event in a dynamical

Saddle-node equilibrium (LP):

an unstable one, merge and

Subcritical Hopf bifurcation:

A system transition where an

unstable limit cycle exists before

the bifurcation point and a stable equilibrium point loses stability

A critical point in a dynamical system

where a small change in system

qualitative change in its behavior.

parameter values results in a

annihilate each other.

after it.

Bifurcation:

(SNIC) bifurcation:

equilibrium point.

RESULTS

Model Dynamics Explaining the Tonic Spiking

Let us recall basic neuron spiking mechanisms that lead to tonic spiking clearly described in Prescott, De Koninck, and Sejnowski (2008). Class I neurons continuously increase their firing rate in response to a steady increase in input current without any abrupt change in their behavior. This type of behavior arises from a saddle-node on an invariant curve (SNIC) bifurcation, characterized by a break of a limit cycle on a saddle-node equilibrium as the input current is varied. On the contrary, class II neurons exhibit a discontinuous increase in their firing rate in response to a change in the input current, with a threshold for tonic spiking emergence. This behavior occurs near a subcritical Hopf bifurcation when a stable spiking orbit already exists due to a saddle-node of limit cycles with respect to the input current. The bifurcation diagram presented in Figure 1 demonstrates that the Morris-Lecar system Equation 11 shows both bifurcations at onset and offset as we vary the external input external current I_{extr} as mentioned. Note that class III neurons do not possess tonic spiking dynamics with a given natural frequency; hence, we will not focus on their type of mechanism. From now on, we will assume the neuron exhibits tonic spiking dynamics with a certain given natural frequency under a suitable external stimulus current.

External current and parameter changes influence the neuron natural frequency; see Figure 1. Membrane capacitance depends on the cell's and membrane's biophysical properties, including fluidity, thickness, permeability, and so forth. (Bakhtiari, Manshadi, Candas, & Beskok, 2023). Moreover, recently published studies (Patel, Tewari, Chaunsali, & Sontheimer, 2019; Tewari et al., 2018) propose pathological changes in the effective capacitance of neurons that may contribute to epilepsy. Therefore, the capacitance appears to be an appropriate parameter for the bifurcation analysis in relation to the emergence of HFOs, VHFOs, and UFOs in epilepsy patients. Similarly, one can investigate the dependence on other model parameters influencing the neuron frequency, for example, φ and I_{ext} (e.g., see Liu, Liu, & Liu, 2014; Tsumoto et al., 2006; Xing et al., 2022) or g_{Ca} , g_{K} , and g_L that can also be influenced by drugs (see Macdonald, 1989; Nicholson, Blanche, Mansfield, & Tran, 2002).



Figure 1. Morris–Lecar model dynamics with respect to the applied external current Iext. Left: The bifurcation diagram shows a limit point (LP) for $I_{ext} \approx 40$ where a saddle-node on an invariant curve (SNIC) leads to a stable cycle, which exists for a wide range of external current values (tonic spiking region) until the limit point of cycles near $I_{ext} = 140$. The branch of cycles turns and becomes unstable, and ends at a subcritical Hopf bifurcation (H). Beyond the LPC, neuronal excitation is impossible. The other parameter values are listed in Table 1, and the membrane capacitance is C = 5 for simplicity. Right: The natural firing frequency as a function of external current I_{ext} for various membrane capacitances C. As the capacitance increases, only the frequency of the oscillation decreases but does not affect the existence of the limit cycle. Other parameter values are listed in Table 1.

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Two Coupled Morris-Lecar Neuron Models

Two-dimensional invariant torus: A set of solutions in a dynamical system that form a toroidal shape, wherein trajectories neither converge nor diverge but remain on the torus.

Quasi-periodic:

A behavior in a dynamical system with oscillations at two or more incommensurable (irrational ratio) frequencies, creating a nonrepeating pattern.





Figure 2. Dynamics in the system (Equation 15) for $\varepsilon = 0.05$, various values of C_1 , and fixed $C_2 = 1$ (see Figure 4). *Left*: projections of orbits (blue dotted line), and stable (blue solid line) and unstable (red dashed line) cycles onto the state space $V_1 \times V_2$ found by continuation. *Right*: time course of V_1 (blue) and V_2 (red) of the blue orbits in the left panel. Other parameter values are listed in Table 1. Choosing C_1 affects the dynamics: (A) $C_1 = 2.75$, quasi-periodicity. (B) $C_1 = 1.60$, a stable in-phase periodic solution (IPS; the bold blue cycle on the left state space $V_1 \times V_2$). (C) $C_1 = 0.90$, bistability, the system possesses stable in-phase and anti-phase periodic solutions (IPS and APS; the bold blue cycles on the left state space $V_1 \times V_2$). (D) $C_1 = 0.581$, bistability, anti-phase cycle lost stability through a period-doubling bifurcation (PD).

Period-doubling bifurcation (PD): A transition in a dynamical system where a small change in parameters causes the period of oscillations to double, often preceding chaos.

Phase Shift for Two Weakly Coupled Morris-Lecar Neuron Models

Two identical oscillators form a symmetrical dynamical system with a symmetrical periodic solution. The symmetry of the system is roughly preserved even for non-identical oscillators due to the continuous dependence of the solutions on parameters (see Perko, 2001), that is, coupled neurons with similar phase dynamics form a nearly symmetrical system with a stable limit cycle such that the phases of both tonically spiking neurons are mutually shifted, although having a common frequency roughly the magnitude of the natural frequencies of the individual neurons. This phenomenon, so-called phase-locking, and the theoretical background for this mechanism is based on Golubitsky and Stewart's results of equivariant bifurcation theory allowing groupoid formalism for networks of systems of differential equations (Golubitsky et al., 2012, 2016; Golubitsky & Stewart, 2006, 2016; Golubitsky et al., 2005; Nijholt, Rink, & Sanders, 2019). The symmetry of the network with a T-periodic solution implies the existence and persistence of a rigid phase-shift synchrony. In our case of two nearly identical weakly coupled oscillators (Equation 15), there may also exist solutions synchronized with time shift T/2 for appropriate system parameters: the attracting stable cycle, corresponding to the presence of an anti-phase synchronous state (see Figure 2C). Simultaneously, the system possesses a solution reflecting in-phase synchrony where the neurons oscillate almost identically; see Figures 2B, 2C (middle), and 2D (middle).

Both branches of stable periodic solutions, that is, in-phase and anti-phase synchrony, persist inside relatively wide regions of the parameter space; see Figure 3. This fact is also depicted in Figure 4 where the bifurcation diagrams with respect to C_1 and C_2 , and ε and C_1 , respectively, are shown. More accurately, Figure 3 precisely corresponds to the dashed sections at $C_2 = 1$



Figure 3. Frequency of the limit cycles in the system (Equation 15) of two coupled Morris–Lecar neuron models with respect to C_1 . The remaining parameter values are identical (see Table 1), $C_2 = 1$, and $\varepsilon = 0.05$. The upper and bottom blue branches correspond to the stable in-phase and anti-phase synchrony, respectively. The dashed sections at the given parameter values in Figure 4 correspond to these depicted branches. Notice the bistability, present for a relatively wide range of C_1 . For completeness, we remark that there exist more and more complex periodic solutions of the system (Equation 15) for C_1 between the LPC₃ and PD₁ points, and between PD₂ and PD₃ points that are not depicted in this figure. The dashed line at $C_1 = 1.2$ denotes the frequencies at IPS near 30 Hz and APS near 26 Hz that reappear in the simulations depicted in Figures S1 and S2 in the Supporting Information.



Figure 4. Arnold tongues in parameter spaces (A) $C_1 \times C_2$ and (B) $C_1 \times \varepsilon$ for two coupled Morris–Lecar neuron models (Equation 15). The regions are delineated by blue and black solid LPC curves corresponding to the IPS and APS, respectively. The blue dash-dotted curves refer to PD of the anti-phase cycle. The background color indicates the dominant frequency in a simulated composed signal $V = V_1 + V_2$. In the region bounded by the PD curves, the system may exhibit more complex, even chaotic dynamics. The red region corresponds to the stable anti-phase cycle, while the other solutions (yellow) have a frequency close to the natural frequency. Compare with Figure 2: (A) quasi-periodicity outside Arnold tongues; (B) IPS inside the outer Arnold tongue; (C) bistability in the red region delineated by LPC curves and PD curves; (D) APS lost stability due to PD, frequency halves.

Arnold tongues:

Regions in parameter space indicating synchronization or frequency-locking ranges in dynamical systems, typically with a characteristic 'tongue' shape. and $\varepsilon = 0.05$ in Figure 4. The regions, so-called Arnold tongues or resonance tongues (Kuznetsov, 2023; Wiggins, 2003), are delineated by borders belonging to the limit point of cycle (LPC) manifolds. Outside the Arnold tongues, the difference in capacitances C_1 and C_2 implies a synchronous state with heterogeneous frequencies (see Figure 1), quasi-periodic oscillations, or more complex dynamics. The LPC manifolds were computed for parameters listed in Table 1. Since interchanging C_1 and C_2 mirrors the synchronization regions, the black and blue solid curves in Figure 4A are symmetrical. Cusps in the intersections of two branches of LPC manifolds are co-dimension two bifurcation points (Kuznetsov, 2023). Although the LPC curves in Figure 4 were computed for given parameters from Table 1, the generality of bifurcation theory guarantees that these LPC manifolds persist for nearby parameter values.

Similarly, the bifurcation diagram in Figure 4B shows the Arnold tongues with respect to the coupling strength ε and membrane capacitance C_1 . In this case, they emanate from the point $C_1 = C_2$ (other parameters are identical) since the limit case $\varepsilon = 0$ belongs to the uncoupled pair of neurons with identical frequencies. Generally, nonidentical similar neurons will have similar intrinsic frequencies, and the Arnold tongue would emanate from a cusp point with the same frequency, but the membrane capacitances C_1 , C_2 , as well as other parameters, would differ slightly. Moreover, results presented in Kobelevskiy (2008), which deals with Morris–Lecar models that incorporate time-delayed gap-junctional coupling, suggest that similar dynamics will emerge in such models. Furthermore, an increase in delay may even play a stabilizing role.

The background color in the bifurcation diagrams in Figure 4A and 4B indicates the dominant frequency detected in a composed signal $V = V_1 + V_2$ simulated on a time interval [200, 1,200], discarding 200 ms transients, for parameter values from a 401 × 401 grid and appropriate initial conditions. These regions may not match precisely since the two-cycle created by crossing the PD curves has a similar spike shape to the cycle corresponding to the APS (see Figure 2D). Furthermore, around the point [1, 0] in Figure 4B, that is, for very weak coupling

• /			
	Setting	Unit	Meaning
С	0.1–2.6	μ F/cm ²	membrane capacitance
g_{L}	2	mS/cm ²	maximum leak conductance
g Ca	4	mS/cm ²	maximum Ca ²⁺ conductance
gк	8	mS/cm ²	maximum K ⁺ conductance
V_{L}	-60	mV	equilibrium potential of leak channel
V_{Ca}	120	mV	equilibrium potential of Ca ²⁺ channel
$V_{\rm K}$	-80	mV	equilibrium potential of K ⁺ channel
β_1	-1.2	mV	tuning parameters for Ca ²⁺ activation function
β_2	18	mV	
β_3	10	mV	tuning parameters for K ⁺ activation function and time function τ_w
β_4	17.4	mV	
φ	1/15	s^{-1}	reference frequency
<i>I</i> _{ext}	43	μ A/cm ²	externally applied direct current

 Table 1.
 Typical physiological parameter values for the Morris–Lecar neuron model of a hippocampal pyramidal cell (Gutkin & Ermentrout, 1998)

and similar membrane capacitances, the observed dominant frequency of \sim 60 Hz in the composition is caused by long-lasting transient dynamics.

Anti-Phase Synchrony in a Model of Morris–Lecar Neuronal Network Leading to Higher Collective Frequency

A similar approach can be used for more complex networks of (nearly) identical neurons with symmetry. The phenomenon leads to complete collective synchrony or phase-shift synchrony in the network of neurons.

Experiments, simulations, and theory show that anti-phase collective synchronization is possible (Chowdhury, Rakshit, Buldu, Ghosh, & Hens, 2021; Kawamura, Nakao, Arai, Kori, & Kuramoto, 2010; Sebek, Kawamura, Nott, & Kiss, 2019). Ermentrout and Terman (2010) previously reported on the anti-phase dynamics of two Morris–Lecar neurons with synaptic coupling; however, our findings demonstrate that this phenomenon is not exceptional and has the potential to lead to a rise of higher frequency in the summed signal. Furthermore, we have observed similar collective dynamics in a network, highlighting the robustness of this phenomenon across different scales of neuronal systems.

Consider a neuronal network model (Equation 15) of N = 50 weakly coupled Morris–Lecar neurons. Based on the results presented in the previous subsection, one can expect the presence of multistability in this system, that is, the coexistence of multiple stable solutions corresponding to various synchronous states depending on the initial conditions and varied model parameters ε and C_i , as is depicted in Figures S1 and S2 in the Supporting Information.

Specifically, for the demonstration of the collective anti-phase behavior, let us suppose identical coupling strength $\varepsilon = 0.1/50 = 0.002$. Further, we will consider that the heterogeneity

Synaptic coupling:

The process where activity in one neuron influences another through synaptic transmission, facilitating communication and synchronization between neurons. of neurons results from differences in their membrane capacitances (one may assume similar heterogeneity in other parameters, as mentioned before). Let C_i be independent and follow a truncated normal (TN) distribution defined on the interval [0.75, 1.05] to avoid unrealistic or extreme values, that is, $C_i \sim \text{TN}(0.90, 0.05^2, 0.75, 1.05)$, while the remaining parameters stay identical (see Table 1). We base our choice for the mean and standard deviation of neuron membrane capacitances C_i on direct measurements from Gentet, Stuart, and Clements (2000). To account for possible pathological changes near the epileptic focus, we also explored high values of the standard deviation. Finally, let the population consist of two sub-populations of roughly the same size, $\mathcal{P}_1 = \{1, ..., 25\}$, $\mathcal{P}_2 = \{26, ..., 50\}$, that have random initial conditions that partially overlap

$$V_{i}(0) \sim \mathsf{TN}(-35, 5^{2}, -50, -20), i \in \mathcal{P}_{1} \cup \mathcal{P}_{2}, w_{i}(0) \sim \begin{cases} \mathsf{TN}(0.01, 0.025^{2}, 0, 0.085), & i \in \mathcal{P}_{1}, \\ \mathsf{TN}(0.06, 0.025^{2}, 0, 0.135), & i \in \mathcal{P}_{2}. \end{cases}$$
(1)

Also, we set $I_{\text{ext}} \sim N(43, 1^2)$.

Figure 5 shows that this setting leads to anti-phase synchronous oscillations of the two clusters, resulting in a double frequency in the composed signal, as one can see in the periodograms of the clusters and the composition, respectively. Namely, the observed frequency increased to \sim 56 Hz. Notice that individual neurons can jump from one cluster to the other due to the noise, while the global behavior remains unchanged.



Figure 5. (A) Collective APS of two subpopulations in a neuronal network model (Equation 15) composed of 50 all-to-all coupled Morris– Lecar neurons with heterogeneous membrane capacitances $C_i \sim \text{TN}(0.90, 0.05^2, 0.75, 1.05)$ and random initial conditions in Equation 1. This setting leads to a doubled dominant frequency in (B) the composed signal $V = \sum_{i=1}^{50} V_i$. Zooming in on the dynamics of (C) all neurons, (D) composed signal, and the corresponding periodograms of (E) individual neurons and (F) the composed signal. The robustness of the anti-phase behavior is demonstrated by a noisy applied current $I_{\text{ext}} \sim \text{N}(43, 1^2)$ at each simulation step $\Delta t = 0.01$. The remaining parameter values are identical (see Table 1), $\varepsilon = 0.002$.

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We acknowledge that the very high-frequency events are only transient phenomena in the experimental observations. We have just shown that with small levels of noise, the anti-phase solution persists. However, higher levels typically lead to complete synchronization (Neiman, Schimansky-Geier, Cornell-Bell, & Moss, 1999; Pikovsky et al., 2001; Zhou & Kurths, 2003). Nevertheless, Figure 6 demonstrates that higher frequencies may temporally appear through external inputs. Let us change the applied current to $I_{ext} = 43 + p_i(t) + \sigma dW$ with $\sigma = 2.0$ and $p_i = 40$ if $t \in [716, 718]$ ms in Equation 15 for $i \in \mathcal{P}_1$. This stimulus is applied only to half of the population, that is, $p_i = 0$ for $i \in \mathcal{P}_2$. We initialize the population into the anti-phase behavior and subsequently observe that the population synchronizes within 100 ms. Looking at the same time interval as before, the external stimulus arrives to halve the population evoking anti-phase behavior that lasts for ~200 ms; then the population synchronizes again. As the periodograms and time series show, it is evident that the higher frequencies are temporally present. This transient phase solution also appears if we stimulate fewer neurons but then lasts only a few cycles (data not shown). This transient phenomenon depends on the timing of the stimulus as a stimulus during different phases of the period does not induce the anti-phase solution.

Model Dynamics for Two Coupled Interneurons

The dynamics of two interacting interneurons coupled through gap-junctional connections has already been studied in Chow and Kopell (2000) and Skinner, Zhang, Velazquez, and Carlen (1999). At low coupling strengths and very high firing rates, the synchronous state is unstable, and a pair of cells fires in anti-phase synchrony, while for a lower range of frequencies, the in-phase and anti-phase synchronies may be bistable (Chow & Kopell, 2000). Moreover, Skinner



Figure 6. (A) Transient collective APS in a neuronal network model (Equation 15) composed of 50 all-to-all coupled Morris–Lecar neurons with heterogeneous membrane capacitances $C_i \sim \text{TN}(1.04, 0.03^2, 0.95, 1.13)$ and random initial conditions in Equation 1. The parameter values remain unchanged from the previous setting, with $\varepsilon = 0.002$. The APS is evoked by 2 ms lasting additional stimulus at t = 716 (arrow). Zooming in on the dynamics of (B) all neurons, (D) composed signal, and the corresponding periodograms of (C) individual neurons and (E) the composed signal.

et al. (1999) found that blocking the inhibition in a network of interneurons coupled by gap junctions and inhibitory synapses can lead to anti-phase bursting with higher frequency.

Similarly, as in the case of Morris–Lecar neuron, let us focus briefly on the dynamics of the interneuron model with respect to parameters C_1 , C_2 , and ε . One can proceed analogously with respect to other parameters affecting the frequency of the membrane potential. Figure 7 shows Arnold tongues in parameter spaces $C_1 \times C_2$ and $C_1 \times \varepsilon$ (*f–I* curves for a single interneuron for various values of *C* are provided in the Supporting Information (see Figure S4); the default parameter choice of $I_{ext} = 24$ leads to a frequency of 335 Hz). The blue LPC curves delineate a pair of Arnold tongues corresponding to two symmetrical in-phase solutions; the black tongue refers to the anti-phase synchrony. For the sake of completeness, the red dashed line denotes the LPC curves of two unstable periodic solutions related to the APS. The background color illustrates the dominant frequency detected in a composed signal $V = V_1 + V_2$ simulated on a time interval [500, 1,500], discarding 500 ms transients, for parameter values from a 401 × 401 grid and appropriate initial conditions. These regions do not match precisely, for example, around the point [1, 0] in Figure 7B, that is, for very weak coupling and similar membrane capacitances, where the observed dominant frequency of ~670 Hz in the composition is caused by transient dynamics (similar to the behavior depicted in Figure 8).

Simulation of the Transient Collective Anti-Phase Behavior in a Neuronal Network of Coupled Interneurons

Consider the neuronal network (Equation 17 and 19) composed of 50 interacting interneurons formed into two clusters, $P_1 = \{1, ..., 25\}$ and $P_2 = \{26, ..., 50\}$, respectively. Further, for the demonstration of the transient anti-phase collective behavior in this system, let the coupling strength take the form

$$\varepsilon_{ij} = \begin{cases} 0.001, & (i,j) \in \mathcal{P}_1 \times \mathcal{P}_1 \cup \mathcal{P}_2 \times \mathcal{P}_2, \\ 0.0002, & \text{else}, \end{cases}$$
(2)



Figure 7. Arnold tongues in parameter spaces (A) $C_1 \times C_2$ and (B) $C_1 \times \varepsilon$ for two coupled interneuron models (Equation 17 and 19) with $I_{ext} = 24$. The regions are delineated by blue and black LPC curves corresponding to the IPS and APS, respectively. The red dashed curves refer to the LPC bifurcation of two unstable periodic solutions. The background color indicates the dominant frequency in a simulated composed signal $V = V_1 + V_2$, reaching up to ~750 Hz. The yellow region inside the APS tongue delimited by black curves belongs to the signals where the multiple frequency persists, but is not dominant.



Figure 8. (A) Transient anti-phase dynamics of two subpopulations in a neuronal network model (Equation 17 and 19) composed of 50 all-toall coupled interneurons with heterogeneous membrane capacitances $C_i \sim \text{TN}(1, 0.03^2, 0.91, 1.09)$, coupling (Equation 2), and initial conditions (Equation 3). This configuration leads to the rise of VHFOs in (B) the composed signal $V = \sum_{i=1}^{50} V_i$. Dynamics of (C) all neurons are zoomed, (D) composed, and depicted with periodograms of (E) individual neurons and (F) the composed signal. The robustness of the antiphase behavior is demonstrated by a noisy applied direct current $I_{\text{ext}} \sim \text{N}(20, 1^2)$ at each simulation step $\Delta t = 0.01$. The remaining parameter values are identical.

where the first row refers to the coupling within each subpopulation, while the second one indicates a weaker connection between them.

Finally, assume that the clusters start in roughly opposite phases, specifically,

$$V_i(0) = \begin{cases} 40, & i \in \mathcal{P}_1, \\ -40, & i \in \mathcal{P}_2, \end{cases} \quad h_i(0) = 0.25, \quad n_i(0) = 0.5, \quad i \in \mathcal{P}_1 \cup \mathcal{P}_2.$$
(3)

Figure 8 reveals the transient anti-phase behavior of these clusters. The periodograms suggest that this dynamics may result in the emergence of VHFOs in the simulation of the composed EEG signal. Namely, summing the signals of individual interneurons resulted in an oscillation with a frequency of ~610 Hz. Moreover, although transient, the VHFOs may sustain for a relatively long time interval, significantly exceeding the duration of VHFOs observed in real EEG signals (Brázdil et al., 2017; Cimbalnik et al., 2020; Travnicek et al., 2021).

Transient UFRs and UFOs in a Two-Level Neuronal Network of Coupled Interneurons

Once more, consider the neuronal network (Equation 17 and 19) composed of four interacting interneurons formed into two clusters, $P_1 = \{1,2\}$ and $P_2 = \{3,4\}$, respectively. Suppose that the network connectivity is given by the coupling

$$\varepsilon_{ij} = \begin{cases} 0.0125, & (i,j) \in \mathcal{P}_1 \times \mathcal{P}_1 \cup \mathcal{P}_2 \times \mathcal{P}_2, \\ 0.0025, & \text{else}; \end{cases}$$
(4)



Figure 9. Network topology leading to apparent UFRs; see Figure 10.

see Figure 9. And finally, assume the following initial conditions:

$$V_i(0) = \begin{cases} 40, & i \in \{1,3\}, \\ -40, & i \in \{2,4\}, \end{cases} \quad h_i(0) = \begin{cases} 0.25, & i \in \mathcal{P}_1, \\ 0.27, & i \in \mathcal{P}_2, \end{cases} \quad n_i(0) = \begin{cases} 0.50, & i \in \mathcal{P}_1, \\ 0.52, & i \in \mathcal{P}_2. \end{cases}$$
(5)

As one can see in Figure 10, this configuration leads to the emergence of transient UFRs in the composed signal $V = \sum_{i=1}^{4} V_i$. Specifically, in spite of the noisy applied direct current, the signal simulation contains a segment reporting a frequency of ~1335 Hz. Despite being transient, the simulated UFRs exhibit a prolonged duration compared to the UFRs and UFOs observed in actual EEG records (Brazdil et al., 2023; Travnicek et al., 2021).



Figure 10. (A) Transient anti-phase dynamics of two subpopulations in a neuronal network model (Equation 17 and 19) composed of four allto-all coupled interneurons with heterogeneous membrane capacitances $C_1 = 0.998$, $C_2 = 0.999$, $C_3 = 1$, $C_4 = 1.001$, coupling (Equation 4), and initial conditions (Equation 5). This configuration results in the emergence of UFRs in (B) the composed signal $V = \sum_{i=1}^{4} V_i$. Dynamics of (C) all neurons are zoomed, (D) composed, and depicted with periodograms of (E) individual neurons and (F) the composed signal. The robustness of the anti-phase behavior is demonstrated by a noisy applied direct current $I_{ext} \sim N(24, 1^2)$ at each simulation step $\Delta t = 0.01$. The remaining parameter values are identical.

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Figure 11. Network topology leading to apparent UFOs; see Figure 12.

Moreover, let us extend the previous neuronal network model by a third pair of interneurons. Specifically, consider six interacting interneurons organized into three clusters, $\mathcal{P}_1 = \{1,2\}$, $\mathcal{P}_2 = \{3,4\}$, and $\mathcal{P}_3 = \{5,6\}$, respectively; assume that the network connectivity is determined by the coupling

$$\varepsilon_{ij} = \begin{cases} 0.01, & (i,j) \in \mathcal{P}_1 \times \mathcal{P}_1 \cup \mathcal{P}_2 \times \mathcal{P}_2 \cup \mathcal{P}_3 \times \mathcal{P}_3, \\ 0.001, & \text{else}; \end{cases}$$
(6)

see Figure 11. Finally, let the initial state of the system satisfy

$$V_i(0) = \begin{cases} 40, & i \in \{1,3,5\}, \\ -40, & i \in \{2,4,6\}, \end{cases} \quad h_i(0) = \begin{cases} 0.25, & i \in \mathcal{P}_1, \\ 0.26, & i \in \mathcal{P}_2, \\ 0.27, & i \in \mathcal{P}_3, \end{cases} \quad n_i(0) = \begin{cases} 0.50, & i \in \mathcal{P}_1, \\ 0.51, & i \in \mathcal{P}_2, \\ 0.52, & i \in \mathcal{P}_3, \end{cases}$$
(7)

characterizing anti-phase synchrony within each cluster and slightly shifted phases between clusters.

As Figure 12 shows, this setting results in the emergence of UFOs in the composed signal $V = \sum_{i=1}^{6} V_i$. Namely, despite the noisy applied direct current, after a transient period, the simulated signal composition oscillates with a frequency of ~2 kHz.

Simulation of the Collective Anti-Phase Behavior in a Model of Destexhe–Paré Neuronal Network

Let us take into account the neuronal network (Equation 20 and 21) comprised of 50 coupled Destexhe–Paré neurons formed into two clusters, $\mathcal{P}_1 = \{1,...,25\}$ and $\mathcal{P}_2 = \{26,...,50\}$, respectively. Unlike previous cases, here we assume that the distribution of capacitances differs in the mean value between these two groups. Namely, let

$$C_i \sim \text{TN}(0.95, 0.03^2, 0.86, 1.04), \quad i \in \mathcal{P}_1, \text{ and } C_i \sim \text{TN}(1, 0.03^2, 0.91, 1.09), \quad i \in \mathcal{P}_2.$$
 (8)

Let the network connectivity take the form

$$\varepsilon_{ij} = \begin{cases} 0.005, & (i,j) \in \mathcal{P}_1 \times \mathcal{P}_1 \cup \mathcal{P}_2 \times \mathcal{P}_2, \\ 0.001, & \text{else}, \end{cases}$$
(9)

characterizing the coupling strength within each subpopulation and between them, respectively.

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Figure 12. Stable dynamics of three subpopulations in a neuronal network model (Equation 17 and 19) comprising six all-to-all coupled interneurons with heterogeneous membrane capacitances $C_i \sim \text{TN}(1, 0.001^2, 0.997, 1.003)$, coupling (Equation 6), and initial conditions (Equation 7). This configuration results in the birth of UFOs in (A), (B) the composed signal $V = \sum_{i=1}^{6} V_i$. Dynamics of (C) all neurons are shown in detail, (D) composed, and depicted with periodograms of (E) individual neurons and (F) the composed signal. The robustness of the regular shift-phase behavior is demonstrated by a noisy applied direct current $I_{\text{ext}} \sim \text{N}(24,0.03^2)$ at each simulation step $\Delta t = 0.01$. The remaining parameter values are identical.

And finally, for the demonstration of transient anti-phase behavior in this network, suppose that the clusters start in roughly opposite phases; specifically, let

$$V_i(0) = \begin{cases} -75, & i \in \mathcal{P}_1, \\ 35, & i \in \mathcal{P}_2, \end{cases} \quad m_i(0) = 0.5, \quad h_i(0) = 0.2, \quad n_i(0) = 0.4, \quad m_{\mathsf{M},i}(0) = 0.24, \quad i \in \mathcal{P}_1 \cup \mathcal{P}_2.$$

$$(10)$$

The transient anti-phase behavior of these clusters is presented in Figure 13. At the bottom, one can see signal segments with the examined dynamics. The periodograms corresponding to these segments demonstrate that the VHFOs are present in the composed EEG signal simulation. To be more specific, the combination of signals from each neuron leads to an oscillation with a frequency of ~740 Hz (*f–I* curves for a single neuron for various values of *C* are provided in the Supporting Information (see Figure S5); the default parameter choice of I_{ext} = 40 leads to a frequency of 360 Hz). Although transient, the VHFOs can persist for a relatively long period, significantly surpassing the length of VHFOs detected in actual EEG signals (Brázdil et al., 2017; Brazdil et al., 2023; Cimbalnik et al., 2020; Travnicek et al., 2021).



Figure 13. (A) Transient anti-phase dynamics of subpopulations \mathcal{P}_1 , \mathcal{P}_2 in a neuronal network model Equation 20 and 21 composed of 50 allto-all coupled Destexhe–Paré neurons with heterogeneous membrane capacitances (Equation 8), coupling (Equation 9), and initial conditions (Equation 10). Dynamics of (B) composed signal, zoomed dynamics of (C) individual neurons, and (D) composed signal are depicted with periodograms of (E) individual neurons and (F) the composed signal. The robustness of the anti-phase behavior is demonstrated by a noisy applied direct current $I_{\text{ext}} \sim N(40, 1^2)$ at each simulation step $\Delta t = 0.01$. The remaining parameter values are identical.

DISCUSSION AND CONCLUSIONS

The primary objective of our paper was to establish a rigorous basis for the possibility that a reduction in the strength of the connections between neurons in an epileptic focus could produce high, very high-frequency, and ultra-fast oscillations measured at microelectrodes in its vicinity. Although such oscillations are biomarkers used for diagnostic purposes in presurgical evaluation, there is still no definitive way to differentiate between physiological (most commonly associated with the normal physiological function of cognitive or sensory processing (see Pail et al., 2020) and pathological HFOs in EEG signals (Frauscher et al., 2018). There are recent studies (Cimbalnik et al., 2018, 2020; Nejedly et al., 2019) that introduced algorithms for the detection of pathological HFOs using pathological events visually marked by expert reviewers inside the seizure onset zone of patients with focal epilepsy (detection training and machine learning), but finding a way to distinguish between physiological and pathological HFOs remains challenging. In contrast, VHFOs and UFOs are likely to be important markers of epileptogenicity (Brázdil et al., 2017), and a plausible mechanistic explanation of such high frequencies in the LFP signal adds to the understanding of the pathophysiology and improving the treatment of drug-resistant focal epilepsy or the development of new less invasive tools for its treatment.

We established that synchronization of a weakly coupled neuronal network occurs in a large parameter region. The phenomenon of multistability allows for the synchronization of in-phase, anti-phase, or other spatiotemporally symmetric phase shifts. This enables the network to generate tonic spiking at its own frequency and simultaneously produce spikes at a

regularly shifted phase. This phenomenon potentially creates an event that can be recorded as a very high-frequency or ultra-fast oscillation by nearby electrodes. One assumes the recorded signal is a weighted sum of the nearby activity (see Katzner et al., 2009; Mitzdorf, 1985), but here, for simplicity, all neurons contribute equally as this choice does not affect the dominant frequency. We simulated the network for different parameters in physiological ranges and subjected the system to an external current with random noise around a reference value. We found that synchronized VHFOs and UFOs are possible for very weakly coupled nearly identical neurons, persisting even with small random noise affecting the magnitude of the external current input. Our findings resonate with prior work highlighting already fast ripples as a network phenomenon in epileptic rodents, driven by out-of-phase synchronized firing across neuron pools (see Jefferys et al., 2012).

In this study, we implemented gap-junctional coupling into our model because the reactions of the dynamics would manifest more slowly if chemical synapses were utilized, particularly in the context of modeling very high-frequency oscillations. Gap junctions, as electrical synapses, are known for their ability to support rapid and bidirectional communication between neurons, which makes them suitable for modeling high-frequency oscillatory dynamics. However, we acknowledge the intricate, multifaceted nature of neuronal networks and therefore concede that both gap junctions and chemical synapses might play significant roles in the overall dynamic behavior. Nevertheless, we have shown that the phenomenon of frequency multiplication is generic and model-independent. There are other, more complex models with gap-junctional coupling but random (Erdös-Renyi graph) connectivity that could show the same (see Ghosh et al., 2020). Initial simulations showed emergent network oscillations with higher frequencies (up to 70 Hz instead of the individual 10 Hz) as the coupling strength increased. We observed that the network fired in clusters, but neurons did not always participate in the active cluster. So, while our mechanism seems relevant, a complete characterization requires a separate study. Future studies might benefit from models that concurrently integrate both types of synapses to capture a broader range of neuronal dynamics and interactions.

We used gap-junctional coupling of near-identical neurons of the same type for our analysis, but other parameters can also be varied with similar results. Specifically, we have already examined rigorously that slight changes in the sodium conductance g_{Na} and reversal potential V_{Na} in the interneuron model do not affect the obtained results. This will be reported elsewhere. Moreover, we found that excitation with a noisy external current gives analogous results. The approach is applicable to a multilevel network comprising subgroups of neurons that already exhibit VHFOs through anti-phase spiking within each subgroup. As a result, the presence of weak coupling between these subgroups gives rise to transient UFRs and UFOs in the simulated EEG signal. However, there is still much to be explored, including whether VHFOs and UFOs can be simulated in the EEG signal for a more complex network composed of multiple different types of neurons and how the network connectivity matrix affects the dynamics. Moreover, the persisting stability of anti-phase oscillations in a multilevel network is currently undetermined, as their stability and resilience have been established in single-level and two-level networks with very weak coupling. However, it is likely that the stability region will diminish in the multilevel scenario.

Our results support the hypothesis of a spatially limited pathological phenomenon, which is also manifested in VHFOs and UFOs. In addition, the coupling type and its strength can affect the dynamics of the coupled neurons, with strong enough coupling leading to in-phase synchrony or small phase-shift synchrony and very weak coupling allowing stable anti-phase regime, that may, in that case, temporally appear in the EEG signal. This result is consistent with the observation of altered connectivity in epileptic brain networks (see DeSalvo, Douw, Tanaka, Reinsberger, & Stufflebeam, 2014).

In conclusion, our study provides a mathematical framework for investigating the occurrence of HFOs, VHFOs, and UFOs in the EEG signals of patients with focal epilepsy. Our findings suggest that a reduction in the strength of the connections between neurons in an epileptic focus can produce very high-frequency signals, which could aid in understanding the pathology and improving the treatment of focal epilepsy. However, there is still much to be explored regarding the coupling types, their strength, and other parameters that could affect the dynamics of the coupled neurons.

METHODS

Morris-Lecar Neuron Model

The Morris–Lecar model was developed by Morris and Lecar (1981) to reproduce the variety of oscillatory behavior in relation to Ca²⁺ and K⁺ conductance (van Putten, 2020). Although it was originally created to model the potential in the muscle fiber of the giant barnacle, the Morris–Lecar model is today used for modeling brain pyramidal neurons due to its simplicity and ability to model various dynamics (Lecar, 2007; Prescott, De Koninck, & Sejnowski, 2008; Prescott, Ratté, et al., 2008). The model is described by the following two-dimensional system of differential equations

$$C\frac{\mathrm{d}}{\mathrm{d}t}V = I_{\mathrm{ext}} - g_{\mathrm{L}}(V - V_{\mathrm{L}}) - g_{\mathrm{C}a}m_{\infty}(V)(V - V_{\mathrm{C}a}) - g_{\mathrm{K}}w(V - V_{\mathrm{K}}),$$

$$\frac{\mathrm{d}}{\mathrm{d}t}w = \frac{w_{\infty}(V) - w}{\tau_{w}(V)},$$
(11)

where V is the membrane potential [mV] and w represents the activation variable for K⁺, that is, the probability that the K⁺ channel is conducting. The parameter C corresponds to the cell membrane capacitance.

In comparison to the K^+ current, the Ca^{2+} current changes rapidly, and its activation is assumed to be instantaneous. The activation function is modeled as

$$m_{\infty}(V) = \frac{1}{2} \left[1 + \tanh\left(\frac{V - \beta_1}{\beta_2}\right) \right], \tag{12}$$

where β_1 and β_2 denote the potential at which the Ca²⁺ current is half-activated and the slope of the activation voltage dependence, respectively.

Similarly, for the voltage-dependent steady state w_{∞} , we use the form

$$w_{\infty}(V) = \frac{1}{2} \left[1 + \tanh\left(\frac{V - \beta_3}{\beta_4}\right) \right],\tag{13}$$

where β_3 and β_4 have analogous meanings to β_1 and β_2 , respectively. The timescale of the *w* variable is given by

$$\tau_w(V) = \frac{1}{2} \left[\varphi \cosh\left(\frac{V - \beta_3}{2\beta_4}\right) \right]^{-1},\tag{14}$$

where the parameter φ is a temperature factor adjusting the relative timescale of *V* and *w*.

The system parameters g_L , g_{Ca} , and g_K represent the maximum leak, Ca^{2+} , and K^+ electrical conductances through membrane channels, respectively, and V_L , V_{Ca} , and V_K are reversal potentials of the specific ion channels. Finally, I_{ext} denotes the externally applied direct current,

whichdirect can be used as a single parameter determining a class of the studied cell, that is, whether we deal with class I or class II neuron according to the classification proposed by Hodg-kin (1948).

Unless otherwise stated, we make the parameter choices as listed in Table 1. These physiological parameter values were taken from Gutkin and Ermentrout (1998). Characteristic values for the hippocampal cell membrane capacitance *C* are found to be around 1 μ F/cm² (Destexhe & Paré, 1999; Ermentrout & Terman, 2010; Hartveit, Veruki, & Zandt, 2019; Keener & Sneyd, 2009; Tewari et al., 2018). Therefore, within the framework of this article, we consider these values.

Model of Morris-Lecar Neuronal Network

Timescales of synapses of neurons and gap junctions differ. Specifically, synaptic coupling refers to the mechanism whereby neurotransmitters are released from a presynaptic neuron and subsequently bind to receptors located on a postsynaptic neuron, inducing changes in the latter's membrane potential. This process is relatively slow, taking several milliseconds to occur. In contrast, gap junctions provide direct electrical communication between neurons, allowing for very rapid transmission of electrical signals with virtually no delay in the communication between the coupled neurons. Given our objective to model high frequencies, we opted for a network model that incorporates gap junctions, which are known to be present in the mammalian central nervous system, including the hippocampus (Draguhn, Traub, Schmitz, & Jefferys, 1998; Fukuda & Kosaka, 2000). Morris-Lecar neuronal networks with gap junctions have recently been studied with regard to epilepsy (Volman, Perc, & Bazhenov, 2011; Naze, Bernard, & Jirsa, 2015). HFOs have also been reported in a computational study with Hodgkin–Huxley type models with gap junctions by Helling, Koppert, Visser, and Kalitzin (2015). Here, we examine a small population of neurons with gap junction coupling as it has been found that the epileptic brain tissue may be functionally isolated from surrounding brain regions (see Klimes et al., 2016; Warren et al., 2010). Moreover, the review of Jin and Zhong (2011) focuses on gap junctions in the pathophysiology of epilepsy, describing their role in the generation, synchronization, and maintenance of seizures, and points to gap junctions as promising targets for the development of antiseizure medication, emphasizing the need for further research in this field to refine our understanding and treatment of epilepsy.

Let us examine a neuronal network model of N electrically coupled Morris–Lecar neurons (Equation 11) described by

$$C_{i}\frac{d}{dt}V_{i} = I_{ext} - g_{L}(V_{i} - V_{L}) - g_{Ca}m_{\infty}(V_{i})(V_{i} - V_{Ca}) - g_{K}w_{i}(V_{i} - V_{K}) - \sum_{j=1}^{N}\varepsilon_{ij}(V_{i} - V_{j}),$$

$$\frac{d}{dt}w_{i} = \frac{w_{\infty}(V_{i}) - w_{i}}{\tau_{w}(V_{i})},$$
(15)

where $i \in \{1, ..., N\}$ and $K = (\varepsilon_{ij})_{i,j=1}^{N}$ represents a coupling matrix determining the network topology, that is, its connectivity. Within this article, concerning the Morris–Lecar model, we focus on the case of an all-to-all coupled neuronal network with identical coupling strength

$$\varepsilon_{ij} = \varepsilon = \frac{0.1}{N}.$$
(16)

The functions $m_{\infty}(V)$, $w_{\infty}(V)$, and $\tau_w(V)$ are defined in Equation 12, 13, and 14, respectively. Initial conditions for gating variable: $w_1(0) = w_2(0) = 0.04$ unless specified otherwise.

ε

Gap junction: A specialized intercellular

connection that allows direct electrical and chemical communication between neighboring cells, facilitating synchronization of activity. Consider a neuronal network composed of N electrically coupled interneurons (White, Chow, Rit, Soto-Treviño, & Kopell, 1998). Let the membrane potential V_i [mV] of neuron i satisfy the equation

$$C_{i}\frac{\mathrm{d}}{\mathrm{d}t}V_{i} = I_{\mathrm{ext}} - I_{\mathrm{L}} - I_{\mathrm{Na}} - I_{\mathrm{K}} - \sum_{j=1}^{N} \varepsilon_{ij} (V_{i} - V_{j}), \qquad (17)$$

where C_i represents the membrane capacitance, I_{ext} is the externally applied current, and I_L , I_{Na} , and I_K stand for the Hodgkin–Huxley type leak, Na⁺, and K⁺ currents, respectively. The last term represents the gap-junctional coupling between neuron *i* and all other neurons in the network, where ε_{ij} is the gap junction conductance. The Na⁺ and K⁺ currents are given by

$$I_{Na} = g_{Na}m_i^3 h_i (V_i - V_{Na}) \quad \text{and} \quad I_K = g_K n_i^4 (V_i - V_K),$$
(18)

where g_{Na} and g_K are the maximum conductances, m_i and h_i represent gating variables for Na⁺ channels, and n_i is a gating variable for K⁺ channels. The leak current is given by $I_L = g_L(V_i - V_L)$, where g_L stands for the leak conductance and V_L is the leak reversal potential.

The activation variable m_i is assumed to tend rapidly to its steady state. Hence one can substitute it by its asymptotic value $m_i = m_{\infty}(V_i) = (1 + \exp[-0.08(V_i + 26)])^{-1}$. Further, the gating variables h_i and n_i obey

$$\frac{\mathrm{d}}{\mathrm{d}t}h_i = \frac{h_{\infty}(V_i) - h_i}{\tau_h(V_i)} \quad \text{and} \quad \frac{\mathrm{d}}{\mathrm{d}t}n_i = \frac{n_{\infty}(V_i) - n_i}{\tau_n(V_i)}$$
(19)

with

Interplay of synchronized phase shifts in neurons

$$h_{\infty}(V) = \frac{1}{1 + \exp[0.13(V + 38)]}, \quad \tau_h(V) = \frac{0.6}{1 + \exp[-0.12(V + 67)]},$$
$$n_{\infty}(V) = \frac{1}{1 + \exp[-0.045(V + 10)]}, \quad \tau_n(V) = 0.5 + \frac{2}{1 + \exp[0.045(V - 50)]},$$

respectively.

In this article, we consider parameter values $C \approx 1 \,\mu\text{S/cm}^2$, $g_L = 0.1 \,\text{mS/cm}^2$, $g_{Na} = 30 \,\text{mS/cm}^2$, $g_K = 20 \,\text{mS/cm}^2$, $V_L = -60 \,\text{mV}$, $V_{Na} = 45 \,\text{mV}$, and $V_K = -80 \,\text{mV}$. These values are within physiological ranges and give the high-frequency firing rates typical of hippocampal interneurons (White et al., 1998).

Destexhe-Paré Neuron Model

The last model of a hippocampal pyramidal cell we consider within this paper comes from Destexhe and Paré (1999). Currents dynamics are described by Hodgkin–Huxley type models with kinetics based on a model of hippocampal pyramidal cells from Traub and Miles (1991), adjusted to match voltage-clamp data of cortical pyramidal cells (Huguenard, Hamill, & Prince, 1988), and a noninactivating K⁺ current was described in Mainen, Joerges, Huguenard, and Sejnowski (1995).

Once again, we examine a neuronal network comprising N coupled neurons. Let the membrane potential V_i [mV] of neuron i be described by equation

$$C_{i}\frac{d}{dt}V_{i} = I_{\text{ext}} - I_{L} - I_{Na} - I_{Kdr} - I_{M} - \sum_{j=1}^{N} \varepsilon_{ij} (V_{i} - V_{j}), \qquad (20)$$

where C_i denotes the membrane capacitance; I_{ext} is the externally applied direct current, and I_L , I_{Na} , I_{Kdr} , and I_M represent the leak, Na⁺, and K⁺ currents. The sum term stands for the gap junction coupling between neuron *i* and all other neurons in the network with the gap junction conductance ε_{ij} . As usual, the leak current is given by $I_L = g_L(V_i - V_L)$, where g_L is the leak conductance, and V_L denotes the leak reversal potential. As mentioned, all gating variables $p \in \{m, h, n, m_M\}$ obey first-order kinetics

$$\frac{\mathrm{d}}{\mathrm{d}t}p_i = \alpha_p(V_i)(1-p_i) - \beta_p(V_i)p_i \tag{21}$$

with functions $\alpha_p(V_i)$ and $\beta_p(V_i)$ taking the form specific for each variable.

The voltage-dependent Na^+ current was described by Traub and Miles (1991) and is given by

$$U_{\rm Na} = g_{\rm Na} m_i^3 h_i (V_i - V_{\rm Na}), \tag{22}$$

where g_{Na} stands for its maximum conductance, V_{Na} is its reversal potential, and m_i and h_i represent gating variables satisfying Equation 21 with

$$\alpha_m(V) = \frac{-0.32(V - V_{\rm T} - 13)}{\exp[-(V - V_{\rm T} - 13)/4] - 1}, \quad \beta_m(V) = \frac{0.28(V - V_{\rm T} - 40)}{\exp[(V - V_{\rm T} - 40)/5] - 1},$$

$$\kappa_h(V) = 0.128 \exp[-(V - V_{\rm T} - V_{\rm S} - 17)/18], \quad \beta_h(V) = \frac{4}{1 + \exp[-(V - V_{\rm T} - V_{\rm S} - 40)/5]}$$

for $V_{\rm T} = -58$ mV and $V_{\rm S} = -10$ mV fitting the voltage-clamp data.

The current corresponding to the delayed rectifier K^+ channel was studied by Traub and Miles (1991) and is given by

$$I_{Kdr} = g_{Kdr} n_i^4 (V_i - V_K),$$
(23)

where g_{Kdr} is the maximum conductance, V_K represents its reversal potential, and n_i obeys Equation 21 with

$$\alpha_n(V) = \frac{-0.032(V - V_{\rm T} - 15)}{\exp[-(V - V_{\rm T} - 15)/5] - 1}, \quad \beta_n(V) = 0.5 \exp[-(V - V_{\rm T} - 10)/40].$$

And finally, the noninactivating current was described in Mainen et al. (1995) and takes the form

$$I_{\mathcal{M}} = g_{\mathcal{M}} m_{\mathcal{M},i} (V_i - V_{\mathcal{K}}), \tag{24}$$

where g_M denotes the maximum conductance and m_M obeys Equation 21 with

$$\alpha_{m_{\rm M}}(V) = \frac{0.0001(V+30)}{1-\exp[-(V+30)/9]}, \quad \beta_{m_{\rm M}}(V) = \frac{-0.0001(V+30)}{1-\exp[(V+30)/9]}.$$

Within this paper, we consider the following physiological parameter values for a hippocampal pyramidal cell taken from Destexhe and Paré (1999). Specifically, $C \approx 1 \,\mu\text{S/cm}^2$, $g_L = 0.019 \,\text{mS/cm}^2$, $g_{Na} = 120 \,\text{mS/cm}^2$, $g_{Kdr} = 100 \,\text{mS/cm}^2$, $g_M = 2 \,\text{mS/cm}^2$, $V_L = -65 \,\text{mV}$, $V_{Na} = 55 \,\text{mV}$, and $V_K = -85 \,\text{mV}$.

NUMERICAL METHODS

α

We used the standard ode45 solver in matlab for deterministic settings, while for simulations with noisy input, we add noise to the external current at each simulation step with $\Delta t = 0.01$

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using the Euler–Maruyama method. Bifurcation curves have been computed using the numerical bifurcation package MatCont (Dhooge, Govaerts, & Kuznetsov, 2003; Dhooge, Govaerts, Kuznetsov, Meijer, & Sautois, 2008).

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SUPPORTING INFORMATION

Supporting information for this article is available at https://doi.org/10.1162/netn_a_00351. Code is available at https://gitlab.ics.muni.cz/9607/UFOs.

AUTHOR CONTRIBUTIONS

Lenka Přibylová: Conceptualization; Formal analysis; Funding acquisition; Methodology; Project administration; Supervision; Validation; Writing – original draft. Jan Ševčík: Data curation; Formal analysis; Methodology; Software; Visualization; Writing – original draft. Veronika Eclerová: Conceptualization; Methodology; Validation; Writing – review & editing. Petr Klimeš: Validation; Writing – review & editing. Milan Brázdil: Supervision; Validation; Writing – review & editing. Hil Meijer: Methodology; Supervision; Validation; Writing – review & editing.

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REFERENCES

- Abrams, D. M., & Strogatz, S. H. (2004). Chimera states for coupled oscillators. *Physical Review Letters*, 93(17), 174102. https://doi .org/10.1103/PhysRevLett.93.174102, PubMed: 15525081
- Aihara, K., Takabe, T., & Toyoda, M. (1990). Chaotic neural networks. *Physics Letters A*, 144(6–7), 333–340. https://doi.org/10 .1016/0375-9601(90)90136-C
- Bakhtiari, S., Manshadi, M. K., Candas, M., & Beskok, A. (2023). Changes in electrical capacitance of cell membrane reflect drug partitioning-induced alterations in lipid bilayer. *Micromachines*, *14*(2), 316. https://doi.org/10.3390/mi14020316, PubMed: 36838014
- Boccaletti, S., Pisarchik, A. N., Del Genio, C. I., & Amann, A. (2018). Synchronization: From coupled systems to complex

networks. Cambridge University Press. https://doi.org/10.1017 /9781107297111

- Brázdil, M., Pail, M., Halámek, J., Plešinger, F., Cimbálník, J., Roman, R., ... Jurák, P. (2017). Very high-frequency oscillations: Novel biomarkers of the epileptogenic zone. *Annals of Neurology*, *82*(2), 299–310. https://doi.org/10.1002/ana.25006, PubMed: 28779553
- Brazdil, M., Worrell, G. A., Travnicek, V., Pail, M., Roman, R., Plesinger, F., ... Jurák, P. (2023). Ultra fast oscillations in the human brain and their functional significance. *medRxiv*. https:// doi.org/10.1101/2023.02.23.23285962
- Calim, A., Hövel, P., Ozer, M., & Uzuntarla, M. (2018). Chimera states in networks of type-I Morris–Lecar neurons. *Physical*

Review E, 98(6), 062217. https://doi.org/10.1103/PhysRevE.98 .062217

- Cessac, B., & Samuelides, M. (2007). From neuron to neural networks dynamics. *European Physical Journal Special Topics*, *142*(1), 7–88. https://doi.org/10.1140/epjst/e2007-00058-2
- Chow, C. C., & Kopell, N. (2000). Dynamics of spiking neurons with electrical coupling. *Neural Computation*, *12*(7), 1643–1678. https://doi.org/10.1162/089976600300015295, PubMed: 10935921
- Chowdhury, S. N., Rakshit, S., Buldu, J. M., Ghosh, D., & Hens, C. (2021). Antiphase synchronization in multiplex networks with attractive and repulsive interactions. *Physical Review E*, *103*(3), 032310. https://doi.org/10.1103/PhysRevE.103.032310, PubMed: 33862752
- Cimbalnik, J., Brinkmann, B., Kremen, V., Jurak, P., Berry, B., Gompel, J. V., ... Worrell, G. (2018). Physiological and pathological high frequency oscillations in focal epilepsy. *Annals of Clinical and Translational Neurology*, *5*(9), 1062–1076. https://doi .org/10.1002/acn3.618, PubMed: 30250863
- Cimbalnik, J., Pail, M., Klimes, P., Travnicek, V., Roman, R., Vajcner, A., & Brazdil, M. (2020). Cognitive processing impacts high frequency intracranial EEG activity of human hippocampus in patients with pharmacoresistant focal epilepsy. *Frontiers in Neurology*, *11*, 578571. https://doi.org/10.3389/fneur.2020 .578571, PubMed: 33193030
- DeSalvo, M. N., Douw, L., Tanaka, N., Reinsberger, C., & Stufflebeam, S. M. (2014). Altered structural connectome in temporal lobe epilepsy. *Radiology*, *270*(3), 842–848. https://doi.org /10.1148/radiol.13131044, PubMed: 24475828
- Destexhe, A., & Paré, D. (1999). Impact of network activity on the integrative properties of neocortical pyramidal neurons in vivo. *Journal of Neurophysiology*, *81*(4), 1531–1547. https://doi.org /10.1152/jn.1999.81.4.1531, PubMed: 10200189
- Dhooge, A., Govaerts, W., & Kuznetsov, Y. A. (2003). MatCont: A MATLAB package for numerical bifurcation analysis of ODEs. *ACM Transactions on Mathematical Software*, *29*(2), 141–164. https://doi.org/10.1145/779359.779362
- Dhooge, A., Govaerts, W., Kuznetsov, Y. A., Meijer, H. G., & Sautois, B. (2008). New features of the software MatCont for bifurcation analysis of dynamical systems. *Mathematical and Computer Modelling of Dynamical Systems*, *14*(2), 147–175. https://doi.org/10.1080/13873950701742754
- Draguhn, A., Traub, R., Schmitz, D., & Jefferys, J. (1998). Electrical coupling underlies high-frequency oscillations in the hippocampus *in vitro*. *Nature*, *394*(6689), 189–192. https://doi.org/10.1038 /28184, PubMed: 9671303
- Duan, L., & Lu, Q. (2006). Codimension-two bifurcation analysis on firing activities in Chay neuron model. *Chaos, Solitons & Fractals, 30*(5), 1172–1179. https://doi.org/10.1016/j.chaos.2005.08.179
- Ermentrout, B., & Terman, D. H. (2010). *Mathematical foundations* of neuroscience (Vol. 35). Springer. https://doi.org/10.1007/978-0 -387-87708-2
- Frauscher, B., von Ellenrieder, N., Zelmann, R., Rogers, C., Nguyen, D. K., Kahane, P., ... Gotman, J. (2018). High-frequency oscillations in the normal human brain. *Annals of Neurology*, *84*(3), 374–385. https://doi.org/10.1002/ana.25304, PubMed: 30051505

- Fukuda, T., & Kosaka, T. (2000). Gap junctions linking the dendritic network of gabaergic interneurons in the hippocampus. *Journal of Neuroscience*, 20(4), 1519–1528. https://doi.org/10 .1523/JNEUROSCI.20-04-01519.2000, PubMed: 10662841
- Gabbiani, F., & Cox, S. J. (2017). *Mathematics for neuroscientists*. Academic Press.
- Gentet, L. J., Stuart, G. J., & Clements, J. D. (2000). Direct measurement of specific membrane capacitance in neurons. *Biophysical Journal*, *79*(1), 314–320. https://doi.org/10.1016/S0006 -3495(00)76293-X, PubMed: 10866957
- Ghosh, S., Mondal, A., Ji, P., Mishra, A., Dana, S. K., Antonopoulos, C. G., & Hens, C. (2020). Emergence of mixed mode oscillations in random networks of diverse excitable neurons: The role of neighbors and electrical coupling. *Frontiers in Computational Neuroscience*, 14, 49. https://doi.org/10.3389/fncom.2020 .00049, PubMed: 32581757
- Golubitsky, M., Messi, L. M., & Spardy, L. E. (2016). Symmetry types and phase-shift synchrony in networks. *Physica D: Non-linear Phenomena, 320*, 9–18. https://doi.org/10.1016/j.physd .2015.12.005
- Golubitsky, M., Romano, D., & Wang, Y. (2012). Network periodic solutions: Patterns of phase-shift synchrony. *Nonlinearity*, *25*(4), 1045. https://doi.org/10.1088/0951-7715/25/4/1045
- Golubitsky, M., & Stewart, I. (2006). Nonlinear dynamics of networks: The groupoid formalism. *Bulletin of the American Mathematical Society*, *43*(3), 305–364. https://doi.org/10.1090 /S0273-0979-06-01108-6
- Golubitsky, M., & Stewart, I. (2016). Rigid patterns of synchrony for equilibria and periodic cycles in network dynamics. *Chaos: An Interdisciplinary Journal of Nonlinear Science*, *26*(9), 094803. https://doi.org/10.1063/1.4953664, PubMed: 27781462
- Golubitsky, M., Stewart, I., & Török, A. (2005). Patterns of synchrony in coupled cell networks with multiple arrows. *SIAM Journal on Applied Dynamical Systems*, *4*(1), 78–100. https:// doi.org/10.1137/040612634
- Gutkin, B. S., & Ermentrout, G. B. (1998). Dynamics of membrane excitability determine interspike interval variability: A link between spike generation mechanisms and cortical spike train statistics. *Neural Computation*, *10*(5), 1047–1065. https://doi .org/10.1162/089976698300017331, PubMed: 9654767
- Hao, J., Cui, Y., Niu, B., Yu, L., Lin, Y., Xia, Y., ... Guo, D. (2021).
 Roles of very fast ripple (500–1000 Hz) in the hippocampal network during status epilepticus. *International Journal of Neural Systems*, *31*(4), 2150002. https://doi.org/10.1142/S0129065721500027, PubMed: 33357153
- Hartveit, E., Veruki, M. L., & Zandt, B.-J. (2019). Capacitance measurement of dendritic exocytosis in an electrically coupled inhibitory retinal interneuron: An experimental and computational study. *Physiological Reports, 7*(15), e14186. https://doi.org/10.14814/phy2.14186, PubMed: 31379117
- Helling, R., Koppert, M., Visser, G., & Kalitzin, S. (2015). Gap junctions as common cause of high-frequency oscillations and epileptic seizures in a computational cascade of neuronal mass and compartmental modeling. *International Journal of Neural Systems*, *25*(6), 1550021. https://doi.org/10.1142/S0129065715500215, PubMed: 26058401

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12024

- Hodgkin, A. L. (1948). The local electric changes associated with repetitive action in a non-medullated axon. *Journal of Physiology*, *107*(2), 165–181. https://doi.org/10.1113/jphysiol.1948 .sp004260, PubMed: 16991796
- Hoppensteadt, F. C., & Izhikevich, E. M. (1997). Weakly connected neural networks (Vol. 126). Springer Science & Business Media. https://doi.org/10.1007/978-1-4612-1828-9
- Huguenard, J., Hamill, O., & Prince, D. (1988). Developmental changes in Na⁺ conductances in rat neocortical neurons: Appearance of a slowly inactivating component. *Journal of Neurophysiology*, *59*(3), 778–795. https://doi.org/10.1152/jn.1988 .59.3.778, PubMed: 2452862
- Izhikevich, E. (2006). Dynamical systems in neuroscience. MIT Press. https://doi.org/10.7551/mitpress/2526.001.0001
- Jacobs, J., LeVan, P., Chander, R., Hall, J., Dubeau, F., & Gotman, J. (2008). Interictal high-frequency oscillations (80–500 Hz) are an indicator of seizure onset areas independent of spikes in the human epileptic brain. *Epilepsia*, 49(11), 1893–1907. https:// doi.org/10.1111/j.1528-1167.2008.01656.x, PubMed: 18479382
- Jacobs, J., Staba, R., Asano, E., Otsubo, H., Wu, J., Zijlmans, M., ... Gotman, J. (2012). High-frequency oscillations (HFOs) in clinical epilepsy. *Progress in Neurobiology*, *98*(3), 302–315. https://doi.org/10.1016/j.pneurobio.2012.03.001, PubMed: 22480752
- Jefferys, J. G., De La Prida, L. M., Wendling, F., Bragin, A., Avoli, M., Timofeev, I., & da Silva, F. H. L. (2012). Mechanisms of physiological and epileptic HFO generation. *Progress in Neurobiol*ogy, 98(3), 250–264. https://doi.org/10.1016/j.pneurobio.2012 .02.005, PubMed: 22420980
- Jin, M.-M., & Zhong, C. (2011). Role of gap junctions in epilepsy. *Neuroscience Bulletin*, 27(6), 389–406. https://doi.org/10.1007 /s12264-011-1944-1, PubMed: 22108816
- Jiruska, P., Alvarado-Rojas, C., Schevon, C. A., Staba, R., Stacey, W., Wendling, F., & Avoli, M. (2017). Update on the mechanisms and roles of high-frequency oscillations in seizures and epileptic disorders. *Epilepsia*, *58*(8), 1330–1339. https://doi.org/10.1111 /epi.13830, PubMed: 28681378
- Katzner, S., Nauhaus, I., Benucci, A., Bonin, V., Ringach, D. L., & Carandini, M. (2009). Local origin of field potentials in visual cortex. *Neuron*, *61*(1), 35–41. https://doi.org/10.1016/j.neuron .2008.11.016, PubMed: 19146811
- Kawamura, Y., Nakao, H., Arai, K., Kori, H., & Kuramoto, Y. (2010). Phase synchronization between collective rhythms of globally coupled oscillator groups: Noiseless nonidentical case. *Chaos: An Interdisciplinary Journal of Nonlinear Science*, *20*(4), 043110. https://doi.org/10.1063/1.3491346, PubMed: 21198080
- Keener, J., & Sneyd, J. (2009). *Mathematical physiology: I: Cellular physiology*. Springer. https://doi.org/10.1007/978-0-387-75847-3
- Klimes, P., Duque, J., Brinkmann, B., Van Gompel, J., Stead, M., St. Louis, E., ... Worrell, G. (2016). The functional organization of human epileptic hippocampus. *Journal of Neurophysiology*, *115*(6), 3140–3145. https://doi.org/10.1152/jn.00089.2016, PubMed: 27030735
- Kobelevskiy, I. (2008). *Bifurcation analysis of a system of Morris-Lecar neurons with time delayed gap junctional coupling.* (Unpublished master's thesis). University of Waterloo.

- Kuznetsov, Y. A. (2023). *Elements of applied bifurcation theory,* (4th ed., Vol. 112). New York: Springer. https://doi.org/10.1007/978-3-031-22007-4
- Lang, X., Lu, Q., & Kurths, J. (2010). Phase synchronization in noise-driven bursting neurons. *Physical Review E*, *82*(2), 021909. https://doi.org/10.1103/PhysRevE.82.021909, PubMed: 20866839
- Lecar, H. (2007). Morris–Lecar model. *Scholarpedia*, *2*(10), 1333. https://doi.org/10.4249/scholarpedia.1333
- Liu, C., Liu, X., & Liu, S. (2014). Bifurcation analysis of a Morris-Lecar neuron model. *Biological Cybernetics*, 108(1), 75–84. https://doi.org/10.1007/s00422-013-0580-4, PubMed: 24435761
- Macdonald, R. L. (1989). Antiepileptic drug actions. *Epilepsia, 30,* S19–S28. https://doi.org/10.1111/j.1528-1157.1989.tb05810.x, PubMed: 2550216
- Mainen, Z. F., Joerges, J., Huguenard, J. R., & Sejnowski, T. J. (1995). A model of spike initiation in neocortical pyramidal neurons. *Neuron*, *15*(6), 1427–1439. https://doi.org/10.1016/0896 -6273(95)90020-9, PubMed: 8845165
- Majhi, S., Bera, B. K., Ghosh, D., & Perc, M. (2019). Chimera states in neuronal networks: A review. *Physics of Life Reviews, 28*, 100–121. https://doi.org/10.1016/j.plrev.2018.09.003, PubMed: 30236492
- Mitzdorf, U. (1985). Current source-density method and application in cat cerebral cortex: Investigation of evoked potentials and EEG phenomena. *Physiological Reviews*, *65*(1), 37–100. https://doi.org /10.1152/physrev.1985.65.1.37, PubMed: 3880898
- Morris, C., & Lecar, H. (1981). Voltage oscillations in the barnacle giant muscle fiber. *Biophysical Journal*, *35*(1), 193–213. https://doi.org/10.1016/S0006-3495(81)84782-0, PubMed: 7260316
- Nagai, K. H., & Kori, H. (2010). Noise-induced synchronization of a large population of globally coupled nonidentical oscillators. *Physical Review E*, *81*(6), 065202. https://doi.org/10.1103 /PhysRevE.81.065202, PubMed: 20866467
- Naze, S., Bernard, C., & Jirsa, V. (2015). Computational modeling of seizure dynamics using coupled neuronal networks: Factors shaping epileptiform activity. *PLOS Computational Biology*, *11*(5), e1004209. https://doi.org/10.1371/journal.pcbi.1004209, PubMed: 25970348
- Neiman, A., Schimansky-Geier, L., Cornell-Bell, A., & Moss, F. (1999). Noise-enhanced phase synchronization in excitable media. *Physical Review Letters*, *83*(23), 4896. https://doi.org/10 .1103/PhysRevLett.83.4896
- Nejedly, P., Cimbalnik, J., Klimes, P., Plesinger, F., Halamek, J., Kremen, V., ... Jurák, P. (2019). Intracerebral EEG artifact identification using convolutional neural networks. *Neuroinformatics*, *17*(2), 225–234. https://doi.org/10.1007/s12021-018-9397-6, PubMed: 30105544
- Nicholson, G. M., Blanche, T., Mansfield, K., & Tran, Y. (2002). Differential blockade of neuronal voltage-gated Na+ and K+ channels by antidepressant drugs. *European Journal of Pharmacology*, 452(1), 35–48. https://doi.org/10.1016/S0014 -2999(02)02239-2, PubMed: 12323383
- Nijholt, E., Rink, B., & Sanders, J. (2019). Center manifolds of coupled cell networks. *SIAM Review*, *61*(1), 121–155. https://doi.org/10.1137/18M1219977

- Pail, M., Cimbálník, J., Roman, R., Daniel, P., Shaw, D. J., Chrastina, J., & Brázdil, M. (2020). High frequency oscillations in epileptic and non-epileptic human hippocampus during a cognitive task. Scientific Reports, 10(1), 18147. https://doi.org /10.1038/s41598-020-74306-3, PubMed: 33097749
- Patel, D. C., Tewari, B. P., Chaunsali, L., & Sontheimer, H. (2019). Neuron-glia interactions in the pathophysiology of epilepsy. Nature Reviews Neuroscience, 20(5), 282-297. https://doi.org /10.1038/s41583-019-0126-4, PubMed: 30792501
- Perko, L. (2001). Differential equations and dynamical systems (Vol. 7). Springer Science & Business Media. https://doi.org/10 .1007/978-1-4613-0003-8
- Pikovsky, A., Rosenblum, M., & Kurths, J. (2001). Synchronization: A universal concept in nonlinear sciences. Cambridge University Press. https://doi.org/10.1017/CBO9780511755743
- Prescott, S. A., De Koninck, Y., & Sejnowski, T. J. (2008). Biophysical basis for three distinct dynamical mechanisms of action potential initiation. PLoS Computational Biology, 4(10), e1000198. https://doi.org/10.1371/journal.pcbi.1000198, PubMed: 18846205
- Prescott, S. A., Ratté, S., De Koninck, Y., & Sejnowski, T. J. (2008). Pyramidal neurons switch from integrators in vitro to resonators under in vivo-like conditions. Journal of Neurophysiology, 100(6), 3030-3042. https://doi.org/10.1152/jn.90634.2008, PubMed: 18829848
- Purves, D., Augustine, G. J., Fitzpatrick, D., Hall, W., LaMantia, A.-S., & White, L. (2019). Neuroscience (6th ed.). Oxford Universitv Press.
- Řehulka, P., Cimbalnik, J., Pail, M., Chrastina, J., Hermanová, M., & Brázdil, M. (2019). Hippocampal high frequency oscillations in unilateral and bilateral mesial temporal lobe epilepsy. Clinical Neurophysiology, 130(7), 1151-1159. https://doi.org/10.1016/j .clinph.2019.03.026, PubMed: 31100580
- Sebek, M., Kawamura, Y., Nott, A. M., & Kiss, I. Z. (2019). Anti-phase collective synchronization with intrinsic in-phase coupling of two groups of electrochemical oscillators. Philosophical Transactions of the Royal Society A, 377(2160), 20190095. https://doi.org/10.1098/rsta.2019.0095, PubMed: 31656145
- Shilnikov, A. (2012). Complete dynamical analysis of a neuron model. Nonlinear Dynamics, 68(3), 305-328. https://doi.org/10 .1007/s11071-011-0046-y
- Skinner, F., Zhang, L., Velazquez, J. P., & Carlen, P. (1999). Bursting in inhibitory interneuronal networks: A role for gap-junctional coupling. Journal of Neurophysiology, 81(3), 1274-1283. https://doi.org/10.1152/jn.1999.81.3.1274, PubMed: 10085354
- Staba, R. J., & Bragin, A. (2011). High-frequency oscillations and other electrophysiological biomarkers of epilepsy: Underlying mechanisms. Biomarkers in Medicine, 5(5), 545-556. https:// doi.org/10.2217/bmm.11.72, PubMed: 22003903
- Strogatz, S. H. (2018). Nonlinear dynamics and chaos: With applications to physics, biology, chemistry, and engineering. CRC Press. https://doi.org/10.1201/9780429492563
- Tewari, B. P., Chaunsali, L., Campbell, S. L., Patel, D. C., Goode, A. E., & Sontheimer, H. (2018). Perineuronal nets decrease membrane capacitance of peritumoral fast spiking interneurons

in a model of epilepsy. Nature Communications, 9(1), 4724. https://doi.org/10.1038/s41467-018-07113-0, PubMed: 30413686

- Traub, R. D., & Miles, R. (1991). Neuronal networks of the hippocampus (Vol. 777). Cambridge University Press. https://doi.org /10.1017/CBO9780511895401
- Travnicek, V., Jurak, P., Cimbalnik, J., Klimes, P., Daniel, P., & Brazdil, M. (2021). Ultra-fast oscillation detection in EEG signal from deep-brain microelectrodes. In 2021 43rd annual international conference of the IEEE Engineering in Medicine & Biology Society (EMBC) (pp. 265-268). https://doi.org/10.1109 /EMBC46164.2021.9629481, PubMed: 34891287
- Tsumoto, K., Kitajima, H., Yoshinaga, T., Aihara, K., & Kawakami, H. (2006). Bifurcations in Morris-Lecar neuron model. Neurocomputing, 69(4-6), 293-316. https://doi.org/10.1016/j.neucom.2005.03.006
- van Putten, M. (2020). Dynamics of neural networks. Springer. https://doi.org/10.1007/978-3-662-61184-5
- Vasickova, Z., Klimes, P., Cimbalnik, J., Travnicek, V., Pail, M., Halamek, J., ... Brazdil, M. (2023). Shadows of very high-frequency oscillations can be detected in lower frequency bands of routine stereoelectroencephalography. Scientific Reports, 13(1), 1065. https://doi.org/10.1038/s41598-023-27797-9, PubMed: 36658267
- Volman, V., Perc, M., & Bazhenov, M. (2011). Gap junctions and epileptic seizures-Two sides of the same coin? PLoS One, 6(5), e20572. https://doi.org/10.1371/journal.pone.0020572, PubMed: 21655239
- Wang, B., Ke, W., Guang, J., Chen, G., Yin, L., Deng, S., ... Shu, Y. (2016). Firing frequency maxima of fast-spiking neurons in human, monkey, and mouse neocortex. Frontiers in Cellular Neuroscience, 10, 239. https://doi.org/10.3389/fncel.2016 .00239, PubMed: 27803650
- Warren, C., Hu, S., Stead, M., Brinkmann, B., Bower, M., & Worrell, G. (2010). Synchrony in normal and focal epileptic brain: The seizure onset zone is functionally disconnected. Journal of Neurophysiology, 104(6), 3530-3539. https://doi.org/10.1152/jn .00368.2010, PubMed: 20926610
- White, J. A., Chow, C. C., Rit, J., Soto-Treviño, C., & Kopell, N. (1998). Synchronization and oscillatory dynamics in heterogeneous, mutually inhibited neurons. Journal of Computational Neuroscience, 5(1), 5-16. https://doi.org/10.1023/A:1008841325921, PubMed: 9580271
- Wiggins, S. (2003). Introduction to applied nonlinear dynamical systems and chaos (2nd ed.). New York: Springer. https://doi .org/10.1007/b97481
- Worrell, G., & Gotman, J. (2011). High-frequency oscillations and other electrophysiological biomarkers of epilepsy: Clinical studies. Biomarkers in Medicine, 5(5), 557-566. https://doi.org /10.2217/bmm.11.74, PubMed: 22003904
- Xing, M., Song, X., Wang, H., Yang, Z., & Chen, Y. (2022). Frequency synchronization and excitabilities of two coupled heterogeneous Morris-Lecar neurons. Chaos, Solitons & Fractals, 157, 111959. https://doi.org/10.1016/j.chaos.2022.111959
- Zhou, C., & Kurths, J. (2002). Noise-induced phase synchronization and synchronization transitions in chaotic oscillators. Physical Review Letters, 88(23), 230602. https://doi.org/10.1103 /PhysRevLett.88.230602, PubMed: 12059346

- Zhou, C., & Kurths, J. (2003). Noise-induced synchronization and coherence resonance of a Hodgkin-Huxley model of thermally sensitive neurons. *Chaos: An Interdisciplinary Journal of Nonlinear Science*, *13*(1), 401–409. https://doi.org/10.1063/1.1493096, PubMed: 12675446
- Zijlmans, M., Jacobs, J., Zelmann, R., Dubeau, F., & Gotman, J. (2009). High frequency oscillations and seizure frequency in patients with focal epilepsy. *Epilepsy Research*, *85*(2–3), 287–292. https://doi.org/10.1016/j.eplepsyres.2009.03.026, PubMed: 19403269